

The Effect of Poloxamer 407-Based Hydrogel on the Osteoinductivity of Demineralized Bone Matrix

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Background: Demineralized bone matrix (DBM) is used for bone healing due to its osteoinductivity, but it requires a carrier for clinical application. Here, we report the effects on the osteoinductivity of DBM by use of a poloxamer 407-based hydrogel as the carrier, compared to sterile water.

Methods: DBM-W and DBM-H represent 27 wt% of DBM with sterile water and DBM with a poloxamer 407-based hydrogel, respectively. Both of the compositions were applied to human mesenchymal stem cell (MSC) cultures, and monitored for alkaline phosphatase (ALP) staining and ALP activity. Six 10-week-old athymic nude rats were used for abdominal muscle grafting with either DBM-W or DBM-H, and were tested by plane radiography, microfocus X-ray computed tomography (CT), and decalcified histology to evaluate ectopic bone formation.

Results: The DBM-W group showed stronger ALP staining at 7, 14, and 21 days of treatment, and significantly higher ALP activity at 7 and 14 days of treatment, compared to the DBM-H group. Plane radiography could not confirm the radio-opaque lesions in the rat ectopic bone formulation model. However, ectopic bone formation was observed in both groups by micro-CT. Compared to the DBM-H group, the DBM-W group showed higher bone volume, percent bone volume and trabecular number, and the difference in percent bone volume was statistically significant. Decalcified histology found bony tissue with lamellation in both groups.

Conclusions: Our results suggest that poloxamer 407-based hydrogel has efficacy as a DBM carrier since it shows ectopic bone formation, but its effects on the quality and quantity of osteoblastic differentiation in rat abdominal ectopic bone and MSC are considered negative.

Keywords: Demineralized bone matrix, Osteoinductivity, Carrier, Poloxamer 407-based hydrogel

Bone fusion surgery is required for the surgical treatment of degenerative spinal diseases, bone fractures, and tumors. To date, autogenous bone has been considered the

Received July 9, 2013; Accepted November 23, 2013

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most ideal bone graft for bone fusion. However, due to its quantitative limits and donor site morbidity,¹⁾ the usage of autogenous bone is gradually decreasing, and various bone graft substitutes have been developed. Bioactive ceramics such as hydroxyapatite, tricalcium phosphate, and calcium pyrophosphate are representative bone graft substitutes which are clinically applied to areas such as spine fusion.²⁻⁶⁾ While bioactive ceramics have appealing advantages such as mass production and low risk of inflammation and virus infection, they are osteoconductive materials with a negligible level of osteoinductivity, and thus need to be

mixed with autogenous bone for application. To complement this limitation, bone morphogenetic proteins (BMPs) with high osteoinductivity have been developed and applied in various areas, including spinal fusion and tibial non-union.^{7,8)} BMPs are gradually replacing autogenous bone grafts, since they are known to facilitate bone healing and improve fusion rate. However, their many advantages are also accompanied by some disadvantages. They are costly, and are known to cause side effects including ectopic bone formation, formation of seroma, 9) neuropathy by neuritis, osteolysis, and soft tissue swelling. 10) Moreover, the risk of retrograde ejaculation, 11) male infertility and malignancy were also recently reported. Thus, the risk of possible side effects is now advised, rather than their usefulness. Due to its osteoinductivity, demineralized bone matrix (DBM) was frequently applied clinically, before BMP was actively used. However, the usage of DBM is also decreasing, since it has significantly lower osteoinductivity than BMP and mainly functions through osteoconductivity. Thus, it cannot be applied for bone fusion by itself.¹²⁾ Despite the limitations, use of DBM is still advantageous to reduce autogenous bone graft without using BMP, as DBM rarely develops side effects.

DBM is provided in a powder form, which is hard to pattern and fix to the right position. Thus, it requires various types of carriers such as calcium sulfate, hydroxyapatite, tricalcium phosphate, carboxymethylcellulose, glycerol, hyaluronic acid, and bovine gelatin.

