



Extracellular HSPs: The Complicated Roles of Extracellular HSPs in Immunity

Stuart K. Calderwood1*, Jianlin Gong2 and Ayesha Murshid1

- Department of Radiation Oncology, Beth Israel Deaconess Medical Center, Harvard Medical School, Boston, MA, USA,
- ²Department of Medicine, Boston University Medical Center, Boston, MA, USA

Extracellular heat-shock proteins (HSPs) interact with the immune system in a very complex manner. Many such HSPs exert powerful effects on the immune response, playing both stimulatory and regulatory roles. However, the influence of the HSPs on immunity appears to be positive or negative in nature – rarely neutral. Thus, the HSPs can act as dominant antigens and can comprise key components of antitumor vaccines. They can also function as powerful immunoregulatory agents and, as such, are employed to treat inflammatory diseases or to extend the lifespan of tissue transplants. Small modifications in the cellular milieu have been shown to flip the allegiances of HSPs from immunoregulatory agents toward a potent inflammatory alignment. These mutable properties of HSPs may be related to the ability of these proteins to interact with multiple receptors often with mutually confounding properties in immune cells. Therefore, understanding the complex immune properties of HSPs may help us to harness their potential in treatment of a range of conditions.

Keywords: heat, shock, protein, immunity, immunosuppression, surface, receptors, scavenger

OPEN ACCESS

Edited by:

Swapan K. Ghosh, Indiana State University, USA

Reviewed by:

Willem Van Eden,
Utrecht University, Netherlands
Sylvie Fournel,
Strasbourg University, France
Robert T. Wheeler,
University of Maine, USA
Xiang-Yang Wang,
Virginia Commonwealth
University, USA

*Correspondence:

Stuart K. Calderwood scalderw@bidmc.harvard.edu

Specialty section:

This article was submitted to Immunotherapies and Vaccines, a section of the journal Frontiers in Immunology

> Received: 27 January 2016 Accepted: 11 April 2016 Published: 25 April 2016

Citation:

Calderwood SK, Gong J and Murshid A (2016) Extracellular HSPs: The Complicated Roles of Extracellular HSPs in Immunity. Front. Immunol. 7:159. doi: 10.3389/fimmu.2016.00159

INTRODUCTION

Molecular chaperones are proteins that permit the maturation and correct folding of most of the proteome (1, 2). As such, they are found in all cellular organisms and seem essential for cellular life. Protein folding seems to require chaperones from a number of different gene families that appear to function at various stages in a concerted folding cascade. These proteins belong to the small heat-shock protein (HSP) family including Hsp27 and the larger 70 kDa HSP family including Hsp70 as well as Hsp60, Hsp90, and Hsp110 families (3) (**Table 1**). We will discuss here, the mammalian immune responses to both prokaryotic (eubacterial) and eukaryotic HSPs under a range of contexts. The acronym HSP is derived from the early findings that some of these proteins are massively induced during proteotoxic stresses such as heat shock (4). Thus, the canonical functions of the HSP chaperones are in the folding of proteins during mRNA translation and in responding to protein unfolding crises in stressed cells (5).

However, HSPs also appear to possess functions outside the realm of protein folding, some of them acquired when they are released from cells to become extracellular HSPs (5, 6, 30, 31). HSPs have been observed in serum from human patients, pointing to their existence outside of cells, in living organisms (9). Among the first functions mooted for extracellular HSPs were in inflammation and immunity (23, 32). HSPs of each of the classes appeared to function in influencing the inflammatory

1

TABLE 1 | Immune/inflammatory roles for extracellular chaperones.

Chaperone	Pro/anti-inflammatory	Adaptive immunity?	Reference
Hsp27	Context	_	(5)
Hsp60	Context	_	(1, 6–8)
Hsp70	Context	+	(9-14)
Hsp90	Context	++	(14-18)
Hsp110	Pro	+++	(19-22)
Grp94	Pro	+	(23, 24)
Grp170	Pro	+++	(25-29)
Calreticulin	Pro	+	(23, 24)

and immunological balance in tissues (Table 1). The hypothesis of a pro-immune function for extracellular HSPs was derived primarily from studies utilizing molecular chaperone vaccines in cancer treatment (10, 23, 33). It was shown that HSPs from a number of chaperone families could be extracted from cancer cells while they were associated with a range of tumor peptide antigens (11, 33, 34). These HSP-peptide complexes could then be injected into hosts as anticancer vaccines, delivering a range of tumorderived antigens to the immune system and promoting antitumor immunity (19-22, 25-29). HSPs were, by the proponents of this approach, conventionally, viewed as playing a dominant role as promoters of immunity (32). In addition, a number of studies showed them to be pro-inflammatory mediators, and extravagant claims were made for molecular chaperones as activators of multiple facets of immunity. However, other investigators have demonstrated powerful anti-inflammatory roles for HSPs that we will discuss more fully, later in this manuscript (12, 35, 36). In addition, the properties of extracellular HSPs have now expanded to include powerful roles in processes outside the immune response. For instance, secreted Hsp90 has been shown to mediate wound healing and tumor metastasis (36, 37). Thus, extracellular HSPs appear to have come of age as major intercellular signaling molecules in biology and medicine.

Some of the issues discussed here, particularly the role of HSPs in antigen presentation, have been mentioned in a previous review (38). Here, however, we focus mainly on the potentially confound pro- and anti-inflammatory roles of HSPs and discuss how these properties can be manipulated toward clinically useful outcomes in both treatment of autoimmune conditions and in the deployment of chaperone anticancer vaccines.

RELEASE OF HSPs INTO THE EXTRACELLULAR MICROENVIRONMENT

Structural considerations would tend to make one skeptical regarding the possibility of HSP secretion into the extracellular milieu. HSP family proteins lack an N-terminal hydrophobic signal sequence, characteristic of most secreted proteins, and thus, cannot be released from cells by the conventional secretion pathways. However, a number of non-canonical secretion pathways exist, many of which are employed by cytokines to gain access to the extracellular milieu. These eccentric mechanisms include release of the polypeptides via secretory lysosomes, a pathway utilized in the release of IL-1 β from inflammatory cells (39).

