# **Original Article**





# Comparison of Snail1, ZEB1, E-Cadherin Expression Levels in HPV-Induced Cervical Cancer

# Mahdieh FARZANEHPOUR<sup>1,2</sup>, Ebrahim FAGHIHLOO<sup>3</sup>, Vahid SALIMI<sup>1</sup>, Somayeh JALILVAND<sup>1</sup>, Setareh AKHAVAN<sup>4</sup>, Ahad MUHAMMADNEJAD<sup>5</sup>, Amir Nader EMAMI RAZAVI<sup>5</sup>, Ehsan KAKAVANDI<sup>1</sup>, \*Talat MOKHTARI AZAD<sup>1</sup>

Department of Virology, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran
 Applied Virology Research Center, Bagiyatallah University of Medical Sciences, Tehran, Iran

3. Department of Microbiology, School of Medicine, Shahid Beheshti University of Medical Sciences, Tehran, Iran

4. Department of Gynecology Oncology, Imam Khomeini Hospital Complex, Valiasr Hospital, Tehran University of Medical Scienc-

es, Tehran, Iran

5. Cancer Biology Research Center, Cancer Institute of Iran, Tehran University of Medical Sciences, Tehran, Iran

\*Corresponding Author: Email: Mokhtari@tums.ac.ir

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#### Abstract

**Background:** Molecular profiling techniques are the rapid detection of biomarkers in the human papillomavirus (HPV) infected cells. We aimed to measure the expression level of three cell factors including Snail1, ZEB-1, and E-cadherin in cervical cancer (CC), precancerous and healthy samples, simultaneously, to find potential biomarkers. **Methods:** The expression level of the mentioned cell factors were investigated in 72 CC patients, precancerous patients, and healthy controls by using Real-Time PCR.

**Results:** The results demonstrated a significant reduction in the expression level of E-cadherin in cancer and precancerous cases than that in healthy cases; whereas the expression level of ZEB-1 and Snail1 were upregulated in cancer and precancerous samples. The receiver operating characteristic (ROC) analyses shows the highest AUC value emerged for Snail1: 1(95% CI: 1-1) in comparing CC and healthy groups with a sensitivity of 100.0 % and specificity of 100.0%.

**Conclusion:** The molecular biomarker Snail1 may be helpful to early diagnosis and prognosis of CC in the HPV-infected human populations. Considering the increased expression level of Snail1 in cancer and precancerous tissue compared to healthy tissue as well as the area under the ROC curve, Snail1 can be used for early detection of CC.

Keywords: Cervical cancer; Human papillomavirus; E-cadherin; Snail1

## Introduction

Cervical cancer (CC) is the third widespread cancer, which has the fourth rank among cancer mortality causes of women worldwide (1, 2). The anticipated outbreak is estimated at 21.7 million until 2030, which occasions the death of 13 million of the infected cases due to the population aging (3). CC is 12th leading cause of female disease in Iran (4).

The viral infections are the causative agents of almost 15% of all cancers (5, 6). Human papillo-



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mavirus (HPV) performs its life cycle in either mucosal or cutaneous stratified squamous epithelia (7, 8). The persistent high-risk human papillomaviruses (HR-HPVs) are the principal etiologic agent in the CC pathogenesis (9-11). Conversely, HR-HPVs, and especially HPV-16 as the most prevalent virus infecting the cervix, are accompanied by the entire spectrum of cervical intraepithelial neoplasia (CIN) lesions as well as, invasive squamous carcinomas (12, 13).

After HR-HPV infection, several cellular changes associated with the epithelial-mesenchymal transition (EMT) were observed (14, 15). EMT turns epithelial cells into the mesenchymal cells that can invade and migrate. In addition, it contributes to the metastatic progression in human cancer cells (16). One of the significant characteristics of EMT is the functional loss of E-cadherin (17, 18).

E-cadherin is a transmembrane glycoprotein encoded by the *CDH1* gene that its low expression is associated with increased invasiveness and metastasis in several cancers (19, 20). It is inactivated by multiple mechanisms, probably because of the genetic alteration, reduced gene expression, changes of another cadherin–catenin complexes or posttranslational modification of the protein leading to cytoplasmic delocalization (21-23).

