

Manipulating the Gut Microbiome as a Therapeutic Strategy to Mitigate Late Effects in Childhood Cancer Survivors

Technology in Cancer Research & Treatment
Volume 22: 1-13
© The Author(s) 2023
Article reuse guidelines:
sagepub.com/journals-permissions
DOI: 10.1177/15330338221149799
journals.sagepub.com/home/tct


Lixian Oh, PhD¹, Syaza Ab Rahman, MBBCh¹, Kailey Dubinsky, BSc¹,
Mohamad Shafiq Azanan, PhD¹, and Hany Ariffin, MBBS, PhD¹ 

Abstract

Recent studies have identified causal links between altered gut microbiome, chronic inflammation, and inflammation-driven conditions such as diabetes and cardiovascular disease. Childhood cancer survivors (CCS) show late effects of therapy in the form of inflammaging-related disorders as well as microbial dysbiosis, supporting a hypothesis that the conditions are interconnected. Given the susceptibility of the gut microbiome to alteration, a number of therapeutic interventions have been investigated for the treatment of inflammatory conditions, though not within the context of cancer survivorship in children and adolescents. Here, we evaluate the potential for these interventions, which include probiotic supplementation, prebiotics/fiber-rich diet, exercise, and fecal microbiota transplantation for prevention and treatment of cancer treatment-related microbial dysbiosis in survivors. We also make recommendations to improve adherence and encourage long-term lifestyle changes for maintenance of healthy gut microbiome in CCS as a potential strategy to mitigate treatment-related late effects.

Keywords

childhood cancer, survivors, inflammaging, microbiome, dysbiosis, inflammation

Abbreviations

BBCT, bacteria-based cancer therapy; BMI, body mass index; CCS, childhood cancer survivors; CDI, *Clostridium difficile* infection; CRP, C-reactive protein; FMT, fecal microbiota transplantation; FOS, fructo-oligosaccharides; GOS, galactooligosaccharides; HDL, high-density lipoprotein; LPS, lipopolysaccharides; RCT, randomized controlled trials; SCFAs, short-chain fatty acids; T1DM, type 1 diabetes mellitus; T2DM, type 2 diabetes mellitus; WMT, washed microbiota transplantation.

Received: August 3, 2022; Revised: October 10, 2022; Accepted: December 19, 2022.

Background

In recent years, research has illuminated the critical role of the host microbiome in a number of essential physiological operations, including but not limited to digestion, immune regulation, and central nervous system function.¹ Microbiota composition can be perturbed by various external factors. In cancer patients, changes in the gut and oral microbiota may result from exposure to chemotherapeutic agents, radiotherapy, and antibiotics.^{2–8} Not unexpectedly, microbiota profile disparities between children and adolescents with cancer compared to healthy peers have been observed. Numerous studies reported reduced microbial diversity and treatment-induced alterations in the abundance of certain bacteria in the cancer group, particularly for the gut microbiome.^{9–12} For example, Fijlstra et al²

demonstrated in rat models that methotrexate, a cytotoxic agent associated with gut mucositis, depletes gut bacteria, reduces microbial diversity, and modifies bacterial composition. Additionally, patients with cancer are predisposed to episodes of neutropenic sepsis which are often treated with broad-spectrum antibiotics. The use of antibiotics has a long-term impact on gut microbiota, including increased prevalence

¹ University of Malaya, Kuala Lumpur, Malaysia

Corresponding Author:

Hany Ariffin, Department of Pediatrics, University of Malaya, 50603 Kuala Lumpur, Malaysia.
Email: hany@ummc.edu.my



Creative Commons Non Commercial CC BY-NC: This article is distributed under the terms of the Creative Commons Attribution-NonCommercial 4.0 License (<https://creativecommons.org/licenses/by-nc/4.0/>) which permits non-commercial use, reproduction and distribution of the work without further permission provided the original work is attributed as specified on the SAGE and Open Access page (<https://us.sagepub.com/en-us/nam/open-access-at-sage>).

of antibiotic-resistant microorganisms.^{5,13} Thus, the cumulative effect of cancer treatment is a significant dysbiosis in gut bacteria.

Early childhood constitutes a “critical period” during which the microbiome is particularly susceptible to alteration; gut flora then stabilizes over the ensuing decades.¹⁴ Children undergoing cancer treatment within this critical period may therefore develop microbial dysbiosis which remains long after completion of cancer therapy. This observation has been reported in children with acute myeloid leukemia, in whom *Bacteroides* and *Bifidobacterium* remained depleted 6 weeks after completing anti-leukemia treatment when compared to controls, despite apparent restoration of other genera.¹⁵ We conducted a longitudinal observational study in 7 children with acute lymphoblastic leukemia and demonstrated similar findings: gut microbiota profiles remained marginally different (both in beta diversity and abundance) 3 months after chemotherapy cessation.⁹ A follow-up study revealed that gut microbiome dysbiosis persisted into adulthood, suggesting a more chronic pathology.¹⁶ Thus, this review will focus on gut microbiome reconstitution as a potential therapeutic intervention to mitigate late effects in childhood cancer survivors (CCS).

Link Between Microbiome Dysbiosis and Chronic Inflammation

Gut microbiota play a key role in immune activation through a variety of mechanisms, many of which are metabolic. The interaction between nutrition, gut flora, and regulation of inflammatory pathways is mediated by short-chain fatty acids (SCFAs), which are produced via the metabolism of complex carbohydrates by gut bacteria. Certain SCFAs have known anti-inflammatory effects. For example, the SCFA butyrate is a metabolite that increases lipolysis and promotes mitochondrial activity in adipocytes, decreasing risk of obesity and ultimately blocking the generation of pro-inflammatory cytokines.^{17–20} Butyrate and several other SCFAs also play a crucial role in the maintenance of the gut mucosal barrier, which prevents the translocation of inflammation-inducing microbes/microbial components into the host bloodstream.²¹

Given their crucial role in maintaining immune balance, changes in bacterial metabolites and microbial translocation actively contribute to systemic immune activation and inflammation. Both these latter conditions are increasingly recognized in CCS.^{22–24} Gut microbiota dysbiosis and subsequent translocation have also been linked to elevated levels of systemic inflammatory markers (serum interleukin 6 [IL-6] and C-reactive protein [CRP]) in rats, thought to be caused by the presence of pro-inflammatory pathogen-associated molecular patterns.^{25–27} Finally, unmethylated CpG motifs, which are abundant in bacterial DNA, as well as lipopolysaccharides (LPS) in the bacterial cell wall have been shown to promote host immune activation.^{28–31} In short, these studies support the current understanding that microbial dysbiosis, disruption of gut mucosa, and chronic inflammation are intimately related as components of a self-perpetuating loop.

Association Between Microbial Dysbiosis, Chronic Inflammation, and Late Effects in Childhood Cancer Survivors

Given the cumulative evidence of treatment-mediated microbial dysbiosis in patients with cancer, as well as the link between gut microbiome perturbations and chronic inflammation, it is unsurprising that a large proportion of CCS goes on to develop inflammation-driven diseases in adulthood. Indeed, CCS are highly susceptible to late effects, a term that refers to therapy-related complications or adverse outcomes that persist or arise years after completion of treatment for a pediatric malignancy.³² CCS have elevated risks of metabolic and cardiovascular disease, neurological complications, and endocrine dysfunction compared to their peers (Figure 1).^{33–37} A large cohort study in the United States which included over 10 000 adult survivors of childhood cancer found that approximately 60% of these survivors reported at least 1 chronic health condition within 30 years of diagnosis, of which 28% were classified as severe, life-threatening, or disabling.³⁸ Prevalent health conditions within the CCS cohort included obesity, diabetes mellitus, and various cardiovascular conditions, with a large increase in the risk of ischemic heart disease in particular.³⁸

While there have been large advances in our understanding of microbiome-mediated chronic inflammation and thus, late effects in CCS, there have not yet been compelling studies looking at interventions focused on gut flora restitution in this population. However, a myriad of studies has evaluated gut microbial interventions for cardiometabolic conditions such as obesity, diabetes mellitus, and ischemic heart disease, all of which constitute prevalent late effects in CCS. Trends of microbial dysbiosis in CCS and patients with cardiometabolic disease appear to overlap, including changes in the relative ratio of *Bacteroidetes*/*Firmicutes*, the 2 most prevalent phyla in the gut.³⁹ Our group reported a significant decrease of *Bacteroidetes* in CCS cohorts compared to controls.¹⁶ Similarly, studies have documented a decreased *Bacteroidetes*/*Firmicutes* ratio in obesity, coronary artery disease, and ischemic stroke.^{39–41} Several other microbes have additionally been implicated in childhood cancer survivorship and inflammation-related diseases. For example, *Faecalibacterium*, which has been called a “master regulator” of inflammatory factors IL-2, IL-6, and IL-8 in the gut, has been found to be depleted both during therapy and as a long-term effect of cancer survivorship.^{42–44} Reduced *Faecalibacterium* has also been documented in coronary artery disease and was found to be inversely correlated with levels of HbA1c in patients with type 1 diabetes mellitus (T1DM).⁴⁵ Notably, *Faecalibacterium* depletion has also been associated with ischemic stroke.⁴⁶ In comparison to healthy peers, our cohort of 73 young adults who survived childhood leukemia had depletion of *Faecalibacterium*, and this finding was significantly correlated with elevated inflammatory and immune activation markers, namely plasma IL-6, CRP, as well as HLA-DR⁺ CD4⁺ and HLA-DR⁺CD8⁺ T cells.¹⁶ On the other hand, both CCS cohorts and patients with type 2 diabetes mellitus (T2DM) have been shown to have an enrichment of *Actinobacteria*, although the direct consequences of this are less

