

SCIENTIFIC REPORTS



OPEN

Brain multiplexes reveal morphological connectional biomarkers fingerprinting late brain dementia states

Ines Mahjoub^{1,2}, Mohamed Ali Mahjoub², Islem Rekik¹ ¹ & Alzheimer's Disease Neuroimaging Initiative*

Accurate diagnosis of mild cognitive impairment (MCI) before conversion to Alzheimer's disease (AD) is invaluable for patient treatment. Many works showed that MCI and AD affect functional and structural connections between brain regions as well as the shape of cortical regions. However, 'shape connections' between brain regions are rarely investigated -e.g., how morphological attributes such as cortical thickness and sulcal depth of a specific brain region change in relation to morphological attributes in other regions. To fill this gap, we unprecedentedly design morphological brain multiplexes for late MCI/AD classification. Specifically, we use structural T1-w MRI to define morphological brain networks, each quantifying similarity in morphology between different cortical regions for a specific cortical attribute. Then, we define a brain multiplex where each intra-layer represents the morphological connectivity network of a specific cortical attribute, and each inter-layer encodes the similarity between two consecutive intra-layers. A significant performance gain is achieved when using the multiplex architecture in comparison to other conventional network analysis architectures. We also leverage this architecture to discover morphological connectional biomarkers fingerprinting the difference between late MCI and AD stages, which included the right entorhinal cortex and right caudal middle frontal gyrus.

Alzheimer Disease (AD) is one of the most devastating neurodegenerative diseases, affecting memory as well as cognitive functions of the human brain. With the absence of immediate treatment for patients diagnosed with AD, an accurate diagnosis of AD in an earlier stage propels early clinical interventions that could help slow down irreversible cognitive decline. Specifically, an intermediate stage exists between AD and normal control (NC) which is Mild Cognitive Impairment (MCI), where unlike AD, the memory deficits in MCI patients may remain relatively stable for years. The MCI stage is regarded as very critical as patients can still benefit from adequate clinical interventions before conversion to AD.

Considering the increasing number of brain imaging datasets on dementia and particularly AD, several methods based on neuroimaging processing and machine-learning have been developed in the purpose of early detection of AD conversion at the MCI stage. Hence, detecting brain biomarkers in the stage of MCI may allow the individualization of effective treatment to demented patients. Remarkably, the majority of these methods have extensively relied on resting state functional and diffusion magnetic resonance imaging (MRI)¹⁻⁸. Some works proposed brain network analysis methods using noninvasive diffusion MRI for AD diagnosis, where structural connectivities were measured using the degree of white matter connectivity between the associated pairs of ROIs^{4,8}. On the other hand, several studies used functional brain networks which mostly focused on characterizing the pairwise correlation (e.g., Pearson Correlation) between ROIs. Recently, more advanced studies proposed novel functional connectivity (FC) representations to model brain networks at different connectional levels. For example, Yu *et al.* proposed a novel method to construct brain FC by taking advantage of both Pearson Correlation and sparse learning¹. While some works only used high-order FC³ considering the relationships

¹BASIRA lab, CVIP group, School of Science and Engineering, Computing, University of Dundee, Dundee, UK. ²LATIS lab, ENISo – National Engineering School of Sousse, Sousse, Tunisia. *A comprehensive list of consortium members appears at the end of the paper. Correspondence and requests for materials should be addressed to I.R. (email: irekik@dundee.ac.uk)

between pairs of ROIs, more recent studies integrated both low-order and high-order FC networks along with interactions between the two levels⁷. However, analysis of functional networks is typically limited by the choice of a single or multiple thresholds for examining network topology, which may discard many important and discriminative brain connectivities. Moreover, while functional MRI can produce spurious and noisy connectomes, diffusion MRI can produce biased and largely variable structural connectomes depending on the employed fiber tractography method⁹. Besides, both structural and functional modalities are rarely acquired in a conventional clinical routine. Additionally, distinguishing between late MCI (LMCI) patients, who might be on the verge to convert to AD, and AD patients is a much more challenging classification task than that of AD vs. NC or early MCI (EMCI) vs. AD. Due to the very subtle brain changes between LMCI and AD brain changes, LMCI/AD classification task remains a hard problem to solve, that has been hardly addressed in the AD literature^{10,11}.

On the other hand, many other studies have demonstrated the importance of considering cortical measures derived from the multi-folded surface of the cerebral cortex for AD diagnosis, such as the cortical thickness^{12–15}. Specifically, cortical thickness is considered as a biomarker of AD progression, which provides insight into normal brain development and neurodegenerative disorders since it is correlated with changes in cognitive performance^{15–18}. For instance, Frisoni *et al.* showed reduction in cortical thickness in AD subjects compared with control subjects¹⁵. Thus, many voxel-based methods^{16,17} or region-based methods¹⁸ heavily relied on morphological features, including volumetric cortical thickness measurements from MRI, for AD diagnosis. However, all these methods were based on volumetric cortical thickness analysis, while there is evidence that AD alters not only volume-based cortical measures, but also the shape of cortical regions -e.g., cortical thinning at different levels¹⁹. For this reason, other studies explored cortical thickness using surface-based methods involving spectral shape description²⁰, or combining shape-derived features with voxel features^{21,22}. However, these approaches considered the morphological features at only the vertex-level. To the best of our knowledge, none of existing network-based analysis methods for disentangling late AD states investigated the *morphological* connectivities between ROIs using structural T1-w MRI -i.e., modeling how the morphology of different brain regions may be affected in relation to one another. Moreover, since AD may affect the complex relationships between a set of attributes in different cortical regions, one cannot rely on a single cortical attribute to examine how the brain is progressively altered by different stages of AD. A more comprehensive approach would consider multiple cortical attributes (e.g., sulcal depth, cortical thickness), each of these representing a single view of cortex morphology to quantify brain morphology. In this paper, we propose the first multi-view morphological brain connectivity using four different cortical attributes: cortical thickness network, sulcal depth network, average curvature network, and principal curvature network. Then, based on this multi-view connective representation of brain morphology, we further propose novel network architecture that would allow us to investigate the complex relationship between these views for identifying late MCI morphological connective biomarkers distinguishing between LMCI and AD.

Typically, the majority of network-based methods developed for MCI/AD classification diagnosis overlooked the high-order relationship between different brain connective layers. A few recent network-analysis works proposed for classification tasks between different AD stages (e.g., early MCI, late MCI), used one-layer network representation²³, a multi-layer (i.e. set of concatenated networks) network²⁴ or high-order networks^{10,11}. Specifically, the recently proposed high-order functional connectivity networks for MCI/AD diagnosis²³ integrated new high-level features that encode how different brain region pairs, instead of two brain regions, functionally interact with each other. Nevertheless, it will be possible to further consider other new connections through exploring how different network pairs interact with each another (and not only brain region pairs). This nicely led us to the concept of a *multiplex network*, which was historically coined to indicate the presence of more than one relationship between the same actors of a social network²⁵. Some previous methods^{26–30} have explored multiplexes to study brain networks (e.g., structural, functional). These multiplex networks (or multiplexes) allow multiple types of relationships to be represented in modelling brain connectivities, thus capturing higher levels of complexity between brain regions. However, all the mentioned studies investigated multiplex as a multi-layer network without exploring similarity networks that encode the relationship between consecutive brain connectivity layers. For instance, Battiston *et al.* used multiplexes as a two-layer network (functional and anatomical) to extract brain subgraphs while overlooking the inter-layer that perform high-order connectivities²⁷. Moreover, these approaches either relied on fMRI, combine fMRI with structural MRI, or used different modalities such as MRI with PET²⁶; but none explored morphological brain network each based on a specific attribute of the cortical surface, with the notable exception of recent works^{31,32} targeting early dementia and autism spectrum disorder diagnosis.

To address this limitation, we further propose a morphological brain multiplex interleaving a set of two different layers: an intra-layer which represents the morphological connectivity network of a specific cortical attribute, and an inter-layer (or a similarity layer) which computes the Pearson Correlation between two consecutive intra-layers. The proposed architecture leverages both morphological networks and the correlational relationship between each two consecutive layers. However, different similarity networks can be extracted by varying the order of layers. Hence, we define an ensemble of morphological brain multiplexes, each capturing complex network-to-network relationships for predefined set of cortical attributes by reordering at each time different intra-layers, with the exception of the first intra-layer, to capture new similarity networks. We aim by this architecture to discover morphological connective biomarkers distinguishing between AD and LMCI patients, which can be clinically useful for early detection of AD conversion at MCI stage.

Results

Data processing and parameters. In our study, we used 77 subjects (41 AD and 36 LMCI) from ADNI GO public dataset, each with structural T1-w MR image³³. Data used in the preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). The ADNI

	AD	LMCI
M	23	20
F	18	16
Total	41	36
Mean age	75.27	72.54
Std age	8.72	6.10

Table 1. Data distribution. M: male. F: female. Total: total number of subjects in each group. Std: standard deviation.