Poloxamer 407 is a tri-block copolymer consisting of a central hydrophobic polypropylene oxide block flanked by two hydrophilic ethylene oxide blocks. Poloxamer 407 shows thermoreversible properties, forming a gel-like composite at room temperature to facilitate the solubilization of poorly water-soluble drugs, and forming a gel state at body temperature. Poloxamer 407 has been used in the pharmaceutical field because of the properties such as solubilization promotion, stabilization and bio-adhesiveness. 14,15)

The putty type of DBM with a carrier improves handling during surgery, but effects on osteoinductivity or osteogenesis have not yet been well examined. This study evaluates the effect on osteogenesis of using poloxamer 407-based hydrogel as a carrier, through various *in vitro* and *in vivo* experiments.

METHODS

Materials

DBM-W and DBM-H were synthesized from CG Bio (Seongnam, Korea). DBM-W consists of 27 wt% DBM and 73 wt% sterile water. DBM-H is a putty form mixed with 75

wt% temperature-sensitive poloxamer 407-based hydrogel as a carrier, and 25 wt% DBM.

In Vitro Study

Cell culture, ALP staining and ALP assay

Alkaline phosphatase (ALP) staining and ALP assay were performed to examine the effects of DBM-W and DBM-H on bone differentiation of messenchymal stem cells (MSCs). 3×10^4 of bone-marrow derived MSCs (BM-MSCs) were seeded on 24-well plates and incubated in a CO₂ incubator. After 24 hours of incubation, transwells with either DBM-W or DBM-H were inserted into the plates of BM-MSCs. The treated cells were further incubated with differentiation media including 10 nM dexamethasone, 10 mM beta-glycerophosphate, and 100 μM ascorbic acid for 7, 14, or 21 days. For ALP staining, the differentiation medium was discarded and the cells were washed with 1X Dulbecco's Phosphate-Buffered Saline (DPBS, Life Technology Korea, Seoul, Korea) twice, fixed with 4% paraformaldehyde for 10 minutes, and washed with dH₂O. The fixed cells were stained with 0.25% naphthol AS-MX phosphate alkaline solution containing fast blue RR salt (Sigma-Aldrich, Brodby, Denmark) for 10 minutes, washed with dH₂O, and subjected to optical microscopy. For ALP assay, the differentiated cells were washed with 1X DPBS (Life Technology Korea) twice and lysed with 500 µL of 0.2% Triton X-100 (Sigma-Aldrich). Equal volumes (20 μ L) of p-nitrophenylphosphate and diethanolamine buffer with 0.5 mM MgCl₂ (pH 9.8, Sigma-Aldrich) were mixed and reacted for 20 minutes. The reactions were stopped by the addition of 0.2 N NaOH, and monitored by measuring the absorbance at 405 nm with an enzyme-linked immunosorbent assay plate reader.

In Vivo Study

Animals and experimental design

A total of 6 male athymic nude rats (Hsd:RH-Foxn1, 10 weeks, 280–290 g) were used for this study, with approval from the Standing Ethical Committee at the Laboratory for Animal Research in the Clinical Research Institute of the Seoul National University Hospital (IACUC No. 10-0083). This study followed the 'Guiding Principles for Research Involving Animals and Human Beings' by the American Physiological Society. Rats in this study were given at least a week for adaptation. They were housed in a specific pathogen-free environment with a 12-hour light/12-hour dark cycle at 24°C. Sterile water was available at all times as drinking water, and the animals were fed commercial diets. Three out of 6 rats were randomly selected for the test with DBM-W, and the rest were assigned to the DBM-

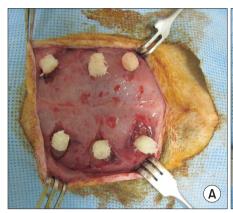




Fig. 1. Operation process. Three incisions and pouches were made per side (right and left) at the abdominal muscle of athymic nude rats. (A) A picture of fixed demineralized bone matrix (DBM). (B) A picture of the sutured pouches after DBM implantation.