The principal mechanism for E-cadherin loss during EMT is transcriptional repression. Several repressors comprise zinc-finger E-box-binding homeobox 1 (ZEB-1), ZEB2/SIP1, Twist, Snail1, and Snail2 can bind to E-box motifs and repress E-cadherin transcription (24-26).

EMT is dynamically regulated during CC progression and the expression level of EMT markers such as E-cadherin and vimentin, change in this dynamic regulation. Transcription factors have a key role in EMT induction. The Snail has been demonstrated to regulate E-cadherin expression and is associated with CC development (27). Studies into the ZEB family in CC are relatively few, and the role of the ZEB family in CC is currently unknown (28).

Among these, Snail1 has a critical role, since its expression is widely observed in EMT processes preceding the remaining EMT-TFs; moreover, ectopic Snail1 induces other EMT-TFs such as Zeb1/2 and Snail2 (29). Also, Snai1 depletion severely impacts mesoderm formation during embryogenesis (30,31).

Snail1 acts as a crucial factor for cellular motion through the progression and metastasis of cancer cells. Moreover, it was found that epidermal growth factor (EGF) stimulation causes the upregulation and accumulation of Snail1 protein in CC cells (32, 33).

We investigate the biomarker potential of the three cell factors, including E-cadherin, ZEB-1, and Snail1 in CC subjects. For this purpose, the expressions of E-cadherin, ZEB-1 and Snail1 were evaluated in the cervical, precancerous, and healthy tissue samples. Also, the receiver operating characteristic curve analysis was employed to compare selected groups for the diagnosis of CC.

# Materials and Methods

## Tissue sample collection

Seventy-two fresh uterine cervix biopsies were collected and kept in RNAlater (Qiagen) at -80 °C to stabilize RNA. Patients that received any chemo/radiation therapy were excluded. The routine hematoxylin-eosin stain on 5 µm paraffin sections was utilized to make the biopsies at colposcopy and surgery and assessed in participating hospitals and classified as healthy, precancer (CIN1, CIN2, CIN3), or invasive cancer according to international criteria (34). All subjects filled consent before operations at Imam Khomeini Complex Hospital (Tehran, Iran) from 2016 to 2018. This study was approved by the Ethics Committee of Tehran University of Medical Sciences (IR.TUMS.SPH.REC.1395.838).

## Total RNA extraction

Total RNA was extracted from tissues with TRIzol (Invitrogen, Carlsbad, CA, USA). The extracted RNAs were stored at – 80 °C until cDNA synthesis. Extraction quality was evaluated by 28s:18s rRNA evaluation using agarose gel electrophoresis stained with SYBR Safe dye (Invitrogen, Carlsbad, CA, The USA) (35).

#### cDNA synthesis

The reverse transcription (Fermentas, Vilnius, Lithuania) was performed at 42 °C for 60 min, followed by 70 °C for 5 min according to the manufacturer's instructions (36).

#### qRT-PCR

The expression of ZEB-1 (37), Snail1 (38), and E-cadherin (39) were analyzed by quantitative Real-Time PCR using the SYBR Green (TAKARA Bio INC., Otsu, Japan) with Applied Biosystems®StepOnePlus<sup>™</sup> (Applied Biosystems, Foster City, CA, USA). The human Glyceraldehyde 3-phosphate dehydrogenase (GAPDH) was applied to normalize the relative quantity (40- 43).

#### Statistical analysis

The Mann-Witney non-parametric test and oneway ANOVA were performed to analyze the statistical discrepancy between groups using GraphPad Prism (7.0.1). A *P*-value of < 0.05 was considered remarkable. The area under receiver operating characteristic (ROC) curves were calculated using R software (version 3.4.4).

### Results

#### Patient and control data

Overall 72 fresh specimens (36 normal, 18 CC and 18 precancerous) were prepared for Snail1, ZEB-1, and E-cadherin expression analysis. The mean age of CC, precancerous, and healthy groups were 61 yr (45-81 yr), 47 yr (27–57 yr), and 36 yr (23–49 yr), respectively with no significant discrepancy.

# The expression levels of Snail1, ZEB-1, and E-cadherin

The expression levels of Snail1, ZEB-1 and Ecadherin were determined in cervical samples (Fig. 1).



