Exosome: Function and Role in Cancer Metastasis and Drug Resistance

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Abstract

As a kind of nanometric lipidic vesicles, exosomes have been presumed to play a leading role in the regulation of tumor microenvironment through exosomes-mediated transfer of proteins and genetic materials. Tumor-derived exosomes are recognized as a critical determinant of the tumor progression. Intriguingly, some current observations have identified that exosomes are essential for several intercellular exchanges of proteins, messenger RNAs, noncoding RNAs (including long noncoding RNAs and microRNAs) as well as to the process of cancer metastasis and drug resistance. Herein, we review the role of exosomes and their molecular cargos in cancer invasion and metastasis, summarize how they interact with antitumor agents, and highlight their translational implications.

Keywords

Exosomes, contents, metastasis, drug resistance, cancer

Abbreviations

CRC, colorectal cancer; CRNDE-h, colorectal neoplasia differentially expressed-h; ECM, extracellular matrix; EMT, epithelialmesenchymal transition; HCC, hepatocellular carcinoma; ITG, integrins; IncRNAs, long noncoding RNAs; MET, mesenchymalepithelial transition; miRNAs, microRNAs; mRNAs, messenger RNAs; MVBs, multivesicular bodies; ncRNAs, noncoding RNAs; PCR, polymerase chain reaction; SR, serine/arginine-rich; TEM, transmission electron microscope; TGF- β , transforming growth factor β ; UCA1, urothelial cancer associated-1

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Introduction

Exosomes are spherical bilayered membrane vesicles with an average diameter of 30 to 100 nm and "saucer-shape" morphology.¹ They are the only type of extracellular vesicles formed from endosomal compartment invaginations, which are called multivesicular bodies (MVBs) and are released in the endosomal network.²⁻⁴ After the fusion of MVBs with plasma membrane, the internal contents are released into the extracellular space as the form of "exosomes" (Figure 1). Exosomes can be found in a range of fluids, including blood, plasma, saliva, urine, synovial fluid, amniotic fluid, malignant ascite, and pleural effusions.⁵⁻⁹ The biogenesis mechanisms of exosomes have not been fully elucidated, but they are secreted from nearly all cell types including normal cells and diseased cells.¹⁰⁻¹² In particular, tumor cells possess more exosomes-releasing properties when compared to normal cells.¹³

Exosomes from different cell phenotypes and body fluids contain various bioactive molecules such as proteins (including oncoproteins, tumor suppressor proteins, and transcriptional

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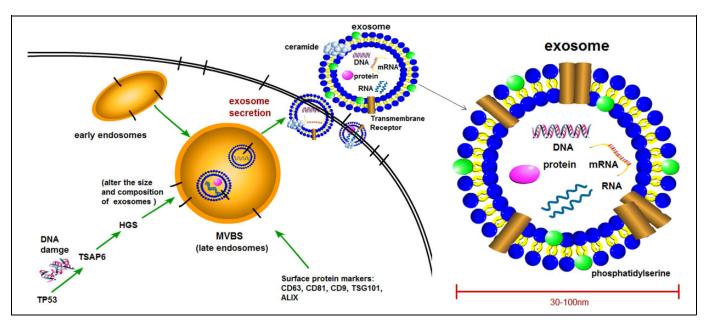


Figure 1. Generation and the structure of exosome. Exosomes are vesicles with a phospholipid bilayer membrane; they are considered to be secreted from intracellular multivesicular bodies (MVBs or late endosomes) into extracellular space. The exosomes contain a range of proteins, RNAs, mRNAs, and DNA molecular cargoes, with surface protein markers including CD9, CD63, CD81, TSG101, and ALIX, and there are equal or asymmetrical distributions of phospholipids between the 2 leaflets of exosome membrane. mRNAs indicates messenger RNAs; TBS, Tris-buffered saline; TSG101, tumor-susceptibility gene-101; ALG-2-interacting protein X (ALIX).

regulators), lipids (including phosphatidylcholines, phosphatidylethanolamines, and phosphatidylserines),¹⁴⁻¹⁶ DNA (including single-strand DNA, genomic DNA, XX DNA, and retrotransposon elements),¹⁷⁻²⁰ and RNAs (including messenger RNAs [mRNAs], long noncoding RNAs [lncRNAs], microRNAs [miRNAs], and other non-coding RNAs [ncRNAs]).²¹⁻²³ Circulating exosomes are complex molecular assemblies of these bioactive molecules, and these biomolecules permit horizontal transfer of oncogenic traits from the primary tumor to recipient target cells located in distant organs. Pioneering works call this phenomenon "genometastasis"²⁴ as an implication of the intercellular trafficking of oncogenic macromolecules via exosomes to explain cancer metastasis.²⁵ Since Abdouh et al²⁶ confirmed the validity of the genometastatic theory in human cells for the first time, recent studies provide more evidences to support this idea, and experimental data demonstrate the role of circulating factors carried by exosomes in conferring mesenchymal-epithelial transition (MET),²⁷ mediating oncogenic transformation²⁸ and contributing to cancer metastasis.

Although it is possible that horizontal transformation may require preexisting alterations in recipient cells,²⁹ Abdouh *et al*³⁰ also reported that human cells carrying a single oncosuppressor mutation would show an increased uptake of exosomes when exposed to blood-circulating cancer factors and were capable of integrating cancer factors at metastatic sites. This further demonstrates that oncogenic factors transferred via circulating cancer exosomes induce malignant transformation of target cells even at distant distance.

The constituent information about the exosomal molecular cargos can be obtained in public access databases, such as ExoCarta³¹ and EVPedia.³² Under the stimulation of external extreme conditions such as oxidative stress and serum starvation, most cells can produce exosomes containing altered RNA contents.^{33,34}

In addition, mutated oncogenes may also alter exosome secretion.^{1,35,36} Tumor cells release exosomes into the circulation, leading to significant increase of exosomes in patients with cancer compared to the healthy controls,¹³ and the abnormal exosomes may be associated with decreased overall survival.³⁷ Tumor-secreted exosomes can even influence organotropism. It is proved that tumor-derived exosomal molecules guide exosomes to specific organs and promote organ-specific metastasis because exosomes from different cancer recapitulate the organ specificity of their cell of origin and prepare premetastatic niches as well.^{38,39}

There are several approaches for the isolation of exosomes: Ultracentrifugation^{1,40-42} and commercial kits (ExoQuick kit, System Biosciences, USA; Total Exosome Isolation Reagent, Invitrogen, USA; Qiagen miRNeasy Kit, Qiagen, Germany) are usually used. Generally, traditional ultracentrifugation-based approach can provide cleaner exosomes, but this method is labor intensive and time consuming and in need of expensive laboratory equipment and larger sample volumes. For the commercial kit approach, the acquired exosomes contain additional molecular complexes that are unnecessary. Therefore, an improved isolation technology is required to obtain exosomes effectively.³⁹

In this article, we review the role of exosomes and its molecular cargos in tumor invasion and metastasis, summarize how they interact with antitumor drugs, and show the potential of clinical application.

Exosomes in Tumor Metastasis and Drug Resistance

Exosomes were considered "garbage bag" of unnecessary cellular materials when they were first discovered.^{11,43} However, many important biological functions of exosomes have been convincingly demonstrated in the recent years. They are involved in intercellular communication, immune system modulation, cell growth enrichment, energy pathways maintenance, and propagation of viruses via releasing intracellular cargoes (eg, proteins, lipids, DNA, and RNAs including tumor-derived ncRNAs).^{40,44-46} One type of the exosomes, tumor-derived exosomes, have been found to apparently facilitate tumor growth and metastasis.⁴⁷ Rather than simple cellular debris, tumor-derived exosomes act as extracellular organelles with roles in remodeling tumor microenvironment by delivering messages for cell-cell communication.⁴⁸⁻⁵¹ It is a well-built means of intercellular exchange via exosomes, and especially, the exosomes transport the nucleic acids and proteins from tumor cells to neighboring cells in tumor microenvironments.52