H group. Zoletil (0.4 mL/kg, Virbac Laboratories, Carros, France) and rompun (10 mg/kg, Bayer Korea Ltd., Seoul, Korea) were used for intraperitoneal anesthesia. After depilation and skin preparation, a longitudinal skin incision was made in the center of the abdominal region. Three pouches were made in the right and left sides of the abdominal muscle. One milliliter of DBM was inserted into the pouches, and the facial layer and skin were sutured (Fig. 1). Immediately after the operation, the rats were intravenously administered with 100 mg of cephazolin, and were then raised without any intervention. Eight weeks later, the animals were sacrificed with an intravenous injection of KCl, after abdominal anesthesia with zoletil and rompun for further tests.

Radiological study

To evaluate the ectopic bone formation, plane radiography was performed under the conditions of a tube to cassette distance of 60 cm, 45 kV, 2.5 mA, and 12 ms for all animal samples. The prepared specimens were subjected to microfocus X-ray computed tomography (Skyscan 1174, Bruker, Kontich, Belgium) with an aluminium filter at 130 kV and 30 μ A. Scanned images were reconstructed with sagittal and coronal plane, and the formation of newly calcified tissue was evaluated quantitatively by analyzing the bone surface/volume ratio, percent bone volume, trabecular pattern factor, structure model index, trabecular thickness, trabecular number, trabecular separation, and degree of anisotropy.

Histological study

Decalcified slides were prepared for hematoxylin and eosin staining to monitor the ectopic bone formation.

Statistical Analyses

The non-parametric Mann-Whitney U-test was used for the analysis. A p-value less than 0.05 was accepted as significant (p < 0.05).

RESULTS

In Vitro Results

The DBM-W group showed stronger ALP staining at 7, 14, and 21 days of treatment, although the staining had faded at 21 days in both of the groups (Fig. 2). ALP activity was also significantly higher in the DBM-W group for the 7- and 14-day treatments (p < 0.001 for DBM-W, p = 0.0003 for DBM-H). The same pattern was observed at 21 days, but no statistical significance was found (p = 0.166).

In Vivo Results

There were no complications such as infection or death throughout the study with the 6 rats.

Radiological results

Plane radiography could not detect any clear radio-opaque lesions in either of the groups (Fig. 3). However, micro-CT confirmed distinct calcified tissues in both of the groups by coronal, axial and sagittal imaging (Fig. 4). Quantitative analysis further revealed that the DBM-W group had a higher bone volume and trabecular number, but with no statistical significance. However, the DBM-W group had a significantly higher percentage of bone volume (p = 0.015) than the DBM-H group (Table 1).

Histological results

Under low magnification, decalcified histology found bony tissue, which was clearly bounded and surrounded by pre-existing abdominal muscle in both of the groups (Fig. 5). Under higher magnification, the lamellate tissue and lacunar space were observed to be arranged concentrically, and a haversian canal was observed in the center. These results indicated that the newly formed calcified tissue was ectopic bone, undergoing remodeling.