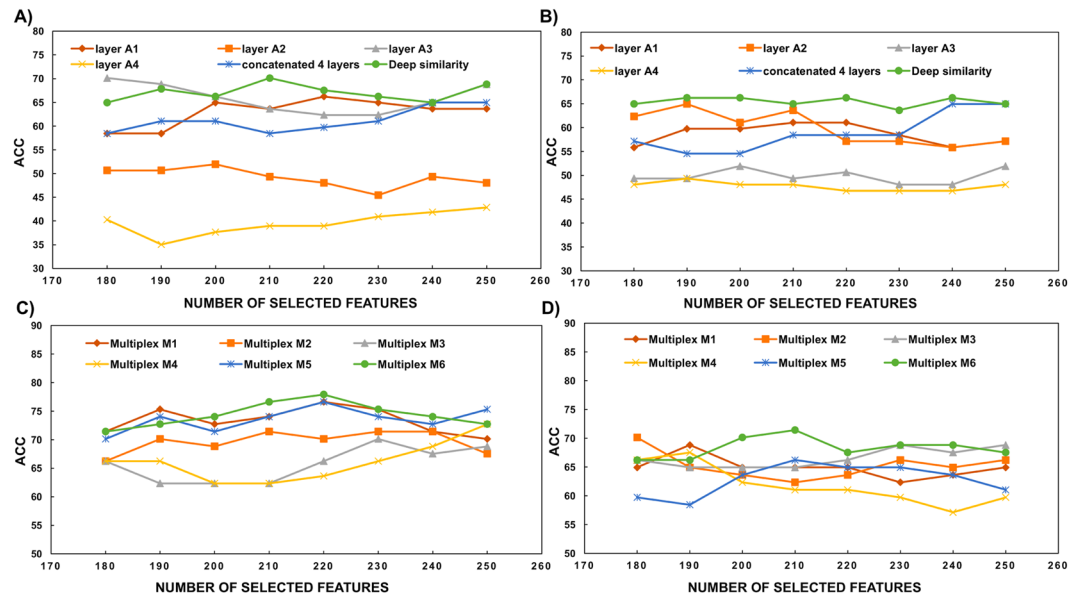


Figure 1. Influence of the selected features number on the accuracy of both baseline and proposed methods. (A) and (B) plot the accuracy curves against the number of selected features for deep similarity network compared with baseline methods for RH and LH, respectively. (C) and (D) plot the accuracy curves against the number of selected features for the 6 proposed multiplexes of right and left hemispheres respectively.

was launched in 2003 as a public-private partnership, led by Principal Investigator Michael W. Weiner, MD. The primary goal of ADNI has been to test whether serial magnetic resonance imaging (MRI), positron emission tomography (PET), other biological markers, and clinical and neuropsychological assessment can be combined to measure the progression of mild cognitive impairment (MCI) and early Alzheimer's disease (AD). We used FreeSurfer processing pipeline³⁴ to reconstruct both right (RH) and left (LH) cortical hemispheres for each subject from T1-w MRI³⁵. Then we parcellated each cortical hemisphere into 35 cortical regions using Desikan-Killiany cortical atlas³⁵. Using FreeSurfer pipeline, each vertex on the cortical surface was assigned four cortical attributes: maximum principal curvature, cortical thickness, sulcal depth, and average curvature.

For the deep similarity network architecture, we used two levels ($l=0, l=1$). We defined $K=6$ multiplexes using 4 cortical attributes, where multiplex \mathcal{M}_1 includes morphological networks generated using different cortical attributes $\{N_1, N_2, N_3, N_4\}$, \mathcal{M}_2 includes $\{N_1, N_2, N_4, N_3\}$, \mathcal{M}_3 includes $\{N_1, N_3, N_4, N_2\}$, \mathcal{M}_4 includes $\{N_1, N_3, N_2, N_4\}$, \mathcal{M}_5 includes $\{N_1, N_4, N_2, N_3\}$, and \mathcal{M}_6 includes $\{N_1, N_4, N_3, N_2\}$. For each cortical region, N_1 denotes the mean maximum principal curvature, N_2 denotes the mean cortical thickness, N_3 denotes the mean sulcal depth and N_4 denotes the mean of average curvature.

Data distribution. Table 1 displays the gender/age distribution for both AD and LMCI groups. Both groups were matched in gender and age.

Comparison methods. We compared our proposed architectures with two conventional methods: (1) one-layer network architecture, and (2) concatenated multi-layer network architecture. For the first baseline method, we used the designed M morphological brain networks. For the second baseline method, we constructed the multi-layer network through concatenating all morphological networks in a large network $\mathcal{N} = \{N_1, \dots, N_M\}$ of size $R \times R \times M$ ($R=35, M=4$).

Evaluation. We evaluated our framework through varying the number of K_l selected features from 180 to 250 by adding 10 features at each evaluation step (Fig. 1). We noted that for the majority of the selected features' dimensions, the deep similarity architecture increased the classification performance in comparison with

	ACC		SEN		SPE	
	LH	RH	LH	RH	LH	RH
Layer A_1	57.69	61.15	56.59	57.73	58.53	67.37
Layer A_2	59.74	51.35	55.21	41.91	66.76	50.55
Layer A_3	49.43	66.47	47.22	63.39	51.22	66.46
Layer A_4	47.18	40.34	43.40	41.27	50.30	41.86
$A_1 - A_4$	59.03	61.51	55.90	59.02	61.89	63.10
Deep similarity	65.32	64.84	67.70	68.05	63.71	66.15
Multiplex \mathcal{M}_1	65.06	71.42	66.32	75.00	65.54	71.95
Multiplex \mathcal{M}_2	65.70	68.47	68.40	66.66	64.02	72.25
Multiplex \mathcal{M}_3	66.38	66.58	69.09	61.68	65.24	69.20
Multiplex \mathcal{M}_4	63.61	66.47	65.97	67.36	62.49	64.93
Multiplex \mathcal{M}_5	61.93	72.25	56.25	72.56	66.76	74.39
Multiplex \mathcal{M}_6	68.61	70.95	73.61	74.65	65.45	74.08

Table 2. LMCI/AD average classification accuracy for the proposed morphological network architectures by varying the number of top selected features from 180 to 250, with an incremental step of 10 features. ACC: accuracy. SEN: sensitivity. SPE: specificity.

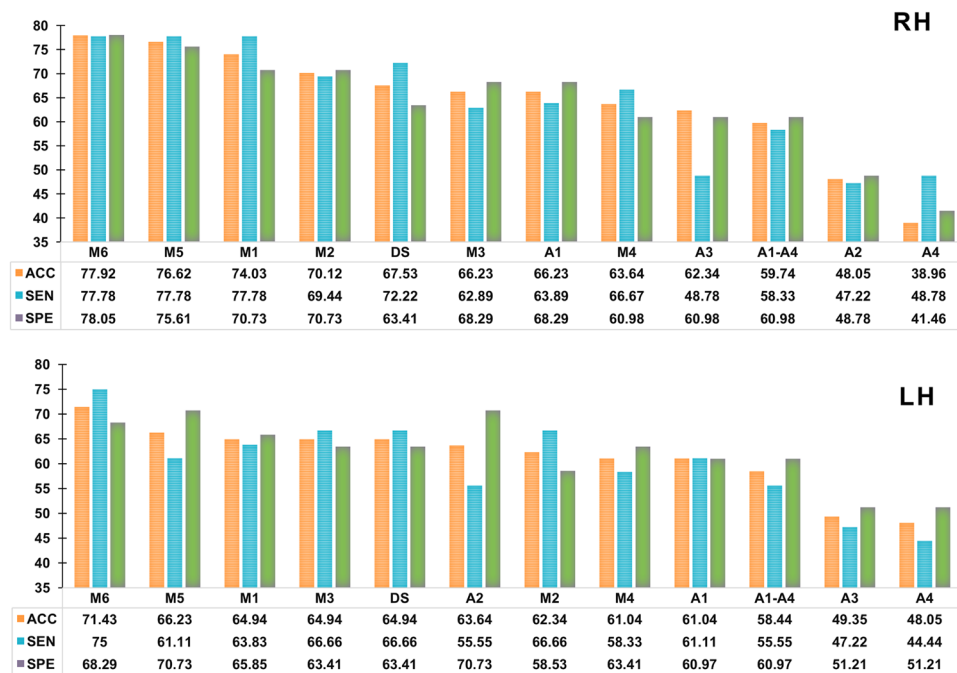


Figure 2. Best performances reached respectively by right hemisphere RH and left hemisphere LH, using different architecture networks. 'A' denotes cortical attribute, 'DS' denotes deep similarity, and 'M' denotes Multiplex.

the conventional methods (i.e., one-layer network and concatenated 4-layer network), for both left and right hemispheres. Remarkably, the classification accuracies highly improved when using particular brain multiplexes. Table 2 displays the average classification accuracies for baseline methods as well as for the proposed architectures. The best average accuracy was achieved by multiplex 6 (respectively including $\{N_1, N_4, N_3, N_2\}$ cortical attributes as intra-layers) for LH (68.61%), while it was achieved by multiplex 5 (respectively including $\{N_1, N_4, N_2, N_3\}$ cortical attributes as intra-layers) for RH (72.25%).