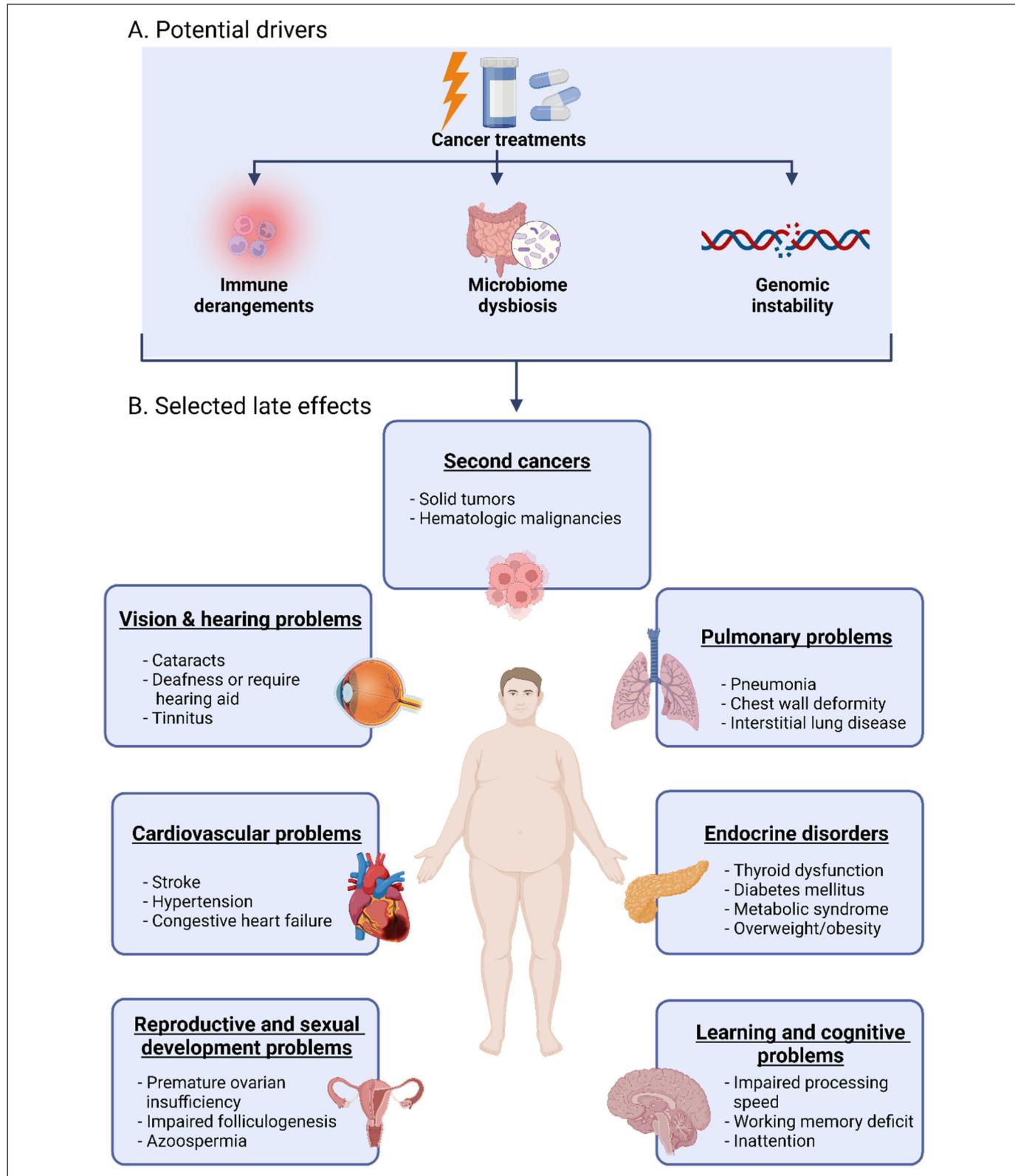


Figure 1. Potential drivers and late effects in childhood cancer survivors (Figure created using BioRender under Academic License).

well known.^{16,47} Given these shared associations, dysbiosis resulting from cancer treatment is a likely driver of inflammation and late effects in CCS.

Therapeutic Approaches to Modulate Microbial Dysbiosis

The studies reviewed above suggest a shared mechanism of gut microbial dysbiosis between CCS and patients with inflammation-related diseases. As previously discussed, many of these conditions are also highly prevalent as late effects in CCS cohorts. Therefore, we posit that microbiome-focused interventions which have been successful in adults with cardiometabolic diseases may also help ameliorate late effects in CCS (Figure 2).

Reviewing recent literature, we identified 4 primary approaches which have shown promising therapeutic applications to address CCS late effects, namely supplementation with oral probiotics, consumption of a ‘healthy’ diet rich in prebiotics, exercise, and fecal microbial transplantation.

Probiotic Supplementation

Probiotics are a promising intervention for the restitution of dysbiotic microbial communities. Probiotic strains confer health benefits through a number of mechanisms, including the restoration of host microbial communities, the competitive exclusion of pathogens, improvements to gut barrier integrity, and the generation of metabolites that can influence host immunity and metabolism.^{48,49} Typical probiotic supplements consist of either homogenous or heterogenous *Bifidobacterium*, *Lactobacillus*, and *Saccharomyces* spp. Several additional species—including *Faecalibacterium prausnitzii*, *Akkermansia muciniphila*, and *Clostridia* spp.—are currently under evaluation for therapeutic use.⁴⁸

Probiotics have demonstrated therapeutic potential for a number of inflammation-related metabolic conditions including obesity, insulin resistance, and T2DM.^{50–52} Numerous randomized controlled trials (RCT) investigating the effect of probiotic supplementation in patients with cardiometabolic risk factors have reported reduction in fasting glucose and HbA1c in diabetic subjects, as well as body fat reduction and improved lipid metabolism in obese/overweight subjects (Table 1 and Supplemental material, Appendix 1).^{52–58} For example, probiotic supplementation of *Lactobacillus acidophilus*, *Bifidobacterium lactis*, and *Lactobacillus casei* over the course of 8 weeks significantly reduced body fat and increased fat-burning adropin in 30 obese adults.⁵³ Similarly, a 6-month intake of *L. acidophilus*, *B. lactis*, *B. bifidum*, and *B. longum* in prediabetic adults ($n = 40$) increased *Bacteroidetes/Firmicutes* ratio, reduced blood glucose and blood pressure, and increased high-density lipoprotein (HDL).^{54,59} Many studies also reported a significant reduction in inflammatory markers in treatment cohorts, a finding which is particularly relevant for inflamming-prone CCS populations.⁵² Considering the increased prevalence of metabolic

disorders in CCS, these interventions are highly promising for the treatment of late effects. Recent clinical trials are summarized in Supplemental material, Appendix 1.

Diet and Prebiotics

Animal studies indicate that diet—particularly intake of fats and complex carbohydrates—may modulate gut microbiota composition and consequently, metabolic health.^{84,85} Human observational studies across globally distinct populations have demonstrated that diets high in animal protein and saturated fats correlate with reduced microbiota richness and diversity.⁸⁶ Alternatively, plant-based diets high in complex carbohydrates were associated with high microbial diversity and increased abundance of *Prevotella* and butyrate-producing bacteria (*Faecalibacterium*, *Roseburia* sp., *Eubacterium rectale*, *Ruminococcus bromii*).⁸⁷ Within the context of the gut microbiome, these complex carbohydrates are often referred to as prebiotics, or “a substrate that is selectively utilized by host microorganisms, conferring a health benefit.”⁸⁸ Prebiotics such as inulin, fructo-oligosaccharides (FOS), and galactooligosaccharides (GOS) work symbiotically to nurture the growth of beneficial bacteria/probiotics.⁸⁹

Sufficient prebiotic intake can improve microbial environment through promotion of *Bifidobacteria*, *Lactobacillus*, and butyrate-producing bacteria (Table 1).^{65–67,71,72,90,91} Such microbial modulation has been correlated with an increase in SCFAs and decrease in inflammatory response, as evident in overweight/obese children and adults with metabolic risk factors or T2DM (Table 1 and Supplemental material, Appendix 2).^{67,71,91–97} In a RCT of 22 overweight/obese children, 16-week FOS-enriched inulin supplementation reduced levels of IL-6 while ameliorating obesity-related microbial dysbiosis.⁶⁷ Likewise, an 8-week whole grain diet reduced serum CRP, IL-1B, and IL-6 in adults with metabolic risk factors ($n = 50$).⁷¹ Similar effects were also shown in individuals with T2DM.^{94–97}

With regards to the efficacy of prebiotics in directly improving clinical outcomes, studies have been generally promising, albeit varied. Prebiotics such as FOS and/or inulin were shown to reduce body fat and body mass index (BMI) while improving satiety in obese/overweight individuals.^{98,99} Moreover, prebiotic-mediated reduction in serum triglycerides, cholesterol, and low-density lipoprotein, as well as elevation of HDL has been reported in individuals with T2DM.^{94,100} Conversely, some studies noticed modulation in microflora compositions without changes in metabolic parameters.^{71,72,101,102} The diverse impacts of prebiotic supplementation could be explained by variations in basal metabolic alteration within patient cohorts, as well as type of prebiotic intervention, dosing, and duration.

