

**Supplementary information for
'Results of a worldwide survey on the currently used histopathological diagnostic criteria for invasive lobular
breast cancer' by De Schepper et al.**

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Supplementary Table 1: survey questionnaire

In which country do you practice pathology?
What is your field of interest in pathology?
In which center do you perform your activities:
What is the average volume/year of breast cancer samples in your center?
How many pathologists handle breast cancer specimens in your laboratory
Do you perform consensus diagnosis for difficult cases in your center?
Are you aware of national/international guidelines recommending the use of IHC for the diagnosis of ILC?
How do you diagnose an invasive lobular carcinoma?
Which subtypes of ILC do you report? (Multiple options possible):
Classic
Non-classic
Mixed non-classic
Alveolar
Solid
Trabecular
Histiocytoid/Apocrine
Pleomorphic
Tubulo-lobular
Invasive carcinoma with mixed ductal and lobular features
Mucinous
All of the above
Other
What is the proportion of ILCs diagnosed with ancillary IHC in your institution?
If you perform IHC, which antibody/antibodies do you use?
E-cadherin
Beta- catenin
p120 catenin
Other (specify later)
Double staining (please specify)
Which patterns of staining's do you use to support the diagnosis of ILC by E-cadherin? (multiple options possible)
Complete absence of membranous E-cadherin staining
Dot-like, perinuclear Golgi-type pattern
Fragmented, focal, or beaded expression of E-cadherin
A complete, but weak membranous E-cadherin
All of the above
Other
Do you use upfront E-cadherin IHC for every breast cancer sample?
When E-cadherin is positive but the H&E pattern is lobular: do you use p120 catenin or / and beta-catenin?
Which pattern do you consider for ILC diagnosis? (Multiple options possible):
cytoplasmic or negative beta catenin
cytoplasmic or nuclear p120 staining
Do you use an external control for staining?
If you use E-cadherin, what are its specifications?
Clone:
Firm:
Concentration:

Tissue for validation:
How do you perform antigen retrieval for E-cadherin?
If you perform HIER, how is it performed?
Which buffer is used?
At which pH?
If you use enzymatic retrieval, which agent do you use?
If you use beta-catenin, what are its specifications?
Clone:
Firm:
Concentration:
Tissue for validation:
How do you perform antigen retrieval for beta-catenin?
If you perform HIER, how is it performed?
Which buffer is used?
At which pH?
If you use enzymatic retrieval, which agent do you use?
If you use p120-catenin, what are its specifications?
Clone:
Firm:
Concentration:
Tissue for validation:
How do you perform antigen retrieval for p120-catenin?
If you perform HIER, how is it performed?
Which buffer is used?
At which pH?
If you use enzymatic retrieval, which agent do you use?
Do you use another antibody other than E-cadherin, beta-catenin and/or p120-catenin?
If you use another antibody, what are its specifications?
Specify antibody:
Clone:
Firm:
Concentration:
Tissue for validation:
How do you perform antigen retrieval for this antibody?
If you perform HIER, how is it performed?
Which buffer is used?
At which pH?
If you use enzymatic retrieval, which agent do you use?
Do you use DNA sequencing for cases with inconclusive features?

Supplementary Table 2: characteristics of participants per continent

		Africa n (%)	Asia n (%)	Europe n (%)	North America n (%)	Oceania n (%)	South America n (%)	p- value
Center of activities	Large tertiary hospital	3 (75.0)	11 (40.7)	26 (28.3)	2 (18.2)	1 (50.0)	1 (9.1)	0.1424
	Private laboratory	0 (0.0)	1 (3.7)	9 (9.8)	0 (0.0)	0 (0.0)	4 (36.4)	
	Small community hospital	0 (0.0)	2 (7.4)	4 (4.3)	0 (0.0)	0 (0.0)	1 (9.1)	
	University Hospital	1 (25.0)	13 (48.1)	53 (57.6)	9 (81.8)	1 (50.0)	5 (45.5)	
Average volume breast cancer samples/year	0-150 samples	1 (25.0)	4 (14.8)	1 (1.1)	0 (0.0)	0 (0.0)	2 (18.2)	0.0005
	151-300 samples	2 (50.0)	10 (37.0)	15 (16.3)	1 (9.1)	0 (0.0)	1 (9.1)	
	301-500 samples	1 (25.0)	3 (11.1)	22 (23.9)	0 (0.0)	2 (100.0)	2 (18.2)	
	>500 samples	0 (0.0)	10 (37.0)	54 (58.7)	10 (90.9)	0 (0.0)	6 (54.5)	
Number of pathologist handling breast cancer specimen	1	0 (0.0)	7 (25.9)	1 (1.1)	0 (0.0)	0 (0.0)	2 (18.2)	0.0005
	1 to 4	0 (0.0)	11 (40.7)	51 (55.4)	4 (36.4)	0 (0.0)	8 (72.7)	
	5 or more	4 (100.0)	9 (33.3)	40 (43.5)	7 (63.6)	2 (100.0)	1 (9.1)	
Total		4	27	92	11	2	11	