Fig. 1: The relative expression level of Snail1, ZEB-1, and E-cadherin between cervical cancer samples, Precancerous samples, and healthy samples. Each sample was analyzed in triplicate and normalized to GAPDH

The expression of E-cadherin was significantly decreased in cancer and precancer groups in comparison to the healthy group (P < 0.0001, P = 0.026), also, in cancer group in comparison to the precancer group (P = 0.02). The expression of ZEB-1 and Snail1 in cervical tissue were remarkably higher in the CC group compared with the

healthy group (P = 0.0003, P < 0.0001, respectively). In addition, the expression of ZEB-1 and Snail1 in the cancer group, compared to the precancer group, was not significant for ZEB-1 and was significant for Snail1 (P < 0.0001). Finally, the expression of ZEB-1 and Snail1 in the cancer group was significantly increased in the precancer group compared with the healthy group (P = 0.0003, P = 0.026, respectively).

# The correlation analysis between Snail1, ZEB-1, and E-cadherin expression

To comprehend the associations between Snail1, ZEB-1, and E-cadherin expressions, the correlation values were explored. A significant and inverse correlation was found between ZEB-1 and E-cadherin in healthy group (r = -0.432, *P*-value = 0.05). The ZEB 1 and Snail1 had an inverse and significant relationship in cancer group (r = -0.94, *P*-value<0.00001). Finally, a meaningful and inverse correlation was observed between ZEB-1 and E-cadherin in cancer group (r = -0.703, *P*-value = 0.005). Any considerable correlation was not observed.

# Receiver operating characteristic (ROC) curve analysis

The ROC curves were generated and they are under area analyses were performed to evaluate the diagnostic value of Snail1, ZEB-1, and E- cadherin expression levels in the samples of CC, precancerous and healthy (Table. 1).

ROC curves showed that the area under the curve (AUC) values in CC and healthy groups were Snail1: 1(95% CI: 1-1), ZEB-1: 0.99 (95%) CI: 0.97-1) and E-cadherin: 0.90 (95% CI: 0.81-1) (Fig. 2). The AUC value in CC and precancerous groups were Snail1: 0.94 (95% CI: 0.88-1), ZEB-1: 0.67(95% CI: 0.48-0.85) and E-cadherin: 0.71 (95% CI: 0.54-0.89). Finally, the AUC in the precancerous and healthy groups were Snail1: 0.88 (95% CI: 0.78-0.98), ZEB-1: 0.74 (95% CI: 0.58-0.90) and E-cadherin: 0.71 (95% CI: 0.55-0.87). So, the highest AUC value was obtained for Snail1: 1(95% CI: 1-1) in comparing CC and healthy groups. It indicated that Snail1 has a strong potential diagnosis value for differentiating of CC from precancerous and healthy groups. Moreover, the sensitivity of Snail1 was higher than two other proteins in comparing CC and healthy groups (100%), precancerous and healthy groups (73.7), and CC and precancerous groups (78.9) (Table 2).

Table 1: ROC curve analysis. Area under the curve (AUC) value of Snail1, ZEB-1, and E-cadherin

Cell Factors	Cervical cancer and Normal groups	Precancerous cervical and Normal groups	Cervical cancer and Precancerous cervical groups
E-cadherin	AUC	AUC	AUC
	95% CI :(0.81-1)	95% CI : (0.55-0.87)	95% CI : (0.54-0.89)
ZEB-1	AUC	AUC	AUC
	95% CI : (0.97-1)	95% CI : (0.58-0.90)	95% CI : (0.48-0.85)
Snail1	AUC	AUC	AUC
	95% CI : (1-1)	95% CI : (0.78-0.98)	95% CI : (0.88-1)

 Table 2: The Snail1, ZEB-1, and E-cadherin cellular factors and their sensitivity and specificity estimation according to the ROC Curve results

Cell Factors	Sensitivity and	Cervical cancer and	Precancerous cervical and	Cervical cancer and Precan-
	specificity	i voimai gioups	i voimai gioups	cerous cervicar groups
E-cadherin	Sensitivity	75	30	65
	Specificity	90.4	90	80
ZEB-1	Sensitivity	100	45	20
	Specificity	95	95	95
Snail1	Sensitivity	100	73.7	78.9
	Specificity	100	84.2	94.7



Fig. 2: (a) Receiver-operating characteristics (ROC) curve analysis using Snail1, ZEB-1, and E-cadherin for discriminating cervical cancer and healthy groups in tissue. The green, blue and violet lines represent Snail1, ZEB-1, and Ecadherin, respectively.