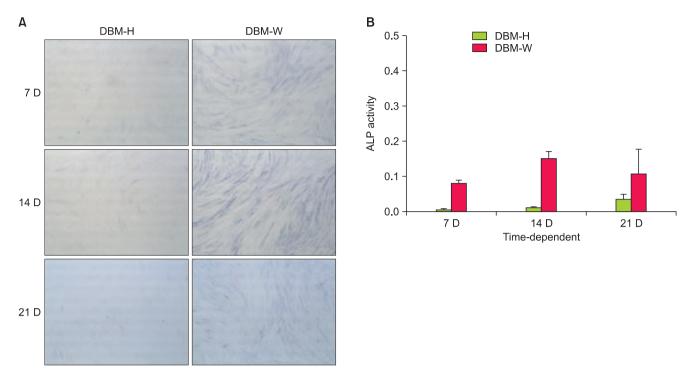


Fig. 2. Alkaline phosphatase (ALP) staining and ALP activity assay. (A) DBM-W group showed stronger ALP staining at 7, 14, and 21 days of treatment, although the staining faded in both groups at 21 days. (B) ALP activity was significantly higher in the DBM-W group at 7 and 14 days of treatment (ρ < 0.001 for DBM-W; ρ = 0.0003 for DBM-H). Although 21 days of treatment showed the same pattern, no statistical significance was found (ρ = 0.166). DBM: demineralized bone matrix, DBM-W: DBM with sterile water, DBM-H: DBM with poloxamer 407-based hydrogel, D: day.



Fig. 3. Anteroposterior abdominal radiographs obtained from DBM-W group (A) and DBM-H group (B) after 8 weeks of implantation. Easily identifiable radio-opaque lesions were not observed in either group. DBM: demineralized bone matrix, DBM-W: DBM with sterile water, DBM-H: DBM with poloxamer 407-based hydrogel.

DISCUSSION

DBM requires various types of carrier for handling during clinical application. The typical carriers are ceramics such as glycerol, hyaluronic acid, and calcium phosphate. ¹⁶⁾ The efficacy of DBM varies based on the level of growth factors such as BMP-2 and BMP-7, thus it is affected by the donor, manufacturing company, vendor, and lot. ^{17,18)} One of the reasons for the various efficacies of DBM obtained from different companies could be that each company uses their own carrier.

Both micro-CT and histology confirmed ectopic bone formation in the abdominal muscles of the nude rats from both of the groups. However, the quantitative analysis revealed that the level of newly formed bony tissue was relatively low compared to that obtained with the hydroxyapatite carrier which was used in our previous study. Done or plane radiography could not confirm the formation of bony tissue. Compared with our previous study using rhBMP-2 in the same ectopic bone formation model, while the rhBMP-2/type I collagen displayed definite ectopic bone formation in 100% of the rats, the two kinds of DBM in this study showed only tiny amounts of calcified tissue, not real ectopic bone formation. These



Fig. 4. Microfocus X-ray computed tomography of a specimen from the DBM-W group (A) and DBM-H group (B) after 8 weeks of implantation. For both A and B, each panel represents a transaxial image, sagittal image, and coronal image (from left to right). DBM: demineralized bone matrix, DBM-W: DBM with sterile water, DBM-H: DBM with poloxamer 407-based hydrogel.

data suggest that DBM alone may play a role in extending the bone graft, rather than inducing osteoinductivity.

Given that hydroxyapatite itself is radio-opaque, both DBM-W and DBM-H were considered to induce very weak osteoinductivity based on the ectopic bone formation confirmed through micro-CT and histology, even though the bone formation was not found by plane radiography. DBM facilitates bone healing through osteoconductivity and endochondral osteogenesis, which induces local cell transformation. For ectopic bone formation, the quantity and release of growth factors (BMP-2, epidermal growth factor, fibroblast growth factor, platelet-derived growth factor, vascular endothelial growth factor) is more essential than osteoconductivity.

DBM-W showed significantly higher ALP activity, although it contains just 8% more DBM than DBM-H. This implies that the poloxamer 407-based hydrogel itself has no toxicity, but it may inhibit the MSC osteoblastic differentiation by filling up the spaces between the DBM powders, which negatively affects the release of growth factors. Micro-CT of the specimens implanted for 8 weeks showed consistent results. Although there was no statisti-

Table 1. Microfocus X-ray Computed Tomography Results after 8 Weeks of Implantation*

Variable	DBM-W (n = 10)	DBM-H (n = 10)	<i>p</i> -value
Bone volume	0.35 ± 0.4	0.11 ± 0.06	0.079
Percent bone volume	2.9 ± 2.2	0.92 ± 0.50	0.015
Trabecular pattern factor	15.0 ± 8.4	18.0 ± 4.10	0.470
Structure model index	3.4 ± 0.94	3.8 ± 0.72	0.300
Trabecular thickness	0.26 ± 0.09	0.24 ± 0.04	0.590
Trabecular number	0.12 ± 0.15	0.04 ± 0.02	0.100
Trabecular separation	0.92 ± 0.23	1.04 ± 0.13	0.160
Degree of anisotropy	0.49 ± 0.15	0.53 ± 0.11	0.560

DBM: demineralized bone matrix, DBM-W: DBM with sterile water, DBM-H: DBM with poloxamer 407-based hydrogel.