Recently, there has been an increasing interest in whether synbiotics—a combination of probiotics and prebiotics—may serve as a potential therapeutic approach for inflammation-related and metabolic diseases. Results from animal studies and clinical trials demonstrated that synbiotics may improve obesity, diabetes, and cardiovascular conditions (Table 1 and Supplemental material, Appendix 3).^{76–83,103} Of note, the positive effect of synbiotic supplementation in improving glycemic control among individuals

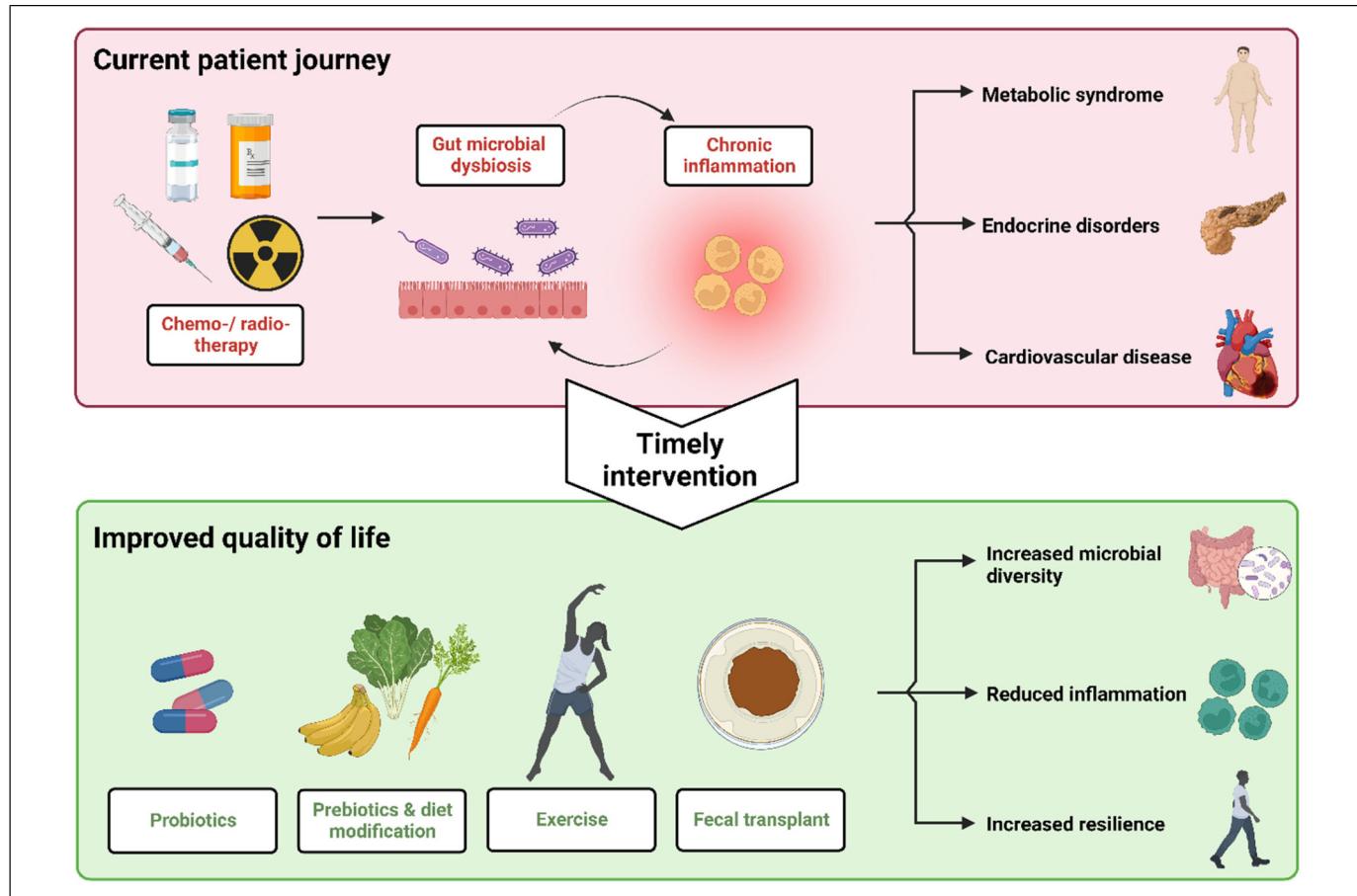


Figure 2. Potential health journey of childhood cancer survivors using interventions to address microbial dysbiosis and late effects (Figure created using BioRender under Academic License).

with diabetes appears to surpass the benefit achieved with probiotics or prebiotics alone.^{55,64,67,71,72,79,80,82,100,104}

Exercise

Exercise has been shown to alter both the gut microbiome and intestinal permeability, which in turn, control immune-inflammatory pathways and energy homeostasis.^{105–107} Sohail et al¹⁰⁸ reported that across several studies, exercise produced consistent global changes at a phylogenetic level (improvement in *Bacteroidetes/Firmicutes* ratio) with increased bacterial diversity and species richness. In a case-control experiment comparing sedentary to physically active rats, Queipo-Ortuno et al¹⁰⁹ reported increases in *Lactobacillus*, *Bifidobacterium*, and *Blautia coccoides-Eubacterium rectale* species with concomitant decrease in *Clostridium* and *Enterococcus* in the exercise group. Notably, both *Bifidobacterium* and *B. coccoides-E. rectale* have been associated with anti-inflammatory butyrogenic effects in the colon.^{109,110} Studies have also repeatedly demonstrated increased production of the beneficial SCFAs acetate, propionate, and butyrate by bacteria following exercise.^{111,112}

In humans, a few observational studies have looked at differences in bacterial composition, inflammatory, and metabolic

markers between athletes and controls, or among individuals with different exercise patterns. Clarke et al¹¹³ demonstrated increased gut microbial diversity in professional rugby players compared to controls, and a significantly greater abundance of *Akkermansia muciniphila* in athletes with low BMI. *Akkermansia* is a mucin-degrading bacteria whose abundance is inversely correlated to obesity and metabolic syndrome.¹¹⁴ In a large study involving 1493 participants from the American Gut Project, individuals who exercised regularly had significantly greater alpha diversity and increased prevalence of *Faecalibacterium*.¹¹⁵ Comparing exercise interventions in healthy elderly women, Morita et al¹¹⁶ found that aerobic exercise training through brisk walking (1 hour daily for 12 weeks) resulted in a significant increase in the relative abundance of intestinal *Bacteroides* compared to those who received trunk muscle training. In summary, the studies reviewed above support the use of exercise to improve gut microbial health and present a particularly attractive intervention for CCS.

Fecal Microbiota Transplantation

Gut microbiota may be directly altered through a therapeutic method known as fecal microbiota transplantation (FMT).

Table 1. Outcomes of probiotic, prebiotic and synbiotic oral supplementation in healthy adults and in patients with cardiometabolic conditions.

Intervention	Subjects	Outcomes				
		Microbial Health	Inflammatory Markers	Glycemic Control	Lipid Control	Body Mass Index/Obesity
Probiotic supplements	Healthy Obese	- No change ⁶¹	Improved ⁶⁰ -	Not applicable No change ⁶¹	Not applicable Improved ⁵³	Not applicable Improved ⁵³ or worsened ⁶¹
	Diabetic/prediabetic/insulin-resistant	Improved ^{56,59,62}	Improved ^{57,63} or no change ⁵⁶	Improved ⁵⁴⁻⁵⁷	Improved ^{54,55} or no change ^{56,57}	No change ⁵⁴⁻⁵⁷
	Obese and insulin resistant	-	-	No change ⁶⁴	Improved ⁶⁴	No change ⁶⁴
	Obese, insulin resistant with metabolic syndrome	No change ⁵⁸	Improved ⁵⁸	Improved ⁵⁸	Improved ⁵⁸	Improved ⁵⁸
Prebiotic supplements	Healthy	Improved ^{65,66}	-	Not applicable	Not applicable	Not applicable
	Obese (including cancer survivors)	Improved ⁶⁷	Improved ^{67,68}	No change ⁶⁷	Improved ⁶⁷	Improved ⁶⁷ or no change ⁶⁹
	Diabetic	-	-	Improved ⁷⁰	No change ⁷⁰	Improved ⁷⁰
Synbiotic supplements	Metabolic syndrome	Improved ^{71,72}	Improved ⁷¹ or no change ⁷²	No change ^{71,72}	No change ^{71,72}	Improved ⁷¹ or no change ⁷²
	Overweight/obese (including cancer survivors)	Improved ^{73,74} or no change ⁷⁵	Improved ^{73,76,77}	No change ^{78,79}	Improved ^{75,78}	Improved ^{73,78} or no change ^{74,79}
	Diabetes/prediabetes	No change ⁵⁹	Improved ⁸⁰	Improved ⁸⁰⁻⁸²	No change ⁸¹	Improved ⁸¹ or worsened ⁸⁰
	Obese and diabetic	No change ⁸³	-	No change ⁸³	-	Improved ⁸³
	Overweight, diabetic with coronary heart disease	-	-	Improved ⁸²	Improved ⁸²	No change ⁸²