First, tumor-derived exosomes induce proinflammatory phenomena. They interact with their proteins and miRNAs to result in an inflammatory behavior, and this is essential for recruiting inflammatory CCR6⁺CD4⁺ Th17⁺ into specific tumor sites⁵³ to contribute to proliferation, angiogenesis, and metastasis of malignant cells in tumor microenvironment. 54,55 For instance, when taken up by resident cells, tumor-derived exosomal integrins (ITG) increase proinflammatory S100 gene expression.³⁸ Second, tumor-derived exosomes mediate vascular leakiness as a key feature of premetastatic niche formation.⁵⁶ Similar to cytokines, tumor-derived exosomes can recruit bone marrow-derived cells to premetastatic tumor tissue, thus contributing to creating a permissive microenvironment for tumor metastases.⁵⁷ In this way, they could remodel the extracellular matrix (ECM) to support tumor growth and a prometastatic phenotype.^{58,59} Third, tumor-derived exosomes are involved in immune response. They suppress the activation of effector T-cells, trigger the apoptosis of activated T-cells, and help metastatic cells to escape tumor immune surveillance.^{60,61} Fourth, tumor-derived exosomes promote tumor progression via regulating drug resistance: (1) tumor cells can utilize the exosome secretion to extrude anticancer drugs; (2) exosomal molecular cargoes can compete with anticancer drugs, especially antibody-based drugs, to bind with oncogenic targets, thus lowering their therapeutic effect^{62,63}; and (3) exosomes contribute to the lateral transfer of drug resistance from drug-resistant cells to drug-sensitive cells.64

Exosomes and Tumor Metastasis

Exosomes can mediate important cancer-related pathways to enhance tumor invasion and metastasis through different manners, including controlling angiogenesis,⁶⁵⁻⁶⁹ modulating stromal cells,⁷⁰⁻⁷² remodeling the ECM,⁷³ transferring malignant

traits,³⁰ and establishing the premetastatic niche.⁷⁴⁻⁷⁷ Exosomes containing ncRNAs and proteins play significant roles in these pathways (Table 1 and Figure 2). However, the biological function of exosomal ncRNAs and proteins in tumor invasion and metastasis remains unclear.

Association between exosomal proteins and tumor metastasis. Exosomes contain numerous proteins, which may be associated with the growth and survival of metastatic tumor cells. Chen et al⁴⁰ revealed some upregulated exosomal proteins from patients with colorectal cancer (CRC) are involved in the modulation of pretumorigenic microenvironment for metastasis and invasiveness. These exosomal proteins, such as fibronectin-1, mediate cytoskeletal organization and actin dynamics to enable cell adhesion and motility.⁸⁰ Matrix metalloproteinase-9 (MMP9), a protein found to be overexpressed in exosomes of patients with CRC, plays a key role in ECM degradation via the activation of transforming growth factor β (TGF- β)/Smad signaling pathway.⁸¹ Additionally, some exosomal protein markers could be used as predictive markers for organ-specific metastasis. For example, epidermal growth factor (EGF)-like repeats and discoidin I-like domain 3 protein from urinary exosomes of patients with bladder cancer can be used for predicting muscle metastasis.^{78,79} Furthermore, surface proteins of exosomes are also involved in metastatic tropism. Hoshino et al revealed a close correlation between exosomal ITG and metastatic tropism.³⁸ They found that exosomal ITG could regulate local microenvironments within future metastatic organs. They also reveal that the expression of exosomal ITGa6 and its partners ITGB4 and ITGB1 was associated with lung metastasis: $ITG\alpha_v$ and ITGB5 were linked to liver metastasis, while ITGβ3 underlied organotropism to the brain.³⁸ Recent studies show that the metastatic tumor cells-derived exosomes are enriched with proteins, such as S100A8 and S100A9.99 Exosomal S100A8/A9 proteins can be upregulated from tumor cells with high activity of Wnt/ β -catenin pathway, and these proteins are able to recruit leukocytes and probably indirectly involved in the formation of premetastatic niches in distal metastatic organs.82

Association between exosomal miRNAs and tumor metastasis. Micro-RNAs, a large class of ncRNAs, play pivotal roles in various cancers through the repression of downstream cancerassociated mRNAs.¹⁰⁰ Since miRNAs are detected in serum from patients with diffuse large B-cell lymphoma first,¹⁰¹ different exosomal miRNAs have been reported in distinct types of malignancies, including tongue cancer,¹⁰² breast cancer,¹⁰³ lung cancer,¹⁰⁴ ovarian cancer,¹⁰⁵ prostate cancer,¹⁰⁶ CRC,^{107,108} and gastric cancer.⁸³ These findings support that these miRNAs are potential biomarkers for cancer diagnosis and prognosis.¹⁰⁹ There are abundant miRNAs in the exosomes. Notably, the expression and distribution exosomal miRNAs are often dysregulated in cancerous tissue. Cancerderived exosomes transfer miRNAs into target cell cytoplasm and then result in downregulation of expression of specific target genes, thus affecting cell function. This process is

	Tumor Types	Oncogene/ Suppressor	Functions in Tumors	Reference
Exosomal proteins				
EDIL-3	Urinary bladder cancer	Oncogene	High-grade muscle invasive	Beckham <i>et al</i> ⁷⁸ Street <i>et al</i> ⁷⁹
FN1	Colorectal cancer	Oncogene	Tumor progression and metastasis; ECM degradation; EMT	Akakura et al ⁸⁰
MMP9	Colorectal cancer	Oncogene	ECM degradation; EMT by TGF-β/Smad signaling pathway	Lampropoulos et al ⁸
S100A8 and S100A9 H-ras	Colorectal cancer Intestinal	Oncogene Oncogene	Involves in distal metastasis by Wnt/β-catenin pathway Exhibit EV uptake and transforming effects	Ichikawa <i>et al⁸²</i> Lee <i>et al²⁹</i>
ITGα6, ITGβ4, and ITGβ1	epithelial cells Breast cancer	Oncogene	Lung metastasis	Hoshino et al ³⁸
ITG α_v and ITG β 5	Pancreatic cancer	Oncogene	Liver metastasis	Hoshino et al ³⁸
ITGβ3 Exosomal miRNA	Breast cancer	Oncogene	Brain metastasis	Hoshino et al ³⁸
let-7 miRNA family	Gastric cancer	Oncogene	Promote metastasis	Ohshima et al ⁸³
miR-31	Breast cancer	Tumor suppressor	Opposing local invasion and metastatic colonization	Ragusa et al ⁴¹
miR-105	Breast cancer	Oncogene	Promote metastasis	Zhou et al ⁸⁴
miR-130	Breast cancer	Oncogene	Enhances cell proliferation and migration	Ragusa <i>et al⁴¹</i> Kruger <i>et al⁸⁵</i>
miR-328	Breast cancer	Oncogene	Promote metastasis	Kruger et al ⁸⁵
miR-150	Monocytic	Oncogene	Cell migration	Zhang et al ⁸⁶
miR-210	Colorectal cancer	Oncogene	Promote EMT, metastasis	Bigagli et al ⁸⁷
miR-200c, miR-141	Colorectal cancer	Tumor suppressor	Cell proliferation, invasion, migration, EMT	Tanaka <i>et al</i> ⁸⁸
miR-125b, miR-130b, miR-155	Prostate cancer	Oncogene	Triggering neoplastic transformation and MET	Abd Elmageed <i>et al</i> ²
Exosomal IncRNA				
MALAT1	Liver cancer	Oncogene	Regulate alternate splicing, cancer metastasis, and tumor recurrence	Tripathi <i>et al</i> ⁸⁹ Yang <i>et al</i> ⁹⁰
ATB	Liver cancer	Oncogene	Mediate TGF-β signaling pathway, promote the invasion-metastasis cascade	Yuan <i>et al</i> ⁹¹
CRNDE-h	Colorectal cancer	Oncogene	Reduce the sensitivity to the cytostatic effect of TGF- β	Liu et al ⁹²
IncRNA-TUC339	Hepatic cancer	Oncogene	Promote the growth and spread	Kogure <i>et al</i> ⁹³
lncRNA-H19	Liver cancer	Oncogene	EMT, hepatic metastases	Conigliaro et al ⁹⁴
lncRNA-HOTAIR	Urothelial bladder cancer		Tumor migration and invasion	Berrondo <i>et al</i> ⁹⁵ Yan <i>et al</i> ⁹⁶
lncRNA-p21	Multiple cancers	Oncogene	Affect global gene expression	Gezer <i>et al</i> ⁹⁷ Dimitrova <i>et al</i> ⁹⁸

Table 1. Functions of Exosomal Noncoding RNAs and Proteins in Tumor Metastasis and Invasion.