*Showing the DBM-W group had significantly higher percent bone volume ratio (p = 0.015) than the DBM-H group.

cal significance, the DBM-W group had about 16% higher bone volume, the volumetric parameter of newly formed bony tissue. The DBM-W group also showed 40% higher trabecular number, a parameter describing the quality of the newly formed bony tissue, indicating the effects on the quality of bony tissue as well as quantity. Thus, this study implies that the efficient release of growth factors in DBM is more essential than the quantity of DBM for enhancing osteoblastic differentiation. Collectively, poloxamer 407-based hydrogel induced ectopic bone formation, which represents the efficacy of DBM, but showed lower osteoinductivity compared to sterile water.

The limitation of our study is that the two groups did not contain the same amount of DBM, which introduces the possibility that the results seen could be, at least in part, due to the amount of DBM delivered in the model, regardless of the carrier. However, the osteoinductivity of the DBM-W group was much higher than that of the DBM-H group, the difference of the amount of DBM does not alter the results.

In conclusion, the efficacy of both DBM-W and DBM-H was evaluated by confirming the ectopic bone formation in the two groups. The treatment with DBM-W showed significantly higher MSC osteoblastic differentiation than DBM-H. Both quality and quantity of the ectopic bone were also higher in the DBM-W group, indicating the negative role of the poloxamer 407-based hydrogel on the osteoinductivity of DBM.

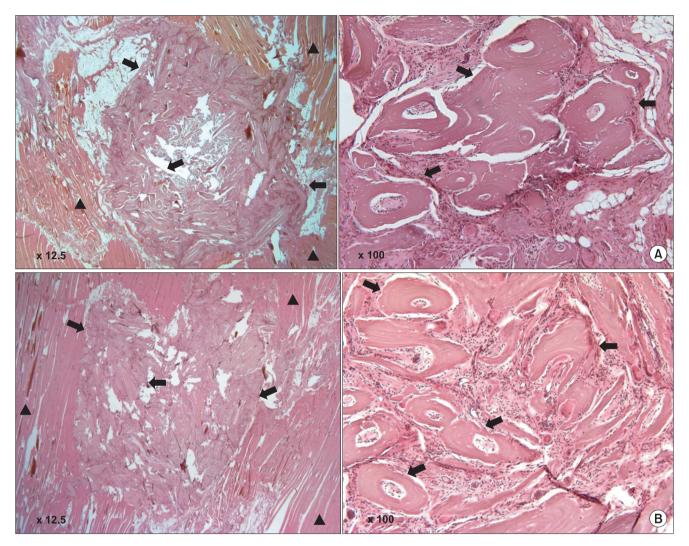


Fig. 5. Decalcified histology of the rats in the DBM-W group (A) and DBM-H group (B) after 8 weeks of implantation (H&E, × 12.5 and × 100). Under low magnification, both of the groups showed newly formed bone surrounded by abdominal muscle. Under higher magnification, the lamellate tissue and lacunar space were arranged concentrically, and a haversian canal was observed in the center, confirming bone remodeling. DBM: demineralized bone matrix, DBM-W: DBM with sterile water, DBM-H: DBM with poloxamer 407-based hydrogel, arrow heads: pre-existing abdominal muscle, arrows: newly formed ectopic calcified tissue.

CONFLICT OF INTEREST

No potential conflict of interest relevant to this article was reported.

ACKNOWLEDGEMENTS

This study was supported by a Grant-in-Aid (No. 03-2012-150) from the Seoul National University Hospital Research Fund.

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