The successful use of FMT was first reported in adult patients with *Clostridium difficile* infection (CDI) with restoration of healthy microbial flora in recipients and subsequent resolution of symptoms.^{117,118} Beyond CDI, FMT has also been implemented in patients with functional gut disorders, inflammatory bowel disease, and conditions associated with alteration in gut microbiota such as obesity, metabolic syndrome, and T2DM.¹¹⁹⁻¹²³ Studies on the use of FMT in patients with obesity and metabolic syndrome have shown a positive association between increased abundance of butyrate-producing bacteria and insulin sensitivity, although neither were sustained long term.^{119,120} Additionally, a recent study examined the use of washed microbiota transplantation (WMT)—a method involving microfiltration and repeated centrifugation and suspension. A significant short and medium-term improvement in lipid profile was reported in 90 human subjects with hyperlipidemia without additional dietary interventions.¹²⁴

A synergistic effect of FMT with diet alteration appears promising in sustaining engraftment of donor fecal microbiota. Mocanu et al¹²⁵ showed that low-fermentable fiber supplementation following a single dose of oral FMT in 17 individuals with severe obesity and metabolic syndrome significantly improved insulin sensitivity at 6 weeks, resulting in increased microbial diversity and composition. Interestingly, patients who received high-fermentable fiber supplementation alone had reduced TNF- α and IL-8, but these markers returned to baseline after fiber cessation and there was no association with glycaemic outcomes.¹²⁵ In a randomized study in which 16 patients with T2DM received a specially designed diet of prebiotics, probiotics, and whole grains, either with or without the addition of FMT for 90 days, Su et al¹²⁶

successfully demonstrated improvements in glycaemic control and blood pressure as well as early weight loss in the treatment cohort. The authors also noted a change in the dominant intestinal microbiota genera from *Bacteroides* to *Prevotella*, regardless of dominant bacterium genera in donors. The overall effect was an increase in the abundance of *Bifidobacterium* and a decrease in prevalence of *Bilophila* and *Desulfovibrio*, which have been positively correlated with hyperglycemia. In summary, FMT with the addition of a healthy diet appears to be promising in addressing gut microbial dysbiosis and its consequences, particularly for CCS populations experiencing metabolic late effects.

Conclusion

Late effects faced by CCS pose a major health challenge. In the United States, for example, approximately 1 in 640 young adults in the 20- to 39-year age group are survivors of childhood cancer.¹²⁷ Thus, management of chronic comorbidities arising from late effects presents an enormous burden to national healthcare expenditure as well as personal well-being.

Basic lifestyle advice, such as exercise and healthy diet, may seem redundant to patients who have completed their cancer journey. However, microbial reconstitution as a targeted therapeutic intervention requires a highly nuanced understanding of the mechanisms driving chronic health problems in CCS. Inflammaging may start well before the diagnosis of CCS-prevalent diseases: for example, in a CCS cohort with a median age of 25, we detected evidence of inflammaging and significant leukocyte telomere shortening in the majority of patients despite the absence of overt T2DM or hypertension.¹²⁸

These findings suggest that the optimal window of intervention may be significantly earlier than the current standard, which is instituting treatment upon clinical manifestation of late effects.

Since the timeline of microbiome-driven inflammaging in CCS remains largely unknown, the possibility of gut bacterial dysbiosis should be considered from the moment of diagnosis. Moreover, early intervention—even while undergoing treatment—could preempt late effects. Thus, we recommend implementing all feasible interventions across the timeline of cancer treatment and recovery: for example, judicious use of broad-spectrum antibiotics, as well as encouraging patient consumption of prebiotic-rich foods from the onset of cancer diagnosis. For safety reasons, low-risk interventions such as exercise and probiotic regimens may be better implemented after the completion of cancer therapy. Moreover, these practices have the potential to improve treatment-related dysbiosis in other parts of the alimentary canal: the oral cavity, for example.^{129–135}

There is the potential that treatment-induced microbial dysbiosis could eventually be avoided altogether. Many emerging cancer therapeutics promise targeted mechanisms and a reduction in adverse effects, which may eventually include perturbations to gut flora. Notably, bacteria-based cancer therapy (BBCT) has the potential to target tumor microenvironments and increase the precision of drug delivery.^{136,137} Genetic engineering of nonpathogenic bacteria in particular may lower host pathogenicity and increase antitumor efficacy without driving late effects in CCS.^{138,139} On the other hand, BBCT inherently activates host antitumor immune response, which may trigger inflammation and disturb gut microbiota.^{136,137} Furthermore, 2 relevant complications of BBCT—high levels of bacterial toxins and undesired colonization of implanted medical devices¹⁴⁰—may drive increased use of antibiotics which exacerbate microbial dysbiosis in cancer patients.¹⁴¹ Long-term clinical trials are needed to evaluate BBCT as well as other targeted cancer therapeutics to determine impact on late effects.

The main challenge of microbiome-focused interventions is that of sustainability, which was a common theme for all 4 approaches examined. Changes to the microbiome through probiotic and prebiotic supplementation, healthy diet, exercise, and FMT all require long-term commitment in order to achieve clinical benefits.^{59,65,68,75,115,119,120} Additionally, the success of each approach was found to be highly dependent on a number of host factors, including age, ethnicity, and health conditions, as well as methodological factors: for example, type and volume of exercise, or quantity/strain of probiotic.^{61,65,68,108,116,142} In many cases, these factors could explain the variability that we noted in study outcomes, particularly when it came to pre-, pro-, and symbiotic supplementation. Altogether, these findings suggest that restoration of the microbiome requires a treatment plan that is intentional, highly targeted, and with the long term in mind.

As such, measures would need to be taken to encourage strict treatment plan adherence by CCS. Exercise and a healthy diet, which promote gut health, are typically considered “lifestyle preferences,” as opposed to “therapy,” and thus may not compel adherence. Specificity in treatment plans—such as the optimal amount of time/type of exercise (eg, ≥30-minute

sessions, thrice weekly) and the number of servings of fruits and vegetables as well as components such as flavonoids may encourage the formation of long-term habits, particularly in adolescents or young adults.^{143,144} Monitoring/tracking apparatuses, such as electronic diet diaries and wearable tracking devices, should appeal to young adults and may prove more accurate than self-reporting.^{145,146} Overall, we believe that reviewing diet, exercise, and biomarkers of inflammaging such as plasma CRP could be the new paradigm for late effects’ surveillance in CCS attending long-term follow-up clinics.¹²⁸ Focused strategies to address microbial dysbiosis, most of which are easily implementable, should be incorporated into the holistic management of children who have been diagnosed with or survived malignancy.

Declaration of Conflicting Interests

The authors declared no potential conflicts of interest with respect to the research, authorship, and/or publication of this article.

Funding

The authors disclosed receipt of the following financial support for the research, authorship, and/or publication of this article: This work is supported by a grant from the Ministry of Higher Education, Malaysia (grant number FRGS/FP006-2021).

Supplemental Material

Supplemental material for this article is available online.

ORCID iD

Hany Ariffin  <https://orcid.org/0000-0002-4607-8837>

References

1. Valdes AM, Walter J, Segal E, Spector TD. Role of the gut microbiota in nutrition and health. *Br Med J*. 2018;361:k2179. doi:10.1136/bmj.k2179
2. Fijlstra M, Ferdous M, Koning AM, Rings EH, Harmsen HJ, Tissing WJ. Substantial decreases in the number and diversity of microbiota during chemotherapy-induced gastrointestinal mucositis in a rat model. *Support Care Cancer*. 2015;23(6):1513–1522. doi:10.1007/s00520-014-2487-6
3. Montassier E, Gastinne T, Vangay P, et al. Chemotherapy-driven dysbiosis in the intestinal microbiome. *Aliment Pharmacol Ther*. 2015;42(5):515–528. doi:10.1111/apt.13302
4. Hu YJ, Shao ZY, Wang Q, et al. Exploring the dynamic core microbiome of plaque microbiota during head-and-neck radiotherapy using pyrosequencing. *PLoS One*. 2013;8(2):e56343. doi:10.1371/journal.pone.0056343
5. Jakobsson HE, Jernberg C, Andersson AF, Sjolund-Karlsson M, Jansson JK, Engstrand L. Short-term antibiotic treatment has differing long-term impacts on the human throat and gut microbiome. *PLoS One*. 2010;5(3):e9836. doi:10.1371/journal.pone.0009836
6. Wipperman MF, Fitzgerald DW, Juste MAJ, et al. Antibiotic treatment for Tuberculosis induces a profound dysbiosis of the