Notably, the best accuracies were obtained using a number of features equal to 220 and 210 for RH and LH, respectively (Fig. 2). When using one-layer morphological networks, the classification accuracy only reached 63.64% for LH and 66.23% for RH. These classification results decreased when concatenating features extracted from all morphological networks (*level 0*), as the classification rate was limited to 59.74% for RH and 58.44% for LH. However, when we further integrated the similarity networks between all pairs of layers in the proposed deep similarity network architecture (*level 1*), the performance significantly increased for both RH (67.53%) and LH (64.94%) compared to one-layer network and concatenated one-layer networks architectures. For both hemispheres, the highest accuracies were achieved using multiplex \mathcal{M}_6 , multiplex \mathcal{M}_5 , and multiplex \mathcal{M}_1 . Specifically,

	% of intra-layers		% of inter-layers	
	LH	RH	LH	RH
Multiplex \mathcal{M}_1	54.12	59.44	45.88	40.56
Multiplex \mathcal{M}_2	50.27	35.52	49.73	64.74
Multiplex \mathcal{M}_3	43.66	39.59	56.34	60.41
Multiplex \mathcal{M}_4	38.15	51.75	61.85	48.24
Multiplex \mathcal{M}_5	42.00	56.00	58.00	44.00
Multiplex \mathcal{M}_6	53.54	62.77	46.45	37.23

Table 3. Percentage of discriminative features belonging to the similarity inter-layers and the intra-layers for each of the proposed multiplexes.

multiplex \mathcal{M}_6 achieved the best accuracy among all architecture networks with a classification accuracy peaking at 77.92% for RH and 71.43% for LH. This shows that the proposed similarity networks allow to better discriminate between LMCI and AD subjects. This was also reflected by the percentages of discriminative features belonging to the similarity inter-layer networks for each multiplex as shown in Table 3. We note that for some multiplexes, more than 50% of K_f discriminative features lied in the network *inter-layers* -i.e., similarity networks.

Identified morphological connective biomarkers for LMCI/AD classification. We further explored our multiplex architecture and morphological networks to identify morphological connective biomarkers that discriminate between LMCI and AD patients. Since we aimed to find the most discriminative morphological connections, we chose the brain multiplex with the highest discriminative power. For LH, we found that the discriminative power of multiplex \mathcal{M}_6 was the most reproducible since it gave the best average accuracy across different numbers of selected features as well as the best accuracy (for $K_f=210$) in comparison with all other network architectures. As for RH, multiplex \mathcal{M}_5 achieved the best mean average across different numbers of selected features, while we noted the highest accuracy reached by multiplex \mathcal{M}_6 for a number of features equal to 220. Hence, we selected multiplex \mathcal{M}_6 achieving the best accuracies for both hemispheres to discover morphological connective biomarkers, using the specific number of discriminative features 220 (77.92%) and 210 (71.43%) for RH and LH, respectively.

In Fig. 3, we visualized using circular graphs the top most frequently selected morphological brain connectivities in multiplex \mathcal{M}_6 . Circular graphs were plotted for the top 10, 15 and 20 discriminative features, respectively. The thickness of each edge connecting a pair of ROIs represents the normalized rank of the discriminative brain connection. The most discriminative connections with the highest normalized ranks have thick edges, while those with less discriminative power have thinner edges. Blue edges denote connections belonging to a multiplex inter-layer, while red edges denote connections falling into a multiplex intra-layer.

We noted that 20% (resp. 50%) of the top 10 discriminative features were located in the multiplex inter-layers for RH (resp. LH). Using the normalized ranks, the most discriminative connective features for RH connected the Entorhinal Cortex (EC) (region 6) and the Caudal Middle Frontal Gyrus (CMFG) (region 3), EC and Temporal Pole (TP) (region 33), EC and Frontal Pole (FP) (region 32), EC and Bank of the Superior Temporal Sulcus (BSTS) (region 1) and the fifth connectivity was between the Paracentral Lobule (PL) (region 17) and Caudal Anterior-cingulate Cortex (CAC) (region 2), respectively. As for the LH, the most discriminative features connected the EC and the Rostral Middle Frontal Gyrus (RMFG) (region 27), EC and Lingual Gyrus (LG) (region 13), EC and Postcentral Gyrus (region 22), EC and CAC, and PL and Precentral Gyrus (region 24), respectively. We noted that the most discriminative morphological hub node in multiplex \mathcal{M}_6 for both hemispheres was the entorhinal cortex, where the top four discriminative connections with the highest normalized ranks branched from it.

Besides, even when we increased the number of discriminative features from 10 to 20, new connectivities appeared, most of them emerged from the EC. Moreover, the percentage of the top discriminative features belonging to inter-layers was low for RH (~15%) compared to LH (~45%). We also note that the majority of the most discriminative morphological brain connections fell into the 5th layer (A_3), which represents the mean sulcal depth attribute (Tables 4 and 5).

More importantly, while most discriminative features belonging to intra-layers emerged from EC for both left and right hemispheres, those belonging to inter-layers emerged from the fusiform gyrus and the paracentral lobule for RH. The same regions were present in LH with new other hub nodes including the precentral gyrus, transverse temporal cortex, rostral anterior cingulate cortex, and isthmus-cingulate cortex.

Discussion

We proposed a novel representation of brain connectivity to identify connective biomarkers based on the *morphology* of the cerebral cortex for distinguishing between late mild cognitively impaired patients and Alzheimer's disease patients. In this study, we unprecedentedly investigated the role of several morphological connectivity networks as well as the correlation between them to discover morphological connective brain biomarker fingerprinting the difference between LMCI and AD states. In particular, we proposed two brain architectures: the deep similarity network and the multiplex network. While in the first architecture we simply concatenated all possible similarity networks with the main morphological network, in the second one we constructed similarity networks only between successive layers, and generated different multiplexes by reordering the morphological layers.

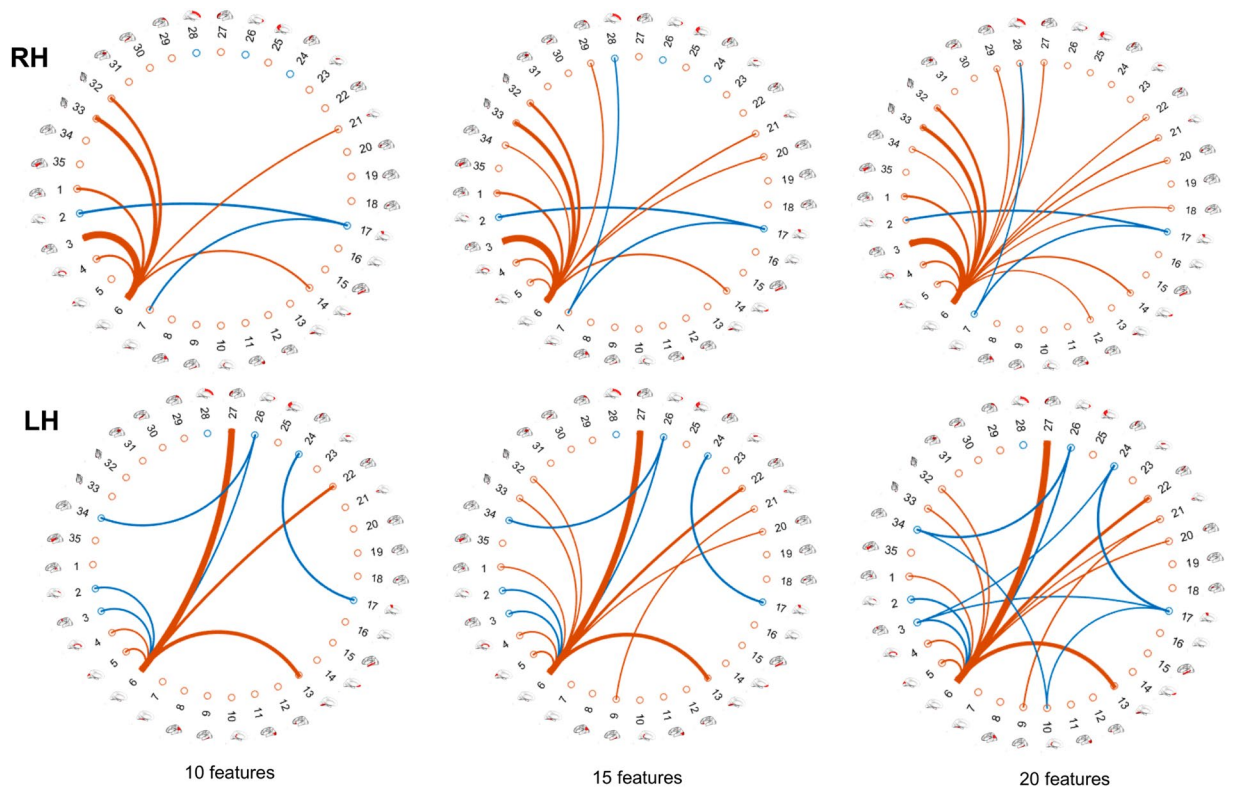


Figure 3. Most discriminative morphological cortical network connections between LMCI and AD for RH and LH, respectively.

Rank	First region	Second region	Layer
1	Entorhinal Cortex	Caudal Middle Frontal Gyrus	Mean sulcal depth
2	Temporal Pole	Entorhinal Cortex	Mean sulcal depth
3	Frontal Pole	Entorhinal Cortex	Mean sulcal depth
4	Entorhinal Cortex	Bank of the superior Temporal Sulcus	Mean sulcal depth
5	Paracentral Lobule	Caudal interior cingulate cortex	Similarity between mean sulcal depth and mean maximum principal curvature
6	Entorhinal Cortex	Unmeasured Corpus Callosum	Mean sulcal depth
7	Medial Orbital Frontal Cortex	Entorhinal Cortex	Mean sulcal depth
8	Paracentral Lobule	Fusiform Gyrus	Mean sulcal depth
9	Pericalcarine Cortex	Entorhinal Cortex	Mean sulcal depth
10	Superior Frontal Gyrus	Entorhinal Cortex	Mean sulcal depth

Table 4. Top 10 discriminative morphological connections in the cortex with their corresponding layers for the right hemisphere.

