mRNAs in the uninduced cells. The situation during myogenesis in vitro appears to be largely comparable (Affara et al., 1977).

The third class includes those cases where a change in growth potential has occurred, for example, normal vs regenerating rat liver (Wilkes et al., 1979) and untransformed cells vs the same cells after viral transformation (Rolton et al., 1977; Williams et al., 1977) or a chemical carcinogen (Getz et al., 1977). Despite the very obvious changes in morphology, the differences between the mRNA populations are very small. Either no (Wilkes et al., 1979) or very small (Rolton et al., 1977; Williams et al., 1977; Getz et al., 1977) qualitative differences are found, while the changes in relative abundance of the mRNAs can represent no more than 4-fold changes in concentration for the bulk of the mRNA sequences. Thus it appears that pathological responses leading to apparently large morphological and metabolic changes are marked by changes in mRNA populations which are subtle by comparison with those which accompany normal differentiation processes.

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AVIAN LEUKAEMIA VIRUSES AND HAEMATOPOIETIC CELL DIFFERENTIATION

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As IN MAN, several distinct types of leukaemia occur in animals, such as lymphoid, myeloid and erythroid leukaemia. In the three best-examined animal systems, chickens, mice and cats, these disorders are usually caused by the infection or activation of C-type retroviruses. Generally, two categories of leukaemia viruses can be distinguished: (a) replication-competent viruses with a long period of latency and causing predominantly lymphatic leukaemia and (b) replicationdefective viruses (DLVs) causing acute leukaemia with a short latent period and capable of inducing in vitro transformation.

Viruses from the latter group have been intensively investigated in the last few years. In the murine system, the best studies are the Friend and the Abelson leukaemia virus. In the avian system, 7 strains have been analysed. These can be subdivided into three categories, according to the types of neoplasms they induce. Their properties are summarized in the Table (Graf & Beug, 1978). AEV-type viruses cause erythroblastosis but also sarcomas. MC29-type viruses cause myelocytomatosis but also carcinomas, and AMVtype viruses cause myeloblastosis. An analysis of haemopoietic cells transformed by these

Virus	Strain	Type of neoplasm induced	Type of haemopoietic cell transformed <i>in vitro</i> and <i>in vivo</i>	Transforming sequences	Candidate- transforming proteins	Haemopoietic target cell
AEV	f R ES4	Erythroblastosis, sarcoma	Erythroblast	erb	p751 p751	Erythroid cell
MC29	MC29 CMII OK10 MH2	Myelocytomatosis, carcinoma	Macrophage	mac	p110 ² p903 p2104 p100 ⁵	Myeloid cell
AMV	BAI/A E26	Myeloblastosis	Myeloblast	myb	?	Myeloid cell
RSV		Sarcomas	_	src	$p60^{6}$	
RAV		Lymphatic leukaemia, osteopetrosis, erythroblastosis*	B lymphoblasts		—	Bursa cell

TABLE.—Avian oncoviruses

* Long latency. ¹Hayman et al., 1979a; ²Bister et al., 1977; ³Hayman et al., 1979b; ⁴Ramsay & Hayman, in preparation; ⁵Hu et al., 1978; ⁶Brugge & Erikson, 1977.

viruses in vitro and in vivo with regard to their differentiation phenotype, revealed that cells transformed by AEV-viruses resemble erythroblasts, by MC29-viruses macrophages, and by AMV-viruses myeloblasts (Beug et al., 1979). The parameters tested included haemoglobin, histone H5, carbonic anhydrase activity and erythroid-specific cell-surface antigens as erythroid markers. As myeloid parameters, phagocytic capacity, Fc receptors, ATPase activity and myeloid-specific cell-surface antigens were tested. Cells transformed by viruses of the same type are essentially alike in all parameters tested, e.g., cells transformed by MC29, CMII, OK10 and MH2 viruses are macrophage-like. Our finding that cells transformed in vitro resembled those from leukaemic animals indicates that our marrow transformation system represents a valid model system to study virus-induced leukaemogenesis (Beug et al., 1979; Graf et al., 1979a).