Hsp70 has been shown to be secreted from a number of cells in free form by a similar pathway, through a mechanism requiring the lysosomal pH gradient (31, 40). Indeed, Hsp70 is cosecreted from cells along with the lysosome resident protein LAMP1 (31). Hsp70 is also released from a range of other cells including tumor cells, reticulocytes, peripheral blood mononuclear cells, B cells, and dendritic cells in various types of lipid vesicles [reviewed by De Maio (41) and Vega et al. (42)]. These vesicles may include a variety of lipid-bounded structures, including ectosomes that are vesicles derived from the plasma membrane and that may contain cytosolic proteins as well as exosomes. Formation of exosomes is a complex process including the internalization of portions of the plasma membrane and subsequent release of exosomes containing a variety of previously intracellular proteins, including HSPs (43). The exosomal pathway is also utilized by some cells for IL-1ß secretion (44). HSP-containing exosomes have a wide array of properties including both immunostimulatory and immunosuppressive functions, depending on the protein content of the exosome, cell of origin, and target cell (45-47).

Heat-shock proteins, therefore, can be secreted from a variety of cells in free form and in membrane-bounded particles. In addition, they can be released from cell undergoing necrotic death when membranes are disrupted, and the HSP can leak passively out of the cells (48). Hsp70 released in such a way has been shown to be strongly immunostimulatory.

HSPs AS CARRIERS OF TUMOR ANTIGENS AND MEDIATORS OF IMMUNITY

Adaptive Immunity

Molecular chaperones are unique immune modulators in that they can associate with a wide range of antigenic peptides and facilitate their delivery to antigen-presenting cells (APCs) (11, 13, 23, 33, 34). This property has proven to be desirable in the preparation of anticancer vaccines. Only a relatively small number of tumor antigens have been characterized, and we presume that this group represents a small minority of the real repertoire of unique cancer-derived antigens. Thus, chaperones, such as Hsp70, can be considered to "sample" the antigenic milieu of the malignant cell on encountering processed peptides in vivo and can be used to carry this sample into the APC during immunization (Figure 1). Such HSP-containing vaccines have proven to be highly effective in studies in experimental tumor systems in mice, in which they can lead to tumor regression associated with the generation of specific immunity (10, 13, 14, 20, 49-51). Issues in the preparation of the vaccines, which may influence the clinical effectiveness of vaccines, include the degree to which antigens can be retained by the chaperone and the affinity for the peptide during immunization and entry into APC (11, 14, 52).

Cross-presentation is a process by which extracellular antigens can gain access to the MHC class I pathway, a mechanism normally reserved for processing and presenting endogenous antigens (38). Efficient antigen cross-presentation is very important for vaccine effectiveness as MHC-I-peptide complexes permit recognition of cells bearing the complexes and killing by CD8+ cytotoxic

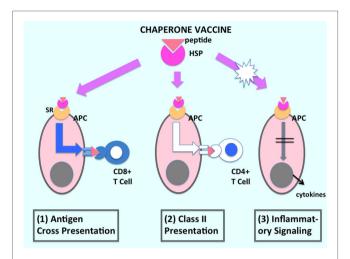


FIGURE 1 | Immune activation by HSP-based anticancer vaccines. The HSP-peptide complexes that comprise the anticancer vaccine are shown to interact with APC after vaccination of host. The vaccines can efficiently (1) cause cross-presentation of tumor antigen and, thus, prime CD8+T lymphocytes as well as activating CD4+T cells through the (2) class II pathway. However, many investigations suggest that HSPs may not have major effects on (3) inflammatory signaling and may require combination with agents with adjuvant activity or inflammatory cell killing. Gray spheres indicate nuclei.

T lymphocytes (53). Interestingly, Hsp90 appears to protect the integrity of internalized antigens associated with it, to trigger cross-presentation, and to carry antigens deep into the cell, penetrating the plasma membrane and endosomal membranes, and delivers chaperoned peptides to cytoplasmic proteasomes for processing (15, 16).

Although CD8+ T cells can be triggered by DC to recognize antigens after cross-presentation, in the absence of further signals, such T cells are unable to kill their targets. Other inputs are required for full activation (54, 55). The principal pathway used by APC for sampling external antigens is the MHC class II pathway. MHC class II molecules are found only on the surfaces of immune cells. The class II pathway involves the uptake of antigens by receptors on DC, processing of such antigens in the lysosomal compartment, transport of vesicles containing antigen-MHC-II complexes to the cell surface, and presentation to CD4+ T lymphocytes. Hsp90 is able to carry associated antigens into APCs and direct them into the class II pathway as well as facilitate crosspresentation. The choice of direction regarding entry of antigens into the class I/cross-presentation or class II pathways appears to usually depend upon the antigen-binding receptor that mediates triage between the two presentation systems (56, 57). However, Hsp90 appears neutral in this regard and increase penetration of associated peptides into either pathway (17, 18). One important activity governed by the MHC class II pathway is a process called dendritic cell licensing (54). In this mechanism, CD4+ T cells that recognize the antigen on a particular DC produce a reaction in the APC that permits it to activate CD8+ cells that interact with the same APC. Interaction of the T cell receptor on the CD4+ T cell triggers the expression of CD40 ligand (CD40L) that can bind the CD40 counter receptor on the DC and induce expression

of inflammatory cytokines, such as TNF α and IL-12 as well as stimulatory coreceptors, like CD80 and CD86 (58, 59). These coreceptors cooperate with MHC class I in fully activating the CD8+ T cell through the T cell receptor. Thus, HSP-peptide complexes become internalized and trigger both the MHC class I and II pathways and may permit DC licensing to occur (17, 18). Our findings that HSPs can facilitate uptake of individual Ova antigens through the MHC-I and MHC-II pathways suggest the possibility of HSP-antigen complex could mediate DC licensing, although this has not yet been formally proven. Homing of CD8+ T cells toward licensed DC may involve surface chemokine receptor CCR5, a process strongly stimulated by chemokines, CCL3 and CCL4 (54). It has been shown in Lewis lung carcinoma cells, in vivo, that antitumor immunity was activated along with release of chemokines CCL2, CCL5, and CCL10, by a mechanism dependent on Hsp70 and TLR4 (60).