Our proposed architectures achieved better performances than one-layer morphological network and concatenated 4-layer networks. This shows that the aggregation of different similarities between morphological brain connections helps better discriminate between AD and LMCI patients compared to using a single morphological network or even all morphological networks without exploring their relationships (Fig. 2). Moreover, our multiplex architecture achieved the highest accuracies, which indicates that the similarity inter-layers between morphological networks are able to capture a higher-level discriminative information. This demonstrates that disease-driven changes in the cortical shape quantified using a specific cortical attribute can be also influenced by shape changes measured using a different cortical attribute.

Through using different cortical attributes and identifying the most of discriminative features by multiplex \mathcal{M}_6 , we found that the mean sulcal depth has the highest discriminative power (Tables 4 and 5). Sulcal depth has been identified in the literature as one of the quantitative measures of cerebral cortex, representing an important morphological biomarker for AD^{36,37}. Im *et al.* presented a surface-based method that investigated changes of sulcal shape in MCI and AD, using sulcal depth and average mean curvature³⁶. They showed that the progression of disease from NC to MCI and MCI to AD was coupled with shallowness in sulcal depth. The same finding³⁷ was

Rank	First region	Second region	Layer
1	Rostral Middle Frontal Gyrus	Entorhinal Cortex	Mean sulcal depth
2	Lingual Gyrus	Entorhinal Cortex	Mean sulcal depth
3	Postcentral Gyrus	Entorhinal Cortex	Mean sulcal depth
4	Entorhinal Cortex	Caudal Anterior-Cingulate Cortex	Similarity between mean maximum principal curvature and mean of average curvature
5	Precentral Gyrus	Paracentral Lobule	Similarity between mean of average curvature and mean sulcal depth
6	Transverse Temporal Cortex	Rostral Anterior Cingulate Cortex	Similarity between mean of average curvature and mean sulcal depth
7	Entorhinal Cortex	Cuneus Cortex	Mean sulcal depth
8	Entorhinal Cortex	Caudal Middle Frontal Gyrus	Similarity between mean maximum principal curvature and mean of average curvature
9	Rostral Anterior Cingulate Cortex	Entorhinal Cortex	Similarity between mean maximum principal curvature and mean of average curvature
10	Entorhinal Cortex	Unmeasured Corpus Callosum	Mean sulcal depth

Table 5. Top 10 discriminative morphological connections in the cortex with their corresponding layers for the left hemisphere.

replicated by Yun *et al.*, which proposed an automated sulcal depth measurement on cortical surface and highlighted that mean sulcal depth in MCI was lower than in NC.

The most discriminative morphological connectivities with the highest normalized ranks were established between EC and CMFG for RH, and EC and RMFG for LH. Many studies highlighted that RMFG is a discriminative region in AD diagnosis as well as CMFG³⁸. It was also noted that about 18% of the CMFG atrophies in AD patients³⁸.

One of the major findings of our study is the detection of morphological brain connectional biomarkers fingerprinting the distinction between LMCI and AD dementia brain states. We found that 85% (resp. 65%) of most RH (resp. LH) discriminative regions connected to the EC fingerprint LMCI/AD classification (Fig. 3). The EC has a major role in working memory processing^{39–43}. Its importance was revealed due to its anatomical interconnection with the hippocampus, which is the major region responsible of memory formation^{43,44}. EC role consists of generating coding schemes for new memories and storing them temporarily. It has numerous reciprocal connections with the hippocampus, specifically an effective connectivity in the hippocampus strongly depends on the connectivity among EC layers⁴⁵.

Our findings based on cortical morphological connectivity were in line with previous studies, since the EC has been considered as a good biomarker for AD and MCI in the literature^{46–50}. It has a great potential for detecting early memory decline and is considered as the region of early neurodegeneration caused by dementia. Velayudhan *et al.* examined the relationship between EC thickness, hippocampal volume and the whole brain volume, and showed that AD patients have thinner EC thickness and smaller hippocampal volume compared with MCI subjects⁴⁶. The same hypothesis about the role of EC was demonstrated by the work of Thaker *et al.*⁴⁷, which considered EC thickness as a marker of medial temporal and neocortical AD neuropathology. The review paper⁴⁸ also highlighted the early EC atrophy detection as an important anatomical marker for MCI and AD, since it was remarkably highly correlated with the early pathological changes in AD. Besides, greater changes in the right EC were present compared with the left one, which substantiates our results, since we achieved the best multiplex-based LMCI/AD classification performance using the right hemisphere.

Our study has a few limitations. *First*, although we used different types of morphological attributes, we simply concatenated all derived connectivities to extract features without creating fused predictors of disease diagnosis. *Second*, though we identified key morphological connectional biomarkers for LMCI stage, mainly involving the entorhinal cortex, we did not investigate the connection of the discovered cortical regions to other non-cortical regions (e.g., EC to hippocampus). It is still not clear how the shape-based morphological connectivities of EC can be altered with the hippocampus connectivities such as functional or structural. *Third*, since MCI is a progressive disease, tracking the discriminative power of the identified morphological biomarkers can help better understand how the morphology of a specific discriminative region (e.g., EC thickness) gets altered progressively with time in cognition in relation to other cortical attributes^{42,44}. *Fourth*, although the structural underpinning of morphological networks remains unclear, the co-vary brain regions were suggested as a result of mutually trophic influences or common experience related plasticity^{51,52}. In particular, it was noted that the pattern of cortical thickness correlation of certain brain regions is similar to the underlying fiber connections from DTI tractography⁵³. Gong *et al.* also pointed out that approximately 35–40% convergent connections exist between brain networks using thickness and diffusion measurements, which suggests that thickness correlations include exclusive information⁵⁴.

In our future work, we will investigate longitudinal morphological connectivities to improve our framework as well as longitudinal morphological changes in the EC. Besides, we will use advanced methods for different morphological and similarity networks fusion⁵⁵ while integrating other multimodal brain networks (e.g., resting-state functional networks and structural diffusion networks) into our proposed brain multiplex architectures²⁶.

Methods

We first introduce our morphological brain network construction strategy from structural T1-w MRI. Then, we propose two different architectures to explore the relationship between multiple brain connectivity morphological

views: (1) a deep multi-level similarity network that aggregates different morphological brain networks with hierarchical combinations of similarity networks between them; and (2) morphological brain multiplex network, which is defined through inserting additional inter-layers between the aggregated networks. Last, we perform feature extraction and selection to classify a testing subject, and morphological biomarker identification. Figure 4 displays the key steps of the proposed framework.

Morphological Brain Network Definition. Following the cortical surface parcellation into R anatomical regions, for each ROI R_p , we average the cortical attribute a across all vertices v in R_i as follows:

$$\frac{1}{\#\{v \in R_i\}} \sum_{v \in R_i} a(v),$$

where $\#\{v \in R_i\}$ denotes the number of vertices v belonging to ROI R_p , and $a(v)$ the cortical attribute value assigned to vertex v . Ultimately, to define the morphological connection $N_a(i, j)$ in network N_a between ROIs R_i and R_j , we compute the absolute difference between averaged cortical attributes in both ROIs:

$$N_a(i, j) = \left| \frac{1}{\#\{v \in R_i\}} \sum_{v \in R_i} a(v) - \frac{1}{\#\{v \in R_j\}} \sum_{v \in R_j} a(v) \right|.$$

Given R cortical regions in each hemisphere, the size of each fully connected morphological network is $R \times R$. We note that according to our definition, as two ROIs R_i and R_j become more similar in morphology, $N_a(i, j)$ tends to 0.

Proposed Morphological Network Architectures. To extract relevant and high-order morphological features from a set of M morphological cortical brain networks $\{N_1, \dots, N_M\}$, each encoding a specific shape attribute of the cortical surface, we propose ‘simple-to-complex’ strategies for building network architectures that capture different characteristics of how these networks interact with one another. In particular, high-order network architectures aim to reflect how these networks are nested with respect to one another in a high dimensional manifold of networks.

Proposed deep similarity network architecture construction. We first propose a deep multi-level network architecture, where each level integrates the similarity networks between all pairs of networks in the previous level. The relationship between pairs of networks is defined by the measure of Pearson correlation (Fig. 5A). Thus, we define the degree of correlation between different cortical networks at each level. The baseline level ($l=0$) is composed of all concatenated networks $\mathcal{N}^0 = \{N_1^0, \dots, N_M^0\}$. To build the next level, we create a larger multi-layer network through concatenating n_s similarity networks, where $n_s = C_M^2 = M!/(M-2)!$, representing the number of possible pairwise combinations between M networks. This produces a new deeper network $\mathcal{N}^1 = \mathcal{N}^0 \cup \{S_{1,2}, \dots, S_{pq}, \dots, S_{M-1,M}\}$, where p and q represent the indices of two different networks in \mathcal{N}^0 . For brevity, we note the baseline network at a specific level l as $\mathcal{N}^l = \{N_1^l, \dots, N_{M_l}^l\}$, where M_l represents the total number of level l networks. Hence, in the next level ($l+1$), we consider \mathcal{N}^l as the baseline network, and add the similarity networks at a specific level $l+1$ as: $\mathcal{S}^{l+1} = \{S_{1,2}, \dots, S_{pq}, \dots, S_{M_l-1, M_l}\}$, where S_{pq} represents the similarity network between networks N_p^l and N_q^l . From level to level, we gradually add similarity networks between networks in the previous level (including similarity networks), thereby producing deeper networks from one level to the next one, where $\mathcal{N}^{l+1} = \mathcal{N}^l \cup \mathcal{S}^{l+1}$ (Fig. 5C). The deep multi-level similarity network architecture is thus constructed in a hierarchical way, which captures not only network-to-network similarities, but also ‘similarity-to-similarity’ similarities.

