

1 Opposite-sex pairing alters social interaction-induced GCaMP and dopamine activity in the insular cortex
2 of male prairie voles

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21

22 Abstract

23 The prairie vole (*Microtus ochrogaster*) is a monogamous rodent species which displays selective
24 social behaviors to conspecifics after establishing a pair bonded relationship, specifically partner-directed
25 affiliation and stranger-directed aggression. This social selectivity relies on the ability of an individual to
26 respond appropriately to a social context and requires salience detection and valence assignment. The
27 anterior insular cortex (aIC) has been implicated in stimulus processing and categorization across a
28 variety of contexts and is well-situated to integrate environmental stimuli and internal affective states to
29 modulate complex goal-directed behaviors and social decision-making. Surprisingly, the contribution of
30 the aIC to the expression of pair bond-induced social selectivity in prairie voles has been drastically
31 understudied. Here we examined whether neural activity and gene expression in the aIC change in
32 response to opposite-sex pairing and/or as a function of pairing length in male prairie voles. Opposite-sex
33 pairing was characterized by changes to calcium and dopamine (DA) transients in the aIC that
34 corresponded with the display of social selectivity across pair bond maturation. Furthermore, D1 and D2
35 receptor mRNA expression was significantly higher in males after 48 hrs of cohabitation with a female
36 partner compared to same-sex housed males, and D2 mRNA remained significantly higher in males with a
37 female partner compared to same-sex housed males after a week of cohabitation. Together, these results
38 implicate a role for DA and its receptors in the aIC across the transition from early- to late-phase pair
39 bonding.

40

41 For virtually all social beings, ranging from insect to primate species, navigating a social
42 environment is commonplace and often necessary for both individual and species survival. Effective
43 social communication relies on one's ability to integrate and process multiple social and environmental
44 modalities to drive an appropriate behavioral response [1, 2]. Examples of such modalities include the
45 external environmental context, the internal emotional and motivational state of the individual, and the
46 perceived internal state(s) of other social conspecifics [3]. Furthermore, social encounters are dyadic and
47 often involve in-the-moment adaptations to the behavioral responses of social partners throughout the
48 length of an interaction session. Social neuroscience has uncovered a network of brain regions that work
49 together to attune, process, and respond appropriately to specific social encounters, and this collection of
50 regions is known as the social decision-making network [4, 5]. The insular cortex (IC) and other "higher
51 order" cortical regions have functional-structural relationships to regions of the social decision-making
52 network but have received less attention regarding its regulation of social behavior [6]. Structural
53 connectivity patterns implicate the posterior IC (pIC) as the main sensory "detector" and relays this
54 information directly to the amygdala for immediate survival-dependent action [7-9]. In contrast, the
55 anterior IC (aIC) receives cortical information, interoceptive information from the hindbrain, and sensory
56 information relayed from the pIC and integrates these signals together to create a wholistic framework for
57 a specific context, then "superimposes" this framework over the motivation-centered striatum to adjust
58 stimuli valence and salience for more nuanced control of reward-based decision-making [10]. Thus, it is
59 no surprise that the IC has been implicated in a wide range of context-dependent learning, memory, and
60 decision-making such as drug and alcohol abuse, taste recognition memory, and social recognition
61 memory [11-16].

62 Given the structural and functional intricacy of the IC and the fact that activity of the IC has been
63 extensively linked to the expression of a variety of human attachments, including mother-infant bonds and
64 adult romantic relationships [17], it is surprising that the IC has been largely unexplored in basic research
65 using model animal species that form similar attachments. The prairie vole (*Microtus ochrogaster*) is a
66 socially monogamous rodent species that is often used to study the neurobiology of adult social
67 attachments [18, 19]. While researchers have spent decades exploring the neurochemicals and circuits that
68 underly the formation and maintenance of such attachments in prairie voles (termed pair bonds), no
69 studies have focused on the IC. Furthermore, the mesolimbic reward system, both at the level of the cell
70 bodies in the ventral tegmental area (VTA) [20, 21] and downstream in the ventral striatum [22], is crucial
71 for pair bonding. The aIC would be well situated to modulate the behavioral expression of pair-bonding,
72 characterized by partner-specific affiliation and stranger-specific aggression, by receiving contextual
73 information from the pIC and relaying it to the ventral striatum and influence social decision-making.

74 We hypothesized that the aIC would be differentially recruited during social encounters in
75 bachelor vs pair bonded prairie voles. Furthermore, we predicted the pattern of aIC activity in pair bonded
76 voles would vary based on the length of pairing (short-term vs long-term) and social stimulus type
77 (familiar vs unfamiliar). We used fiber photometry to assess calcium and DA transients in the aIC during
78 partner and stranger encounters. To determine whether pair bonding is associated with post-synaptic
79 changes in the aIC, we used qRT-PCR to analyze mRNA expression of DA receptor types 1 (DRD1) and
80 2 (DRD2) in short-term and long-term pair bonded prairie voles.

81

82 **Methods**

83 *Animals*

84 Prairie voles were lab-bred from a population captured in southern Illinois. Voles were weaned at
85 21 ± 3 days of age and were housed in same-sex, non-sibling pairs in microisolator cages (29.2L x 19.1W
86 x 12.7H cm) with corn cob bedding, crinkle nesting material, and ad libitum access to food (Tekland
87 global rabbit diet 2030) and water. Colony rooms were maintained at $21 \pm 1^\circ\text{C}$ with a 14L:10D
88 photoperiod (lights on at 0600 h). Male subjects were between 90 and 120 days of age at the start of the
89 experiment. Female prairie voles (also between 90 and 120 days of age) were used as partners and had
90 been tubal ligated at least one week before pairing (see [23] for description of tubal ligation procedure).
91 All behavior testing was performed between 0900-1700h, and all procedures were conducted in
92 accordance with the National Institutes of Health Guide for the Care and Use of Laboratory Animals and
93 the Institutional Animal Care and Use Committee at the University of Kansas.

94

95 Experiment 1: Effects of same- vs opposite-sex cohabitation on social stimuli-induced aIC activity

96 *Experimental Design*

97 Sixteen male prairie voles were used as subjects ($n = 8/\text{group}$). Subjects received stereotaxic
98 surgery (see below for surgery details) then remained housed with their same-sex, non-sibling cage mate
99 throughout the recovery and viral incubation period of three weeks. For subjects in the same-sex (SS)
100 paired group, only one animal from the cage received surgery and they recovered and remained with their
101 same-sex cage mate throughout the entirety of the experiment. Subjects in the opposite-sex (OS) paired
102 group were housed with a non-related, tubal ligated female immediately following the social exposure test
103 on Pairing Day (Experimental Day 0).