Inflammation and Innate Immunity

On exposure to prokaryotic cells or cell products, a separate branch of immunity known as innate immunity is stimulated. In this process, molecules characteristic of individual pathogens including contrasting types of viruses and bacteria, known as pathogen-associated molecular patterns (PAMPs) herald the infection and prime the immune response (61). Then, PAMPs interact with specific receptors on macrophages or DC, known as pattern recognition receptors (PRR), and trigger innate immunity. Best known among the PRR are the toll-like receptors (TLR) that can couple binding of individual PAMPs to intracellular signaling pathways and gene expression programs (62, 63). Most notable among the mechanisms triggered by PRR occupation are the NKK and MAP-kinase pathways that influence inflammatory transcription through activation of factors, such as NFkB and IRF3 (64). This process can lead to synthesis of costimulatory molecules, such as CD80, and activating cytokines such as TNF α and IL-12 that synergize with MHC class I signaling in generation of active and long lived CTL (54). It is not clear to what extent HSPs derived from prokaryotes might function as PAMPs, although their extreme conservation across all cellular species would seem to argue against this. HSPs derived from mycobacteria are, however, recognized by the mammalian immune response and invoke powerful immunity to the extent that they have been described as superantigens (65). The mechanisms by which prokaryotic Hsp60 activates immunity are not clear but could involve PRR, such as TLRs, or other mechanisms.

It has also been shown that some molecules released from damaged and dying cells, such as uric acid and high mobility group box 1 protein (HMGM1), may trigger a form of sterile innate immunity, and such molecules are referred to as damage-associated molecular patterns (DAMPs), in order to suggest a functional similar to PAMPs (66, 67). Thus, DAMPs are thought to trigger innate immunity by binding to PRR and triggering inflammatory signaling cascades. Hsp70 was widely reported to function as a DAMP and to trigger innate immunity through the TLR2 and TLR4 pathways (32). Although this field has run into some controversy, the majority of findings in studies carried out *in vivo* over the past 15 years suggested that Hsp70, through interaction with TLR4, could potentially act as

a DAMP (68). This area has been recently reviewed in depth (69). In addition to Hsp70, extracellular Hsp27 has recently been shown to cause both inflammatory and anti-inflammatory effects (5).

SOME ANTI-INFLAMMATORY AND IMMUNOREGULATORY EFFECTS OF HSPs

Although prokaryotic HSPs can trigger a powerful immunodominant response in animals, most reports indicate that their effects are generally not pro-inflammatory, and the antibodies and T cells activated in the response had anti-inflammatory properties (69-71). Curiously, the epitopes that T cells respond to in mycobacterial Hsp60 were conserved with mammalian HSPs, and such cells recognized and responded to epitopes in the mammalian proteins. Prokaryotic Hsp60, therefore, did not seem to act as a PAMP (7). In addition, although HSPs were shown to interact with TLRs, such PRR often provoked anti-inflammatory signaling (8). For instance, Hsp60-derived peptides interacted with TLR2 on regulatory T cells (Tregs) leading to an immunosuppressive response. In addition, purified mycobacterial Hsp70 inhibited the maturation of DC (72, 73). Intracellular HSP levels were shown to increase in inflamed tissues and HSP-derived peptides expressed on the cell surface and appeared to activate Treg responses, thus mediating immunoregulatory functions (74). No studies, to date, have shown direct interactions between HSPs and TLRs, and in fact, attempts to show such binding have been negative (73, 75). The extracellular influences on TLR activity that have been reported may, therefore, be indirect and likely dependent on primary interactions of the HSP with other receptors on immune cells, such as the scavenger receptors (SR), followed by recruitment of TLR (64). The powerful immune effects of non-mammalian Hsp60 may also involve mechanisms independent of TLRs, and it has been suggested that the immune response may be genetically programmed to respond to such chaperones (76).

Interestingly, some of the studies applying HSP vaccines to cancer therapy indicated that, although there was significant activation of antitumor CTL by these agents, these were followed by a delayed Treg response. These findings suggest contrasting effects of the vehicle (HSP) and cargo (antigenic peptide) components of chaperone vaccines on immunity. These data might be interpreted as, suggesting that, while tumor antigens chaperoned by the HSPs trigger antitumor immunity, processed peptides from the HSP component of the vaccine led to a suppression of immunity. Using the chaperone vaccines at lower doses appeared to favor induction of CTL over the immunoregulatory response, perhaps by reducing the levels of HSP-derived peptides below a threshold (77, 78). It would seem that most chaperone vaccines, although efficiently triggering external tumor antigen presentation, do not deliver the inflammatory signal required to overcome antigenic tolerance (Figure 1). Such vaccines might be improved by use of adjuvants or pro-inflammatory forms of therapy, as discussed below.

WHEN HSPs BECOME PRO-INFLAMMATORY FACTORS

One notable finding observed in multiple investigations of HSPs was that, even in studies where an inflammatory response to HSPs was not detected, the chaperones could strongly amplify responses to PAMPs, such as LPS (79). Mycobacterial Hsp65, a protein discussed in the last section as provoking generally immunomodulatory responses, when covalently fused to antigenic polypeptides produced a potent vaccine that generated effective CTL even in the absence of adjuvant (80). In addition, the combination of Hsp70 elevation in target tissues with therapies leading to necrotic cell killing led to a profound stimulation of inflammation and CTL killing that could lead to tumor rejection (81). This approach involved, after elevation of tissue Hsp70, targeting the normal tissues of origin with treatments that led to inflammatory modes of cell killing. This combined treatment resulted in the regression of distant, transplanted tumors (81). The findings observed in these studies were that, for instance, in prostatic tissue, cell killing by fusogenic viruses in the presence of elevated Hsp70 led to induction of the cytokines IL-6 and TGF-β, resulting in generation of highly inflammatory IL-17 and tumor rejection by antigen-specific CTL (82). This effect seemed to depend on generation of IL-6 by the combination of high tissue levels of Hsp70 and inflammatory death. However, in similar studies on pancreatic tissues, combination of Hsp70 and lytic virus failed to generate IL-6 and led to generation of a Treg response and continued growth of pancreatic carcinoma (82). Thus, the balance between immunoregulatory and immunogenic responses of Hsp70 appears to be poised on a knife-edge, influenced by the tissue type and mode of cell killing.