Proposed ensemble multiplex network architecture construction. Although the proposed deep similarity network architecture allows to explore similarities between networks at different hierarchical levels, this aggregates the similarity networks at the end of previous multi-layer network, in an agglomerative manner without enabling us to take account into the most correlated pairs of morphological networks. To enforce a more structured design of networks and their similarities, we propose to use a multiplex network to model the inter-relations between different layers. In a generic way, we define a brain multiplex \mathcal{M} as a set of M intra-layers $\{N_1, \dots, N_M\}$ (i.e., morphological networks), where between two consecutive layers N_i and N_j , we slide an inter-layer $S_{i,j}$. This yields to following multiplex architecture: $\mathcal{M} = \{N_1, S_{1,2}, N_2, \dots, N_j, S_{i,j}, N_j, \dots, N_M\}$ (Fig. 5B). Unlike the previous architecture (Fig. 5C), we note that for a specific multiplex, we are only allowed to explore similarities between consecutive layers. We also use Pearson Correlation to generate inter-layers as the deep similarity network architecture. Hence, to explore the inter-relationship between all possible combinations of layers for each subject, we generate K multiplexes through simply reordering the intra-layer networks while fixing the first intra-layer, thereby generating an ensemble of multiplexes $\mathbb{M} = \{\mathcal{M}_1, \dots, \mathcal{M}_K\}$. Each of these multiplexes captures specific similarities between different kinds of morphological networks (e.g., sulcal depth network and cortical thickness network) that may not be present in another brain multiplex.

Network feature extraction and selection for classification. For each of the proposed subject-specific network architectures in the previous section, we perform feature extraction, selection and LMCI/AD classification as follows.

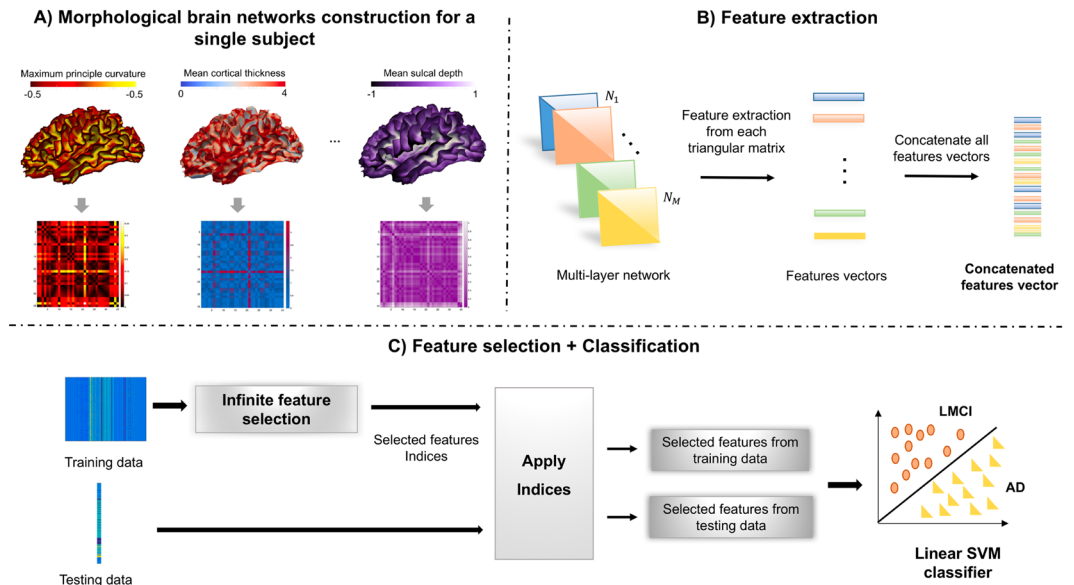


Figure 4. Illustration of AD/LMCI classification framework steps for the proposed cortical morphological network architectures. (A) We generate different morphological networks, each derived from a specific attribute of the cortical surface shape. (B) For each multi-layer network, we extract features from the triangular part of each cortical connectivity matrix. (C) For feature selection, we use IFS strategy (Roffo *et al.*⁵⁶), then we train a linear support vector machine classifier using the selected connectional features.

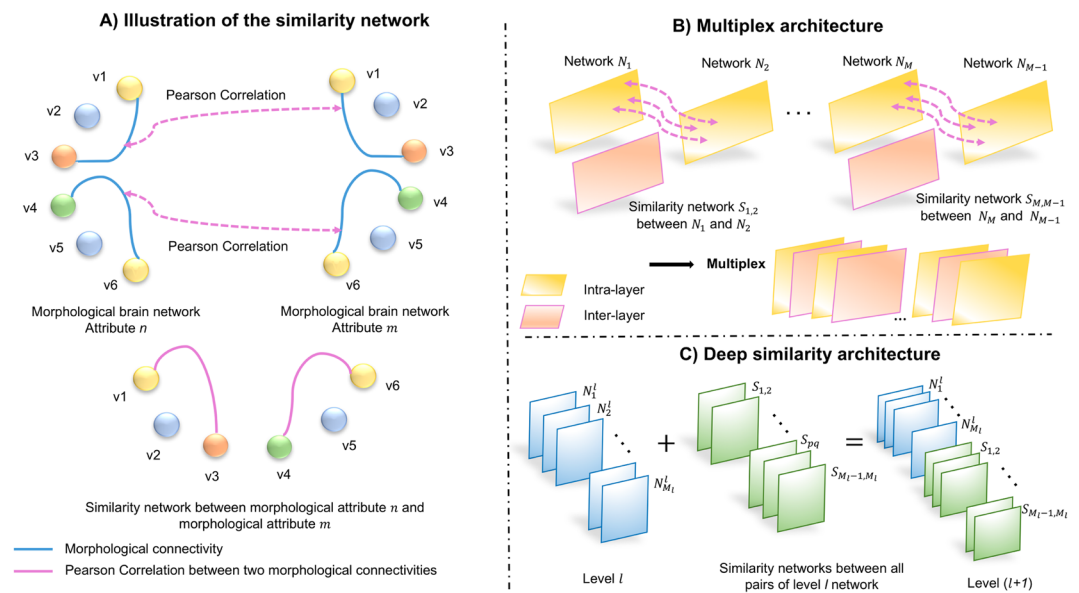


Figure 5. Proposed deep similarity network and multiplex network architectures, with illustration of the similarity network construction step. (A) We generate the similarity network between two morphological attribute networks by computing Pearson Correlation between them. (B) We construct the multiplex architecture where each inter-layer is a similarity network. (C) We consider the inter-relations between \mathcal{N}^l networks through progressively concatenating, from a previous level l to a current level $(l + 1)$, all possible similarity networks between pairs of networks.

Feature extraction. To explore the discriminative power of each region-to-region morphological connectivity in the cortex, we directly use the weights of edges in the morphological network as connectional brain features.

Since the constructed morphological connectivity matrix (or network N_a) is symmetric (Fig. 4A), connectional features are extracted for each subject through concatenating the weights of all connectivities in each triangular matrix. Of note, for each network of size $R \times R$, we extract a feature vector of size $(R(R - 1)/2)$. For the deep similarity network and multiplex architectures, we extract features from each network in the architecture, then

concatenate them all together into a high-dimensional feature vector. For a network architecture comprising M_n networks, the size of the final feature vector is $M_n \times (R(R-1)/2)$.

Feature selection and classification. Due to the high-dimensionality of the extracted feature vectors and the small number of data samples, feature selection is a key step in classification tasks to both reduce the dimension of the training feature vectors and single out the most discriminative features. To this aim, we train a support vector machine (SVM) classifier using leave one-out cross-validation (LOO-CV) strategy. Given P subjects, we apply Infinite Feature Selection (IFS)⁵⁶ in a supervised manner using the $(P-1)$ training subjects to select the top K_f features that significantly distinguish LMCI from AD patients. The most frequently selected features across different cross-validation schemes represent the morphological connective biomarkers that allow to distinguish between AD and LMCI patients.

IFS method is a filter-based algorithm that aims to avoid over-fitting in a high-dimensional data by not considering irrelevant and/or redundant features. Compared with other feature selection methods, IFS has a compelling aspect that allows to efficiently identify reliable distinctive features for classification tasks. Most feature selection methods, which rank and select features, evaluate the importance of each feature individually, usually by neglecting potential interactions among the elements of the joint set. However, IFS performs joint ranking with selection and the score attributed for each feature is influenced by all other features. The idea is based on building a graph for the feature distribution, where the vertices denote the features and the edges represent the pairwise relationships among the feature distribution. Then, the algorithm ranks different morphological features by their importance and discriminative power. It evaluates the importance of a given feature while considering all the possible subsets of features. Given the output indices of the ranked features, we select the top K_f ranked features to train a linear SVM classifier using LOO-CV strategy to assign a label (LMCI or AD) to a new testing subject (Fig. 4C).