(b) Receiver-operating characteristics (ROC) curve analysis using Snail1, ZEB-1, and E-cadherin for discriminating cervical cancer and precancerous groups in tissue. The green, blue, and violet lines represent Snail1, ZEB-1, and E-cadherin, respectively.

(c) Receiver-operating characteristics (ROC) curve analysis using Snail1, ZEB-1, and E-cadherin for discriminating precancerous and healthy groups in tissue. The green, blue, and violet lines represent Snail1, ZEB-1, and E-cadherin, respectively

# Discussion

Finding new biomarkers for CC has a great importance to diagnose disease before the tumor becomes invasive. The HPV infection of cervical cells leads to many cellular alterations. In this study, we surveyed potential value of three candidate genes as CC tissue biomarkers. The outcomes revealed that the E-cadherin downregulation and up-regulation of ZEB-1 and Snail1 in cancerous/precancerous samples in comparison to healthy tissue.

Epithelial to mesenchymal transition (EMT) take places through physiological states e.g. development and tissue damage repair, also in pathological situations like cancer initiation and progression. In this phenomenon, epithelial cells transdifferentiate to mesenchymal cells.

EMT also plays a key role in the tumor progression and metastasis with stem cell properties that render resisting to cancer treatment (18, 31, 44-48).

The EMT procedure involves cellular and cell adhesion abnormalities. It is accompanied by betterment in attacking and dynamic exclusivities (49). There are currently other complementary ways with EMT not yet known. EMT is becoming a chosen aim for anti-cancer treatment (15). Although, some include well-known genes and markers such as E-cadherin and Vimentin that can be helpful in identifying EMT in tumors (50, 51).

The loss of E-Cadherin is a sign of completing EMT in epithelial tumors. E-cadherin, as the prevailing cell-cell adhesion molecule is found in the epithelial cell types (52). Some studies disclosed the important role of E-cadherin during tumor progression and invasion (53, 54). Moreover, they are critical factors in the designation final Ecadherin level via binding to its inhibitory factors such as transcription factors zinc finger E-boxbinding proteins 1 and 2 (ZEB-1 and ZEB2) and Snail1 levels (55-57). Snail1 is a zinc finger protein known as the transcription factor, which is expressed by some epithelial tumor cells and fibroblasts (58). Snail1 straightly binds to the Eboxes present in the proximal E-cadherin pro-

moter and suppresses E-cadherin gene expression (23, 59). Moreover, Snail blocks the progression of cell cycle and participates in cell movement and survival, and transcriptional regulation of cytokines to mediate invasion and inflammation (60). The expression levels of Snail1 and ZEB-1 up-regulate due to abrupt changes in the tumor microenvironment. In cancer, it has been reported that promoting ZEB1 expression plays a vital role in progression and metastasis in the renal cell carcinoma (61), endometrial cancer (62), invasive breast cancer (63) and lung adenocarcinoma (50). Down-regulation of ZEB1 the expression can prevent invasive tumors from converting to mesenchymal phenotype by reducing the proliferation and mobility of CC cells, which suggests that ZEB1 may be a potential therapeutic target for cervical squamous cell carcinoma (64).

The function of E-cadherin in counteracting with the invasive property of cancer cells can lead to design treatment methods, which reduce the suppressor genes and induce the increase of the E-cadherin expression (25).

Overexpression of Snail was reported in association with metastasis and poor prognosis in gastric cancer (65). In addition, the correlation of high expression level of ZEB-1 with loss of Ecadherin expression was found in various cancer cells (66-70). Herein, the similar results as above were obtained, so that the up-regulation of Snail1 and ZEB-1 were along with the down-regulation of E-cadherin in the cervical cancerous cells.