Abbreviations: EDIL-3, epidermal growth factor-like repeats and discoidin I-like domain 3; FN1, fibronectin-1; ECM, extracellular matrix; EMT, epithelialmesenchymal transition; MMP9, matrix metalloproteinase-9; TGF- β , transforming growth factor β ; ITG, integrins; miRNA, microRNA; MET, mesenchymalepithelial transition; lncRNA, long noncoding RNA; MALAT1, metastasis-associated lung adenocarcinoma transcript-1; CRNDE-h, colorectal neoplasia differentially expressed-h.

possibly modulated by important signaling pathways in recipient cells, such as Wnt, Ras, TGF- β , and p53 signaling pathways.^{110,111} Intriguingly, packaging of miRNAs into exosomes is not stochastic, miRNAs are preferentially selected into CD63-positive exosomes via a ceramidedependent pathway, and some oncogenic signaling, such as Kirsten rat sarcoma viral oncogene homolog, may result in selective packaging of miRNA into exosomes, thus generating different cancer-specific exosome profiles.^{112,113}

Among those exosomal miRNAs mentioned earlier, some miRNAs are closely associated with tumor invasion and

metastasis. For example, exosomal miR-31 from breast cancer cells influences metastasis by suppressing local invasion and metastatic colonization, and it is an onco-miR whose overexpression enhances cancer cell proliferation and migration.⁴¹ Similarly, miR-130a and miR-328 are observed to be upregulated in exosomes derived from metastatic MDA-MB-231 cells.⁸⁵ Moreover, the let-7 miRNA family, including let-7a, let-7b, let-7c, let-7d, let-7e, and let-7i, is packaged into exosomes from metastatic AZ-P7a gastric cancer cell to maintain their oncogenesis and invasiveness.⁸³ Thus, this miRNA family has a vital role in the delivery of oncogenic signals to promote

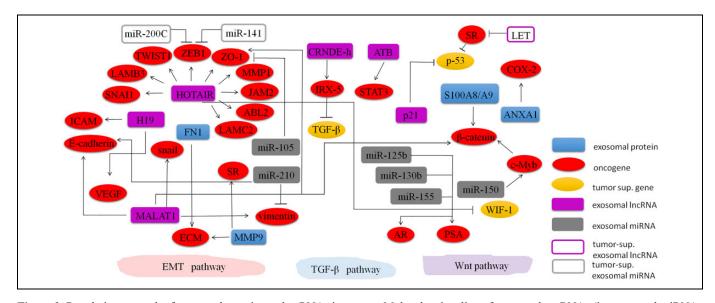


Figure 2. Regulating network of exosomal proteins and ncRNAs in tumors. Molecular signaling of exosomal nc-RNAs (ie, exosomal miRNAs [exo-miRNAs] and exosomal lncRNAs [exo-lncRNAs]) and proteins in most frequent cancer-related pathways involved in cancer metastasis (ie, EMT, TGF- β , and Wnt). These bioactive molecules carried in exosomes take effects in metastasis promotion or repression and interact with others. EMT pathway is a process by which epithelial cells lose their cell–cell adhesion and acquire migratory property to become mesenchymal stem cells; TGF- β pathway is involved in cellular processes including cell growth, differentiation, and invasion; and Wnt pathway regulates crucial aspects of cell fate determination and cell migration during embryonic development. ncRNAs indicates noncoding RNAs; miRNA, microRNA; lncRNAs, long noncoding RNAs; EMT, epithelial–mesenchymal transition; TGF- β , transforming growth factor β .

metastasis. Le et al¹¹⁴ clearly demonstrated that exosomes containing miR-200 could transfer the metastatic capability between metastatic and nonmetastatic cancer cells. Bigagli and her group also found that exosomes containing miR-210 might be considered as epithelial-mesenchymal transition (EMT) promoting signal that guides metastatic cells to free new sites of dissemination with low level of vimentin and high level of E-cadherin.⁸⁷ Zhang et al⁸⁶ showed that exosomes from the human monocytic cells human acute monocytic leukemia cells (THP-1) with a high level of miR-150 could deliver miR-150 into the human endothelial cells human microvascular endothelial cells (HMEC-1) and then affect MYB proto-oncogene (c-Myb) expression and cell migration. Likewise, exosomes derived from MDA-MB-231 cells and Michigan Cancer Foundation-10 (MCF-10) cells harbor plentiful miR-105, which was responsible for promoting metastasis by suppressing the expression of tight junction protein zonula occludens 1 (ZO-1).⁸⁴ Abd Elmageed et al²⁷ revealed a new role for prostate cancer (PC) cell-derived exosomes in triggering neoplastic transformation and MET through trafficking of oncogenic factors (including onco-miRNAs miR-125b, miR-130b, and miR-155) into adipose-derived stem cells of patients with PC. In contrast, some exosomal miRNAs are negatively correlated with invasion ability. Shota et al⁸⁸ observed an increased expression of exosomal miR-200c and miR-141 in SW620 and SW620/OxR CRC cells after decitabine treatment, accompanied by the acquisition of epithelial cell-like characteristics. So exosomal miR-200c and miR-141 may be an indicator for MET and potentially associated with metastasis inhibition.

Association between exosomal IncRNAs and tumor metastasis. The lncRNAs from exosomes may serve as signaling molecules in various biological processes mediating intercellular communication. High levels of exosomal lncRNAs not only provide a novel diagnostic method for early prediction but also show predictive potential for tumor invasion and metastasis.115,116 Notably, quantitative comparisons prove that exosomes lack mRNAs but contain abundant lncRNAs.^{117,118} A subsequent attempt points out that lncRNAs occupy 3.36% of the total exosomal RNA content.¹¹⁹ Liu et al⁹² also indicated that exosomal colorectal neoplasia differentially expressed-h (CRNDE-h) levels were associated with distant metastasis and lymph node metastasis, accompanied by high expression of iroquois homeobox protein-5 in the metastatic sites.¹²⁰ Besides, this exosomal lncRNA decreased the sensitivity to the cytostatic effect of TGF- β and granted a growth performance to tumor cells in TGF-B/SMADS pathway.¹²⁰ Kogure and colleagues⁹⁷ suggested that the exosome-mediated transfer of IncRNA-TUC339 could improve the growth of cancer cells and facilitate the spread of hepatocellular carcinoma (HCC). Through the secretion of exosomes containing lncRNA-H19, CD90+ liver cancer cells can modulate endothelial-mesenchymal phenotype and involve in hepatic metastases by inducing an increase in vascular EGF and intercellular adhesion molecule-1 transcripts,121 thus influencing tumor microenvironment in a prometastatic way.¹²² Through the release of exosomes containing metastasis-associated lung adenocarcinoma transcript-1 (MALAT1), liver cancer cells can modify RNA alternate splicing by adjusting the levels of phosphorylated to dephosphorylated serine/arginine-rich (SR) proteins.⁸⁹ This

	Tumor Types	Function in Drug Resistance	Drug	Reference
Exosomal miRNA miR-433	Ovarian cancer	Promote	Paclitaxel	Weiner-Gorze et al ¹²⁶
Exosomal lncRNA lncRNA-ROR Hepatocellular cancer	Promote	Chemotherapeutic drugs	Takahashi <i>et al</i> ¹²⁷ Nawaz <i>et al</i> ⁹⁸	
IncRNA UCA1	Breast cancer	Promote	Tamoxifen	Xu et al^{96}
IncRNA UCA1	Bladder cancer	Promote	Chemotherapeutic drugs	Fan <i>et al</i> ¹²⁸
HER2	Breast cancer	Promote	Trastuzumab	Ciravolo et al ⁶²
lncARSR	Renal cancer	Promote	Sunitinib	Qu et al ¹²⁹

Table 2. Functions of Exosomal ncRNAs in Tumor Drug Resistance.

Abbreviations: ncRNAs, noncoding RNAs; miRNA, microRNA; lncRNA, long noncoding RNAs; UCA1, urothelial cancer associated-1; HER2, human epidermal growth factor-2; lncARSR.

process is connected with metastasis and recurrence of liver cancer as well as a diverse range of cancers.⁹⁰ It has also been suggested that MALAT1 could alter the protein expression of E-cadherin, ZO-1, β -catenin, vimentin, and snail, which are involved in EMT.⁹³

Through the release of exosomes containing lncRNA-ATB, the TGF- β signaling pathway can be modulated. In detail, IncRNA-ATB binds to interleukin 11 and triggers signal transducer and activator of transcription 3 (STAT3) signaling pathway to promote the invasion-metastasis cascade.⁹¹ Analogously, exosomal lncRNA-p21 has been reported to regulate mRNA translation and to suppress the p53 to affect global gene expression by binding to the heterogeneous nuclear ribonucleoprotein K (hnRNP-K) complex.94,117,123 Besides, exosomal lncRNA-HOTAIR in urothelial bladder cancer cells can regulate the expression of EMT genes, including SNAI1, LAMB3, TWIST1, MMP1, ZEB1, ZO-1, JAM2, ABL2, and LAMC2, thus inducing tumor migration and invasion.¹²⁴ Yan et al¹²⁵ also showed that HOTAIR was involved in migration and invasion through inhibiting the canonical Wnt pathway antagonist protein WIF-1 in vitro.

