



# Complete Genome Sequence of a Zika Virus Strain Isolated from the Serum of an Infected Patient in Thailand in 2006

Narong Nitatpattana,<sup>a</sup> Kumchol Chaiyo,<sup>a</sup> Supoth Rajakam,<sup>a</sup> Kanya Poolam,<sup>a</sup> Kusuma Chansiprasert,<sup>a</sup> Norapath Pesirikan,<sup>b</sup> Supanat Buree,<sup>a</sup> Ekkarat Rodpai,<sup>a</sup> Sutee Yoksan<sup>a</sup>

<sup>a</sup>Institute of Molecular Biosciences, Mahidol University, Nakhon Pathom, Thailand

<sup>b</sup>Government Pharmaceutical Organization Ministry of Public Health, Bangkok, Thailand

**ABSTRACT** The complete genome of Zika virus (ZIKV) strain CVD\_06-274 was isolated from the serum of an infected patient in Thailand in 2006. Phylogenetic analysis showed that this strain belongs to the Asian lineage and also high titers in Vero cells (RCB 10-87). It has potential for development as an inactivated ZIKV vaccine.

Zika virus (ZIKV) is an icosahedral enveloped arbovirus. ZIKV is a member of the genus *Flavivirus*, family *Flaviviridae*. This family also includes dengue virus (DENV), West Nile virus (WNV), and yellow fever virus (YFV) (1). The ZIKV genome is a single-stranded positive-sense RNA approximately 11 kb in length. The viral genome consists of a single open reading frame (ORF) flanked with 5'- and 3'-untranslated regions (UTRs). The ORF encodes 3 structural proteins and 7 nonstructural proteins (2). The structural proteins play a role in viral particle assembly and pathogenicity. The nonstructural proteins and UTRs play a role in viral replication (3). This phylogenetic relationship indicated that ZIKV has 2 major lineages, African and Asian. The Asian-lineage ZIKV was reported in an outbreak in the South Pacific islands, South America, and Asia during 2013 to 2017 (2).

We report the complete genome sequence of ZIKV strain CVD\_06-274, isolated from the serum of an infected patient in Thailand in 2006; this study was approved by the Siriraj Ethics Committee, Mahidol University (Si 180/2006). The serum sample was inoculated into *Toxorhynchites splendens* mosquitoes for 3 passages (4), followed by WHO-certified Vero cells (RCB 10-87) for 3 passages. The infected culture supernatant was positive for flavivirus (5) and ZIKV (6) by real-time quantitative reverse transcription-PCR (qRT-PCR) using the Kapa Probe Fast one-step qRT-PCR kit (Kapa Biosystems, USA) on a Chromo4 system (Bio-Rad, USA). The CVD\_06-274 isolate was titrated by plaque assay in an LLC-MK2 cell (7).

Viral RNA was extracted using the viral RNA minikit (Qiagen, Germany) and converted to cDNA using the Maxima H Minus first-strand cDNA synthesis kit (Thermo Scientific, USA). cDNA was amplified using the Phusion Flash high-fidelity PCR master mix (Thermo Scientific). A set of ZIKV whole-genome sequencing primers (8) was used. PCR products were sequenced using the Sanger method (1st Base, Malaysia). Sequences were analyzed using BioEdit 7.0.5 software (9) and the Basic Local Alignment Search Tool (BLAST; <https://www.ncbi.nlm.nih.gov/BLAST>). The whole-genome sequence of the CVD\_06-274 isolate and other ZIKV strains from GenBank were used for phylogenetic tree analysis. A neighbor-joining tree was constructed with 1,000 bootstrap replicates using MEGA6 (10).