- microbiome that persists long after therapy is completed. *Sci Rep.* 2017;7(1):10767. doi:10.1038/s41598-017-10346-6
7. Hong BY, Sobue T, Choquette L, et al. Chemotherapy-induced oral mucositis is associated with detrimental bacterial dysbiosis. *Microbiome.* 2019;7(1):66. doi:10.1186/s40168-019-0679-5
 8. Triarico S, Agresti P, Rinninella E, et al. Oral microbiota during childhood and its role in chemotherapy-induced oral mucositis in children with cancer. *Pathogens.* 2022;11(4): 448. doi:10.3390/pathogens11040448
 9. Chua LL, Rajasuriar R, Lim YAL, Woo YL, Loke P, Ariffin H. Temporal changes in gut microbiota profile in children with acute lymphoblastic leukemia prior to commencement-, during-, and post-cessation of chemotherapy. *BMC Cancer.* 2020;20(1):151. doi:10.1186/s12885-020-6654-5
 10. Bai L, Zhou P, Li D, Ju X. Changes in the gastrointestinal microbiota of children with acute lymphoblastic leukaemia and its association with antibiotics in the short term. *J Med Microbiol.* 2017;66(9):1297-1307. doi:10.1099/jmm.0.000568
 11. Rajagopala SV, Yooseph S, Harkins DM, et al. Gastrointestinal microbial populations can distinguish pediatric and adolescent acute lymphoblastic leukemia (ALL) at the time of disease diagnosis. *BMC Genomics.* 2016;17(1):635. doi:10.1186/s12864-016-2965-y
 12. Wang Y, Xue J, Zhou X, et al. Oral microbiota distinguishes acute lymphoblastic leukemia pediatric hosts from healthy populations. *PLoS One.* 2014;9(7):e102116. doi:10.1371/journal.pone.0102116
 13. Allos BM. *Campylobacter jejuni* infections: update on emerging issues and trends. *Clin Infect Dis.* 2001;32(8):1201-1206. doi:10.1086/319760
 14. Rodriguez JM, Murphy K, Stanton C, et al. The composition of the gut microbiota throughout life, with an emphasis on early life. *Microb Ecol Health Dis.* 2015;26:26050. doi:10.3402/mehd.v26.26050
 15. van Vliet MJ, Tissing WJ, Dun CA, et al. Chemotherapy treatment in pediatric patients with acute myeloid leukemia receiving antimicrobial prophylaxis leads to a relative increase of colonization with potentially pathogenic bacteria in the gut. *Clin Infect Dis.* 2009;49(2):262-270. doi:10.1086/599346
 16. Chua LL, Rajasuriar R, Azanan MS, et al. Reduced microbial diversity in adult survivors of childhood acute lymphoblastic leukemia and microbial associations with increased immune activation. *Microbiome.* 2017;5(1):35. doi:10.1186/s40168-017-0250-1
 17. Jia Y, Hong J, Li H, et al. Butyrate stimulates adipose lipolysis and mitochondrial oxidative phosphorylation through histone hyperacetylation-associated β_3 -adrenergic receptor activation in high-fat diet-induced obese mice. *Exp Physiol.* 2017;102(2):273-281. doi:10.1113/EP086114
 18. Turnbaugh PJ, Hamady M, Yatsunenko T, et al. A core gut microbiome in obese and lean twins. *Nature.* 2009;457(7228):480-484. doi:10.1038/nature07540
 19. Segain JP, Raingeard de la Bletiere D, Bourreille A, et al. Butyrate inhibits inflammatory responses through NF κ B inhibition: implications for Crohn's disease. *Gut.* 2000;47(3):397-403. doi:10.1136/gut.47.3.397
 20. Luhrs H, Gerke T, Schäuber J, et al. Cytokine-activated degradation of inhibitory κ B protein is inhibited by the short-chain fatty acid butyrate. *Int J Colorectal Dis.* 2001;16(4):195-201. doi:10.1007/s003840100295
 21. Dou X, Ma Z, Yan D, et al. Sodium butyrate alleviates intestinal injury and microbial flora disturbance induced by lipopolysaccharides in rats. *Food Funct.* 2022;13(3):1360-1369. doi:10.1039/d1fo03183j
 22. Andersen K, Kesper MS, Marschner JA, et al. Intestinal dysbiosis, barrier dysfunction, and bacterial translocation account for CKD-related systemic inflammation. *J Am Soc Nephrol.* 2017;28(1):76-83. doi:10.1681/ASN.2015111285
 23. Jin Y, Wu Y, Zeng Z, et al. Exposure to oral antibiotics induces gut microbiota dysbiosis associated with lipid metabolism dysfunction and low-grade inflammation in mice. *Toxicol Sci.* 2016;154(1):140-152. doi:10.1093/toxsci/kfw150
 24. Vujkovic-Cvijin I, Dunham RM, Iwai S, et al. Dysbiosis of the gut microbiota is associated with HIV disease progression and tryptophan catabolism. *Sci Transl Med.* 2013;5(193):193ra91. doi:10.1126/scitranslmed.3006438
 25. Wang F, Zhang P, Jiang H, Cheng S. Gut bacterial translocation contributes to microinflammation in experimental uremia. *Dig Dis Sci.* 2012;57(11):2856-2862. doi:10.1007/s10620-012-2242-0
 26. Lehto M, Groop PH. The gut-kidney axis: putative interconnections between gastrointestinal and renal disorders. *Front Endocrinol.* 2018;9:553. doi:10.3389/fendo.2018.00553
 27. Xi Y, Yan J, Li M, Ying S, Shi Z. Gut microbiota dysbiosis increases the risk of visceral gout in goslings through translocation of gut-derived lipopolysaccharide. *Poult Sci.* 2019;98(11): 5361-5373. doi:10.3382/ps/pez357
 28. Takesita F, Leifer CA, Gursel I, et al. Cutting edge: role of toll-like receptor 9 in CpG DNA-induced activation of human cells. *J Immunol.* 2001;167(7):3555-3558. doi:10.4049/jimmunol.167.7.3555
 29. Ramirez-Ortiz ZG, Specht CA, Wang JP, et al. Toll-like receptor 9-dependent immune activation by unmethylated CpG motifs in *Aspergillus fumigatus* DNA. *Infect Immun.* 2008;76(5):2123-2129. doi:10.1128/IAI.00047-08
 30. Tanaka M, Ishii K, Nakamura Y, et al. Toll-like receptor 9-dependent activation of bone marrow-derived dendritic cells by URA5 DNA from *Cryptococcus neoformans*. *Infect Immun.* 2012;80(2):778-786. doi:10.1128/IAI.05570-11
 31. Molteni M, Gemma S, Rossetti C. The role of toll-like receptor 4 in infectious and noninfectious inflammation. *Mediat Inflamm.* 2016;2016:6978936. doi:10.1155/2016/6978936
 32. Landier W, Bhatia S, Eshelman DA, et al. Development of risk-based guidelines for pediatric cancer survivors: the Children's Oncology Group long-term follow-up guidelines from the Children's Oncology Group late effects committee and nursing discipline. *J Clin Oncol.* 2004;22(24):4979-4990. doi:10.1200/JCO.2004.11.032
 33. Armenian SH, Robison LL. Childhood cancer survivorship: an update on evolving paradigms for understanding pathogenesis and screening for therapy-related late effects. *Curr Opin Pediatr.* 2013;25(1):16-22. doi:10.1097/MOP.0b013e32835b0b6a
 34. Ness KK, Armenian SH, Kadan-Lottick N, Gurney JG. Adverse effects of treatment in childhood acute lymphoblastic leukemia:

- general overview and implications for long-term cardiac health. *Expert Rev Hematol.* 2011;4(2):185-197. doi:10.1586/ehm.11.8
35. Oeffinger KC, Mertens AC, Sklar CA, et al. Chronic health conditions in adult survivors of childhood cancer. *New Engl J Med.* 2006;355(15):1572-1582. doi:10.1056/Nejmsa060185
36. Ou JY, Smits-Seemann RR, Kaul S, Fluchel MN, Sweeney C, Kirchhoff AC. Risk of hospitalization among survivors of childhood and adolescent acute lymphoblastic leukemia compared to siblings and a general population sample. *Cancer Epidemiol.* 2017;49:216-224. doi:10.1016/j.canep.2017.06.005
37. Zhang Y, Lorenzi MF, Goddard K, Spinelli JJ, Gotay C, McBride ML. Late morbidity leading to hospitalization among 5-year survivors of young adult cancer: a report of the childhood, adolescent and young adult cancer survivors research program. *Int J Cancer.* 2014;134(5):1174-1182. doi:10.1002/ijc.28453
38. Hudson MM, Ness KK, Gurney JG, et al. Clinical ascertainment of health outcomes among adults treated for childhood cancer. *JAMA.* 2013;309(22):2371-2381. doi:10.1001/jama.2013.6296
39. Cui L, Zhao T, Hu H, Zhang W, Hua X. Association study of gut Flora in coronary heart disease through high-throughput sequencing. *Biomed Res Int.* 2017;2017:3796359. doi:10.1155/2017/3796359
40. Zhu Q, Gao R, Zhang Y, et al. Dysbiosis signatures of gut microbiota in coronary artery disease. *Physiol Genomics.* 2018;50(10):893-903. doi:10.1152/physiolgenomics.00070.2018
41. Lee YT, Mohd Ismail NI, Wei LK. Microbiome and ischemic stroke: a systematic review. *PLoS One.* 2021;16(1):e0245038. doi:10.1371/journal.pone.0245038
42. Lenoir M, Martin R, Torres-Maravilla E, et al. Butyrate mediates anti-inflammatory effects of *Faecalibacterium prausnitzii* in intestinal epithelial cells through Dact3. *Gut Microbes.* 2020;12(1):1-16. doi:10.1080/19490976.2020.1826748
43. Thomas R, Wong WSW, Saadon R, et al. Gut microbial composition difference between pediatric ALL survivors and siblings. *Pediatr Hematol Oncol.* 2020;37(6):475-488. doi:10.1080/08880018.2020.1759740
44. Hakim H, Dallas R, Wolf J, et al. Gut microbiome composition predicts infection risk during chemotherapy in children with acute lymphoblastic leukemia. *Clin Infect Dis.* 2018;67(4):541-548. doi:10.1093/cid/ciy153
45. Huang Y, Li SC, Hu J, et al. Gut microbiota profiling in Han Chinese with type 1 diabetes. *Diabetes Res Clin Pract.* 2018;141:256-263. doi:10.1016/j.diabres.2018.04.032
46. Yin J, Liao SX, He Y, et al. Dysbiosis of gut microbiota with reduced trimethylamine-n-oxide level in patients with large-artery atherosclerotic stroke or transient ischemic attack. *J Am Heart Assoc.* 2015;4(11):e002699. doi:10.1161/JAHA.115.002699
47. Que Y, Cao M, He J, et al. Gut bacterial characteristics of patients with type 2 diabetes mellitus and the application potential. *Front Immunol.* 2021;12:722206. doi:10.3389/fimmu.2021.722206
48. Wieers G, Belkhir L, Enaud R, et al. How probiotics affect the microbiota. *Front Cell Infect Microbiol.* 2019;9:454. doi:10.3389/fcimb.2019.00454
49. Gomaa EZ. Human gut microbiota/microbiome in health and diseases: a review. *Antonie Van Leeuwenhoek.* 2020;113(12):2019-2040. doi:10.1007/s10482-020-01474-7
50. Green M, Arora K, Prakash S. Microbial medicine: prebiotic and probiotic functional foods to target obesity and metabolic syndrome. *Int J Mol Sci.* 2020;21(8):2890. doi:10.3390/ijms21082890
51. Azad MAK, Sarker M, Li T, Yin J. Probiotic species in the modulation of gut microbiota: an overview. *Biomed Res Int.* 2018;2018:9478630. doi:10.1155/2018/9478630
52. Companys J, Pla-Paga L, Calderon-Perez L, et al. Fermented dairy products, probiotic supplementation, and cardiometabolic diseases: a systematic review and meta-analysis. *Adv Nutr.* 2020;11(4):834-863. doi:10.1093/advances/nmaa030
53. Zarrati M, Raji Lahiji M, Salehi E, et al. Effects of probiotic yogurt on serum omentin-1, adropin, and nesfatin-1 concentrations in overweight and obese participants under low-calorie diet. *Probiotics Antimicrob Proteins.* 2019;11(4):1202-1209. doi:10.1007/s12602-018-9470-3
54. Kassaian N, Feizi A, Aminorroaya A, Amini M. Probiotic and synbiotic supplementation could improve metabolic syndrome in prediabetic adults: a randomized controlled trial. *Diabetes Metab Syndr.* 2019;13(5):2991-2996. doi:10.1016/j.dsx.2018.07.016
55. Razmpoosh E, Javadi A, Ejtahed HS, Mirmiran P, Javadi M, Yousefinejad A. The effect of probiotic supplementation on glycemic control and lipid profile in patients with type 2 diabetes: a randomized placebo controlled trial. *Diabetes Metab Syndr.* 2019;13(1):175-182. doi:10.1016/j.dsx.2018.08.008
56. Palacios T, Vitetta L, Coulson S, et al. Targeting the intestinal microbiota to prevent type 2 diabetes and enhance the effect of metformin on glycaemia: a randomised controlled pilot study. *Nutrients.* 2020;12(7): 2041. doi:10.3390/nu12072041
57. Soleimani A, Zarrati Mojarrad M, Bahmani F, et al. Probiotic supplementation in diabetic hemodialysis patients has beneficial metabolic effects. *Kidney Int.* 2017;91(2):435-442. doi:10.1016/j.kint.2016.09.040
58. Depommier C, Everard A, Druart C, et al. Supplementation with *Akkermansia muciniphila* in overweight and obese human volunteers: a proof-of-concept exploratory study. *Nat Med.* 2019;25(7):1096-1103. doi:10.1038/s41591-019-0495-2
59. Kassaian N, Feizi A, Rostami S, Aminorroaya A, Yaran M, Amini M. The effects of 6 mo of supplementation with probiotics and synbiotics on gut microbiota in the adults with prediabetes: a double blind randomized clinical trial. *Nutrition.* 2020;79-80: 110854. doi:10.1016/j.nut.2020.110854
60. Meng H, Ba Z, Lee Y, et al. Consumption of *Bifidobacterium animalis* subsp. *lactis* BB-12 in yogurt reduced expression of TLR-2 on peripheral blood-derived monocytes and pro-inflammatory cytokine secretion in young adults. *Eur J Nutr.* 2017;56(2):649-661. doi:10.1007/s00394-015-1109-5
61. Jones RB, Alderete TL, Martin AA, et al. Probiotic supplementation increases obesity with no detectable effects on liver fat or gut microbiota in obese Hispanic adolescents: a 16-week, randomized, placebo-controlled trial. *Pediatr Obes.* 2018;13(11):705-714. doi:10.1111/ijpo.12273
62. Sato J, Kanazawa A, Azuma K, et al. Probiotic reduces bacterial translocation in type 2 diabetes mellitus: a randomised controlled study. *Sci Rep.* 2017;7(1):12115. doi:10.1038/s41598-017-12535-9

63. Hajifaraji M, Jahanjou F, Abbasizadeh F, Aghamohammadzadeh N, Abbasi MM, Dolatkhah N. Effect of probiotic supplements in women with gestational diabetes mellitus on inflammation and oxidative stress biomarkers: a randomized clinical trial. *Asia Pac J Clin Nutr.* 2018;27(3):581-591. doi:10.6133/apjcn.082017.03
64. Naito E, Yoshida Y, Kunihiro S, et al. Effect of *Lactobacillus casei* strain Shirota-fermented milk on metabolic abnormalities in obese prediabetic Japanese men: a randomised, double-blind, placebo-controlled trial. *Biosci Microbiota Food Health.* 2018;37(1):9-18. doi:10.12938/bmfh.17-012
65. Tandon D, Haque MM, Gote M, et al. A prospective randomized, double-blind, placebo-controlled, dose-response relationship study to investigate efficacy of fructo-oligosaccharides (FOS) on human gut microflora. *Sci Rep.* 2019;9(1):5473. doi:10.1038/s41598-019-41837-3
66. Healey G, Murphy R, Butts C, Brough L, Whelan K, Coad J. Habitual dietary fibre intake influences gut microbiota response to an inulin-type fructan prebiotic: a randomised, double-blind, placebo-controlled, cross-over, human intervention study. *Br J Nutr.* 2018;119(2):176-189. doi:10.1017/S0007114517003440
67. Nicolucci AC, Hume MP, Martinez I, Mayengbam S, Walter J, Reimer RA. Prebiotics reduce body fat and alter intestinal microbiota in children who are overweight or with obesity. *Gastroenterology.* 2017;153(3):711-722. doi:10.1053/j.gastro.2017.05.055
68. Sheflin AM, Borresen EC, Kirkwood JS, et al. Dietary supplementation with rice bran or navy bean alters gut bacterial metabolism in colorectal cancer survivors. *Mol Nutr Food Res.* 2017;61(1): 1500905. doi:10.1002/mnfr.201500905
69. Pol K, de Graaf C, Meyer D, Mars M. The efficacy of daily snack replacement with oligofructose-enriched granola bars in overweight and obese adults: a 12-week randomised controlled trial. *Br J Nutr.* 2018;119(9):1076-1086. doi:10.1017/S0007114518000211
70. Roshanravan N, Mahdavi R, Alizadeh E, et al. Effect of butyrate and inulin supplementation on glycemic status, lipid profile and glucagon-like peptide 1 level in patients with type 2 diabetes: a randomized double-blind, placebo-controlled trial. *Horm Metab Res.* 2017;49(11):886-891. doi:10.1055/s-0043-119089
71. Roager HM, Vogt JK, Kristensen M, et al. Whole grain-rich diet reduces body weight and systemic low-grade inflammation without inducing major changes of the gut microbiome: a randomised cross-over trial. *Gut.* 2019;68(1):83-93. doi:10.1136/gutjnl-2017-314786
72. Kjolbaek L, Benitez-Paez A, Gomez Del Pulgar EM, et al. Arabinoxylan oligosaccharides and polyunsaturated fatty acid effects on gut microbiota and metabolic markers in overweight individuals with signs of metabolic syndrome: a randomized cross-over trial. *Clin Nutr.* 2020;39(1):67-79. doi:10.1016/j.clnu.2019.01.012
73. Hibberd AA, Yde CC, Ziegler ML, et al. Probiotic or symbiotic alters the gut microbiota and metabolism in a randomised controlled trial of weight management in overweight adults. *Benef Microbes.* 2019;10(2):121-135. doi:10.3920/BM2018.0028
74. Sergeev IN, Aljutaily T, Walton G, Huarte E. Effects of symbiotic supplement on human gut microbiota, body composition and weight loss in obesity. *Nutrients.* 2020;12(1): 222. doi:10.3390/nu12010222
75. Krumbeck JA, Rasmussen HE, Hutkins RW, et al. Probiotic *Bifidobacterium* strains and galactooligosaccharides improve intestinal barrier function in obese adults but show no synergism when used together as synbiotics. *Microbiome.* 2018;6(1):121. doi:10.1186/s40168-018-0494-4
76. Raji Lahiji M, Zarrati M, Najafi S, et al. Effects of symbiotic supplementation on serum adiponectin and inflammation status of overweight and obese breast cancer survivors: a randomized, triple-blind, placebo-controlled trial. *Support Care Cancer.* 2021;29(7):4147-4157. doi:10.1007/s00520-020-05926-8
77. Vafa S, Haghigat S, Janani L, et al. The effects of symbiotic supplementation on serum inflammatory markers and edema volume in breast cancer survivors with lymphedema. *EXCLI J.* 2020;19:1-15. doi:10.17179/excli2019-1876
78. Hadi A, Sepandi M, Marx W, Moradi S, Parastouei K. Clinical and psychological responses to symbiotic supplementation in obese or overweight adults: a randomized clinical trial. *Complement Ther Med.* 2019;47:102216. doi:10.1016/j.ctim.2019.102216
79. Raji Lahiji M, Najafi S, Janani L, Yazdani B, Razmipoosh E, Zarrati M. The effect of symbiotic on glycemic profile and sex hormones in overweight and obese breast cancer survivors following a weight-loss diet: a randomized, triple-blind, controlled trial. *Clin Nutr.* 2021;40(2):394-403. doi:10.1016/j.clnu.2020.05.043
80. Soleimani A, Motamedzadeh A, Zarrati Mojarrad M, et al. The effects of symbiotic supplementation on metabolic status in diabetic patients undergoing hemodialysis: a randomized, double-blinded, placebo-controlled trial. *Probiotics Antimicrob Proteins.* 2019; 11(4):1248-1256. doi:10.1007/s12602-018-9499-3
81. Ebrahimi ZS, Nasli-Esfahani E, Nadjarzade A, Mozaffari-Khosravi H. Effect of symbiotic supplementation on glycemic control, lipid profiles and microalbuminuria in patients with non-obese type 2 diabetes: a randomized, double-blind, clinical trial. *J Diabetes Metab Disord.* 2017;16:23. doi:10.1186/s40200-017-0304-8
82. Tajabadi-Ebrahimi M, Sharifi N, Farrokhan A, et al. A randomized controlled clinical trial investigating the effect of symbiotic administration on markers of insulin metabolism and lipid profiles in overweight type 2 diabetic patients with coronary heart disease. *Exp Clin Endocrinol Diabetes.* 2017;125(1):21-27. doi:10.1055/s-0042-105441
83. Horvath A, Leber B, Feldbacher N, et al. Effects of a multispecies symbiotic on glucose metabolism, lipid marker, gut microbiome composition, gut permeability, and quality of life in diabesity: a randomized, double-blind, placebo-controlled pilot study. *Eur J Nutr.* 2020;59(7):2969-2983. doi:10.1007/s00394-019-02135-w
84. Cani PD, Bibiloni R, Knauf C, et al. Changes in gut microbiota control metabolic endotoxemia-induced inflammation in high-fat diet-induced obesity and diabetes in mice. *Diabetes.* 2008;57(6):1470-1481. doi:10.2337/db07-1403
85. Cani PD, Amar J, Iglesias MA, et al. Metabolic endotoxemia initiates obesity and insulin resistance. *Diabetes.* 2007;56(7):1761-1772. doi:10.2337/db06-1491