104

105 *Stereotaxic Surgery*

106 Subjects were anesthetized with an intraperitoneal injection of a ketamine (75mg/kg) and
107 dexmedetomidine (1mg/kg) cocktail. The head was shaved, and the skin was cleaned with 70% ethanol
108 then betadine solutions and repeated three times. A subcutaneous injection of a local anesthetic (lidocaine,
109 2mg/kg) was administered at the surgical site. The head was fixed into a stereotaxic apparatus (Stoelting),
110 and an incision was made exposing the skull. A hole was drilled above the injection site, and a 1 uL
111 Neuros needle (Hamilton) was slowly lowered to the rostral insular cortex (AP +1.60 mm, ML +3.25 mm,
112 DV -4.00 mm from bregma). A viral cocktail consisting of 150 nL of pAAV1-hSyn-GCaMP6f-WPRE-
113 SV40 (AddGene) and 150 nL of pAAV9-hSyn-GRAB-rDA1m (AddGene) was infused at a rate of 60
114 nL/min followed by a 5 min diffusion period. Two surgical screws were fixed to the skull, and a fiber
115 optic implant (1.25mm ceramic ferrule, 200 μm core, 0.5NA, 6mm length; RWD) was lowered to the
116 same aIC coordinates with the DV adjusted to -3.80 mm. Dental cement was used to secure the implant
117 and screws, then antisedan (2.5mg/kg) was administered to reverse the anesthesia. Subjects received 2
118 doses of meloxicam (2mg/kg) post-operation (over a 24-hour period) and were also permanently re-
119 housed with their cage mate in a slightly larger cage (36.2L x 20.3W x 14.0H cm) with more vertical
120 clearance from the hopper to accommodate the cranial implant. Animals remained unmanipulated aside
121 from regular facility cage changes for 3 weeks to allow for complete viral transfection.

122

123 *Fiber Photometry & Social Exposure Tests*

124 On the first day of experimental testing (Pairing Day = Experimental Day 0), subjects were
125 brought to the behavioral testing suite and allowed to habituate for 1 hour. Next, a subject was removed
126 from the home cage, the optical implant was attached to a fiber optic cable (Plexon) via a ceramic ferrule,
127 and the animal was placed into a clean, novel cage (47.6L x 26.0W x 15.2H cm) and allowed to habituate
128 for 10 min. A baseline (no behavior) recording was captured via a Plexon Multi-Channel photometry
129 system for 5 min before two 20-min social exposure test sessions (i.e. Session A and Session B): SS
130 males interacting with their same-sex cagemate or a novel same-sex conspecific and OS males interacting
131 with two different novel females (one which will serve as their partner moving forward). Stimulus order
132 was randomized between subjects, and there was a 20 min nonsocial “rest” period between each social
133 exposure. Immediately following Session B, males in the OS group were permanently housed with one of
134 the two females they had interacted with during the social exposure test. Social exposure tests were also
135 conducted on Experimental days 2 and 8, with all partner interactions occurring with the same
136 partner/cage mate while all stranger interactions occurred with a conspecific that the subject had never
137 previously interacted with. All social exposure behavior sessions were video recorded at 30 frames per
138 second using a Logitech web camera. Fiber photometry data from the 410, 450, and 560 nm channels
139 were captured at 30 fps. Cameras were connected to an input relay device attached to the Plexon Multi-
140 Channel photometry system that signaled the start of the video recording to the system for time-locking
141 behavioral assessments with the photometry signals. Social behaviors were coded frame-by-frame using
142 Solomon Coder, and timestamps for the onset of all behaviors were extracted and used as events for
143 photometry analysis. Social behaviors were analyzed for frequency and duration, including social
144 proximity, olfactory investigation, affiliative behaviors (side-by-side, huddling, allogrooming), and
145 agonistic behaviors (boxing, defensive treading, fleeing from the stimulus, and chase/tumble sequences).

146

147 *Tissue Collection and Confirmation of Virus Placement*

148 At the end of the experiment, subjects were deeply anesthetized with a ketamine/dexmedetomidine
149 cocktail, then perfused with saline and 4% paraformaldehyde in 0.1M Phosphate Buffer solution (pH =
150 7.4). Brains were extracted, post-fixed overnight, transferred to 30% sucrose, and sectioned at 30 μ m.
151 Every other section through the aIC/mIC (+1.90mm-0.20mm from bregma) was mounted onto glass slides
152 and coverslipped using a hard-set, antifade coverslipping medium (Gelvatol, made in-house). Once dry,
153 sections were viewed under a Leica microscope using a chroma 488 filter (L5-ET, Leica) to visualize the
154 GFP-conjugated GCaMP virus and a 555 nm filter (RHOD-ET, Leica) to visualize the mApple-
155 conjugated GRAB_{DA}. Animals were removed from the study if viral expression or optical fiber implant
156 was found outside of the IC.

157

158 *Fiber Photometry Analysis*

159 Raw fiber photometry output for the 410, 450, and 560 nm channels was exported from Plexon
160 Software, time-locked to the start of video recording, then analyzed using open-source fiber photometry
161 analysis software (Guided Photometry Analysis in Python; GuPPy) [24]. Motion artifact was corrected for
162 in GuPPy by using raw signal from the isobestic (410) channel. Timestamps for each behavior were used
163 as events to examine event-related changes in aIC calcium and DA activity corresponding to specific
164 social behaviors. Area under the curve (AUC) was calculated for both anticipatory (-2 to 0 sec prior to
165 behavior onset) and initiation-induced (0 to 5 sec) brain activity responses to each social behavior.