However, some other exosomal lncRNAs show a function to inhibit tumor metastasis. The lncRNA-LET binds to and destabilizes nuclear factor of activated T-cells 90 kDa (NF90),¹²⁶ thus suppressing cell growth and metastasis in HCC.¹²⁷ With these observations, exosomal lncRNAs may be useful for diagnosis and prevention of metastasis. Exosomal miRNAs have been extensively studied in various cancers,⁹⁸ whereas exosomal lncRNAs are being pursued for similar goals. Although the specific association of exosomes with cancer carcinogenesis and metastasis remains unclear, their stable property makes them to be widely used in cancer research. It is reported that exosomes remain stable even at a temperature range from 4°C to 42°C.⁹⁵

Exosomes and Drug Resistance

Exosomes-mediated drug resistance. Drug resistance is a major challenge for cancer therapy. Several studies reveal that exosomes are involved in the modulation of chemosensitivity by transferring the resistant phenotype to recipient cells.⁶⁴ Transport of ncRNAs, including miRNAs and lncRNAs, mediated by

exosomes, is believed to be an effective mechanism for acquiring drug resistance in cancer cells (Table 2). For example, in ovarian cancer, exosomal transfer of miR-433 can promote paclitaxel resistance through the induction of cellular senescence.¹²⁶ In HCC, exosomal transfer of lncRNA-ROR can increase TGF-B-dependent chemoresistance.^{127,98} In breast cancer, exosomal transfer of lncRNA urothelial cancer associated-1 (UCA1) can increase tamoxifen resistance in estrogen receptor-positive MCF-7 cells through mTOR signaling pathway.⁹⁶ Exosomal transfer of lncRNA UCA1 can also increase chemoresistance of bladder cancer cells via activating the Wnt signaling pathway.¹²⁸ Exosomes expressing fulllength human epidermal growth factor 2 molecules enable them to bind and sequester Trastuzumab, thus lowering the therapeutic effectiveness of Trastuzumab in breast cancer.⁶² In renal cancer, exosome-transmitted lncRNA Activated in RCC with Sunitinib Resistance (IncARSR) can disseminate Sunitinib resistance by acting as a competing endogenous RNA for miR-34 and miR-449, thus facilitating expression of receptor tyrosine kinase (AXL) and c-MET.¹²⁹ Exosomes are speculated as a novel target for cancer therapy because they can promote angiogenic phenotype and cell-to-cell adhesion in target cells.130

A study suggests that the acid microenvironment of the tumor–host interface may stimulate the output of exosomes,¹³¹ and such exosomes could release several contents that transport a resistance phenotype to sensitive tumor cells by changing cell cycle control and stimulating antiapoptosis programs.¹³² It has also been proposed that EMT inducers can increase resistance to chemotherapy.¹³³ Bigagli *et al*¹¹⁴ proved the chemosensitivity of metastatic cells undergoing EMT was diminished compared to adherent HCT-8 colon cancer cells, since exosomes secreted from HCT-8 cells might impact the EMT program and made the metastatic cells spontaneously insensitive to chemotherapeutic strategies. There is also a deduction that vesicles such as exosomes are able to move between cells using specific structures called nanotubes.¹³³ However, it is not clear which of these theories may be correct.

Therapeutic potential of exosomes in drug resistance of cancer. Treatment with locked nucleic acids targeting exosomal IncRNA and miRNAs can restore drug sensitivity. Through the untiring efforts of researchers, the lncRNAs and miRNAs are considered as putative targets for cancer therapy via exosomemediated mechanism, such as lncRNA-H19 and lnc-ARSR9.^{121,134} Exosomal ncRNAs-based therapeutic protocols, such as antisense oligonucleotides, have been proved to be capable of restraining pathological lncRNAs. Therefore, these exosomal ncRNAs are inspiring and appealing for cancer therapy. In addition, there is a speculation that exosomes can deliver drugs to selective targets as a novel platform in a drug delivery system. In particular, exosomes are capable of crossing the blood-brain barrier without inducing an immune response.¹³⁵ However, a set of troubles have been suffered during the clinical development of exosomes-based therapeutic technologies for cancers, including their indefinite functions in the complex networks, the difficulty in accurate quantification of exosomal lncRNAs and miRNAs, the hard transport of several lncRNAs and miRNAs antagonists or mimics as well as the unclear clinical pharmacokinetics and drug toxicity.³⁹ Therefore, more studies are needed to verify the potential of applying exosomes and their contents in cancer therapy. Exosomesbased drugs are expected to enter clinical trials, and their great potential may be revealed.

Prospects and Challenges

From the above, we conclude that tumor-derived exosomes not only discard cellular waste but also trigger signaling pathways and drug resistance in target cells and facilitate the growth of metastatic cells. It is encouraging that nanomaterials or microbodies, such as exosomes-mediated delivery system, provide more possibility. Exosomes containing MAGE family member A3 (MAGE-3) peptides show minimal toxic effects after treatment in patients with stages III/IV melanoma.¹³⁵ However, small RNA-based drugs, such as miRNAs and circRNAs, face different kinds of challenges due to the lack of targeted delivery strategies in cancer-targeted therapy.¹³⁶

Switching to a wider scientific horizon, exosomes are still on initial exploration at present, and only a few molecular mechanisms about exosomes in cancer progression are discovered. In addition, the strategies and methods used for investigating exosomes are still limited and challenging. The identification methods of exosomes are diverse; for instance, the size of pelleted particles can be measured using dynamic light scattering such as Brookhaven instruments BI-9000 digital correlator (Brookhaven instruments, USA), Zetasizer Nano ZS (Malvern, UK), LM10 NanoSight (Malvern, UK), or BI200-SM goniometer (Brookhaven instruments, USA), configured with a solid-state laser tuned at 532 nm.^{122,124} The morphology of exosomes is suitable to be examined under a transmission electron microscope (TEM) at 80 keV, with a diameter of approximately 30 to 100 nm under the TEM ultimately. The typical electron micrographs can be captured by a megapixel digital camera, such as Erlangshen 11 digital camera (Gatan, USA).¹²⁴ Exosomes can also be identified by Western blot analysis with exosomal surface protein markers antibodies

such as anti-CD9, anti-CD63, anti-CD81, anti-CD82, and so on.¹³⁷ However, from the reported studies, we can see that the identification of exosomes is not coherent and comprehensive, which cannot avoid the occurrence of false-positive or false-negative results.

Currently, the large-scale profiles of miRNAs or lncRNAs in exosomes are usually obtained using microarrays of nextgeneration sequencing followed by the extraction of exosomes, which can be customized and time reduced.¹³⁸ Besides, quantitative polymerase chain reaction (PCR), digital PCR, and NanoString nCounter Gene Expression Assay (NanoString nCounter system, USA) can be used for detection of miRNAs or lncRNAs, with features of expensive and uneffective but high sensitivity.¹³⁹ Exosome protein can be isolated by optimized lysis buffer¹⁴⁰ and quantitated by a microBCA Protein Assay kit (Pierce, USA). When the focus is on the number of total exosomal proteins in patients with cancer, a quantitative proteomic analysis can be used.¹⁴¹

In conclusion, more details regarding the functional contents of exosomes involved in cancer progression is highly desired, and exploring their biological mechanism will improve the clinical applicability of exosomes in cancer diagnosis and therapy.

Authors' Note

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References

- Sun Y, Zheng W, Guo Z, et al. A novel TP53 pathway influences the HGS-mediated exosome formation in colorectal cancer. *Sci Rep.* 2016;6:28083. doi:10.1038/srep28083
- Hurley JH, Odorizzi G. Get on the exosome bus with ALIX. Nat Cell Biol. 2012;14(7):654-655. doi:10.1038/ncb2530
- Urbanelli L, Magini A, Buratta S, et al. Signaling pathways in exosomes biogenesis, secretion and fate. *Genes*. 2013;4(2): 152-170. doi:10.3390/genes4020152
- Yanez-Mo M, Siljander PR, Andreu Z, et al. Biological properties of extracellular vesicles and their physiological functions. *J Extracell Vesicles*. 2015;4:27066. doi:10.3402/jev.v4.27066