86. Wolters M, Ahrens J, Romani-Perez M, et al. Dietary fat, the gut microbiota, and metabolic health—a systematic review conducted within the MyNewGut project. *Clin Nutr.* 2019;38(6):2504-2520. doi:10.1016/j.clnu.2018.12.024
87. Fan Y, Pedersen O. Gut microbiota in human metabolic health and disease. *Nat Rev Microbiol.* 2021;19(1):55-71. doi:10.1038/s41579-020-0433-9
88. Gibson GR, Hutkins R, Sanders ME, et al. Expert consensus document: the International Scientific Association for Probiotics and Prebiotics (ISAPP) consensus statement on the definition and scope of prebiotics. *Nat Rev Gastroenterol Hepatol.* 2017;14(8):491-502. doi:10.1038/nrgastro.2017.75
89. Delzenne NM, Neyrinck AM, Cani PD. Modulation of the gut microbiota by nutrients with prebiotic properties: consequences for host health in the context of obesity and metabolic syndrome. *Microb Cell Fact.* 2011;10(Suppl 1):S10. doi:10.1186/1475-2859-10-S1-S10
90. Wang S, Xiao Y, Tian F, et al. Rational use of prebiotics for gut microbiota alterations: specific bacterial phylotypes and related mechanisms. *J Funct Foods.* 2020;66:103838. <https://doi.org/10.1016/j.jff.2020.103838>
91. Watson H, Mitra S, Croden FC, et al. A randomised trial of the effect of omega-3 polyunsaturated fatty acid supplements on the human intestinal microbiota. *Gut.* 2018;67(11):1974-1983. doi:10.1136/gutjnl-2017-314968
92. Hartvigsen ML, Laerke HN, Overgaard A, Holst JJ, Bach Knudsen KE, Hermansen K. Postprandial effects of test meals including concentrated arabinoxylan and whole grain rye in subjects with the metabolic syndrome: a randomised study. *Eur J Clin Nutr.* 2014;68(5):567-574. doi:10.1038/ejcn.2014.25
93. Chen T, Long W, Zhang C, Liu S, Zhao L, Hamaker BR. Fiber-utilizing capacity varies in *Prevotella*- versus *Bacteroides*-dominated gut microbiota. *Sci Rep.* 2017;7(1):2594. doi:10.1038/s41598-017-02995-4
94. O'Connor S, Chouinard-Castonguay S, Gagnon C, Rudkowska I. Prebiotics in the management of components of the metabolic syndrome. *Maturitas.* 2017;104:11-18. doi:10.1016/j.maturitas.2017.07.005
95. Aliasgharzadeh A, Dehghan P, Gargari BP, Asghari-Jafarabadi M. Resistant dextrin, as a prebiotic, improves insulin resistance and inflammation in women with type 2 diabetes: a randomised controlled clinical trial. *Br J Nutr.* 2015;113(2):321-330. doi:10.1017/S0007114514003675
96. Dehghan P, Gargari BP, Jafar-Abadi MA, Aliasgharzadeh A. Inulin controls inflammation and metabolic endotoxemia in women with type 2 diabetes mellitus: a randomized-controlled clinical trial. *Int J Food Sci Nutr.* 2014;65(1):117-123. doi:10.3109/09637486.2013.836738
97. Dehghan P, Pourghassem Gargari B, Asghari Jafar-abadi M. Oligofructose-enriched inulin improves some inflammatory markers and metabolic endotoxemia in women with type 2 diabetes mellitus: a randomized controlled clinical trial. *Nutrition.* 2014;30(4):418-423. doi:10.1016/j.nut.2013.09.005
98. Genta S, Cabrera W, Habib N, et al. Yacon syrup: beneficial effects on obesity and insulin resistance in humans. *Clin Nutr.* 2009;28(2):182-187. doi:10.1016/j.clnu.2009.01.013
99. Parnell JA, Reimer RA. Weight loss during oligofructose supplementation is associated with decreased ghrelin and increased peptide YY in overweight and obese adults. *Am J Clin Nutr.* 2009;89(6):1751-1759. doi:10.3945/ajcn.2009.27465
100. Beserra BT, Fernandes R, do Rosario VA, Mocellin MC, Kuntz MG, Trindade EB. A systematic review and meta-analysis of the prebiotics and synbiotics effects on glycaemia, insulin concentrations and lipid parameters in adult patients with overweight or obesity. *Clin Nutr.* 2015;34(5):845-858. doi:10.1016/j.clnu.2014.10.004
101. Dewulf EM, Cani PD, Claus SP, et al. Insight into the prebiotic concept: lessons from an exploratory, double blind intervention study with inulin-type fructans in obese women. *Gut.* 2013;62(8):1112-1121. doi:10.1136/gutjnl-2012-303304
102. Vulevic J, Juric A, Tzortzis G, Gibson GR. A mixture of trans-galactooligosaccharides reduces markers of metabolic syndrome and modulates the fecal microbiota and immune function of overweight adults. *J Nutr.* 2013;143(3):324-331. doi:10.3945/jn.112.166132
103. Li HY, Zhou DD, Gan RY, et al. Effects and mechanisms of probiotics, prebiotics, synbiotics, and postbiotics on metabolic diseases targeting gut microbiota: a narrative review. *Nutrients.* 2021;13(9): 3211. doi:10.3390/nu13093211
104. Barengolts E, Smith ED, Reutrakul S, Tonucci L, Anothaisintawee T. The effect of probiotic yogurt on glycemic control in type 2 diabetes or obesity: a meta-analysis of nine randomized controlled trials. *Nutrients.* 2019;11(3): 671. doi:10.3390/nu11030671
105. Clarke G, Stilling RM, Kennedy PJ, Stanton C, Cryan JF, Dinan TG. Minireview: gut microbiota: the neglected endocrine organ. *Mol Endocrinol.* 2014;28(8):1221-1238. doi:10.1210/me.2014-1108
106. Marchesi JR, Adams DH, Fava F, et al. The gut microbiota and host health: a new clinical frontier. *Gut.* 2016;65(2):330-339. doi:10.1136/gutjnl-2015-309990
107. Walsh NP, Gleeson M, Shephard RJ, et al. Position statement. Part one: immune function and exercise. *Exerc Immunol Rev.* 2011;17:6-63.
108. Sohail MU, Yassine HM, Sohail A, Thani AAA. Impact of physical exercise on gut microbiome, inflammation, and the pathobiology of metabolic disorders. *Rev Diabet Stud.* 2019;15:35-48. doi:10.1900/RDS.2019.15.35
109. Queipo-Ortuno MI, Seoane LM, Murri M, et al. Gut microbiota composition in male rat models under different nutritional status and physical activity and its association with serum leptin and ghrelin levels. *PLoS One.* 2013;8(5):e65465. doi:10.1371/journal.pone.0065465
110. De Vuyst L, Leroy F. Cross-feeding between *Bifidobacteria* and butyrate-producing colon bacteria explains bifidobacterial competitiveness, butyrate production, and gas production. *Int J Food Microbiol.* 2011;149(1):73-80. doi:10.1016/j.ijfoodmicro.2011.03.003
111. Matsumoto M, Inoue R, Tsukahara T, et al. Voluntary running exercise alters microbiota composition and increases n-butyrate concentration in the rat cecum. *Biosci Biotechnol Biochem.* 2008;72(2):572-576. doi:10.1271/bbb.70474