We used the ROC curves to investigate the predictive power of Snail1 as a diagnostic biomarker for CC. The uttermost AUC value emerged for Snail1: 1(95% CI: 1-1) in comparing CC and the healthy groups with a sensitivity of 100.0% and specificity of 100.0%. Moreover, the highest AUC value and sensitivity were obtained in comparing precancerous cancer and the healthy groups and precancerous cancer compared with the cancer groups. Therefore, the expression level of Snail1 may be tracked as a tissue marker for CC to its prognosis and diagnosis. The overexpression of ZEB1 and Snail1 and the regulation of E-cadherin expression are closely related to the differentiation

status and the invasive capacity of cervical carcinoma. In cervical carcinoma tissue, ZEB1 and Snail1 are highly expressed, which may further enhance the regulation of the expression of E-cadherin. This may cause an increase in malignancy and invasiveness of CC cells. This study also demonstrated that the critical role of ZEB1, Snail1, and E-cadherin in cervical carcinoma, which provides a theoretical basis for the purpose of gene therapy in the metastasis of cervical carcinoma.

Ideal tumor markers should have high sensitivity and specificity to differentiate cancer patients from healthy subjects. They should be secreted into the circulation and activities at a concentration proportional to tumor burden.

### Conclusion

The expression level of several cell factors changes in CC cells. The identification of these factors is beneficial to cancer prognosis and early treatment. We found that up-regulation of the tissue cell factors including Snail1, ZEB-1, and down-regulation of E-cadherin are considerable biomarkers for the diagnosis of the HPV associated CC. Finally, we found out a correlation between expression levels of ZEB-1 and Ecadherin in healthy and cancer groups, which were in agreement with other previous reports. The molecular biomarker Snail1 may be helpful to early diagnosis and prognosis of CC in the HPV-infected human populations. Then Snail1 can be used for early detection of CC.

## Ethical considerations

Ethical issues (Including plagiarism, informed consent, misconduct, data fabrication and/or falsification, double publication and/or submission, redundancy, etc.) have been completely observed by the authors.

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# **Conflict** of interest

The authors declare that there is no conflict of interest.

### References

- 1. Duenas-Gonzalez A, Serrano-Olvera A, Cetina L, et al (2014). New molecular targets against cervical cancer. *Int J Womens Health*, 6: 1023-31.
- 2. Siegel RL, Miller KD, Jemal A (2017). Cancer Statistics. CA Cancer J Clin, 67: 7-30.
- 3. Freddie B, Ahmedin J, Nathan G, et al (2012). Global cancer transitions according to the Human Development Index (2008-2030): a population-based study. *Lancet Oncol*, 13: 790-801.
- Bruni L, Albero G, Serrano B, et al (2014). ICO Information Centre on HPV and Cancer (HPV Information Centre). Human Papillomavirus and Related Diseases in the World. Summary Report 2015. HPV information center.
- zur Hausen H (1999). Papillomaviruses in human cancers. Proc Assoc Am Physicians, 111: 581-7.
- Munoz N, Castellsague X, de Gonzalez AB, et al (2006). HPV in the etiology of human cancer. *Vacine*, 24 Suppl 3:S3/1-10.
- Moody CA, Laimins LA (2010). Human papillomavirus oncoproteins: pathways to transformation. *Nat Rev Cancer*, 10(8):550-60.
- Troha M, sterbenc A, Mlaker M, et al (2018). Human papillomavirus (HPV) infection and vaccination: knowledge and attitudes among healthcare professionals and the general public in Slovenia. *Acta Dermatovenerol Alp Pannonica Adriat*, 27(2):59-64.
- zur Hausen H (2002). Papillomaviruses and cancer: from basic studies to clinical application. Nat Rev Cancer, 2(5): 342-50.
- Walboomers JM, Jacobs MV, Manos MM, et al (1999). Human papillomavirus is a necessary cause of invasive cervical cancer worldwide. J Pathol, 189(1): 12-9.