- Gonzalez-Begne M, Lu B, Han X, et al. Proteomic analysis of human parotid gland exosomes by multidimensional protein identification technology (MudPIT). *J Proteome Res.* 2009;8(3): 1304-1314. doi:10.1021/pr800658c
- Garcia JM, Garcia V, Pena C, et al. Extracellular plasma RNA from colon cancer patients is confined in a vesicle-like structure and is mRNA-enriched. *RNA*. 2008;14(7):1424-1432. doi:10. 1261/rna.755908
- Keller S, Rupp C, Stoeck A, et al. CD24 is a marker of exosomes secreted into urine and amniotic fluid. *Kidney Int.* 2007;72(9): 1095-1102. doi:10.1038/sj.ki.5002486
- Dear JW, Street JM, Bailey MA. Urinary exosomes: a reservoir for biomarker discovery and potential mediators of intrarenal signalling. *Proteomics*. 2013;13(10-11):1572-1580. doi:10.1002/ pmic.201200285
- Gallo A, Tandon M, Alevizos I, Illei GG. The majority of micro-RNAs detectable in serum and saliva is concentrated in exosomes. *PLoS One*. 2012;7(3):e30679. doi:10.1371/journal.pone.0030679
- Yang C, Robbins PD. The roles of tumor-derived exosomes in cancer pathogenesis. *Clin Dev Immunol*. 2011;2011:842849. doi: 10.1155/2011/842849
- Raposo G, Stoorvogel W. Extracellular vesicles: exosomes, microvesicles, and friends. J Cell Biol. 2013;200(4):373-383. doi:10.1083/jcb.201211138
- Pant S, Hilton H, Burczynski ME. The multifaceted exosome: biogenesis, role in normal and aberrant cellular function, and frontiers for pharmacological and biomarker opportunities. *Biochem Pharmacol.* 2012;83(11):1484-1494. doi:10.1016/j.bcp. 2011.12.037
- Taylor DD, Gercel-Taylor C. MicroRNA signatures of tumorderived exosomes as diagnostic biomarkers of ovarian cancer. *Gynecol Oncol.* 2008;110(1):13-21. doi:10.1016/j.ygyno.2008. 04.033
- Subra C, Laulagnier K, Perret B, Record M. Exosome lipidomics unravels lipid sorting at the level of multivesicular bodies. *Biochimie*. 2007;89(2):205-212. doi:10.1016/j.biochi.2006.10.014
- Record M, Carayon K, Poirot M, Silvente-Poirot S. Exosomes as new vesicular lipid transporters involved in cell–cell communication and various pathophysiologies. *Biochim Biophys Acta*. 2014;1841(1):108-120. doi:10.1016/j.bbalip.2013.10.004
- Kahlert C, Melo SA, Protopopov A, et al. Identification of doublestranded genomic DNA spanning all chromosomes with mutated KRAS and p53 DNA in the serum exosomes of patients with pancreatic cancer. *J Biol Chem.* 2014;289(7):3869-3875. doi:10. 1074/jbc.C113.532267
- Balaj L, Lessard R, Dai L, et al. Tumour microvesicles contain retrotransposon elements and amplified oncogene sequences. *Nat Commun.* 2011;2:180. doi:10.1038/ncomms1180
- Cai J, Han Y, Ren H, et al. Extracellular vesicle-mediated transfer of donor genomic DNA to recipient cells is a novel mechanism for genetic influence between cells. J Mol Cell Biol. 2013;5(4): 227-238. doi:10.1093/jmcb/mjt011
- Burger D, Schock S, Thompson CS, Montezano AC, Hakim AM, Touyz RM. Microparticles: biomarkers and beyond. *Clin Sci* (*Lond*). 2013;124(7):423-441. doi:10.1042/cs20120309

- Thakur BK, Zhang H, Becker A, et al. Double-stranded DNA in exosomes: a novel biomarker in cancer detection. *Cell Res.* 2014; 24(6):766-769. doi:10.1038/cr.2014.44
- Mittelbrunn M, Gutierrez-Vazquez C, Villarroya-Beltri C, et al. Unidirectional transfer of microRNA-loaded exosomes from T cells to antigen-presenting cells. *Nat Commun.* 2011;2:282. doi: 10.1038/ncomms1285
- Record M, Subra C, Silvente-Poirot S, Poirot M. Exosomes as intercellular signalosomes and pharmacological effectors. *Biochem Pharmacol.* 2011;81(10):1171-1182. doi:10.1016/j.bcp. 2011.02.011
- Raimondo F, Morosi L, Chinello C, Magni F, Pitto M. Advances in membranous vesicle and exosome proteomics improving biological understanding and biomarker discovery. *Proteomics*. 2011;11(4):709-720. doi:10.1002/pmic.201000422
- Garcia-Olmo D, Garcia-Olmo DC, Ontanon J, Martinez E, Vallejo M. Tumor DNA circulating in the plasma might play a role in metastasis. The hypothesis of the genometastasis. *Histol Histopathol.* 1999;14(4):1159-1164.
- Colombo M, Raposo G, Thery C. Biogenesis, secretion, and intercellular interactions of exosomes and other extracellular vesicles. *Annu Rev Cell Dev Biol.* 2014;30:255-289. doi:10.1146/annurevcellbio-101512-122326
- Abdouh M, Zhou S, Arena V, et al. Transfer of malignant trait to immortalized human cells following exposure to human cancer serum. *J Exp Clin Cancer Res.* 2014;33:86. doi:10.1186/s13046-014-0086-5
- Abd Elmageed ZY, Yang Y, Thomas R, et al. Neoplastic reprogramming of patient-derived adipose stem cells by prostate cancer cell-associated exosomes. *Stem Cells*. 2014;32(4):983-997. doi: 10.1002/stem.1619
- Hamam D, Abdouh M, Gao ZH, Arena V, Arena M, Arena GO. Transfer of malignant trait to BRCA1 deficient human fibroblasts following exposure to serum of cancer patients. *J Exp Clin Cancer Res.* 2016;35:80. doi:10.1186/s13046-016-0360-9
- Lee TH, Chennakrishnaiah S, Meehan B, et al. Barriers to horizontal cell transformation by extracellular vesicles containing oncogenic H-ras. *Oncotarget*. 2016;7(32):51991-52002. doi:10. 18632/oncotarget.10627
- 30. Abdouh M, Hamam D, Gao ZH, Arena V, Arena M, Arena GO. Exosomes isolated from cancer patients' sera transfer malignant traits and confer the same phenotype of primary tumors to oncosuppressor-mutated cells. *J Exp Clin Cancer Res.* 2017; 36(1):113. doi:10.1186/s13046-017-0587-0
- Mathivanan S, Fahner CJ, Reid GE, Simpson RJ. ExoCarta 2012: database of exosomal proteins, RNA and lipids. *Nucleic Acids Res.* 2012;40(Database issue):D1241-D1244. doi:10.1093/nar/ gkr828
- Kim DK, Kang B, Kim OY, et al. EVpedia: an integrated database of high-throughput data for systemic analyses of extracellular vesicles. *J Extracell Vesicles*. 2013;2. doi:10.3402/jev.v2i0.20384
- Valadi H, Ekstrom K, Bossios A, Sjostrand M, Lee JJ, Lotvall JO. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat Cell Biol*. 2007;9(6):654-659. doi:10.1038/ncb1596

- Eldh M, Ekstrom K, Valadi H, et al. Exosomes communicate protective messages during oxidative stress; possible role of exosomal shuttle RNA. *PLoS One*. 2010;5(12):e15353. doi:10.1371/ journal.pone.0015353
- 35. Manohar S, Harlow M, Nguyen H, Li J, Hankins GR, Park M. Chromatin modifying protein 1A (Chmp1A) of the endosomal sorting complex required for transport (ESCRT)-III family activates ataxia telangiectasia mutated (ATM) for PanC-1 cell growth inhibition. *Cell Cycle*. 2011;10(15):2529-2539. doi:10.4161/cc. 10.15.15926
- Hupalowska A, Miaczynska M. The new faces of endocytosis in signaling. *Traffic*. 2012;13(1):9-18. doi:10.1111/j.1600-0854. 2011.01249.x
- Silva J, Garcia V, Rodriguez M, et al. Analysis of exosome release and its prognostic value in human colorectal cancer. *Genes Chromosomes Cancer*. 2012;51(4):409-418.
- Hoshino A, Costa-Silva B, Shen TL, et al. Tumour exosome integrins determine organotropic metastasis. *Nature*. 2015; 527(7578):329-335. doi:10.1038/nature15756
- Qi P, Zhou XY, Du X. Circulating long non-coding RNAs in cancer: current status and future perspectives. *Mol Cancer*. 2016;15(1):39. doi:10.1186/s12943-016-0524-4
- Chen Y, Xie Y, Xu L, et al. Protein content and functional characteristics of serum-purified exosomes from patients with colorectal cancer revealed by quantitative proteomics. *Int J Cancer*. 2017;140(4):900-913. doi:10.1002/ijc.30496
- Ragusa M, Statello L, Maugeri M, et al. Highly skewed distribution of miRNAs and proteins between colorectal cancer cells and their exosomes following Cetuximab treatment: biomolecular, genetic and translational implications. *Oncoscience*. 2014;1(2): 132-157. doi:10.18632/oncoscience.19
- Ahadi A, Khoury S, Losseva M, Tran N. A comparative analysis of lncRNAs in prostate cancer exosomes and their parental cell lines. *Genomics Data*. 2016;9:7-9. doi:10.1016/j.gdata.2016.05. 010
- Johnstone RM, Adam M, Hammond JR, Orr L, Turbide C. Vesicle formation during reticulocyte maturation. Association of plasma membrane activities with released vesicles (exosomes). *J Biol Chem.* 1987;262(19):9412-9420.
- Mathivanan S, Ji H, Simpson RJ. Exosomes: extracellular organelles important in intercellular communication. *J Proteomics*. 2010;73(10):1907-1920. doi:10.1016/j.jprot.2010.06.006
- Nolte-'t Hoen EN, Buschow SI, Anderton SM, Stoorvogel W, Wauben MH. Activated T cells recruit exosomes secreted by dendritic cells via LFA-1. *Blood*. 2009;113(9):1977-1981. doi: 10.1182/blood-2008-08-174094
- Fevrier B, Vilette D, Archer F, et al. Cells release prions in association with exosomes. *Proc Natl Acad Sci U S A*. 2004;101(26): 9683-9688. doi:10.1073/pnas.0308413101
- Filipazzi P, Burdek M, Villa A, Rivoltini L, Huber V. Recent advances on the role of tumor exosomes in immunosuppression and disease progression. *Semin Cancer Biol.* 2012;22(4):342-349. doi:10.1016/j.semcancer.2012.02.005
- Clayton A, Mason MD. Exosomes in tumour immunity. *Curr* Oncol. 2009;16(3):46-49.