It is well known that the mode of cell death has a powerful influence on inflammation and immunity in interacting APC (83). For instance, when cells die by an apoptotic mechanism, their intracellular contents remain enveloped by an external membrane and, thus, are not released into the environment to trigger inflammation (Figure 2). In addition to this, many apoptotic cells expose "eat me" signals, such as the phospholipid phosphatidylserine on the surface, triggering engulfment by macrophages and leading to immunosuppression (84, 85). Additional anti-inflammatory signals emanating from apoptotic cells may include the release of AMP from the apoptotic cell (86). In necrotic cell death, cell membranes become permeabilized, the intracellular milieu becomes externalized, and DAMPs, such as HMGB1, urate, and nucleic acids, released in this way become accessible to detection by neighboring macrophages or DC (85). It should be noted that the response of phagocytic cells to apoptotic bodies is complex and depends on the nature of the dead or dying cells and the surface densities of "eat me" or "don't eat me" signaling molecules that are cell specific. In addition, in late apoptotic cells that have failed to be phagocytosed at an early stage, membranes become permeabilized. Therefore, such late apoptotic cells acquire some of the properties of necrotic cells, permitting release of DAMPs and switching the effects of the cell corpses on engulfing phagocytes toward a more immunogenic influence (85, 87).

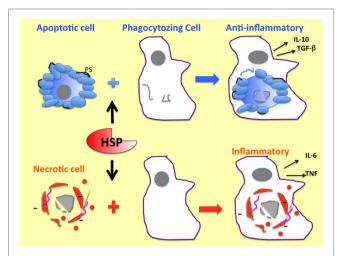


FIGURE 2 | Contrasting effects of cell corpses resulting from apoptotic and necrotic cell killing on immunity. Apoptotic cells, depicted here with surface blebs, externalize the phospholipid phosphatidylserine to the outer leaflet of the plasma membrane leading to anti-inflammatory signaling and suppression of immunity. Necrotic cells depicted here with compromised membranes release DAMPS, such as HMGB1 (small orange spheres), and acquire net negative charge. Nucleic acids (pink lines) become located to the cell surface. Necrotic cells often trigger release of inflammatory cytokines, such as IL-6 and TNF α . HSPs, such as Hsp70, trigger the phagocytosis of dead mammalian cells as shown here, although the mechanisms involved in such stimulation are currently not clear. Gray spheres indicate nuclei.

It is not clear in which way extracellular HSPs might synergize with the extruded contents of necrotic cells in activating immunity, although increased capacity for engulfment by immune cells which is triggered by Hsp70 might amplify inflammatory functions (88-90). The earlier studies of Todryk et al., in fact, indicated that extracellular Hsp70 decrease DC maturation and increase capacity for engulfment of antigenic materials by the immature APC (91). Phagocytosis is a complex process and involves the formation of a broad synapse type structure between the engulfing cell and the mammalian cell body of prokaryotic cells. Multiple receptors appear to make contact between the cells. Interestingly, in *C. elegans* this process has been shown to require the surface receptor CED-1 (92). Both, mammalian scavenger receptor associated with endothelial cells (SREC-I) and cluster of differentiation 91 (CD91) possess sequence similarities to this protein, suggesting some involvement in phagocytosis and a potential mechanism for HSPs in this process (93). In addition, necrotic cells can supply signals to augment the immune effects of HSPs through release of HMGB1, a ligand for TLR4, and externalized nucleic acids that might interact with TLR3 or TLR9 and trigger inflammatory signaling (85) (Figures 2 and 3).

POTENTIAL MECHANISMS – CELL SURFACE RECEPTORS AND CELL SIGNALING PATHWAYS

Most evidence suggests that the biological effects of extracellular HSPs are mediated through cell surface receptors (24, 94)

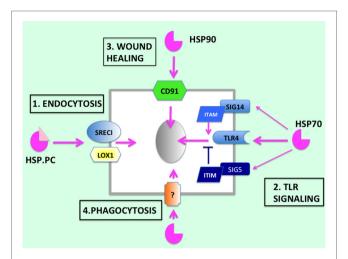


FIGURE 3 | HSP receptors. HSP receptors include SREC-I and LOX-1 that mediate (1) endocytosis and, thus, presentation of antigens to APC. Hsp70 can also (2) trigger signaling through the TLR4 pathway in a range of cells. (It is not clear whether HSPs can interact directly with TLRs or whether the primary interactions are through other receptors.) Effects of Hsp70 on TLR4 signaling can be modulated by binding to either Siglec-14 that activates TLR4 signaling or other Siglec family members, such as Siglec-5, that activate the pathway. Hsp90 can also (3) activate wound healing responses, which may play key roles in inflammation, through binding to CD91. Hsp70 triggers (4) phagocytosis, a property that may be important in its immune functions, through currently unknown mechanisms. Gray ovals indicate nuclei.