- Guo F, Cofie LE, Berenson AB (2018). Cervical Cancer Incidence in Young U.S. Females After Human Papillomavirus Vaccine Introduction. *Am J Prev Med*, 55(2):197-204.
- 12. Stoler MH (2003). Human papillomavirus biology and cervical neoplasia: implications for diagnostic criteria and testing. *Arch Pathol Lab Med*, 127(8): 935-9.
- Li TY, Wu ZN, Jiang MY (2018). Association between high risk human papillomavirus DNA load and cervical lesions in different infection status. *Zhonghua Zhong Liu Za Zhi*, 40(6): 475-480.
- Geiger T, Sabanay H, Kravchenko-Balasha N, et al (2008). Anomalous features of EMT during keratinocyte transformation. *PLoS One*, 3: e1574.
- 15. Marcucci F, Stassi G, De Maria R, et al (2016). Epithelial-mesenchymal transition: a new target in anticancer drug discovery. *Nat Rev Drug Discov*, 15(5):311-25.
- 16. Thiery JP (2002). Epithelial-mesenchymal transitions in tumour progression. *Nat Rev Cancer*, 2(6):442-54.
- Perl AK, Wilgenbus P, Dahl U, et al (1998). A causal role for E-cadherin in the transition from adenoma to carcinoma. *Nature*, 392(6672):190-3.
- Moustakas A, de Herreros AG (2017). Epithelialmesenchymal transition in cancer. *Mol Oncol*, 11: 715-717.
- Derksen PW, Liu X, Saridin F, van der Gulden H, et al (2006). Somatic inactivation of Ecadherin and p53 in mice leads to metastatic lobular mammary carcinoma through induction of anoikis resistance and angiogenesis. *Cancer Cell*, 10(5): 437-49.
- Gould Rothberg BE, Bracken MB (2006). Ecadherin immunohistochemical expression as a prognostic factor in infiltrating ductal carcinoma of the breast: a systematic review and meta-analysis. *Breast Cancer Res Treat*, 100(2): 139-48.
- 21. Hirohashi S (1998). Inactivation of the Ecadherin-mediated cell adhesion system in human cancers. *Am J Pathol*, 153(2): 333-9.
- 22. Mareel M, Berx G, Van Roy F, et al (1996).Cadherin/catenin complex: a target for antiinvasive therapy? *J Cell Biochem*, 61: 524–30.
- 23. Mazzolini R, Gonzalez N, Garcia-Garijo A, et al (2018). Snail1 transcription factor controls

telomere transcription and integrity. Nucleic Acids Res, 46(1): 146-158.

- 24. Barrallo-Gimeno A, Nieto MA (2005). The Snail genes as inducers of cell movement and survival: implications in development and cancer. *Development*, 132: 3151-61.
- Hao F, Liu J, Zhong M et al (2018). Expression of E-cadherin, vimentin and beta-catenin in ameloblastoma and association with clinicopathological characteristics of ameloblastoma. *Int J Clin Exp Pathol*, 11(1): 199–207.
- Yan L, Li Y, Shi Z, et al (2017). The zinc finger E-box-binding homeobox 1 (Zeb1) promotes the conversion of mouse fibroblasts into functional neurons. J Biol Chem, 292(31):12959-12970.
- 27. Yoshida J, Horiuchi A, Kikuchi N, et al (2009). Changes in the expression of E-cadherin repressors, Snail, Slug, SIP1, and Twist, in the development and progression of ovarian carcinoma: the important role of Snail in ovarian tumorigenesis and progression. *Med Mol Morphol*, 42(2):82-91.
- 28. Davidson B, Trope CG, Reich R (2012). Epithelial-mesenchymal transition in ovarian carcinoma. *Front Oncol*, 2: 33.
- 29. Baulida J, Garcia de Herreros A (2015). Snail1driven plasticity of epithelial and mesenchymal cells sustains cancer malignancy. *Biochim Biophys Acta*, 1856(1):55-61.
- Carver EA, Jiang R, Lan Y, et al (2001). The mouse snail gene encodes a key regulator of the epithelial-mesenchymal transition. *Mol Cell Biol*, 21(23): 8184–8188.
- Nieto MA, Huang RY, Jackson RA, et al (2016). EMT: 2016. *Cell*, 166(1):21-45.
- 32. Lee MY, Chou CY, Tang MJ, et al (2008). Epithelial-mesenchymal transition in cervical cancer: correlation with tumor progression, epidermal growth factor receptor overexpression, and snail up-regulation. *Clin Cancer Res*, 14(15):4743-50.
- Liao TT, Yang MH (2017). Revisiting epithelialmesenchymal transition in cancer metastasis: the connection between epithelial plasticity and stemness. *Mol Oncol*, 11(7): 792-804.
- Doutre S, Omar T, Goumbri-Lompo O, et al (2018). Cervical intraepithelial neoplasia (CIN) in African women living with HIV:

role and effect of rigorous histopathological review by a panel of pathologists in the HARP study endpoint determination. *J Clin Pathol*, 71(1):40-45.