Identification of morphological connective biomarker. To identify morphological connective biomarkers, we select the top n_f indices of K_f discriminative features ranked by IFS⁵⁶ across all PLOO-CV. Specifically, we generate a matrix of size $n_f \times P$, where each row represents the top ranked indices of features and each column represents a specific rank. For a given feature f_k , we calculate its normalized rank across different LOO-CV as follows: $r(f_k) = (\sum_{i=1}^P \sum_{j=1}^{n_f} \delta_{ij} w_{ij}(f_k)) / P$, where $\delta_{ij} = 1$ if f_k is selected, $\delta_{ij} = 0$ otherwise. The weight $w_{ij}(f_k)$ denotes the corresponding weight of feature f_k assigned by IFS, at the i^{th} LOO and j^{th} rank. Next, we identify the connective biomarkers as features with the top normalized ranks.

Availability of materials and data. The data that support the findings of this study are available from ADNI data (<http://adni.loni.usc.edu/>). For reproducibility and comparability, the authors will make available upon request all morphological networks generated based on the four cortical attributes (maximum principal curvature, cortical thickness, sulcal depth, and average curvature) for 77 subjects (41 AD and 36 LMCI) following the approval by ADNI Consortium. The Matlab code for generating an ensemble of multiplexes using M brain networks for a single subject (e.g., morphological, structural, or functional) is also available from the authors upon request.

References

1. Yu, R. *et al.* Connectivity strength-weighted sparse group representation-based brain network construction for MCI classification. *Human Brain Mapping* **38**(5), 2370–2383 (2017).
2. Wee, C.-Y., Yang, S., Yap, P.-T. & Shen, D. For the Alzheimer's Disease Neuroimaging Initiative. Sparse Temporally Dynamic Resting-State Functional Connectivity Networks for Early MCI Identification. *Brain Imaging and Behavior* **10**(2), 342–356 (2016).
3. Chen, X. *et al.* High-Order Resting-State Functional Connectivity Network for MCI Classification. *Human Brain Mapping* **37**(9), 3282–3296 (2016).
4. Casanova, R., Hsu, F.-C. & Espeland, M. A. For the Alzheimer's Disease Neuroimaging Initiative. Classification of Structural MRI Images in Alzheimer's Disease from the Perspective of Ill-Posed Problems. *PLoS ONE* **7**(10), e44877 (2012).
5. Fei, F., Jie, B. & Zhang, D. Frequent and Discriminative Subnetwork Mining for Mild Cognitive Impairment Classification. *Brain Connectivity* **4**(5), 347–360 (2014).
6. Jie, B., Zhang, D., Wee, C.-Y. & Shen, D. Topological Graph Kernel on Multiple Thresholded Functional Connectivity Networks for Mild Cognitive Impairment Classification. *Human Brain Mapping* **35**(7), 2876–2897 (2014).
7. Zhang, Y., Zhang, H., Chen, X., Lee, S.-W. & Shen, D. Hybrid High-order Functional Connectivity Networks Using Resting-state Functional MRI for Mild Cognitive Impairment Diagnosis. *Scientific Reports* **7**, 6530 (2017).
8. Ebadi, A. *et al.* Ensemble Classification of Alzheimer's Disease and Mild Cognitive Impairment Based on Complex Graph Measures from Diffusion Tensor Images. *Frontiers in Neuroscience* **11**, 56 (2017).
9. Petrov, D. *et al.* Evaluating 35 Methods to Generate Structural Connectomes Using Pairwise Classification. *International Conference on Medical Image Computing and Computer-Assisted Intervention*, 515–522 (2017).
10. Goryawala, M. *et al.* Inclusion of Neuropsychological Scores in Atrophy Models Improves Diagnostic Classification of Alzheimer's Disease and Mild Cognitive Impairment. *Computational Intelligence and Neuroscience*, 865265 (2015).
11. Weiner, M. W. *et al.* The Alzheimer's Disease Neuroimaging Initiative: A review of papers published since its inception. *Alzheimer's & Dementia: The Journal of the Alzheimer's Association* **8**(10), S1–68 (2012).
12. McEvoy, L. K. *et al.* Alzheimer Disease: Quantitative Structural Neuroimaging for Detection and Prediction of Clinical and Structural Changes in Mild Cognitive Impairment. *Radiology* **251**(1), 195–205 (2009).
13. Ridgway, G. R. *et al.* Early-onset Alzheimer disease clinical variants: Multivariate analyses of cortical thickness. *Neurology* **79**(1), 80–84 (2012).
14. Dickerson, B. C. *et al.* The Cortical Signature of Alzheimer's Disease: Regionally Specific Cortical Thinning Relates to Symptom Severity in Very Mild to Mild AD Dementia and is Detectable in Asymptomatic Amyloid-Positive Individuals. *Cerebral Cortex* **19**(3), 497–510 (2009).
15. Frisoni, G. *et al.* Detection of grey matter loss in mild Alzheimer's disease with voxel based morphometry. *Journal of Neurology, Neurosurgery, and Psychiatry* **73**(6), 657–664 (2002).

16. Chételat, G. *et al.* Using voxel-based morphometry to map the structural changes associated with rapid conversion in MCI: A longitudinal MRI study. *NeuroImage* **27**, 934–46 (2005).
17. Chételat, G. *et al.* Mapping gray matter loss with voxel-based morphometry in mild cognitive impairment. *Neuroreport* **13**(15), 1939–43 (2002).
18. Zhou, L., Wang, Y., Li, Y., Yap, P.-T. & Shen, D., the Alzheimer's Disease Neuroimaging Initiative (ADNI). Hierarchical Anatomical Brain Networks for MCI Prediction: Revisiting Volumetric Measures. *PLoS ONE* **6**(7), e21935 (2011).
19. Källin, A. M. *et al.* Subcortical Shape Changes, Hippocampal Atrophy and Cortical Thinning in Future Alzheimer's Disease Patients. *Frontiers in Aging Neuroscience* **9**, 38 (2017).
20. Shakeri, M., Lombaert, Herve T., Shashank, T. & Kadoury, S. Deep Spectral-Based Shape Features for Alzheimer's Disease Classification. *First International Workshop, SeSAMI 2016, Held in Conjunction with MICCAI 2016*, 15–24 (2016).
21. Liao, W. *et al.* Discerning Mild Cognitive Impairment and Alzheimer Disease from Normal Aging: Morphologic Characterization Based on Univariate and Multivariate Models. *Academic Radiology* **21**(5), 597–604 (2014).
22. Tripathi, S., Hossein, N. S., Shakeri, M. & Kadoury, S. Sub-Cortical Shape Morphology And Voxel-Based Features For Alzheimer's Disease Classification. *Conference: IEEE 14th International Symposium on Biomedical Imaging*, 991–994 (2017).
23. Liu, Y. *et al.* Impaired Long Distance Functional Connectivity and Weighted Network Architecture in Alzheimer's Disease. *Cerebral Cortex* **24**(6), 1422–1435 (2014).
24. Liu, M., Du, J., Jie, B. & Zhang, D. Ordinal Patterns for Connectivity Networks in Brain Disease Diagnosis. In *International Conference on Medical Image Computing and Computer-Assisted Intervention*, 1–9 (2016).
25. Padgett, J. F. & Christopher, K. Ansell. Robust Action and the Rise of the Medici. *American Journal of Sociology* **98**(6), 1259–1319 (1993).
26. Zippo, A. G. & Castiglioni, I. Integration of 18FDG-PET Metabolic and Functional Connectomes in the Early Diagnosis and Prognosis of the Alzheimer's Disease. *Current Alzheimer Research* **13**(5), 487–97 (2016).
27. Battiston, F., Nicosia, V., Chavez, M. & Latora, V. Multilayer motif analysis of brain networks. *Chaos*, **27**(4) (2016).
28. De Domenico, M., Sasai, S. & Arenas, A. Mapping Multiplex Hubs in Human Functional Brain Networks. *Frontiers in Neuroscience* **10**, 326 (2016).
29. Crofts, J. J., Forrester, M. & O'Dea, R. D. Structure-function clustering in multiplex brain networks. *Europhysics Letters* **116**(1), 18003 (2016).
30. La Rocca, M. *et al.* A Multiplex Network Model to Characterize Brain Atrophy in Structural MRI. *XXIII International Conference on Nonlinear Dynamics of Electronic Systems*, 189–198 (2017).
31. Soussia, M. & Rezik, I. High-order Connectomic Manifold Learning for Autistic Brain State Identification. In *International Workshop on Connectomics in Neuroimaging*, 51–59, (2017).
32. Lisowska, A., Rezik, I. & Alzheimers Disease Neuroimaging Initiative. (2017, September). Pairing-based Ensemble Classifier Learning using Convolutional Brain Multiplexes and Multi-view Brain Networks for Early Dementia Diagnosis. In *International Workshop on Connectomics in Neuroimaging*, 42–50, (2017).
33. Mueller, S. G. *et al.* The Alzheimer's Disease Neuroimaging Initiative. *Neuroimaging Clinics of North America*, **15**(4), 869–xii (2005).
34. Bruce, F. "Freesurfer," *Neuroimage* **62**(2), 774–781 (2012).
35. Fischl, B. *et al.* Automatically parcellating the human cerebral cortex. *Cereb Cortex* **14**(1), 11–22 (2004).
36. Im, K. *et al.* Sulcal morphology changes and their relationship with cortical thickness and gyral white matter volume in mild cognitive impairment and Alzheimer's disease. *Neuroimage* **43**(1), 103–13 (2008).
37. Yun, H. J., Im, K., Yang, J.-J., Yoon, U. & Lee, J.-M. Automated Sulcal Depth Measurement on Cortical Surface Reflecting Geometrical Properties of Sulci. *PLoS ONE* **8**(2), e55977 (2013).
38. Wee, C.-Y., Yap, P.-T., Shen, D. & Alzheimer's Disease Neuroimaging Initiative. Prediction of Alzheimer's Disease and Mild Cognitive Impairment Using Baseline Cortical Morphological Abnormality Patterns. *Human Brain Mapping*, **34**(12) (2013).
39. Bakkour, A., Morris, J. C., Wolk, D. A. & Dickerson, B. C. The effects of aging and Alzheimer's disease on cerebral cortical anatomy: Specificity and differential relationships with cognition. *NeuroImage* **76**, 332–344 (2013).
40. Bannerman, D. M. *et al.* The role of the entorhinal cortex in two forms of spatial learning and memory. *Experimental Brain Research* **141**(3), 281–303 (2001).
41. Fransen, E. Functional role of entorhinal cortex in working memory processing. *Neural Networks* **18**(9), 1141–9 (2005).
42. Eichenbaum, H., Ott, T. & Cohen, N. J. Two functional components of the hippocampal memory system. *Behavioral and Brain Sciences* **17**(3), 449–472 (1994).
43. Riedel, G. & Micheau, J. Function Of The Hippocampus In Memory Formation: Desperately Seeking Resolution. *Progress in Neuro-Psychopharmacology and Biological Psychiatry* **25**(4), 835–853 (2001).
44. Teylerand, J. & Rudy, W. The Hippocampal Indexing Theory and Episodic Memory: Updating the Index. *Hippocampus* **17**(12), 1158–69 (2007).
45. Lopez-Madrona, V. J., Matias, F. S., Pereda, E., Canals, S. & Mirasso, C. R. The Role of The Entorhinal Cortex In The Effective Connectivity Of The Hippocampal Formation. *Chaos* **27**(4), 047401 (2017).
46. Velayudhan, L. *et al.* Entorhinal Cortex Thickness Predicts Cognitive Decline in Alzheimer's Disease. *Journal of Alzheimer's Disease* **33**(3), 755–66 (2013).
47. Thaker, A. A. *et al.* Entorhinal Cortex: Antemortem Cortical Thickness and Postmortem Neurofibrillary Tangles and Amyloid Pathology. *American Journal of Neuroradiology* **38**(5), 961–965 (2017).
48. Zhou, M., Zhang, F., Zhao, L., Qian, J. & Dong, C. Entorhinal cortex: a good biomarker of mild cognitive impairment and mild Alzheimer's disease. *Reviews in the Neurosciences* **27**(2), 185–95 (2016).
49. Van Hoesen, G. W., Hyman, B. T. & Damasio, A. R. Entorhinal Cortex Pathology in Alzheimer's Disease. *Hippocampus* **1**(1), 1–8 (1991).
50. Eustache, F., Desgranges, B., Giffard, B., de la Sayette, V. & Baron, J. C. Entorhinal cortex disruption causes memory deficit in early Alzheimer's disease as shown by PET. *Neuroreport* **12**(4), 683–5 (2001).
51. Ferrer, I. *et al.* Transforming growth factor- α immunoreactivity in the developing and adult brain. *Neuroscience* **66**, 189–199 (1995).
52. Mechelli, A., Friston, K., Frackowiak, R. & Price, C. Structural covariance in the human cortex. *Journal of Neuroscience* **25**, 8303–8310 (2005).
53. Lerch, J. *et al.* Mapping anatomical correlations across cerebral cortex (MACACC) using cortical thickness from MRI. *NeuroImage* **31**, 993–1003 (2006).
54. Gong, G., He, Y., Chen, Z. J. & Evans, A. C. Convergence and divergence of thickness correlations with diffusion connections across the human cerebral cortex. *NeuroImage* **59**, 1239–1248 (2012).
55. Singanamalli, A. *et al.* Supervised Multi-View Canonical Correlation Analysis: Fused Multimodal Prediction of Disease Diagnosis and Prognosis. *SPIE Medical Imaging*, 9038 (2014).
56. Roffo, G., Melzi, S. *Infinite Feature Selection*. *ICCV '15 Proceedings of the 2015 IEEE International Conference on Computer Vision (ICCV)*, 4202–4210 (2015).