Studies with cDNAs specific for AEV, MC29 and AMV RNAs demonstrated common transforming sequences in AEV viruses (erb sequences), in MC29-viruses (mac sequences) and in AMV-viruses (myb sequences) (Roussel et al., 1979; Stéhelin et al., 1979). These sequences are different from the src sequences of avian sarcoma viruses and are present in the DNA of normal cells of several avian species (Roussel et al., 1974; Stéhelin et al., 1979). In the cells transformed with DLVs in the absence of helper viruses proteins of various molecular weights have been detected by radioimmune precipitation with antisera to virus structural proteins. These proteins seem to be fusion proteins between part or all of the gag gene product (specifying viral core proteins) and a unique portion which is similar in viruses with similar biological specificity only (Kitchener & Hayman, in press). This, in addition to the co-linearity of the unique ("transforming") sequences in the RNA of AEV and MC29 with the corresponding fusion proteins (Kitchener & Hayman, in press; Mellon et al., 1978; Lai et al., 1979) suggests that they represent the transforming proteins of DLVs.

How do DLVs cause a leukaemic transformation? That they cause an arrest in maturation in their host cells is indicated by the finding that at least AEV-, AMV- and to a lesser extent MC29-transformed cells are rather immature. This concept has been confirmed by the characterization of a temperature-sensitive mutant of AEV. After shift to 41°C of erythroblasts transformed by the mutant at 35°C, there is an increase of haemoglobin synthesis and a change towards maturation in the pattern of erythroid cellsurface antigens (Graf et al., 1978b). Using the differentiation-specific antisera described we could demonstrate that haemopoietic target cells for AEV already express erythroid cell-surface antigens, whereas target cells for MC29 and AMV express myeloid antigens (Graf et al., 1976, 1978b). A simple explanation for the target-cell specificity of DLVs would therefore be that they are capable of infecting only certain types of haemopoietic cells. That this is not the case was shown by the finding that AEV replicates and induces the expression of p75 in macrophages, and that MC29 replicates and is expressed in erythroblasts (Graf *et al.*, in press).

These results are compatible with the following model of DLV-induced leukaemogenesis and target-cell specificity (Graf & Beug, 1978; Graf et al., in press; Graf et al., 1979). (1) DLVs induce a leukaemic transformation by blocking the differentiation of their haemopoietic target cells. (2) The differentiation block is the consequence of a competitive inhibition of the viral transforming protein synthesized in large amounts by a hypothetical homologous but nonidentical cellular differentiation-specific protein. (3) That a competitive inhibition of transforming protein is only possible in those cells expressing the putative cellular counterpart protein (e.g. AEV p75 in erythroblasts and MC29 p110 in immature macrophages) explains the target-cell specificity of DLVs.

We are currently trying to test our hypothesis by searching for the postulated cellular counterpart proteins of the transforming proteins of DLVs, and to study their tissue distribution.

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EARLY CARCINOGENESIS, DIFFERENTIATION AND PROMOTION

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THE NORMAL mammalian urinary bladder is lined by highly differentiated epithelium known as the urothelium, which has been described and illustrated in detail (Hicks, 1975; Hicks et al., 1974; Severs & Hicks, The markers for normal differen-1977). tiation include development of highly specialized, polyploid surface cells, with a uniquely structured surface membrane. This particular form of transitional-cell differentiation is in a state of balance which is very easily disturbed. Furthermore, the urothelium is a multipotential tissue (Hicks, 1975) and has the option of other forms of differentiation. Thus, if subjected to regular irritation

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as from a bladder calculus, the normal differentiation is disturbed and switches from transitional-cell differentiation to epidermalization with the synthesis of gross keratin plaques. Similarly, epidermalization is also produced if the animal is made vitamin Adeficient, when again the genome for normal transitional differentiation is apparently switched off, and that for differentiation to squamous metaplasia is switched on or amplified (Hicks, 1968, 1969, 1975, 1976). The frequency with which epidermalization or squamous metaplasia of the urothelium is seen suggests that the genome responsible for keratinization, though normally repressed,