166

167 Experiment 2: Effects of same- vs opposite-sex cohabitation on DA receptor mRNA expression

168 *Experimental Design, Brain Collection, and qRT-PCR*

169 A separate cohort of male prairie voles (N=36) were divided into same-sex paired (SS, n=11), and
170 opposite-sex paired groups that cohabitated with a tubal ligated female for either 48 h (OS-ST, n=13) or 1
171 week (OS-LT, n=12). Immediately prior to tissue collection, subjects were tested for the expression of a
172 pair bond using the partner preference test (PPT; pair bonding was characterized by an animal spending
173 greater than or equal to a 3:1 ratio of affiliation with their partner vs the stranger, [25]). Subjects were
174 rapidly decapitated, and brains were extracted and stored at -80°C before being sectioned at 200 µm using
175 a cryostat (Leica). Unilateral tissue punches (1.0 mm diameter) were collected from the aIC. Tissue
176 punches were homogenized, and RNA was extracted and purified using a Qiagen RNEasy mini kit
177 following manufacturer's instructions. RNA was quantified using a Qubit 3 fluorometer and a high
178 sensitivity RNA quantification kit (Thermo Fisher Scientific, Waltham, MA). 20-100 ng of mRNA was
179 converted to cDNA using a high-capacity reverse transcription kit (Applied Biosystems, Foster City, CA)
180 per the manufacturer's instructions. mRNA for dopamine receptor 1 (DRD1) and dopamine receptor 2
181 (DRD2) were analyzed using probes designed from target gene sequences of the prairie vole genome [25].
182 Hypoxanthine-guanine phosphoribosyltransferase (HPRT) was used as the comparison "housekeeping"
183 gene based on its relatively constant expression in cells independent of experimental conditions [26, 27].
184 qRT-PCR for each target was run in triplicate for every subject on one plate with wells containing 5 ng
185 cDNA, SYBR green PCR Master Mix (Applied Biosystems, Foster City, CA) and 200 nM of each
186 forward and reverse primer (see [25] for specific sequences). A ThermoFisher StepOnePlus PCR plate
187 reader was used for quantification. A dissociation curve was generated for each sample and used to
188 confirm that only a single product was transcribed. The $\Delta\Delta CT$ method was used to calculate the fold
189 differences between groups [28].

190

191 *Data Analyses*

192 Significant differences were determined by a p-value of <0.05 for all analyses. In Experiment 1,
193 social behaviors and behavior-related AUC results were analyzed using linear mixed modeling (LMM)
194 using test session (Pairing Day, ST, LT) and stimulus type (Partner vs Stranger) as within-groups
195 variables and companion sex (SS vs OS paired) as a between groups variable. Significant main effects and
196 interactions were followed by Bonferroni post-hoc analyses to further elucidate the data patterns. The
197 mRNA results in Experiment 2 violated the One-Way ANOVA assumption of homogeneity of variances
198 between groups, so Kruskal-Wallis was used to analyze group differences in DRD1 and DRD2
199 expression. Significant results were followed by Bonferroni post-hoc analyses.

200

201 **Results**

202 *Experiment 1-1: Companion sex and pairing length influence partner- and stranger-directed social*
203 *behaviors*

204 All subjects spent more time in social proximity (F=8.234, p = 0.008) and side-by-side contact (F
205 = 7.19, p = 0.013) with their partner compared to the stranger conspecific and were more aggressive to the
206 stranger than partner (F = 7.19, p = 0.013). However, there were significant 3-way interactions between

207 time, companion sex, and social stimulus type for each of these behaviors. Specifically, SS males spent
208 significantly more time in proximity (Fig 2A) and side-by-side contact (Fig 4A) with their partner
209 compared to a stranger on the first day of testing but not subsequent test periods. In contrast, OS males
210 were in proximity (Fig 2B) and side-by-side contact (Fig 4A) to the two novel females during the first
211 encounter, but males spent more time in proximity and side-by-side contact to their female partner than
212 the stranger female after cohabitating with their partner for two and seven days. In addition, SS males
213 showed the most stranger-directed aggression during the first social exposure test while OS males were
214 not aggressive before cohabitation then became significantly more aggressive towards the stranger during
215 subsequent social exposure tests (ST and LT timepoints).

216

217 *Experiment 1-2: Companion sex and pairing length influences social behavior-related GCaMP and*
218 *GRAB_{DA} signal in the aIC*

219 There were several behavior-dependent main effects and interactions for GCaMP and GRAB_{DA}
220 responses across different conditions. When subjects initiated proximity with a social stimulus, there was
221 a significant difference in the size (AUC) of GCaMP transients between SS and OS subjects, with
222 GCaMP in the aIC showing a larger proximity-induced response overall in SS subjects compared to OS
223 subjects ($F = 5.170$, $p = 0.026$). However, this main effect of companion sex appears to be driven by an
224 interaction with stimulus type ($F = 8.852$, $p = 0.004$), with SS males showing a substantially higher total
225 aIC GCaMP response to proximity with strangers compared to OS males in proximity to strangers
226 (Bonferroni corrected pairwise comparison $t = 3.695$, $p < 0.001$; Fig 2C-H). There was also a significant
227 interaction between companion sex and test day, such that SS males showed a larger response than OS
228 males on the first (pairing; $t = 3.973$, $p < 0.001$) test day but not the second (short-term paired) or the final
229 (long-term paired) test days. Finally, there was a significant interaction between social stimulus type and
230 testing day ($F = 9.206$; $p < 0.001$), with there being a significantly higher response to proximity with the
231 partner compared to the stranger on the final test day (regardless of SS or OS pairing; $t = 3.954$, $p <$
232 0.001). aIC GCaMP response to partner proximity in OS males increased significantly as pairing length
233 continued, while this pattern did not emerge for OS males in proximity to strangers or SS males in
234 proximity to either their partner or to strangers.

235 GRAB_{DA} transient size (AUC) in the aIC during social proximity changed significantly across
236 testing day ($F = 11.133$, $p < 0.001$). However, this main effect was driven by a significant 3-way
237 interaction between companion sex, stimulus type, and test day ($F = 5.761$, $p = 0.005$; Fig 2I-N).
238 Specifically, SS males showed a greater DA response to stranger proximity than OS males on the first
239 testing day ($t = 2.186$, $p = 0.032$). Furthermore, this DA response to stranger proximity in SS males also
240 significantly differed from their DA response to partner proximity, with an inversion of response
241 occurring where strangers elicited significantly greater DA release on the first test day ($t = 2.019$, $p =$
242 0.047) whereas partners elicited greater DA release on the last test day ($t = 3.507$, $p < 0.001$). DA activity
243 in the aIC of OS males in social proximity did not differ across social stimulus type or testing day.