(Figure 3). Thus, some of the clues to the properties of extracellular HSPs may lie within the interactions of these chaperones with these receptors and the unleashing of their embedded signaling cascades. The first surface receptor reported to bind to HSPs was CD91 also known as Low density lipoprotein receptorrelated protein 1 (LRP1), the alpha-2-macroglobulin receptor (A2MR), and apolipoprotein E receptor (APOER) (24). This high molecular weight receptor was shown originally to bind to LDL and, as its possession of multiple names suggests, a wide range of (over 30) other extracellular structures (94). CD91 is expressed most abundantly in vascular smooth muscle cells and in hepatocytes. There has been some skepticism regarding its expression in APC, such as DC, and, therefore, its potential role in immune responses involving the HSPs (95). However, there seems little doubt that CD91/LRP1 is a bona fide HSP receptor, and this protein appears to play key roles in, for instance, responses to Hsp90α in the wound healing response and in tumor metastasis (36, 37).

It was next shown that Hsp70 could associate with lectintype oxidized LDL receptor (LOX-1), a receptor found on the surfaces of human DC and that such binding could mediate cross-presentation of peptide cargo associated with this HSP, leading to CD8+ lymphocyte-mediated immunity (75). LOX-1 is the product of the oxidized low-density lipoprotein receptor 1 (*ORL1*) gene expressed most abundantly in vascular endothelial cells, macrophages, and DC (96). Although LOX-1 belongs to the C-type lectin receptor family, it also clusters with the SR, a group of proteins that have, in common, the ability to bind to covalently modified LDL molecules, such as oxidized LDL and

acetylated LDL (97). Furthermore, exploration of the SR family indicated significant interaction of Hsp70 with at least two other members, including SREC-I, encoded by the SCARF1 gene and Stabilin-1/FEEL-1 (encoded by the STAB1 gene) (73, 98). SREC-I/SCARF1 was able to bind to Hsp70 and Hsp90 in DC and mediated cross-presentation of associated tumor antigens, leading to activated CTL (13, 17). Murine bone marrow derived DC appeared to utilize both LOX-1 and SREC-I/SCARF1 in interacting with Hsp90-peptide complexes (17, 75). In addition, SREC-I was also shown to mediate the uptake of antigens chaperoned by Hsp90 into the Class II pathway and stimulate activation of CD4+ T lymphocytes (18). The SR are regarded as receptors that respond to cellular debris including cell bodies, the remains of endogenous proteins spilled into the extracellular milieu, and the residue from invading pathogens. As such, the SR might be regarded as good choices for receptors with which APC might sample the extracellular environment (93). Stabilin-1 appears to be only sparsely expressed on the surface of DC and appears to function mostly in intracellular trafficking (99). Thus, there appeared little doubt that that the SR could bind to HSP-PC and mediate presentation of peptide cargo (17, 18). However, the nature of the receptors that might respond to HSPs and modify the inflammatory response still required investigation.

Many reports suggested that TLRs, particularly TLR2 and TLR4, might mediate inflammatory responses to HSPs in an up or down manner (32, 76). However, as mentioned above, attempts to demonstrate direct binding of HSPs to TLRs have not been generally successful (73, 75). Thus, HSP interaction with TLRs is likely to be an indirect one. It was shown, recently, that SREC-I/ SCARF1 could interact with TLR4 on the surface of mouse macrophages (64). Interestingly, both LPS and Hsp90 could mediate this complex interaction and lead to the sequestration of TLR4 in lipid raft domains (64). In the case of LPS, encasement of SREC-I-TLR4 complexes within lipid rafts was required for activation of signaling through the NFκB and MAP-kinase cascades, resulting in inflammatory cytokine secretion. Significantly, exposure to Hsp90, although leading to SREC-I-TLR4 colocalization, failed to trigger the inflammatory cascade through this mechanism. Alternative pathways of inflammatory suppression by Hsp90 may, thus, be involved (64).

Recent studies have also suggested that another receptor family, sialic acid-binding immunoglobulin-like lectins (Siglecs) may participate in inflammatory responses resulting from binding to HSPs (100). These receptors have been shown to bind to conjugated sugar residues in the cell coats of adjacent cells and generally suppress inflammation. Suppression of inflammation by these receptors involves immunoreceptor tyrosine-based inhibitory motif (ITIM) sequences in the intracellular domain that recruit anti-inflammatory proteins after ligand-triggered tyrosine phosphorylation (100). Upon activation, Siglec receptors have been shown to directly associate with TLRs and inhibit TLR-mediated activation of inflammatory signaling cascades, such as the NFκB pathway (101). Interestingly, recently, it was shown that two human Siglecs, Siglec-5 and Siglec-14 each, bind to Hsp70 (102). What was most intriguing about these interactions was

that, while Siglec-5 contained the expected ITIM domain and repressed anti-inflammatory signaling through quenching NF κ B, Siglec-14 appeared to have acquired a domain that binds proteins, such as DAP12, containing an intracellular immunoreceptor tyrosine-based activating motif (ITAM) that stimulated release of inflammatory cytokines (102, 103). Differential expression of members of the Siglec family might, thus, either amplify or suppress the HSP regulated activities of cell surface TLR and inflammatory signaling through mediation of the NF κ B pathway. A further property of extracellular HSPs mentioned above, that may play a key role in their immune functions, is their ability to trigger phagocytosis. However, the mechanisms and receptors involved in this process have not been well characterized and further experiments will be required to understand the mechanisms involved (Figure 3).

Thus, responses to extracellular HSPs appear to involve the cooperative or confounding outputs of a range of cell surface receptors that together may determine their influence on immune reactions within tissues. To date, however, we have not been able to identify a dedicated high-affinity HSP receptor. Dedicated receptors, such as the insulin receptor, are often able to pick up tiny concentrations of ligands, often with affinities in the range of 10^{-9} – 10^{-10} M, with exquisite selectivity. For HSPs, we have identified, so far, only "hand me down" receptors that also bind a large range of other ligands with moderate affinity. It remains to be determined if dedicated HSP receptors will be found in immune cells of mammalian species.