- 49. Corrado C, Raimondo S, Saieva L, Flugy AM, De Leo G, Alessandro R. Exosome-mediated crosstalk between chronic myelogenous leukemia cells and human bone marrow stromal cells triggers an interleukin 8-dependent survival of leukemia cells. *Cancer Lett.* 2014;348(1-2):71-76. doi:10.1016/j.canlet.2014.03. 009
- Kahlert C, Kalluri R. Exosomes in tumor microenvironment influence cancer progression and metastasis. *J Mol Med (Berl)*. 2013; 91(4):431-437. doi:10.1007/s00109-013-1020-6
- Hong BS, Cho JH, Kim H, et al. Colorectal cancer cell-derived microvesicles are enriched in cell cycle-related mRNAs that promote proliferation of endothelial cells. *BMC Genomics*. 2009;10: 556. doi:10.1186/1471-2164-10-556
- Zomer A, Vendrig T, Hopmans ES, van Eijndhoven M, Middeldorp JM, Pegtel DM. Exosomes: fit to deliver small RNA. *Commun Integr Biol.* 2010;3(5):447-450. doi:10.4161/cib.3.5.12339
- Deng ZB, Zhuang X, Ju S, et al. Exosome-like nanoparticles from intestinal mucosal cells carry prostaglandin E2 and suppress activation of liver NKT cells. *J Immunol.* 2013;190(7):3579-3589. doi:10.4049/jimmunol.1203170
- Colotta F, Allavena P, Sica A, Garlanda C, Mantovani A. Cancerrelated inflammation, the seventh hallmark of cancer: links to genetic instability. *Carcinogenesis*. 2009;30(7):1073-1081. doi: 10.1093/carcin/bgp127
- Bondar T, Medzhitov R. The origins of tumor-promoting inflammation. *Cancer Cell*. 2013;24(2):143-144. doi:10.1016/j.ccr. 2013.07.016
- Costa-Silva B, Aiello NM, Ocean AJ, et al. Pancreatic cancer exosomes initiate pre-metastatic niche formation in the liver. *Nat Cell Biol.* 2015;17(6):816-826. doi:10.1038/ncb3169
- Peinado H, Aleckovic M, Lavotshkin S, et al. Melanoma exosomes educate bone marrow progenitor cells toward a prometastatic phenotype through MET. *Nat Med.* 2012;18(6): 883-891. doi:10.1038/nm.2753
- Milane L, Singh A, Mattheolabakis G, Suresh M, Amiji MM. Exosome mediated communication within the tumor microenvironment. *J Control Release*. 2015;219:278-294. doi:10.1016/j. jconrel.2015.06.029
- Vella LJ. The emerging role of exosomes in epithelial– mesenchymal-transition in cancer. *Front Oncol.* 2014;4:361. doi:10.3389/fonc.2014.00361
- Andreola G, Rivoltini L, Castelli C, et al. Induction of lymphocyte apoptosis by tumor cell secretion of FasL-bearing microvesicles. *J Exp Med*. 2002;195(10):1303-1316.
- Huber V, Fais S, Iero M, et al. Human colorectal cancer cells induce T-cell death through release of proapoptotic microvesicles: role in immune escape. *Gastroenterology*. 2005;128(7): 1796-1804.
- Ciravolo V, Huber V, Ghedini GC, et al. Potential role of HER2overexpressing exosomes in countering trastuzumab-based therapy. J Cell Physiol. 2012;227(2):658-667. doi:10.1002/jcp.22773
- Safaei R, Larson BJ, Cheng TC, et al. Abnormal lysosomal trafficking and enhanced exosomal export of cisplatin in drugresistant human ovarian carcinoma cells. *Mol Cancer Ther*. 2005;4(10):1595-1604. doi:10.1158/1535-7163.mct-05-0102

- Sousa D, Lima RT, Vasconcelos MH. Intercellular transfer of cancer drug resistance traits by extracellular vesicles. *Trends Mol Med*. 2015;21(10):595-608. doi:10.1016/j.molmed.2015.08.002
- 65. Kucharzewska P, Christianson HC, Welch JE, et al. Exosomes reflect the hypoxic status of glioma cells and mediate hypoxiadependent activation of vascular cells during tumor development. *Proc Natl Acad Sci U S A*. 2013;110(18):7312-7317. doi:10.1073/ pnas.1220998110
- Ekstrom EJ, Bergenfelz C, von Bulow V, et al. WNT5A induces release of exosomes containing pro-angiogenic and immunosuppressive factors from malignant melanoma cells. *Mol Cancer*. 2014;13:88. doi:10.1186/1476-4598-13-88
- Tadokoro H, Umezu T, Ohyashiki K, Hirano T, Ohyashiki JH. Exosomes derived from hypoxic leukemia cells enhance tube formation in endothelial cells. *J Biol Chem.* 2013;288(48): 34343-34351. doi:10.1074/jbc.M113.480822
- 68. Ji H, Greening DW, Barnes TW, et al. Proteome profiling of exosomes derived from human primary and metastatic colorectal cancer cells reveal differential expression of key metastatic factors and signal transduction components. *Proteomics*. 2013; 13(10-11):1672-1686. doi:10.1002/pmic.201200562
- Lokody I. Genetics: exosomally derived miR-105 destroys tight junctions. *Nat Rev Cancer*. 2014;14(6):386-387. doi:10.1038/ nrc3747
- Rana S, Malinowska K, Zoller M. Exosomal tumor microRNA modulates premetastatic organ cells. *Neoplasia*. 2013;15(3): 281-295.
- Luga V, Zhang L, Viloria-Petit AM, et al. Exosomes mediate stromal mobilization of autocrine Wnt-PCP signaling in breast cancer cell migration. *Cell*. 2012;151(7):1542-1556. doi:10. 1016/j.cell.2012.11.024
- Webber J, Steadman R, Mason MD, Tabi Z, Clayton A. Cancer exosomes trigger fibroblast to myofibroblast differentiation. *Cancer Res.* 2010;70(23):9621-9630. doi:10.1158/0008-5472.can-10-1722
- Roma-Rodrigues C, Fernandes AR, Baptista PV. Exosome in tumour microenvironment: overview of the crosstalk between normal and cancer cells. *BioMed Res Int*. 2014;2014:179486. doi:10.1155/2014/179486
- Hood JL, San RS, Wickline SA. Exosomes released by melanoma cells prepare sentinel lymph nodes for tumor metastasis. *Cancer Res.* 2011;71(11):3792-3801. doi:10.1158/0008-5472.can-10-4455
- Boelens MC, Wu TJ, Nabet BY, et al. Exosome transfer from stromal to breast cancer cells regulates therapy resistance pathways. *Cell*. 2014;159(3):499-513. doi:10.1016/j.cell.2014.09.051
- Bobrie A, Krumeich S, Reyal F, et al. Rab27a supports exosomedependent and -independent mechanisms that modify the tumor microenvironment and can promote tumor progression. *Cancer Res.* 2012;72(19):4920-4930. doi:10.1158/0008-5472.can-12-0925
- 77. Chiba M, Kimura M, Asari S. Exosomes secreted from human colorectal cancer cell lines contain mRNAs, microRNAs and natural antisense RNAs, that can transfer into the human hepatoma HepG2 and lung cancer A549 cell lines. *Oncol Rep.* 2012;28(5): 1551-1558. doi:10.3892/or.2012.1967