244 During social olfactory investigation, SS males showed a significantly greater overall GCaMP
245 response compared to OS males ($F = 6.882$, $p = 0.011$; Fig 3C-H). DA response, however, showed a
246 significant interaction between companion sex and test day ($F = 3.836$, $p = 0.026$), with SS males having
247 an overall larger DA response to olfactory investigation of strangers than OS males did when
248 investigating strangers ($t = 2.030$, $p = 0.046$; Fig 3I-N). Finally, DA response to olfactory investigation
249 between partners and strangers in SS males showed the same pattern as proximity, where sniffing

250 strangers elicited greater DA activity on the first test day ($t = 2.276$, $p = 0.026$), while sniffing partners
251 elicited more DA activity than sniffing strangers did on the subsequent testing days (short-term: $t = 2.034$,
252 $p = 0.046$; long-term: $t = 2.271$, $p = 0.026$).

253 Upon initiation of side-by-side contact with partners, OS males showed a significantly greater aIC
254 GCaMP response compared to SS males (main effect of companion sex: $F = 4.494$, $p = 0.046$). Similarly,
255 there appeared to be a greater anticipatory response in OS males, as indicated by a significantly higher
256 area under the curve during the 2 sec immediately before initiation of side-by-side contact compared to SS
257 males ($F = 8.313$, $p = 0.007$). Although there were no significant effects of side-by-side contact initiation
258 with the partner on DA activity in the aIC based on companion sex or testing day, there was an
259 anticipatory interaction between companion sex and testing day ($F = 12.203$, $p < 0.001$). Specifically, OS
260 males showed significantly greater DA activity just before (2 sec) initiation of side-by-side contact with
261 their partner on the second day of testing (after 48 hrs of pairing) compared to SS males ($t = 3.483$, $p =$
262 0.002). This DA response to anticipation of side-by-side partner contact in OS males after 48 hours of
263 pairing was also significantly higher than the response on pairing day and after one week of pairing.

264 There was a significant effect of companion sex on GCaMP activity to initiation of stranger-
265 directed aggression where SS males showed a significantly higher total response compared to OS males
266 ($F = 5.468$, $p = 0.028$). There were no effects of companion sex or test day on DA activity in response to
267 stranger-directed aggression, nor were there any anticipatory effects for either GCaMP or DA activity.

268

269 *Experiment 2: Opposite-sex cohabitation alters DA receptor mRNA expression in the aIC*

270 There was a significant group effect on mRNA expression of DRD1 ($F_{(33,2)} = 6.911$, $p = 0.032$)
271 and DRD2 ($F_{(32,2)} = 11.096$, $p = 0.004$) receptors in the aIC. Specifically, subjects in the OS-ST pairing
272 group had significantly higher DRD1 mRNA expression compared to the SS group ($t = 11.343$, $p =$
273 0.026), while both opposite-sex pairing groups had significantly higher DRD2 mRNA expression
274 compared to the SS group in the aIC (OS-ST vs SS: $t = 13.90$, $p = 0.004$; OS-LT vs SS: $t = 10.90$, $p =$
275 0.039).

276

277 Discussion

278 Prairie voles increased display of partner-directed affiliation and stranger-directed aggression as
279 the length of opposite-sex pairing increases, findings that replicate previous work [23, 29]. Furthermore,
280 during social encounters with a novel social stimulus, proximity-induced aIC activity was significantly
281 lower in pair bonded males than bachelor males, most notably during the first and second novel encounter
282 sessions. Bachelor males also had significantly greater GCaMP transients to stranger proximity than
283 partner proximity, suggesting that the bachelor aIC responds more to social novelty. A recent study in
284 mice showed that interacting with a novel conspecific increased calcium activity in most socially-
285 responsive aIC neurons, particularly during stationary contact behavior [30]. This aligns with the
286 proposed role of the aIC in social decision-making, as the bulk of its social interaction-induced calcium
287 transients occurs during stationary “processing” phases of the social encounter then relays the behavioral
288 “decision” to other brain regions [10]. Social-related aIC activity also appears to be modulated by pair
289 bonding, as evidenced by significantly greater aIC GCaMP transients to partner proximity compared to
290 stranger proximity in the pair bonded males. Bachelor males also exhibited stable, positive GCaMP
291 transients during peer proximity throughout the experiment. Continuous interaction with a social

292 companion may strengthen the pathways from sensory-related areas to the IC to facilitate recognition of a
293 familiar social companion. It has recently been shown that excitotoxic lesions to the aIC impair social
294 recognition [12, 31, 32]. Thus, the ability to recognize and associate socio-sensory cues with a particular
295 social partner appears to require activity of the aIC.

296 DA release and receptor expression have been previously shown to modulate both pair bond
297 formation and maintenance in prairie vole males [33]. For instance, DRD2 receptors in the NAcSh are
298 critical for partner preference formation [22]. Conversely, DRD1 receptor expression is upregulated in the
299 NAcSh after long-term opposite-sex cohabitation, and their activity is required for stranger-directed
300 aggression [34, 35]. DRD1-expressing neurons are also necessary for the display of appetitive aspects of
301 partner motivation since lever pressing for a pair bonded partner causes increased NAcSh DA signaling
302 and blocking DRD1 receptors significantly reduces lever pressing for a partner [36]. Surprisingly, DA-
303 related mechanisms during the transition period from short-term to long-term pair bonding have rarely
304 been studied. Here, we noticed a significant increase in partner-directed side-by-side contact in OS males
305 after 2 days of pairing, and though this behavior remained high after a week of pairing, the average
306 duration did drop slightly. Interestingly, both aIC GCaMP and GRAB_{DA} transients during initiation of
307 side-by-side contact were significantly higher than aIC transients in SS males to side-by-side contact only
308 at this early pairing timepoint. Agmo et al. [37] noted a similar behavioral pattern in pair bonded black
309 tufted-ear marmosets (*Callithrix jacchus*), with huddling and allogrooming behaviors peaking on days
310 2-6 of pairing and then dropping slightly thereafter. Although this behavioral pattern has not been
311 consistently observed in prairie voles [29], the significant increase in aIC GCaMP activity and DA release
312 during side-by-side contact at this stable but still early phase of a pair bond could signify a
313 neurobiological epoch that forms the foundation of a long-term relationship by linking motivational
314 systems to social contact with their partner. Indeed, both affective touch and physiologic sensory input (or
315 interoception) are relayed from the body to the brain via small, unmyelinated afferents that converge in
316 the mid-posterior IC [38-40], implying that social touch can be a powerful modulator of physiological and
317 emotional state via the IC.