We obtained the genome sequence of CVD\_06-274, with a total length of 10,623 nucleotides. BLAST analysis showed that the highest identity value, at 99.3%, was to Zika virus/H.sapiens-tc/KHM/2010/FSS13025 strain (GenBank accession number KU955593, Cambodia, 2010) and SK403/13AS strain (GenBank accession number KX051561, Thailand, 2013). A phylogenetic tree revealed that the CVD\_06-274 isolate

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Address correspondence to Narong Nitatpattana, [narong.nit@mahidol.ac.th](mailto:narong.nit@mahidol.ac.th).

belongs to the Asian lineage. CVD\_06-274 produces a medium plaque size (2 to 3 mm) in LLC-MK2 cells. This virus showed a high level of replication at a peak titer of  $8 \log_{10}$  PFU/ml in WHO-certified Vero cells, on a par with inactivated-ZIKV vaccine strain PRVABC59 (GenBank accession number KU501215), which was developed by the Walter Reed Army Institute of Research (WRAIR, USA) (11). An inactivated vaccine has high safety and less reactivity, and a high viral titer is required to achieve sufficient immunogenicity. The high level of replication of CVD\_06-274 in a vaccine production cell line indicated a potential for further development into an inactivated Zika vaccine.

**Accession number(s).** The assembled complete genome sequence of the Zika virus CVD\_06-274 was submitted to GenBank under the accession number [MG645981](https://doi.org/10.1093/mbe/mz005).

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## REFERENCES

1. Marano G, Pupella S, Vaglio S, Liumbruno GM, Grazzini G. 2015. Zika virus and the never-ending story of emerging pathogens and transfusion medicine. *Blood Transfus* 14:95–100. <https://doi.org/10.2450/2015.0066-15>.
2. Wang A, Thurmond S, Islas L, Hui K, Hai R. 2017. Zika virus genome biology and molecular pathogenesis. *Emerg Microbes Infect* 6:e13. <https://doi.org/10.1038/emi.2016.141>.
3. Hasan SS, Sevvana M, Kuhn RJ, Rossmann MG. 2018. Structural biology of Zika virus and other flaviviruses. *Nat Struct Mol Biol* 25:13–20. <https://doi.org/10.1038/s41594-017-0010-8>.
4. Jirakanjanakit N, Khin MM, Yoksan S, Bhamarapavati N. 1999. The use of *Toxorhynchites splendens* for identification and quantitation of serotypes contained in the tetravalent live attenuated dengue vaccine. *Vaccine* 17:597–601. [https://doi.org/10.1016/S0264-410X\(98\)00239-4](https://doi.org/10.1016/S0264-410X(98)00239-4).
5. Moureau G, Temmam S, Gonzalez JP, Charrel RN, Grard G, de Lamballerie X. 2007. A real-time RT-PCR method for the universal detection and identification of flaviviruses. *Vector Borne Zoonotic Dis* 7:467–477. <https://doi.org/10.1089/vbz.2007.0206>.
6. Faye O, Faye O, Dupressoir A, Weidmann M, Ndiaye M, Alpha Sall A. 2008. One-step RT-PCR for detection of Zika virus. *J Clin Virol* 43:96–101. <https://doi.org/10.1016/j.jcv.2008.05.005>.
7. Baer A, Kehn-Hall K. 2014. Viral concentration determination through plaque assays: using traditional and novel overlay systems. *J Vis Exp* 93:e52065. <https://doi.org/10.3791/52065>.
8. Leguia M, Cruz CD, Felices V, Torre A, Troncos G, Espejo V, Guevara C, Mores C. 2017. Full-genome amplification and sequencing of Zika viruses using a targeted amplification approach. *J Virol Methods* 248:77–82. <https://doi.org/10.1016/j.jviromet.2017.06.005>.
9. Hall TA. 1999. BioEdit: a user-friendly biological sequence alignment editor and analysis program for Windows 95/98/NT. *Nucleic Acids Symp Ser* 41:95–98.
10. Tamura K, Stecher G, Peterson D, Filipski A, Kumar S. 2013. MEGA6: Molecular Evolutionary Genetics Analysis version 6.0. *Mol Biol Evol* 30:2725–2729. <https://doi.org/10.1093/molbev/mst197>.
11. Larocca RA, Abbink P, Peron JPS, de A Zanotto PM, Lampietro MJ, Badamchi-Zadeh A, Boyd M, Ng'ang'a D, Kirilova M, Nityanandam R, Mercado NB, Li Z, Moseley ET, Bricault CA, Borducchi EN, Giglio PB, Jetton D, Neubauer G, Nkolola JP, Maxfield LF, De La Barrera RA, Jarman RG, Eckels KH, Michael NL, Thomas SJ, Barouch DH. 2016. Vaccine protection against Zika virus from Brazil. *Nature* 536:474–478. <https://doi.org/10.1038/nature18952>.