Acknowledgements

This work was supported in part by the University of Dundee Research Fund. Data collection and sharing for this project was funded by the Alzheimer's Disease Neuroimaging Initiative (ADNI) (National Institutes of Health Grant U01 AG024904) and DOD ADNI (Department of Defense award number W81XWH-12-2-0012). ADNI is funded by the National Institute on Aging, the National Institute of Biomedical Imaging and Bioengineering, and through generous contributions from the following: AbbVie, Alzheimer's Association; Alzheimer's Drug Discovery Foundation; Araclon Biotech; BioClinica, Inc.; Biogen; Bristol-Myers Squibb Company; CereSpir, Inc.; Cogstate; Eisai Inc.; Elan Pharmaceuticals, Inc.; Eli Lilly and Company; EuroImmun; F. Hoffmann-La Roche Ltd and its affiliated company Genentech, Inc.; Fujirebio; GE Healthcare; IXICO Ltd.; Janssen Alzheimer Immunotherapy Research & Development, LLC.; Johnson & Johnson Pharmaceutical Research & Development LLC.; Lumosity; Lundbeck; Merck & Co., Inc.; Meso Scale Diagnostics, LLC.; NeuroRx Research; Neurotrack Technologies; Novartis Pharmaceuticals Corporation; Pfizer Inc.; Piramal Imaging; Servier; Takeda Pharmaceutical Company; and Transition Therapeutics. The Canadian Institutes of Health Research is providing funds to support ADNI clinical sites in Canada. Private sector contributions are facilitated by the Foundation for the National Institutes of Health (www.fnih.org). The grantee organization is the Northern California Institute for Research and Education, and the study is coordinated by the Alzheimer's Therapeutic Research Institute at the University of Southern California. ADNI data are disseminated by the Laboratory for Neuro Imaging at the University of Southern California. Data used in preparation of this article were obtained from the Alzheimer's Disease Neuroimaging Initiative (ADNI) database (adni.loni.usc.edu). As such, the investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at: http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf.

Author Contributions

I.M. implemented the code, designed the experiments, and drafted the manuscript. I.R. designed the framework and revised the manuscript. M.A.M. participated in idea discussion and reviewed the manuscript. The investigators within the ADNI contributed to the design and implementation of ADNI and/or provided data but did not participate in analysis or writing of this report. A complete listing of ADNI investigators can be found at http://adni.loni.usc.edu/wp-content/uploads/how_to_apply/ADNI_Acknowledgement_List.pdf.

Additional Information

Competing Interests: The authors declare no competing interests.

Publisher's note: Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.



Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit <http://creativecommons.org/licenses/by/4.0/>.