318 In mice, the activity of DRD1-expressing neurons in the aIC projecting to the lateral NAcSh
319 predicts both affiliative behaviors during a prosocial interaction and agonistic behaviors during an
320 aversive social experience [41]. Thus, social stimuli-dependent synaptic plasticity originating from aIC^{D1}
321 neurons appears to drive contextually appropriate learned social behaviors. Furthermore, the pair bonding
322 process seems to involve its own form of DA-related neural plasticity since DRD1 and DRD2 mRNA
323 expression were increased in the aIC of pair bonded male prairie voles compared to bachelors. This means
324 that while a similar DA response occurs in the aIC during both partner and novel social encounters after
325 pair bonding, changes in DA receptor expression resulting from pair bonding may bias how aIC neurons
326 “interpret” this DA signal to drive social valence-specific behaviors. aIC DRD1-expressing neurons
327 project to DRD1-expressing neurons in the NAcSh and respond to social stimuli in mice [41], and NAcSh
328 DRD1 receptors are required for the display of bond-specific behaviors in prairie voles [35, 36]. Thus,
329 DA in the aIC may play an important and overlooked role upstream to modulate the pivotal role of NAc
330 DA and its receptors in prairie vole pair bonding. We have previously observed changes in DA receptor
331 expression in the aIC for prairie voles experience loss of a pair bonded partner [25]. These changes may
332 serve an important role in social recognition and salience and valence assignment to specific social
333 encounters or experiences. Proper integration of such socio-contextual information is a crucial first step
334 when engaging in social encounters and forms the framework for establishing and maintaining a variety
335 of social relationships.

337 Figure Captions

338 Figure 1: Schematic of experimental procedures and timelines.

339 (A) GRAB_{DA} and GCaMP expression and optical fiber placement in the anterior insular cortex. (B) Fiber
340 Photometry recording was performed during each social exposure session. Subjects were exposed to one
341 of two potential social conspecifics during Session A (partner or stranger), followed by a non-social “rest”
342 period. During Session B, subjects were exposed to whichever social stimuli was not introduced in
343 Session A. (C) Viral injection and fiber implantation occurred 3 weeks before the start of the experiment.
344 On Day 0, subjects were randomly assigned to either the same-sex (SS) or opposite-sex (OS) paired
345 groups. SS paired males remained with their same-sex cage mate and were exposed to their cage mate and
346 a stranger during the Day 0 social exposure test. OS paired males were exposed to two novel females
347 during the Day 0 social exposure, with one of these females becoming the subjects’ partner immediately
348 following the test. Social exposure tests also occurred on Days 2 (ST pairing) and 8 (LT pairing). (D) A
349 separate cohort of male subjects were randomly divided into 3 groups: SS housed subjects were never
350 paired with a female, while two OS groups were cohabitated with a female for either 2 days (ST-OS) or 1
351 week (LT-OS). Brains were collected from all three groups and tissue was used for qRT-PCR analysis.
352 (E) Tissue punches were collected unilaterally through the anterior IC. (F) A partner preference test was
353 used to confirm pair bonding in both OS groups and occurred several hours before subjects were
354 sacrificed.

355 Figure 2: Social proximity-induced GCaMP and GRAB_{DA} activity in the aIC.

356 SS paired subjects (A) and OS paired subjects (B) show differing patterns of close proximity to partners
357 vs strangers across testing days. GCaMP relative fluorescence during close proximity to partners vs
358 strangers in SS and OS paired subjects during social exposure tests on experimental Day 0 (C), Day 2 (D),
359 and Day 8 (E) with corresponding area under the curve (AUC) calculations (F-H). GRAB_{DA} relative
360 fluorescence during close proximity to partners vs strangers in SS and OS paired subjects during social
361 exposure tests on Day 0 (I), Day 2 (J), and Day 8 (K) with corresponding AUC calculations (L-N).
362 ^denotes $p < 0.05$ for partner vs stranger within pairing group, *denotes $p < 0.05$ for SS vs OS pairing
363 condition within the same social stimulus type.

364 Figure 3: Olfactory investigation-induces GCaMP and GRAB_{DA} activity in the aIC.

365 SS paired subjects (A) and OS paired subjects (B) do not significantly differ in their patterns of olfactory
366 investigation of partners vs strangers across testing days. GCaMP relative fluorescence during olfactory
367 investigation of partners vs strangers in SS and OS paired subjects during social exposure tests on
368 experimental Day 0 (C), Day 2 (D), and Day 8 (E) with corresponding area under the curve (AUC)
369 calculations (F-H). GRAB_{DA} relative fluorescence during olfactory investigation of partners vs strangers
370 in SS and OS paired subjects during social exposure tests on Day 0 (I), Day 2 (J), and Day 8 (K) with
371 corresponding AUC calculations (L-N). ^denotes $p < 0.05$ for partner vs stranger within pairing group,
372 *denotes $p < 0.05$ for SS vs OS pairing condition within the same social stimulus type.

373 Figure 4: Affiliation-induced GCaMP and GRAB_{DA} activity in the aIC.

374 SS and OS paired subjects show selective affiliation with their partners and rarely affiliate with strangers
375 (A). GCaMP relative fluorescence and AUC (B) and GRAB_{DA} relative fluorescence and AUC (C) during
376 side-by-side contact on Day 0 partner exposure session, Day 2 partner exposure session (D-E), and Day 8
377 partner exposure session (F-G). ^denotes $p < 0.05$ for partner vs stranger within pairing group, *denotes

378 p<0.05 for SS vs OS pairing condition within the same social stimulus type, different letters denote
379 p<0.05 for same stimulus type across testing sessions for OS paired subjects.

380 Figure 5: DRD1 and DRD2 mRNA expression in the aIC.

381 OS pairing is characterized by a selective increase DRD1 mRNA after 2 days but not 1 week of opposite-
382 sex cohabitation in comparison to SS paired subjects. DRD2 mRNA expression is significantly higher in
383 both OS pairing groups compared to SS males. Different letters denote p<0.05 between groups.