- Beckham CJ, Olsen J, Yin PN, et al. Bladder cancer exosomes contain EDIL-3/Del1 and facilitate cancer progression. J Urol. 2014;192(2):583-592. doi:10.1016/j.juro.2014.02.035
- Street JM, Yuen PS, Star RA. Bioactive exosomes: possibilities for diagnosis and management of bladder cancer. *J Urol.* 2014; 192(2):297-298. doi:10.1016/j.juro.2014.05.050
- Akakura N, Hoogland C, Takada YK, et al. The COOH-terminal globular domain of fibrinogen gamma chain suppresses angiogenesis and tumor growth. *Cancer Res.* 2006;66(19):9691-9697. doi: 10.1158/0008-5472.can-06-1686
- Lampropoulos P, Zizi-Sermpetzoglou A, Rizos S, Kostakis A, Nikiteas N, Papavassiliou AG. TGF-beta signalling in colon carcinogenesis. *Cancer Lett.* 2012;314(1):1-7. doi:10.1016/j.canlet. 2011.09.041
- Ichikawa M, Williams R, Wang L, Vogl T, Srikrishna G. S100A8/ A9 activate key genes and pathways in colon tumor progression. *Mol Cancer Res.* 2011;9(2):133-148. doi:10.1158/1541-7786. mcr-10-0394
- Ohshima K, Inoue K, Fujiwara A, et al. Let-7 microRNA family is selectively secreted into the extracellular environment via exosomes in a metastatic gastric cancer cell line. *PLoS One*. 2010; 5(10):e13247. doi:10.1371/journal.pone.0013247
- Zhou W, Fong MY, Min Y, et al. Cancer-secreted miR-105 destroys vascular endothelial barriers to promote metastasis. *Cancer Cell*. 2014;25(4):501-515. doi:10.1016/j.ccr.2014.03.007
- Kruger S, Abd Elmageed ZY, Hawke DH, et al. Molecular characterization of exosome-like vesicles from breast cancer cells. *BMC Cancer*. 2014;14:44. doi:10.1186/1471-2407-14-44
- Zhang Y, Liu D, Chen X, et al. Secreted monocytic miR-150 enhances targeted endothelial cell migration. *Mol Cell*. 2010; 39(1):133-144. doi:10.1016/j.molcel.2010.06.010
- Bigagli E, Luceri C, Guasti D, Cinci L. Exosomes secreted from human colon cancer cells influence the adhesion of neighboring metastatic cells: role of microRNA-210. *Cancer Biol Ther*. 2016: 1-8. doi:10.1080/15384047.2016.1219815
- Tanaka S, Hosokawa M, Ueda K, Iwakawa S. Effects of decitabine on invasion and exosomal expression of miR-200c and miR-141 in oxaliplatin-resistant colorectal cancer cells. *Biol Pharm Bull.* 2015;38(9):1272-1279. doi:10.1248/bpb. b15-00129
- Tripathi V, Ellis JD, Shen Z, et al. The nuclear-retained noncoding RNA MALAT1 regulates alternative splicing by modulating SR splicing factor phosphorylation. *Mol Cell.* 2010;39(6): 925-938. doi:10.1016/j.molcel.2010.08.011
- Yang X, Xie X, Xiao YF, et al. The emergence of long non-coding RNAs in the tumorigenesis of hepatocellular carcinoma. *Cancer Lett.* 2015;360(2):119-124. doi:10.1016/j. canlet.2015.02.035
- Yuan JH, Yang F, Wang F, et al. A long noncoding RNA activated by TGF-beta promotes the invasion-metastasis cascade in hepatocellular carcinoma. *Cancer Cell*. 2014;25(5):666-681. doi: 10.1016/j.ccr.2014.03.010
- 92. Liu T, Zhang X, Gao S, et al. Exosomal long noncoding RNA CRNDE-h as a novel serum-based biomarker for diagnosis and prognosis of colorectal cancer. *Oncotarget*. 2016;7(51): 85551-85563. doi:10.18632/oncotarget.13465

- 93. Sun R, Qin C, Jiang B, et al. Down-regulation of MALAT1 inhibits cervical cancer cell invasion and metastasis by inhibition of epithelial-mesenchymal transition. *Mol BioSyst.* 2016; 12(3):952-962. doi:10.1039/c5mb00685f
- 94. Huarte M, Guttman M, Feldser D, et al. A large intergenic noncoding RNA induced by p53 mediates global gene repression in the p53 response. *Cell*. 2010;142(3):409-419. doi:10.1016/j.cell. 2010.06.040
- Malik ZA, Kott KS, Poe AJ, et al. Cardiac myocyte exosomes: stability, HSP60, and proteomics. *Am J Physiol Heart Circ Physiol*. 2013;304(7):H954-H965. doi:10.1152/ajpheart.00835.2012
- Xu CG, Yang MF, Ren YQ, Wu CH, Wang LQ. Exosomes mediated transfer of lncRNA UCA1 results in increased tamoxifen resistance in breast cancer cells. *Eur Rev Med Pharmacol Sci.* 2016;20(20):4362-4368.
- 97. Kogure T, Yan IK, Lin WL, Patel T. Extracellular vesiclemediated transfer of a novel long noncoding RNA TUC339: a mechanism of intercellular signaling in human hepatocellular cancer. *Genes Cancer*. 2013;4(7-8):261-272. doi:10.1177/ 1947601913499020
- Nawaz M, Fatima F, Nazarenko I, et al. Extracellular vesicles in ovarian cancer: applications to tumor biology, immunotherapy and biomarker discovery. *Expert Rev Proteomics*. 2016;13(4): 395-409. doi:10.1586/14789450.2016.1165613
- Coghlin C, Murray GI. Biomarkers of colorectal cancer: recent advances and future challenges. *Proteomics Clin Appl.* 2015; 9(1-2):64-71. doi:10.1002/prca.201400082
- Romero-Cordoba SL, Salido-Guadarrama I, Rodriguez-Dorantes M, Hidalgo-Miranda A. miRNA biogenesis: biological impact in the development of cancer. *Cancer Biol Ther*. 2014; 15(11):1444-1455. doi:10.4161/15384047.2014.955442
- 101. Lawrie CH, Gal S, Dunlop HM, et al. Detection of elevated levels of tumour-associated microRNAs in serum of patients with diffuse large B-cell lymphoma. *Br J Haematol.* 2008; 141(5):672-675. doi:10.1111/j.1365-2141.2008.07077.x
- 102. Wong TS, Liu XB, Wong BY, Ng RW, Yuen AP, Wei WI. Mature miR-184 as potential oncogenic microRNA of squamous cell carcinoma of tongue. *Clin Cancer Res.* 2008;14(9): 2588-2592. doi:10.1158/1078-0432.ccr-07-0666
- King HW, Michael MZ, Gleadle JM. Hypoxic enhancement of exosome release by breast cancer cells. *BMC Cancer*. 2012;12: 421. doi:10.1186/1471-2407-12-421
- 104. Aushev VN, Zborovskaya IB, Laktionov KK, et al. Comparisons of microRNA patterns in plasma before and after tumor removal reveal new biomarkers of lung squamous cell carcinoma. *PLoS One.* 2013;8(10):e78649. doi:10.1371/journal. pone.0078649
- 105. Kobayashi M, Salomon C, Tapia J, Illanes SE, Mitchell MD, Rice GE. Ovarian cancer cell invasiveness is associated with discordant exosomal sequestration of Let-7 miRNA and miR-200. J Transl Med. 2014;12:4. doi:10.1186/1479-5876-12-4
- 106. Westermann AM, Schmidt D, Holdenrieder S, et al. Serum microRNAs as biomarkers in patients undergoing prostate biopsy: results from a prospective multi-center study. *Anticancer Res.* 2014;34(2):665-669.