- 35. Grinstein M, Dingwall HL, Shah RR, et al (2018). A robust method for RNA extraction and purification from a single adult mouse tendon. *PeerJ*, 6: e4664.
- Faghihloo E, Akbari A, Adjaminezhad-Fard F, et al (2016).Transcriptional regulation of Ecadherin and oncoprotein E7 by valproic acid in HPV positive cell lines. *Iran J Basic Med Sci*, 19(6): 601–607.
- 37. Renthal NE, Chen CC, Williams KC, et al (2010). miR-200 family and targets, ZEB1 and ZEB2, modulate uterine quiescence and contractility during pregnancy and labor. *Proc Natl Acad Sci U S A*, 107(48):20828-33.
- 38. Hotz B, Arndt M, Dullat S, Bhargava S, et al (2007). Epithelial to mesenchymal transition: expression of the regulators snail, slug, and twist in pancreatic cancer. *Clin Cancer Res*, 13(16):4769-76.
- 39. Yu Q, Zhang K, Wang X, et al (2010). Expression of transcription factors snail, slug, and twist in human bladder carcinoma. *J Exp Clin Cancer Res*, 29(1):119.
- Jafarian M, Mozhgani SH, Patrad E, et al (2017). Evaluation of INOS, ICAM-1, and VCAM-1 gene expression: A study of adult T cell leukemia malignancy associated with HTLV-1. Arth Vind, 162(4):1009-1015.
- 41. Yang Y, Xie YJ, Xu Q, et al (2015). Downregulation of miR-1246 in cervical cancer tissues and its clinical significance. *Gynecol Oncol*, 138(3):683-8.
- 42. Sossey-Alaoui K, Plow EF (2016). miR-138-Mediated Regulation of KINDLIN-2 Expression Modulates Sensitivity to Chemotherapeutics. *Mol Cancer Res*, 14(2):228-38.
- Sossey-Alaoui K, Pluskota E, Davuluri G, et al (2014). Kindlin-3 enhances breast cancer progression and metastasis by activating Twist-mediated angiogenesis. *FASEB J*, 28(5):2260-71.
- 44. Sabbah M, Emami S, Redeuilh G, et al (2008). Molecular signature and therapeutic perspective of the epithelial-to-mesenchymal transitions in epithelial cancers. *Drug Resist Updat*, 11(4-5):123-51.

- Yilmaz M, Christofori G (2010). Mechanisms of motility in metastasizing cells. *Mol Cancer Res*, 8(5):629-42.
- Yilmaz M, Christofori G (2009). EMT, the cytoskeleton, and cancer cell invasion. *Cancer Metastasis* Rev, 28(1-2):15-33.
- Lambert AW, Pattabiraman DR, Weinberg RA (2017). Emerging Biological Principles of Metastasis. *Cell*, 168(4):670-691.
- Jolly MK, Boareto M, Huang B, et al (2015). Implications of the Hybrid Epithelial/Mesenchymal Phenotype in Metastasis. Front Oncol, 5: 155.
- De Craene B, Berx G (2013). Regulatory networks defining EMT during cancer initiation and progression. Nat Rev Cancer, 13(2):97-110.
- Ohira T, Gemmill RM, Ferguson K, et al (2003). WNT7a induces E-cadherin in lung cancer cells. *Proc Natl Acad Sci U S A*, 100(18):10429-34.
- 51. Takeyama Y, Sato M, Horio M, et al (2010). Knockdown of ZEB1, a master epithelial-tomesenchymal transition (EMT) gene, suppresses anchorage-independent cell growth of lung cancer cells. *Cancer Lett*, 296(2):216-24.
- Jean Paul Thiery, Hervé Acloque, Ruby Y J Huang, M Angela Nieto (2009). Epithelialmesenchymal transitions in development and disease. *Cell*, 139(5):871-90.
- 53. de Boer CJ, van Dorst E, van Krieken H, et al (1999). Changing roles of cadherins and catenins during progression of squamous intraepithelial lesions in the uterine cervix. *Am J Pathol*, 155(2): 505–515.
- Birchmeier W, Behrens J (1994). Cadherin expression in carcinomas: role in the formation of cell junctions and the prevention of invasiveness. *Biochim Biophys Acta*, 1198(1):11-26.
- Bartel DP (2009). MicroRNAs: target recognition and regulatory functions. *Cell*, 136(2):215-33.
- 56. Tan HX, Wang Q, Chen LZ, et al (2010). MicroRNA-9 reduces cell invasion and Ecadherin secretion in SK-Hep-1 cell. Med Oncol, 27(3):654-60.
- Wang B, Herman-Edelstein M, Koh P, et al (2010). E-cadherin expression is regulated by miR-192/215 by a mechanism that is