384

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389 **Author Contributions**

390 EV and AS conceptualized and designed all experiments; EV collected and analyzed all data with
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398

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401 **Competing Interests**

402 The authors have nothing to disclose.

403

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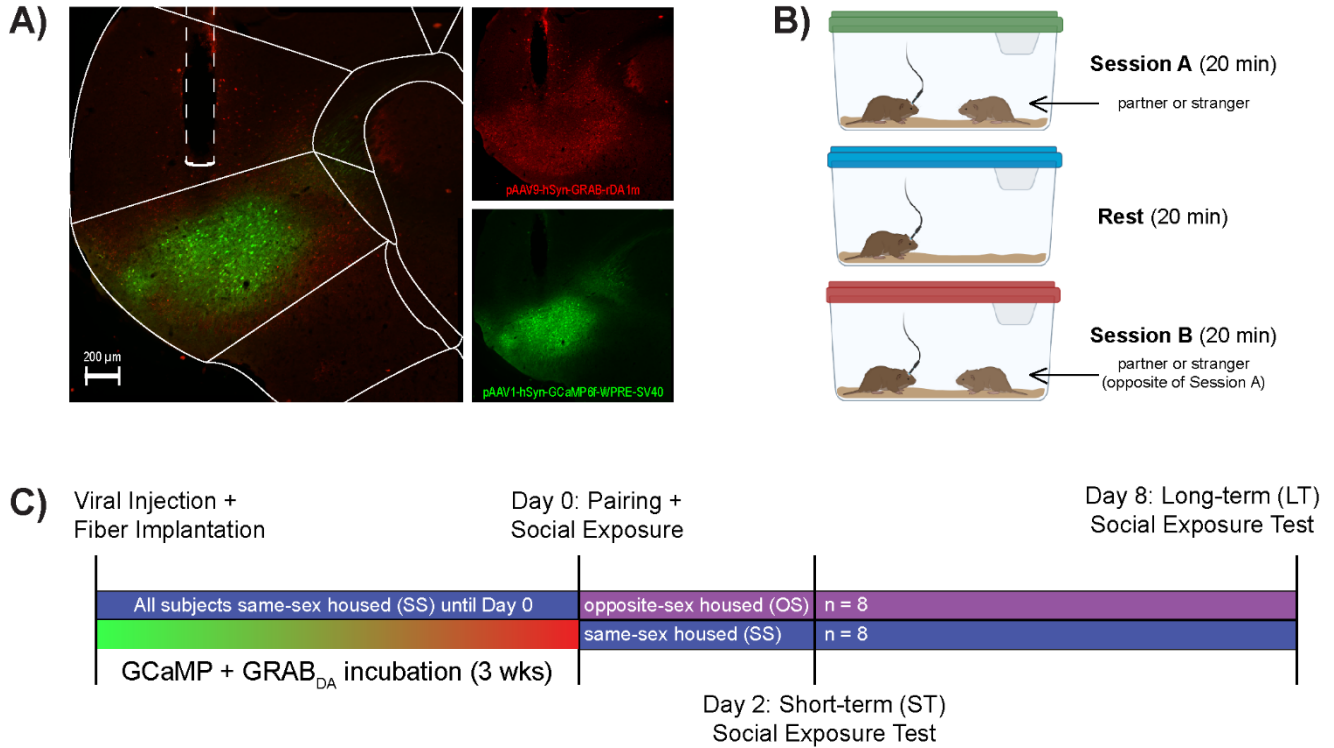
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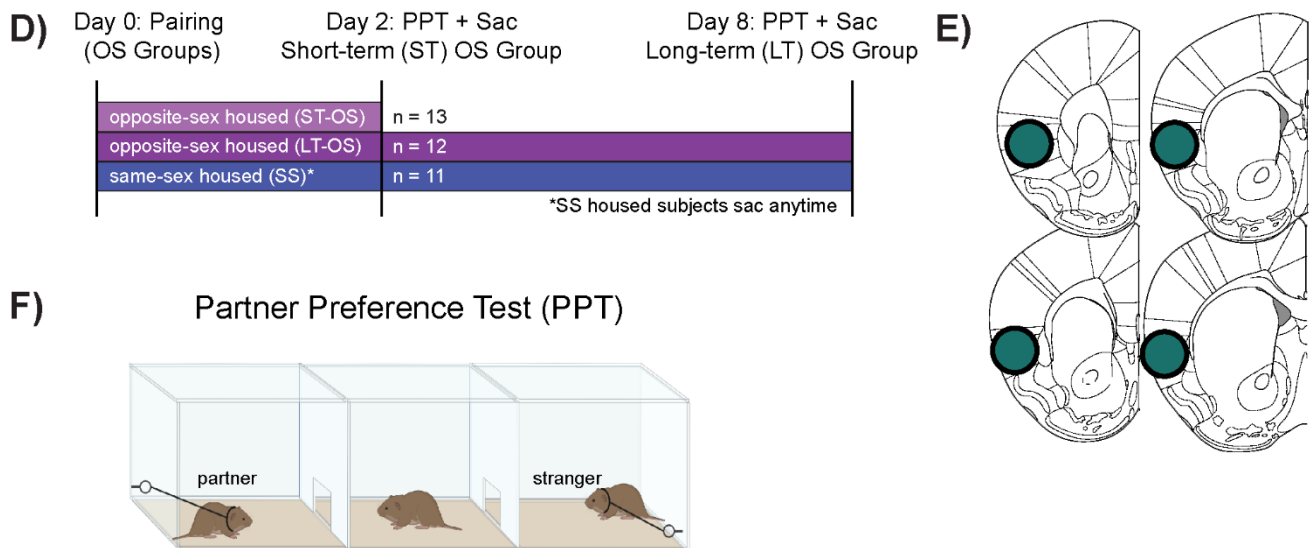
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Experiment 1