CONCLUSION

The intracellular roles of HSPs in protein folding have been conserved since the dawn of cellular life (3). However, the HSPs also appear to have acquired key roles in the immune systems of animals early during evolution, and these roles are preserved in modern animal species (104). Such HSPs can capture intracellular antigens and present them to APC, mediating the crosspriming of recipient cells. In addition, HSPs, when processed and presented on the APC cell surface, can activate Treg cells and inhibit immunity and inflammation. Their effects on the immune system are, thus, bivalent. In the presence of PAMPs or tissues undergoing necrosis, Hsp70, in particular, becomes a strong inflammatory agent. The precise nature of the responses elicited by extracellular HSPs may, therefore, depend upon the particular tissue milieu within which they are released and the identities of the receptors on the surfaces of immune cells that encounter them.

OUTSTANDING QUESTIONS AND POTENTIAL ADVANCES

(1) Many of the immune-active HSPs are members of different gene families, with only a modicum of structural similarity (Table 1). However, each chaperone appears to be recognized by similar families of receptors, including SR and the LDL receptors, despite their sequence dissimilarity (94). Understanding the structural basis for extracellular HSP

- recognition by these and potentially new classes of receptors would be an advance in determining the roles of the chaperones in immunity and predicting novel recognition structures. However, this has been an open question for a number of years now and research is ongoing.
- (2) It is becoming apparent that members of individual HSP gene families have distinct properties. For instance, only Hsp90 α is secreted from cells and the other major isoform, Hsp90 β , is mostly retained within the cell (36). It will be enlightening to learn whether such specificity in extracellular function holds for other HSP families.
- (3) We have, in recent years, acquired some understanding of how these "leaderless" proteins gain access to the extracellular microenvironment in intact cells (see Release of HSPs into the Extracellular Microenvironment), although it will be invaluable to gain a more concerted understanding of mechanisms of HSP secretion.
- (4) We will continue the search for HSP receptors in immune cells, their expression patterns and their connection to inflammatory and anti-inflammatory responses to both endogenous HSPs and prokaryotic paralogs (94). It is likely that a major key to understanding how chaperones trigger immune responses and the direction of such responses depends on understanding the combinatorial effects of the multiple receptor families.
- (5) It seems clear that many HSPs, although capable of efficient transport of antigenic peptides within chaperone vaccines, do not on their own deliver a second signal

REFERENCES

- Haldar S, Gupta AJ, Yan X, Miličić G, Hartl FU, Hayer-Hartl M. Chaperoninassisted protein folding: relative population of asymmetric and symmetric GroEL:GroES complexes. J Mol Biol (2015) 427:2244–55. doi:10.1016/j. imb.2015.04.009
- Ellis RJ. Protein misassembly: macromolecular crowding and molecular chaperones. Adv Exp Med Biol (2007) 594:1–13. doi:10.1007/978-0-387-39975-1_1
- 3. Lindquist S, Craig EA. The heat-shock proteins. *Annu Rev Genet* (1988) **22**:631–77. doi:10.1146/annurev.ge.22.120188.003215
- Richter K, Haslbeck M, Buchner J. The heat shock response: life on the verge of death. Mol Cell (2010) 40:253–66. doi:10.1016/j.molcel.2010.10.006
- Salari S, Seibert T, Chen YX, Hu T, Shi C, Zhao X, et al. Extracellular Hsp27 acts as a signaling molecule to activate NF-kappaB in macrophages. *Cell Stress Chaperones* (2013) 18:53–63. doi:10.1007/s12192-012-0356-0
- Tian J, Guo X, Liu XM, Liu L, Weng QF, Dong SJ, et al. Extracellular Hsp60 induces inflammation through activating and up-regulating TLRs in cardiomyocytes. *Cardiovasc Res* (2013) 98:391–401. doi:10.1093/cvr/cvt047
- Anderton SM, van der Zee R, Prakken B, Noordzij A, van Eden W. Activation of T cells recognizing self 60-kD heat shock protein can protect against experimental arthritis. *J Exp Med* (1995) 181:943–52. doi:10.1084/ jem.181.3.943
- Zanin-Zhorov A, Cahalon L, Tal G, Margalit R, Lider O, Cohen IR. Heat shock protein 60 enhances CD4+ CD25+ regulatory T cell function via innate TLR2 signaling. J Clin Invest (2006) 116:2022–32. doi:10.1172/ JCI28423
- Wright BH, Corton JM, El-Nahas AM, Wood RF, Pockley AG. Elevated levels
 of circulating heat shock protein 70 (Hsp70) in peripheral and renal vascular
 disease. Heart Vessels (2000) 15:18–22. doi:10.1007/s003800070043
- Weng D, Song B, Koido S, Calderwood SK, Gong J. Immunotherapy of radioresistant mammary tumors with early metastasis using molecular chaperone vaccines combined with ionizing radiation. *J Immunol* (2013) 191:755–63. doi:10.4049/jimmunol.1203286

for APC maturation, thus reducing their effectiveness as stand-alone agents. In order to break immune tolerance to tumor antigens, chaperone vaccines might be best deployed in cancer therapy in combination with PAMPs, such as double stranded RNA or unmethylated CpG motifs or checkpoint inhibitors such as anti-CTLA4 (105, 106). Alternatively, vaccines could be used with agents, such as ionizing radiation, that can cause necrotic killing and subsequent inflammatory effects in tumors (including release of intracellular DAMPs) thus leading to immune rejection of the cancers (10).

AUTHOR CONTRIBUTIONS

SC, JG, and AM contributed in writing the text of the review.

ACKNOWLEDGMENTS

The authors thank the Department of Radiation Oncology, BIDMC, Boston, for generous support. The authors also thank Roxanne Calderwood for preparation of cartoon figures. This work was supported by NIH research grants, RO-1CA119045 and RO-1CA094397.

FUNDING

This study was supported by a grant (RO-1CA119045) from the Office of Extramural Research, National Institutes of Health.