© The Author(s) 2018

Consortia

Alzheimer's Disease Neuroimaging Initiative

Michael Weiner³, Paul Aisen⁴, Ronald Petersen⁵, Clifford Jack⁶, William Jagust⁷, John Trojanowki⁸, Arthur Toga⁹, Laurel Beckett¹⁰, Robert Green¹¹, Andrew Saykin¹², John Morris¹³, Leslie Shaw¹³, Jeffrey Kaye¹⁴, Joseph Quinn¹⁴, Lisa Silbert¹⁴, Betty Lind¹⁴, Raina Carter¹⁴, Sara Dolen¹⁴, Lon Schneider¹⁴, Sonia Pawluczyk⁹, Mauricio Beccera⁹, Liberty Teodoro⁹, Bryan Spann⁹, James Brewer¹⁵, Helen Vanderswag¹⁵, Adam Fleisher^{15,54}, Judith Heidebrink¹⁶, Joanne Lord¹⁶, Sara Mason⁶, Colleen Albers⁶, David Knopman⁶, Kris Johnson⁶, Rachelle Doody¹⁷, Javier Villanueva-Meyer¹⁷, Munir Chowdhury¹⁷, Susan Rountree¹⁷, Mimi Dang¹⁷, Yaakov Stern¹⁸, Lawrence Honig¹⁸, Karen Bell¹⁸, Beau Ances¹³, Maria Carroll¹³, Mary Creech³, Erin Franklin¹³, Mark Mintun¹³, Stacy Schneider¹³, Angela Oliver¹³, Daniel Marson¹⁹, Randall Griffith¹⁹, David Clark¹⁹, David Geldmacher¹⁹, John Brockington¹⁹, Erik Roberson¹⁹, Marissa Natelson Love¹⁹, Hillel Grossman²⁰, Efe Mitsis²⁰, Raj Shah²¹, Leyla de Toledo-Morrell²¹, Ranjan Duara²², Daniel Varon²², Maria Greig²², Peggy Roberts²², Marilyn Albert²³, Chiadi Onyike²³, Daniel D'Agostino²³, Stephanie Kielb²³, James Galvin²⁴, Brittany Cerbone²⁴, Christina Michel²⁴, Dana Pogorelec²⁴, Henry Rusinek²⁴, Mony de Leon²⁴, Lidia Glodzik²⁴, Susan De Santi²⁴, P. Doraiswamy²⁵, Jeffrey Petrella²⁵, Salvador Borges-Neto²⁵, Terence Wong²⁵, Edward Coleman²⁵, Charles Smith²⁶, Greg Jicha²⁶, Peter Hardy²⁶, Partha Sinha²⁶, Elizabeth Oates²⁶, Gary Conrad²⁶, Anton Porsteinsson²⁷, Bonnie Goldstein²⁷, Kim Martin²⁷, Kelly Makino²⁷, M. Ismail²⁷, Connie Brand²⁷, Ruth Mulnard²⁸, Gaby Thai²⁸, Catherine Mc-Adams-Ortiz²⁸, Kyle Womack²⁹, Dana Mathews²⁹, Mary Quiceno²⁹, Allan Levey³⁰, James Lah³⁰, Janet Cellar³⁰, Jeffrey Burns³¹, Russell Swerdlow³¹, William Brooks³¹, Liana Apostolova³², Kathleen Tingus³², Ellen Woo³², Daniel Silverman³², Po Lu³², George Bartzokis³², Neill Graf-Radford³³, Francine Parfitt³³, Tracy Kendall³³, Heather Johnson³³, Martin Farlow¹², Ann Marie Hake¹², Brandy Matthews¹², Jared Brosch¹², Scott Herring¹², Cynthia Hunt¹², Christopher Dyck³⁴, Richard Carson³⁴, Martha MacAvoy³⁴, Pradeep Varma³⁴, Howard Chertkow³⁵, Howard Bergman³⁵, Chris Hosein³⁵, Sandra Black³⁶, Bojana Stefanovic³⁶, Curtis Caldwell³⁶, Ging-Yuek Robin Hsiung³⁶, Howard Feldman³⁷, Benita Mudge³⁷, Michele Assaly³⁷, Elizabeth Finger³⁸, Stephen Pasternack³⁸, Irina Rachisky³⁸, Dick Trost³⁸, Andrew Kertesz³⁸, Charles Bernick³⁹, Donna Munic³⁹, Marek-Marsel Mesulam⁴⁰, Kristine Lipowski⁴⁰, Sandra Weintraub³⁹, Borna Bonakdarpour³⁹, Diana Kerwin³⁹, Chuang-Kuo Wu³⁹, Nancy Johnson⁴⁰, Carl Sadowsky⁴¹, Teresa Villena⁴¹, Raymond Scott Turner⁴², Kathleen Johnson⁴², Brigid Reynolds⁴², Reisa Sperling⁴³, Keith Johnson⁴³, Gad Marshall⁴³, Jerome Yesavage⁴⁴, Joy Taylor⁴⁴, Barton Lane⁴⁴, Allyson Rosen⁴⁴, Jared Tinklenberg⁴⁴, Marwan Sabbagh⁴⁵, Christine Belden⁴⁵, Sandra Jacobson⁴⁵, Sherye Sirrel⁴⁵, Neil Kowall⁴⁶, Ronald Killiany⁴⁶, Andrew Budson⁴⁶, Alexander Norbash⁴⁶, Patricia Lynn Johnson⁴⁶, Thomas Obisesan⁴⁷, Saba Wolday⁴⁷, Joanne Allard⁴⁷, Alan Lerner⁴⁸, Paula Ogrocki⁴⁸, Curtis Tatsuoka⁴⁸, Parianne Fatica⁴⁸, Evan Fletcher⁴⁹, Pauline Maillard⁴⁹, John Olichney⁴⁹, Charles DeCarli⁴⁹, Owen Carmichael⁴⁹, Smita Kittur⁵⁰, Michael Borrie⁵¹, T-Y Lee⁵¹, Rob Bartha⁵¹, Sterling Johnson⁵², Sanjay Asthana⁵², Cynthia Carlsson⁵², Steven Potkin⁵³, Adrian Preda⁵³, Dana Nguyen⁵³, Pierre Tariot⁵⁴, Anna Burke⁵⁴, Nadira Trncic⁵⁴, Stephanie Reeder⁵⁴, Vernice Bates⁵⁵, Horacio Capote⁵⁵, Michelle Rainka⁵⁵, Douglas Scharre⁵⁶, Maria Katakis⁵⁶, Anahita Adeli⁵⁶, Earl Zimmerman⁵⁷, Dzintra Celmins⁵⁷, Alice Brown⁵⁷, Godfrey Pearlson⁵⁸, Karen Blank⁵⁸, Karen Anderson⁵⁸, Laura Flashman⁵⁹, Marc Seltzer⁵⁹, Mary Hynes⁵⁹, Robert Santulli⁵⁹, Kaycee Sink⁶⁰, Leslie Gordineer⁶⁰, Jef Williamson⁶⁰, Pradeep Garg⁶⁰, Franklin Watkins⁶⁰, Brian Ott⁶¹, Henry Querfurth⁶¹, Geoffrey Tremont⁶¹, Stephen Salloway⁶², Paul Malloy⁶², Stephen Correia⁶², Howard Rosen⁶³, Bruce Miller⁶³, David Perry⁶³, Jacobo Mintzer⁶⁴, Kenneth Spicer⁶⁴, David Bachman⁶⁴, Nunzio Pomara⁶⁵, Raymundo Hernando⁶⁶, Antero Sarrael⁶⁵, Norman Relkin⁶⁶, Gloria Chaing⁶⁶, Michael Lin⁶⁶, Lisa Ravdin⁶⁶, Amanda Smith⁶⁷, Balebail Ashok Raj⁶⁷ & Kristin Fargher⁶⁷

³Magnetic Resonance Unit at the VA Medical Center and Radiology, Medicine, Psychiatry and Neurology, University of California, San Francisco, USA. ⁴San Diego School of Medicine, University of California, California, USA. ⁵Mayo Clinic, Rochester, Minnesota, USA. ⁶Mayo Clinic, Rochester, USA. ⁷University of California, Berkeley, USA. ⁸University of Pennsylvania, Pennsylvania, USA. ⁹University of Southern California, California, USA. ¹⁰University of California, Davis, California, USA. ¹¹MPH Brigham and Women's Hospital/Harvard Medical School, Massachusetts, USA. ¹²Indiana University, Indiana, USA. ¹³Washington University St. Louis, St Louis, Missouri, USA. ¹⁴Oregon Health and Science University, Oregon, USA. ¹⁵University of California—San Diego, California, USA. ¹⁶University of Michigan, Ann Arbor, Michigan, USA. ¹⁷Baylor College of Medicine, Houston, State of Texas, USA. ¹⁸Columbia University Medical

Center, New York, South Carolina, USA. ¹⁹University of Alabama, Birmingham, Alabama, USA. ²⁰Mount Sinai School of Medicine, New York, USA. ²¹Rush University Medical Center, Rush University, Chicago, Illinois, USA. ²²Wien Center, Miami, Florida, USA. ²³Johns Hopkins University, Baltimore, Maryland, USA. ²⁴New York University, New York, NY, USA. ²⁵Duke University Medical Center, Durham, North Carolina, USA. ²⁶University of Kentucky, Lexington, Kentucky, USA. ²⁷University of Rochester Medical Center, Rochester, NY, USA. ²⁸University of California, Irvine, California, USA. ²⁹University of Texas Southwestern Medical School, Dallas, Texas, USA. ³⁰Emory University, Atlanta, Georgia, USA. ³¹University of Kansas, Medical Center, Kansas, USA. ³²University of California, Los Angeles, California, USA. ³³Mayo Clinic, Jacksonville, Jacksonville, USA. ³⁴Yale University School of Medicine, New Haven, Connecticut, USA. ³⁵McGill University, Montreal-Jewish General Hospital, Montreal, Canada. ³⁶Sunnybrook Health Sciences, Ontario, USA. ³⁷U.B.C. Clinic for AD & Related Disorders, Vancouver, BC, Canada. ³⁸Cognitive Neurology - St. Joseph's, Ontario, USA. ³⁹Cleveland Clinic Lou Ruvo Center for Brain Health, Ohio, USA. ⁴⁰Northwestern University, San Francisco, USA. ⁴¹Premiere Research Inst (Palm Beach Neurology), west Palm Beach, USA. ⁴²Georgetown University Medical Center, Washington, DC, USA. ⁴³Brigham and Women's Hospital, Massachusetts, USA. ⁴⁴Stanford University, California, USA. ⁴⁵Banner Sun Health Research Institute, Sun City, AZ, 85351, USA. ⁴⁶Boston University, Massachusetts, USA. ⁴⁷Howard University, Washington, DC, USA. ⁴⁸Case Western Reserve University, Ohio, USA. ⁴⁹University of California, Davis – Sacramento, California, USA. ⁵⁰Neurological Care of CNY, Liverpool, NY, 13088, USA. ⁵¹Parkwood Hospital, Pennsylvania, USA. ⁵²University of Wisconsin, Wisconsin, USA. ⁵³University of California, Irvine, BIC, USA. ⁵⁴Banner Alzheimer's Institute, Phoenix, AZ, 85006, USA. ⁵⁵Dent Neurologic Institute, Amherst, NY, USA. ⁵⁶Ohio State University, Ohio, USA. ⁵⁷Albany Medical College, Albany, NY, USA. ⁵⁸Hartford Hospital, Olin Neuropsychiatry Research Center, Hartford, Connecticut, USA. ⁵⁹Dartmouth-Hitchcock Medical Center, Lebanon, New Hampshire, USA. ⁶⁰Wake Forest University Health Sciences, Winston-Salem, North Carolina, USA. ⁶¹Rhode Island Hospital, state of Rhode Island, Providence, RI, 02903, USA. ⁶²Butler Hospital, Providence, Rhode Island, USA. ⁶³University of California, San Francisco, USA. ⁶⁴Medical University South Carolina, Charleston, SC, 29425, USA. ⁶⁵Nathan Kline Institute, Orangeburg, New York, USA. ⁶⁶Cornell University, Ithaca, New York, USA. ⁶⁷USF Health Byrd Alzheimer's Institute, University of South Florida, Tampa, FL, 33613, USA.