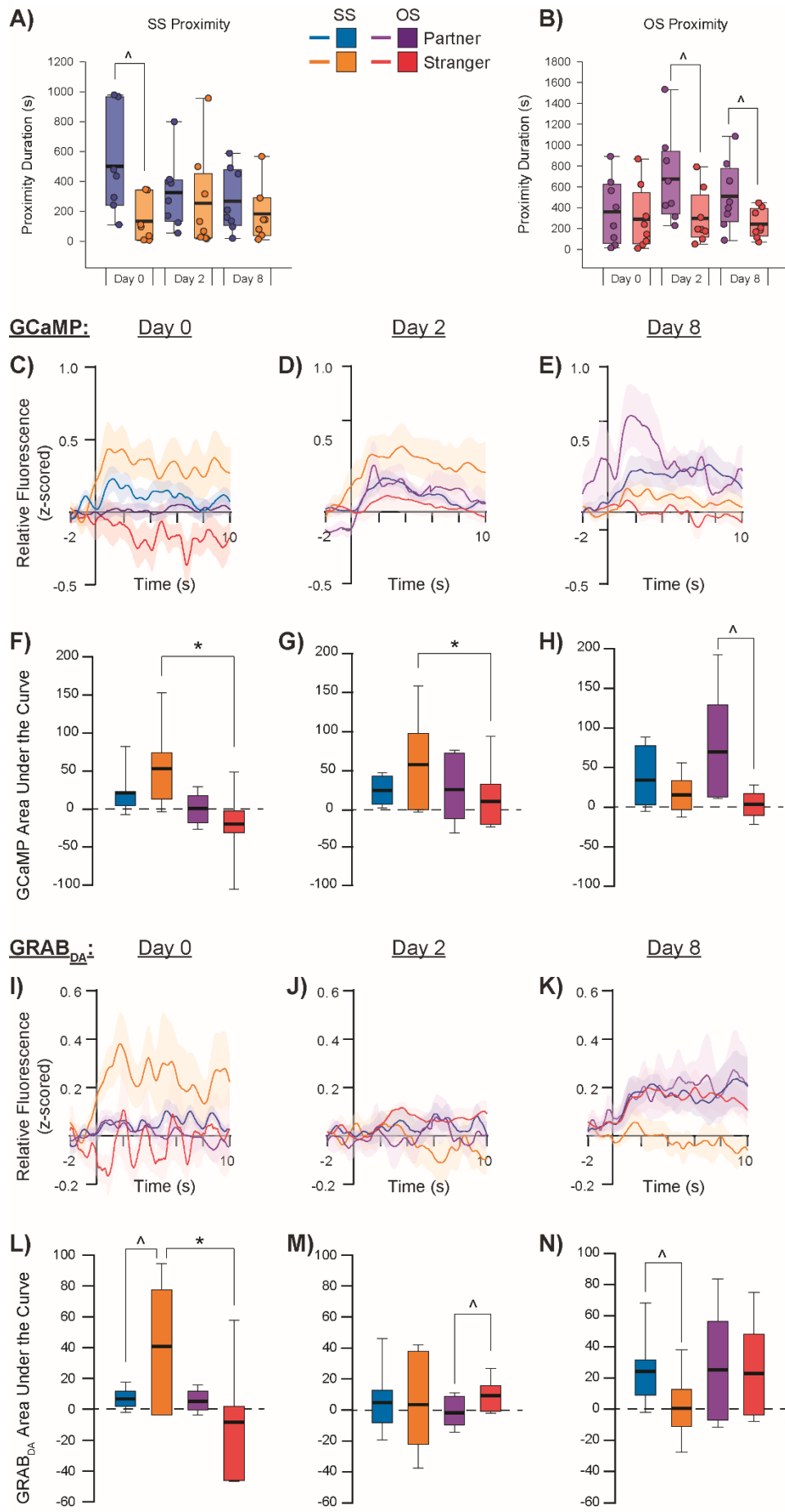


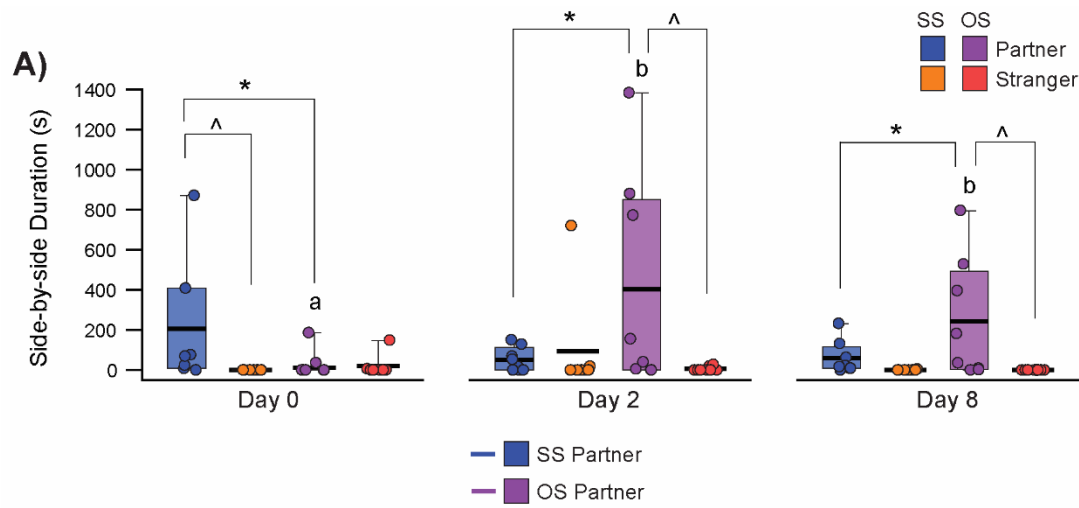
Experiment 2



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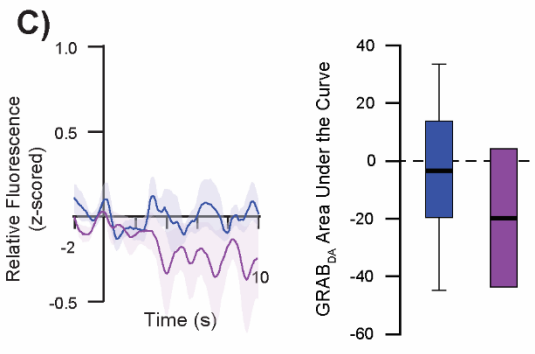
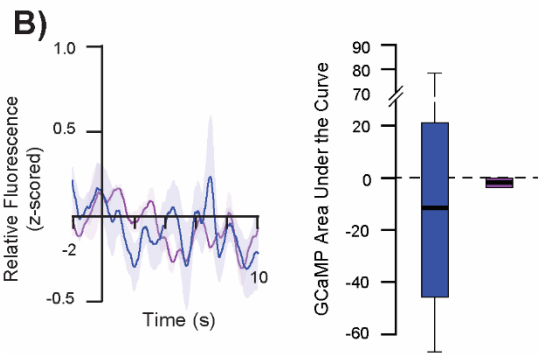
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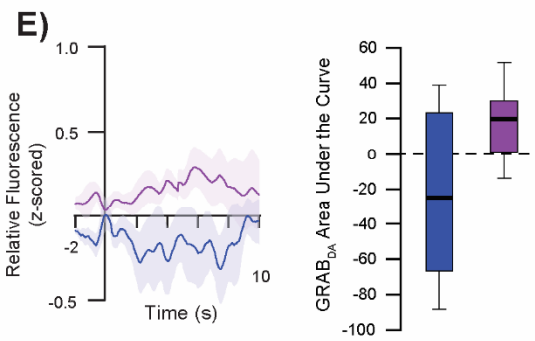
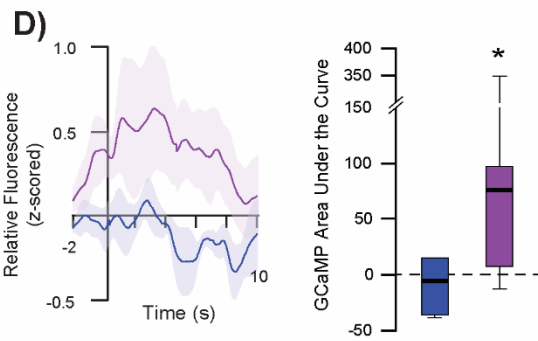
GCaMP: Day 0