Supplementary Table 3: Overview of used concentration per used E-cadherin clone

Concentration per clone	Number of participants
NCH-38	38 (42%)
RTU	12 (13%)
1/100	10 (11%)
1/50	4 (4%)
1/200	3 (3%)
1/300	1 (1%)
1/50 to 1/100	1 (1%)
1/170	1 (1%)
1/25	1 (1%)
Missing data	5 (5%)
Clone 36	15 (16%)
RTU	8 (9%)
0,314 µg/ml	2 (2%)
1/200	1 (1%)
Missing data	4 (4%)
EP700Y	14 (15%)
1/200	4 (4%)
RTU	5 (5%)
0,314 µg/ml	1 (1%)
1/700	1 (1%)
unknown	1 (1%)
Missing data	1 (1%)
Clone 36B5	7 (8%)
RTU	3 (3%)
1/100	1 (1%)
1/40	1 (1%)
Missing data	2 (2%)
unknown	6 (7%)
Missing data	6 (7%)
4A2C7	5 (5%)
57µg/l	1 (1%)
1,25 µg/ml	1 (1%)
1/50	1 (1%)
570µg/l	1 (1%)
Missing data	1 (1%)
HECD1	1 (1%)

1/50	1 (1%)
IHC564	1 (1%)
Unknown	1 (1%)
EP6	1 (1%)
1/1000	1 (1%)
ECH-6	1 (1%)
1/100	1 (1%)
G-10	1 (1%)
1/100	1 (1%)
GM016	1 (1%)
RTU	1 (1%)
Grand Total	91

Supplementary Table 4: E-cadherin antibody clone per continent

Europe	92
Missing data	36
NCH-38	24
EP700Y	11
Clone 36	10
Clone 36B5	5
4A2C7	2
ECH-6	1
HECD1	1
GM016	1
G-10	1
Asia	27
NCH-38	8
Missing data	8
unknown	4
4A2C7	3
Clone 36	2
Clone 36B5	1
IHC564	1
South America	11
NCH-38	4
Missing data	4
EP700Y	2
EP6	1
North America	11
Missing data	7
Clone 36	2
NCH-38	1
Clone 36B5	1
Africa	4
Missing data	2
EP700Y	1
NCH-38	1
Oceania	2
Missing data	1
Clone 36	1
Grand Total	147

Supplementary Table 5: staining procedures per antibody

Epitope retrieval modality	E-cadherin	β-catenin	p120-catenin
Enzymatic retrieval	2	1	0
Heat induced epitope retrieval (HIER)	76	31	23
No Antigen retrieval	10	4	5
Unknown	3	0	1
Grand Total	91	36	29
HIER modality	E-cadherin	β-catenin	p120-catenin
Hot plate	20	9	6
Microwave	7	2	1
Pressure cooker	4	6	2
Stainer	28	11	11
Steamer	2	0	1
Water bath	3	1	0
Unknown	11	1	2
Other	1	1	0
Grand Total	76	31	23
HIER buffer	E-cadherin	β-catenin	p120-catenin
Acidic	3	5	2
Low pH (<7,0)	3	5	2
Basic pH	60	24	19
High pH (>8,0)	38	10	10
slightly basic pH [7,0-8,0]	8	6	4
Unknown	14	8	5
Missing data	13	2	2
Grand Total	76	31	23

Supplementary Table 6: concentration per reported clone

Supplementary Table 6a: concentration per clone for β -catenin

Concentration per clone	Number of participants
Clone 14	20
RTU	6
unknown	5
1/100	2
1/200	2
1/75	1
1/150	1
1/300	1
1.25 μ g/ml	1
1/250	1
β-Catenin-1	12
1/200	3
RTU	2
1/1000	1
1/400	1
2.2mg/L	1
unknown	4
17C2	3
1/100	1
RTU	1
unknown	1
CAT-5H10	2
1/200	1
1/100	1
unknown	2
Grand Total	38

Supplementary Table 6b: concentration per clone for p120-catenin

Concentration per clone	Number of participants
Clone 98	14
unknown	5
RTU	3
1/500	1
1/200	1
1/150	1
0.076 μ g/mL	1
1/250	1
1/300	1
EP66	7
RTU	3

1/1800	1
1/200	1
1/75	1
unknown	1
MRQ-5	5
1/50	2
RTU	1
1/100	1
1/400	1
unknown	4
15D2	1
RTU	1
Grand Total	29

Supplementary Table 6c: concentration per clone for other reported antibodies

Concentration per clone	Number of participants
Catenin Delta-1 (D7S2M)	1
0.7 µg/mL	1
Cytokeratin 34βE12	5
1/100	1
1/300	1
1/50	2
RTU	1
P-cadherin (clone 56)	1
1/100	1
Grand Total	7