- Weng D, Calderwood SK, Gong J. Preparation of a heat-shock protein 70-based vaccine from DC-tumor fusion cells. *Methods Mol Biol* (2011) 787:255–65. doi:10.1007/978-1-61779-295-3_19
- Borges TJ, Wieten L, van Herwijnen MJ, Broere F, van der Zee R, Bonorino C, et al. The anti-inflammatory mechanisms of Hsp70. Front Immunol (2012) 3:95. doi:10.3389/fimmu.2012.00095
- Gong J, Zhu B, Murshid A, Adachi H, Song B, Lee A, et al. T cell activation by heat shock protein 70 vaccine requires TLR signaling and scavenger receptor expressed by endothelial cells-1. *J Immunol* (2009) 183:3092–8. doi:10.4049/ jimmunol.0901235
- Gong J, Zhang Y, Durfee J, Weng D, Liu C, Koido S, et al. A heat shock protein 70-based vaccine with enhanced immunogenicity for clinical use. *J Immunol* (2010) 184:488–96. doi:10.4049/jimmunol.0902255
- Ichiyanagi T, Imai T, Kajiwara C, Mizukami S, Nakai A, Nakayama T, et al. Essential role of endogenous heat shock protein 90 of dendritic cells in antigen cross-presentation. *J Immunol* (2010) 185:2693–700. doi:10.4049/ jimmunol.1000821
- Imai T, Kato Y, Kajiwara C, Mizukami S, Ishige I, Ichiyanagi T, et al. Heat shock protein 90 (Hsp90) contributes to cytosolic translocation of extracellular antigen for cross-presentation by dendritic cells. *Proc Natl Acad Sci U S A* (2011) 108:16363–8. doi:10.1073/pnas.1108372108
- Murshid A, Gong J, Calderwood SK. Heat shock protein 90 mediates efficient antigen cross presentation through the scavenger receptor expressed by endothelial cells-I. *J Immunol* (2010) 185:2903–17. doi:10.4049/ jimmunol.0903635
- Murshid A, Gong J, Calderwood SK. Hsp90-peptide complexes stimulate antigen presentation through the class II pathway after binding scavenger receptor SREC-I. *Immunobiology* (2014) 219:924–31. doi:10.1016/j. imbio.2014.08.001
- Manjili MH, Park J, Facciponte JG, Subjeck JR. Hsp110 induces "danger signals" upon interaction with antigen presenting cells and mouse mammary carcinoma. *Immunobiology* (2005) 210:295–303. doi:10.1016/j. imbio.2005.04.002

- Manjili MH, Wang XY, Chen X, Martin T, Repasky EA, Henderson R, et al. Hsp110-HER2/neu chaperone complex vaccine induces protective immunity against spontaneous mammary tumors in HER-2/neu transgenic mice. *J Immunol* (2003) 171:4054–61. doi:10.4049/jimmunol.171.8.4054
- Wang XY, Chen X, Manjili MH, Repasky E, Henderson R, Subjeck JR. Targeted immunotherapy using reconstituted chaperone complexes of heat shock protein 110 and melanoma-associated antigen gp100. Cancer Res (2003) 63:2553–60.
- Wang XY, Kazim L, Repasky EA, Subjeck JR. Characterization of heat shock protein 110 and glucose-regulated protein 170 as cancer vaccines and the effect of fever-range hyperthermia on vaccine activity. *J Immunol* (2001) 166:490–7. doi:10.4049/jimmunol.166.1.490
- Srivastava P. Roles of heat-shock proteins in innate and adaptive immunity. Nat Rev Immunol (2002) 2:185–94. doi:10.1038/nri749
- Binder RJ, Han DK, Srivastava PK. CD91: a receptor for heat shock protein gp96. Nat Immunol (2000) 1:151–5. doi:10.1038/77835
- Manjili MH, Park JE, Facciponte JG, Wang XY, Subjeck JR. Immunoadjuvant chaperone, GRP170, induces 'danger signals' upon interaction with dendritic cells. *Immunol Cell Biol* (2006) 84:203–8. doi:10.1111/j.1440-1711.2006.01418.x
- Wang XY, Arnouk H, Chen X, Kazim L, Repasky EA, Subjeck JR. Extracellular targeting of endoplasmic reticulum chaperone glucose-regulated protein 170 enhances tumor immunity to a poorly immunogenic melanoma. *J Immunol* (2006) 177:1543–51. doi:10.4049/jimmunol.177.3.1543
- Park JE, Facciponte J, Chen X, MacDonald I, Repasky EA, Manjili MH, et al. Chaperoning function of stress protein grp170, a member of the Hsp70 superfamily, is responsible for its immunoadjuvant activity. *Cancer Res* (2006) 66:1161–8. doi:10.1158/0008-5472.CAN-05-2609
- Gao P, Sun X, Chen X, Wang Y, Foster BA, Subjeck J, et al. Secretable chaperone Grp170 enhances therapeutic activity of a novel tumor suppressor, mda-7/ IL-24. Cancer Res (2008) 68:3890–8. doi:10.1158/0008-5472.CAN-08-0156
- Zuo D, Yu X, Guo C, Yi H, Chen X, Conrad DH, et al. Molecular chaperoning by glucose-regulated protein 170 in the extracellular milieu promotes macrophage-mediated pathogen sensing and innate immunity. FASEB J (2012) 26:1493–505. doi:10.1096/fj.11-197707
- Hance MW, Dole K, Gopal U, Bohonowych JE, Jezierska-Drutel A, Neumann CA, et al. Secreted Hsp90 is a novel regulator of the epithelial to mesenchymal transition (EMT) in prostate cancer. *J Biol Chem* (2012) 287:37732–44. doi:10.1074/jbc.M112.389015
- Mambula SS, Calderwood SK. Heat shock protein 70 is secreted from tumor cells by a nonclassical pathway involving lysosomal endosomes. *J Immunol* (2006) 177:7849–57. doi:10.4049/jimmunol.177.11.7849
- Asea A, Kraeft SK, Kurt-Jones EA, Stevenson MA, Chen LB, Finberg RW, et al. Hsp70 stimulates cytokine production through a CD14-dependant pathway, demonstrating its dual role as a chaperone and cytokine. *Nat Med* (2000) 6:435–42. doi:10.1038/74697
- Yu X, Guo C, Yi H, Qian J, Fisher PB, Subjeck JR, et al. A multifunctional chimeric chaperone serves as a novel immune modulator inducing therapeutic antitumor immunity. *Cancer Res* (2013) 73:2093–103. doi:10.1158/0008-5472.CAN-12-1740
- Ishii T, Udono H, Yamano T, Ohta H, Uenaka A, Ono T, et al. Isolation of MHC class I-restricted tumor antigen peptide and its precursors associated with heat shock proteins Hsp70, Hsp90, and gp96. *J Immunol* (1999) 162:1303–9.
- Spiering R, van der Zee R, Wagenaar J, van Eden W, Broere F. Mycobacterial and mouse Hsp70 have immuno-modulatory effects on dendritic cells. *Cell Stress Chaperones* (2013) 18:439–46. doi:10.1007/s12192-012-0397-4
- Jayaprakash P, Dong H, Zou M, Bhatia A, O'Brien K, Chen M, et al. Hsp90alpha and Hsp90beta together operate a hypoxia and nutrient paucity stress-response mechanism during wound healing. *J Cell Sci* (2015) 128:1475–80. doi:10.1242/jcs.166363
- Annamalai B, Liu X, Gopal U, Isaacs JS. Hsp90 is an essential regulator of EphA2 receptor stability and signaling: implications for cancer cell migration and metastasis. *Mol Cancer Res* (2009) 7:1021–32. doi:10.1158/1541-7786. MCR-08-0582
- Murshid A, Gong J, Calderwood SK. The role of heat shock proteins in antigen cross presentation. Front Immunol (2012) 3:63. doi:10.3389/ fimmu.2012.00063
- Andrei C, Dazzi C, Lotti L, Torrisi MR, Chimini G, Rubartelli A. The secretory route of the leaderless protein interleukin 1beta involves exocytosis of