112. Kang SS, Jeraldo PR, Kurti A, et al. Diet and exercise orthogonally alter the gut microbiome and reveal independent associations with anxiety and cognition. *Mol Neurodegener.* 2014;9:36. doi:10.1186/1750-1326-9-36
113. Clarke SF, Murphy EF, O'Sullivan O, et al. Exercise and associated dietary extremes impact on gut microbial diversity. *Gut.* 2014;63(12):1913-1920. doi:10.1136/gutjnl-2013-306541
114. Everard A, Belzer C, Geurts L, et al. Cross-talk between *Akkermansia muciniphila* and intestinal epithelium controls diet-induced obesity. *Proc Natl Acad Sci U S A.* 2013;110(22):9066-9071. doi:10.1073/pnas.1219451110
115. McFadzean R. *Exercise can help modulate human gut microbiota.* Undergraduate Honors Theses. Paper 155. University of Colorado; 2014.
116. Morita E, Yokoyama H, Imai D, et al. Aerobic exercise training with brisk walking increases intestinal *Bacteroides* in healthy elderly women. *Nutrients.* 2019;11(4): 868. doi:10.3390/nu11040868
117. van Nood E, Vrieze A, Nieuwdorp M, et al. Duodenal infusion of donor feces for recurrent *Clostridium difficile*. *N Engl J Med.* 2013;368(5):407-415. doi:10.1056/NEJMoa1205037
118. Cammarota G, Masucci L, Ianiro G, et al. Randomised clinical trial: faecal microbiota transplantation by colonoscopy vs. vancomycin for the treatment of recurrent *Clostridium difficile* infection. *Aliment Pharmacol Ther.* 2015;41(9):835-843. doi:10.1111/apt.13144
119. Vrieze A, Van Nood E, Holleman F, et al. Transfer of intestinal microbiota from lean donors increases insulin sensitivity in individuals with metabolic syndrome. *Gastroenterology.* 2012; 143(4):913-916.e7. doi:10.1053/j.gastro.2012.06.031
120. Kootte RS, Levin E, Salojarvi J, et al. Improvement of insulin sensitivity after lean donor feces in metabolic syndrome is driven by baseline intestinal microbiota composition. *Cell Metab.* 2017;26(4):611-619 e6. doi:10.1016/j.cmet.2017.09.008
121. Smits LP, Kootte RS, Levin E, et al. Effect of vegan fecal microbiota transplantation on carnitine- and choline-derived trimethylamine-n-oxide production and vascular inflammation in patients with metabolic syndrome. *J Am Heart Assoc.* 2018;7(7):e008342. doi:10.1161/JAHA.117.008342
122. Kang DW, Adams JB, Gregory AC, et al. Microbiota transfer therapy alters gut ecosystem and improves gastrointestinal and autism symptoms: an open-label study. *Microbiome.* 2017;5(1): 10. doi:10.1186/s40168-016-0225-7
123. Bajaj JS, Kassam Z, Fagan A, et al. Fecal microbiota transplant from a rational stool donor improves hepatic encephalopathy: a randomized clinical trial. *Hepatology.* 2017;66(6):1727-1738. doi:10.1002/hep.29306
124. Liang F, Lu X, Deng Z, et al. Effect of washed microbiota transplantation on patients with dyslipidemia in South China. *Front Endocrinol.* 2022;13:827107. doi:10.3389/fendo.2022.827107
125. Mocanu V, Zhang Z, Deehan EC, et al. Fecal microbial transplantation and fiber supplementation in patients with severe obesity and metabolic syndrome: a randomized double-blind, placebo-controlled phase 2 trial. *Nat Med.* 2021;27(7):1272-1279. doi:10.1038/s41591-021-01399-2
126. Su L, Hong Z, Zhou T, et al. Health improvements of type 2 diabetic patients through diet and diet plus fecal microbiota transplantation. *Sci Rep.* 2022;12(1):1152. doi:10.1038/s41598-022-05127-9
127. Mariotto AB, Rowland JH, Yabroff KR, et al. Long-term survivors of childhood cancers in the United States. *Cancer Epidemiol Biomarkers Prev.* 2009;18(4):1033-1040. doi:10.1158/1055-9965.EPI-08-0988
128. Ariffin H, Azanan MS, Abd Ghafar SS, et al. Young adult survivors of childhood acute lymphoblastic leukemia show evidence of chronic inflammation and cellular aging. *Cancer.* 2017; 123(21):4207-4214. doi:10.1002/cncr.30857
129. Bescos R, Brookes ZLS, Belfield LA, Fernandez-Sanjurjo M, Casas-Agustench P. Modulation of oral microbiota: a new frontier in exercise supplementation. *PharmaNutrition.* 2020;14: 100230. <https://doi.org/10.1016/j.phanu.2020.100230>
130. Al-Zahrani MS, Borawski EA, Bissada NF. Periodontitis and three health-enhancing behaviors: maintaining normal weight, engaging in recommended level of exercise, and consuming a high-quality diet. *J Periodontol.* 2005;76(8):1362-1366. doi:10.1902/jop.2005.76.8.1362
131. Omori S, Uchida F, Oh S, et al. Exercise habituation is effective for improvement of periodontal disease status: a prospective intervention study. *Ther Clin Risk Manag.* 2018;14:565-574. doi:10.2147/TCRM.S153397
132. Uchida F, Oh S, Shida T, et al. Effects of exercise on the oral microbiota and saliva of patients with non-alcoholic fatty liver disease. *Int J Environ Res Public Health.* 2021;18(7): 3470. doi:10.3390/ijerph18073470
133. Mohd Fuad AS, Amran NA, Nasruddin NS, Burhanudin NA, Dashper S, Arzmi MH. The mechanisms of probiotics, prebiotics, synbiotics, and postbiotics in oral cancer management. *Probiotics Antimicrob Proteins.* 2022. doi:10.1007/s12602-022-09985-7
134. Baker JL, Edlund A. Exploiting the oral microbiome to prevent tooth decay: has evolution already provided the best tools? *Front Microbiol.* 2018;9:3323. doi:10.3389/fmicb.2018.03323
135. Chugh P, Dutt R, Sharma A, Bhagat N, Dhar MS. A critical appraisal of the effects of probiotics on oral health. *J Funct Foods.* 2020;70:103985. <https://doi.org/10.1016/j.jff.2020.103985>
136. Duong MT, Qin Y, You SH, Min JJ. Bacteria-cancer interactions: bacteria-based cancer therapy. *Exp Mol Med.* 2019; 51(12):1-15. doi:10.1038/s12276-019-0297-0
137. Toso JF, Gill VJ, Hwu P, et al. Phase I study of the intravenous administration of attenuated *Salmonella typhimurium* to patients with metastatic melanoma. *J Clin Oncol.* 2002;20(1):142-152. doi:10.1200/JCO.2002.20.1.142
138. Sasaki T, Fujimori M, Hamaji Y, et al. Genetically engineered *Bifidobacterium longum* for tumor-targeting enzyme-prodrug therapy of autochthonous mammary tumors in rats. *Cancer Sci.* 2006;97(7):649-657. doi:10.1111/j.1349-7006.2006.00221.x
139. Yazawa K, Fujimori M, Amano J, Kano Y, Taniguchi S. *Bifidobacterium longum* as a delivery system for cancer gene therapy: selective localization and growth in hypoxic tumors. *Cancer Gene Ther.* 2000;7(2):269-274. doi:10.1038/sj.cgt.7700122

140. Gupta KH, Nowicki C, Giurini EF, Marzo AL, Zloza A. Bacterial-based cancer therapy (BBCT): recent advances, current challenges, and future prospects for cancer immunotherapy. *Vaccines*. 2021;9(12): 1497. doi:10.3390/vaccines9121497
141. Kramer MG, Masner M, Ferreira FA, Hoffman RM. Bacterial therapy of cancer: promises, limitations, and insights for future directions. *Front Microbiol*. 2018;9:16. doi:10.3389/fmicb.2018.00016
142. Million M, Angelakis E, Paul M, Armougom F, Leibovici L, Raoult D. Comparative meta-analysis of the effect of *Lactobacillus* species on weight gain in humans and animals. *Microb Pathog*. 2012;53(2): 100-108. doi:10.1016/j.micpath.2012.05.007
143. Allen JM, Mailing LJ, Niemiro GM, et al. Exercise alters gut microbiota composition and function in lean and obese humans. *Med Sci Sports Exerc*. 2018;50(4):747-757. doi:10.1249/MSS.0000000000001495
144. Luo J, Lin X, Bordiga M, Brennan C, Xu B. Manipulating effects of fruits and vegetables on gut microbiota—a critical review. *Int J Food Sci Technol*. 2021;56(5):2055-2067. <https://doi.org/10.1111/ijfs.14927>
145. Kerner C, Goodyear VA. The motivational impact of wearable healthy lifestyle technologies: a self-determination perspective on FitBits with adolescents. *Am J Health Educ*. 2017;48(5): 287-297. doi:10.1080/19325037.2017.1343161
146. Ringeval M, Wagner G, Denford J, Pare G, Kitsiou S. Fitbit-based interventions for healthy lifestyle outcomes: systematic review and meta-analysis. *J Med Internet Res*. 2020;22(10): e23954. doi:10.2196/23954