independent of the profibrotic effects of transforming growth factor-beta. *Diabetes*, 59(7): 1794–1802.

- Zeisberg M, Neilson EG (2009). Biomarkers for epithelial-mesenchymal transitions. J Clin Invest, 119(6): 1429–1437.
- 59. Batlle E, Sancho E, Franci C, et al (2000). The transcription factor snail is a repressor of E-cadherin gene expression in epithelial tumour cells. *Nat Cell Biol*, 2(2):84-9.
- 60. Lyons JG, Patel V, Roue NC, et al (2008). Snail up-regulates proinflammatory mediators and inhibits differentiation in oral keratinocytes. *Cancer Res*, 68(12): 4525–4530.
- 61. Weber KL, Doucet M, Price JE (2003). Renal cell carcinoma bone metastasis: epidermal growth factor receptor targeting. *Clin Orthop Relat Res*, (415 Suppl): S86-94.
- 62. Singh M, Spoelstra NS, Jean A, et al (2008). ZEB1 expression in type I vs type II endometrial cancers: a marker of aggressive disease. *Mod Pathol*, 21(7):912-23.
- 63. Karihtala P, Auvinen P, Kauppila S, et al (2013). Vimentin, zeb1 and Sip1 are up-regulated in triple-negative and basal-like breast cancers: association with an aggressive tumour phenotype. *Breast Cancer Res Treat*, 138(1):81-90.
- 64. Ran J, Lin DL, Wu RF, et al (2015). ZEB1 promotes epithelial-mesenchymal transition in cervical cancer metastasis. *Fertil Steril*, 103(6):1606.

- 65. Kudo-Saito C, Shirako H, Takeuchi T, et al (2009). Cancer metastasis is accelerated through immunosuppression during Snailinduced EMT of cancer cells. *Cancer Cell*, 15(3):195-206.
- 66. Singh M, Spoelstra NS, Jean A, et al (2008). ZEB1 expression in type I vs type II endometrial cancers: a marker of aggressive disease. *Mod Pathol*, 21(7):912-23.
- 67. Chua HL, Bhat-Nakshatri P, Clare SE, et al (2007). NF-[kappa] B represses E-cadherin expression and enhances epithelial to mesenchymal transition of mammary epithelial cells: potential involvement of ZEB-1 and ZEB-2. Oncogene, 26(5):711-24.
- Kleer CG, Zhang Y, Pan Q, et al (2004). WISP3 (CCN6) is a secreted tumor-suppressor protein that modulates IGF signaling in inflammatory breast cancer. *Neoplasia*, 6(2): 179–185.
- Burk U, Schubert J, Wellner U, et al (2008). A reciprocal repression between ZEB1 and members of the miR-200 family promotes EMT and invasion in cancer cells. *EMBO Rep*, 9(6): 582–589.
- 70. Wang F, Sloss C, Zhang X, et al (2007). Membrane-Bound Heparin-Binding Epidermal Growth Factor–Like Growth Factor Regulates E-Cadherin Expression in Pancreatic Carcinoma Cells. *Cancer Res*, 67(18):8486-93.