GRAB_{DA}: Day 0



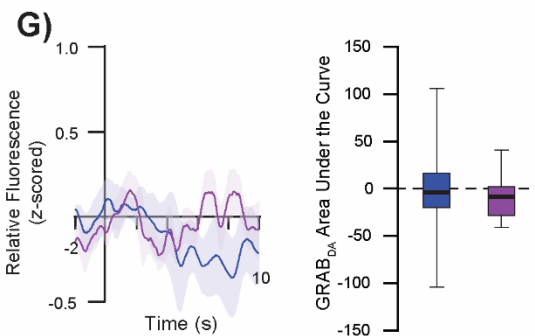
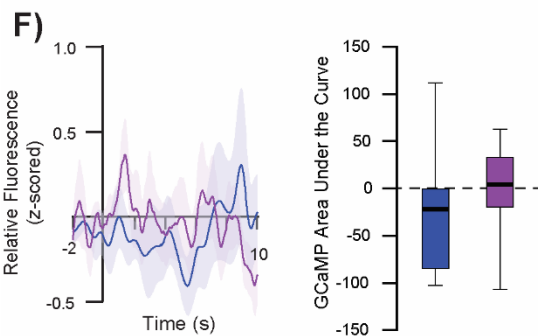
GCaMP: Day 2

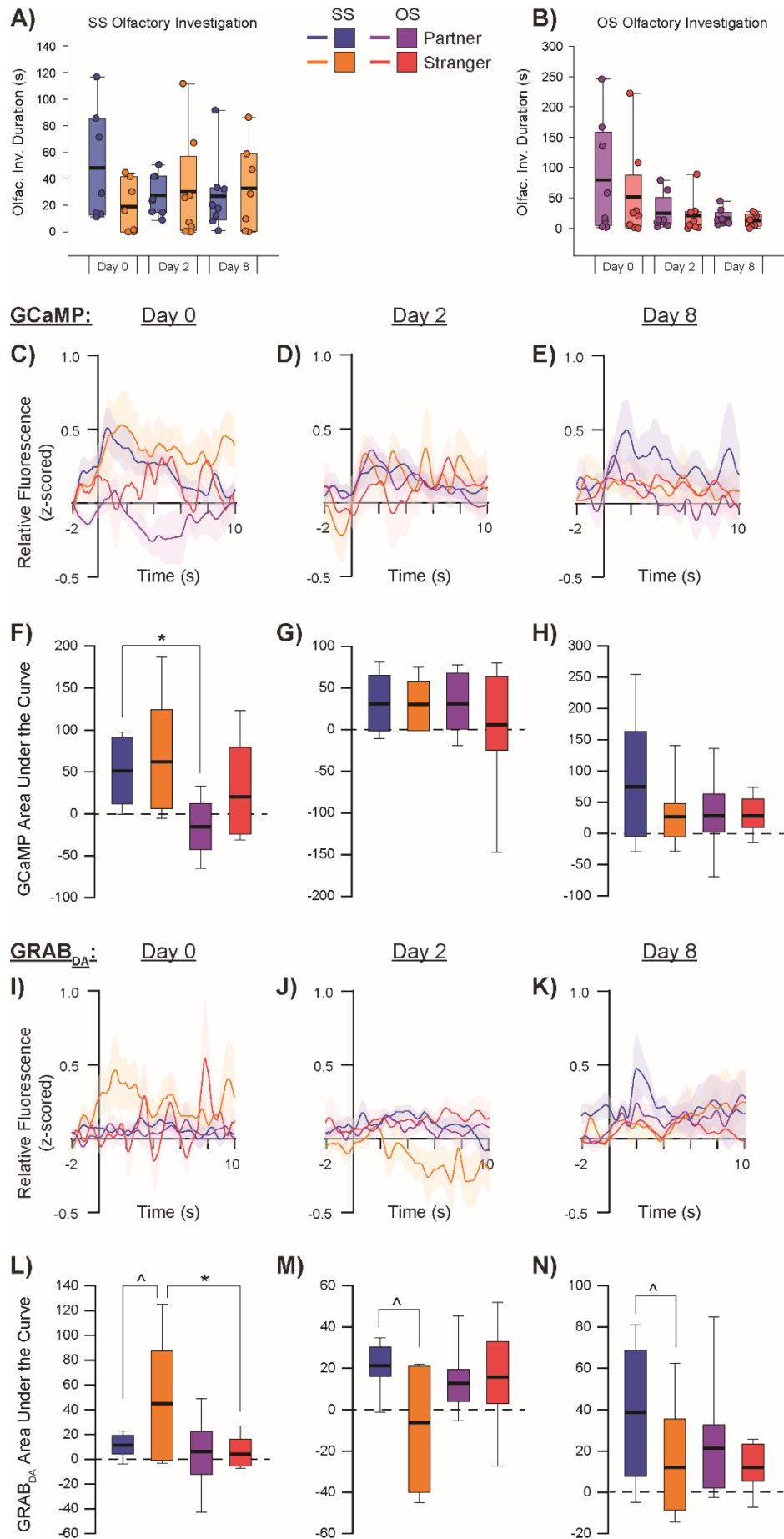
GRAB_{DA}: Day 2



GCaMP: Day 8

GRAB_{DA}: Day 8





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