- 107. Yang Y, Gu X, Zhou M, Xiang J, Chen Z. Serum microRNAs: a new diagnostic method for colorectal cancer. *Biomed Rep.* 2013; 1(4):495-498. doi:10.3892/br.2013.109
- 108. Kalimutho M, Del Vecchio Blanco G, Di Cecilia S, et al. Differential expression of miR-144* as a novel fecal-based diagnostic marker for colorectal cancer. *J Gastroenterol*. 2011;46(12): 1391-1402. doi:10.1007/s00535-011-0456-0
- 109. Simpson RJ, Lim JW, Moritz RL, Mathivanan S. Exosomes: proteomic insights and diagnostic potential. *Expert Rev Proteomics*. 2009;6(3):267-283. doi:10.1586/epr.09.17
- 110. Ji H, Chen M, Greening DW, et al. Deep sequencing of RNA from three different extracellular vesicle (EV) subtypes released from the human LIM1863 colon cancer cell line uncovers distinct miRNA-enrichment signatures. *PloS One.* 2014;9(10): e110314. doi:10.1371/journal.pone.0110314
- 111. Tovar-Camargo OA, Toden S, Goel A. Exosomal microRNA biomarkers: emerging frontiers in colorectal and other human cancers. *Expert Rev Mol Diagn*. 2016;16(5):553-567. doi:10. 1586/14737159.2016.1156535
- Cha DJ, Franklin JL, Dou Y, et al. KRAS-dependent sorting of miRNA to exosomes. *Elife*. 2015;4:e07197. doi:10.7554/eLife. 07197
- 113. Kosaka N, Iguchi H, Yoshioka Y, Takeshita F, Matsuki Y, Ochiya T. Secretory mechanisms and intercellular transfer of microRNAs in living cells. J Biol Chem. 2010;285(23): 17442-17452. doi:10.1074/jbc.M110.107821
- 114. Le MT, Hamar P, Guo C, et al. miR-200-containing extracellular vesicles promote breast cancer cell metastasis. J Clin Invest. 2014;124(12):5109-5128. doi:10.1172/jci75695
- 115. Pandey GK, Mitra S, Subhash S, et al. The risk-associated long noncoding RNA NBAT-1 controls neuroblastoma progression by regulating cell proliferation and neuronal differentiation. *Cancer Cell*. 2014;26(5):722-737. doi:10.1016/j.ccell.2014.09.014
- 116. Veltri WR. Non-coding RNAs as biomarkers for metastatic prostate cancer. *The Lancet Oncol.* 2014;15(13):1412-1413. doi:10. 1016/s1470-2045(14)71124-6
- 117. Gezer U, Ozgur E, Cetinkaya M, Isin M, Dalay N. Long noncoding RNAs with low expression levels in cells are enriched in secreted exosomes. *Cell Biol Int.* 2014;38(9):1076-1079. doi:10. 1002/cbin.10301
- 118. Batagov AO, Kuznetsov VA, Kurochkin IV. Identification of nucleotide patterns enriched in secreted RNAs as putative cisacting elements targeting them to exosome nano-vesicles. *BMC Genomics.* 2011;12(suppl 3):S18. doi:10.1186/1471-2164-12s3-s18
- Huang X, Yuan T, Tschannen M, et al. Characterization of human plasma-derived exosomal RNAs by deep sequencing. *BMC Genomics*. 2013;14:319. doi:10.1186/1471-2164-14-319
- 120. Liu T, Zhang X, Yang YM, Du LT, Wang CX. Increased expression of the long noncoding RNA CRNDE-h indicates a poor prognosis in colorectal cancer, and is positively correlated with IRX5 mRNA expression. *Onco Targets Ther.* 2016;9: 1437-1448. doi:10.2147/ott.s98268
- 121. Fellig Y, Ariel I, Ohana P, et al. H19 expression in hepatic metastases from a range of human carcinomas. *J Clin Pathol.* 2005;58(10):1064-1068. doi:10.1136/jcp.2004.023648

- 122. Conigliaro A, Costa V, Lo Dico A, et al. CD90+ liver cancer cells modulate endothelial cell phenotype through the release of exosomes containing H19 lncRNA. *Mol Cancer*. 2015;14:155. doi:10.1186/s12943-015-0426-x
- 123. Dimitrova N, Zamudio JR, Jong RM, et al. LincRNA-p21 activates p21 in cis to promote Polycomb target gene expression and to enforce the G1/S checkpoint. *Mol Cell*. 2014;54(5):777-790. doi:10.1016/j.molcel.2014.04.025
- 124. Berrondo C, Flax J, Kucherov V, et al. Expression of the long non-coding RNA HOTAIR correlates with disease progression in bladder cancer and is contained in bladder cancer patient urinary exosomes. *PLoS One*. 2016;11(1):e0147236. doi:10. 1371/journal.pone.0147236
- 125. Yan TH, Lu SW, Huang YQ, et al. Upregulation of the long noncoding RNA HOTAIR predicts recurrence in stage Ta/T1 bladder cancer. *Tumour Biol.* 2014;35(10):10249-10257. doi: 10.1007/s13277-014-2344-8
- 126. Weiner-Gorzel K, Dempsey E, Milewska M, et al. Overexpression of the microRNA miR-433 promotes resistance to paclitaxel through the induction of cellular senescence in ovarian cancer cells. *Cancer Med.* 2015;4(5):745-758. doi:10.1002/cam4.409
- 127. Takahashi K, Yan IK, Kogure T, Haga H, Patel T. Extracellular vesicle-mediated transfer of long non-coding RNA ROR modulates chemosensitivity in human hepatocellular cancer. *FEBS Open Bio.* 2014;4:458-467. doi:10.1016/j.fob.2014.04.007
- 128. Fan Y, Shen B, Tan M, et al. Long non-coding RNA UCA1 increases chemoresistance of bladder cancer cells by regulating Wnt signaling. *FEBS J.* 2014;281(7):1750-1758. doi:10.1111/ febs.12737
- 129. Qu L, Ding J, Chen C, et al. Exosome-Transmitted IncARSR Promotes Sunitinib Resistance in Renal Cancer by Acting as a Competing Endogenous RNA. *Cancer cell*. 2016;29(5):653-668. doi: 10.1016/j.ccell.2016.03.004
- Tominaga N, Yoshioka Y, Ochiya T. A novel platform for cancer therapy using extracellular vesicles. *Adv Drug Deliv Rev.* 2015;95:50-55. doi:10.1016/j.addr.2015.10.002
- 131. Taraboletti G, D'Ascenzo S, Giusti I, et al. Bioavailability of VEGF in tumor-shed vesicles depends on vesicle burst induced

by acidic pH. Neoplasia. 2006;8(2):96-103. doi:10.1593/neo. 05583

- 132. Chen WX, Liu XM, Lv MM, et al. Exosomes from drug-resistant breast cancer cells transmit chemoresistance by a horizontal transfer of microRNAs. *PLoS One*. 2014;9(4):e95240. doi:10. 1371/journal.pone.0095240
- 133. Hoshino H, Miyoshi N, Nagai K, et al. Epithelial–mesenchymal transition with expression of SNAI1-induced chemoresistance in colorectal cancer. *Biochem Biophys Res Commun.* 2009;390(3): 1061-1065. doi:10.1016/j.bbrc.2009.10.117
- 134. Qi P, Du X. The long non-coding RNAs, a new cancer diagnostic and therapeutic gold mine. *Mod Pathol.* 2013;26(2):155-165. doi:10.1038/modpathol.2012.160
- 135. Alvarez-Erviti L, Seow Y, Yin H, Betts C, Lakhal S, Wood MJ. Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nat Biotechnol.* 2011;29(4):341-345. doi:10. 1038/nbt.1807
- 136. Gong Z, Yang J, Li J, et al. Novel insights into the role of microRNA in lung cancer resistance to treatment and targeted therapy. *Curr Cancer Drug Targets*. 2014;14(3):241-258.
- 137. Wang J, Zhou Y, Lu J, et al. Combined detection of serum exosomal miR-21 and HOTAIR as diagnostic and prognostic biomarkers for laryngeal squamous cell carcinoma. *Med Oncol.* 2014;31(9):148. doi:10.1007/s12032-014-0148-8
- 138. Huang YK, Yu JC. Circulating microRNAs and long non-coding RNAs in gastric cancer diagnosis: an update and review. World J Gastroenterol. 2015;21(34):9863-9886. doi:10.3748/wjg.v21. i34.9863
- Mohankumar S, Patel T. Extracellular vesicle long noncoding RNA as potential biomarkers of liver cancer. *Brief Funct Genomics*. 2016;15(3):249-256. doi:10.1093/bfgp/elv058
- 140. Alhamdani MS, Schroder C, Hoheisel JD. Analysis conditions for proteomic profiling of mammalian tissue and cell extracts with antibody microarrays. *Proteomics*. 2010;10(17): 3203-3207. doi:10.1002/pmic.201000170
- 141. Lim JW, Mathias RA, Kapp EA, et al. Restoration of full-length APC protein in SW480 colon cancer cells induces exosomemediated secretion of DKK-4. *Electrophoresis*. 2012;33(12): 1873-1880. doi:10.1002/elps.201100687