- endolysosome-related vesicles. *Mol Biol Cell* (1999) **10**:1463–75. doi:10.1091/mbc.10.5.1463
- Murshid A, Calderwood SK, editors. Pathways of Hsp70 Release: Lessons from Cytokine Secretion. Dordrecht: Springer (2012). p. 103–14.
- De Maio A. Extracellular heat shock proteins, cellular export vesicles, and the Stress Observation System: a form of communication during injury, infection, and cell damage. It is never known how far a controversial finding will go! Dedicated to Ferruccio Ritossa. Cell Stress Chaperones (2011) 16:235–49. doi:10.1007/s12192-010-0236-4
- 42. Vega VL, Rodríguez-Silva M, Frey T, Gehrmann M, Diaz JC, Steinem C, et al. Hsp70 translocates into the plasma membrane after stress and is released into the extracellular environment in a membrane-associated form that activates macrophages. *J Immunol* (2008) **180**:4299–307. doi:10.4049/jimmunol.180.6.4299
- Chalmin F, Ladoire S, Mignot G, Vincent J, Bruchard M, Remy-Martin JP, et al. Membrane-associated Hsp72 from tumor-derived exosomes mediates STAT3-dependent immunosuppressive function of mouse and human myeloid-derived suppressor cells. *J Clin Invest* (2010) 120:457–71. doi:10.1172/JCI40483
- 44. Piccioli P, Rubartelli A. The secretion of IL-1beta and options for release. Semin Immunol (2013) 25:425–9. doi:10.1016/j.smim.2013.10.007
- Taieb J, Chaput N, Ménard C, Apetoh L, Ullrich E, Bonmort M, et al. A novel dendritic cell subset involved in tumor immunosurveillance. *Nat Med* (2006) 12:214–9. doi:10.1038/nm1356
- Zitvogel L, Regnault A, Lozier A, Wolfers J, Flament C, Tenza D, et al. Eradication of established murine tumors using a novel cell-free vaccine: dendritic cell-derived exosomes. *Nat Med* (1998) 4:594–600. doi:10.1038/ nm0598-594
- 47. Gastpar R, Gehrmann M, Bausero MA, Asea A, Gross C, Schroeder JA, et al. Heat shock protein 70 surface-positive tumor exosomes stimulate migratory and cytolytic activity of natural killer cells. *Cancer Res* (2005) 65:5238–47. doi:10.1158/0008-5472.CAN-04-3804
- Mambula SS, Calderwood SK. Heat induced release of Hsp70 from prostate carcinoma cells involves both active secretion and passive release from necrotic cells. *Int J Hyperthermia* (2006) 22:575–85. doi:10.1080/02656730 600976042
- Wang XY, Kazim L, Repasky EA, Subjeck JR. Immunization with tumorderived ER chaperone grp170 elicits tumor-specific CD8+ T-cell responses and reduces pulmonary metastatic disease. *Int J Cancer* (2003) 105:226–31. doi:10.1002/ijc.11058
- Yu X, Wang XY. Engineering Grp170-based immune modulators for cancer immunotherapy. *Oncoimmunology* (2013) 2:e24385. doi:10.4161/ onci.24385
- Manjili MH, Henderson R, Wang XY, Chen X, Li Y, Repasky E, et al. Development of a recombinant Hsp110-HER-2/neu vaccine using the chaperoning properties of Hsp110. Cancer Res (2002) 62:1737–42.
- Murshid A, Gong J, Stevenson MA, Calderwood SK. Heat shock proteins and cancer vaccines: developments in the past decade and chaperoning in the decade to come. *Expert Rev Vaccines* (2011) 10:1553–68. doi:10.1586/ erv.11.124
- Rock KL, Shen L. Cross-presentation: underlying mechanisms and role in immune surveillance. *Immunol Rev* (2005) 207:166–83. doi:10.1111/j.0105-2896.2005.00301.x
- Kurts C, Robinson BW, Knolle PA. Cross-priming in health and disease. Nat Rev Immunol (2010) 10:403–14. doi:10.1038/nri2780
- Gottschalk C, Mettke E, Kurts C. The role of invariant natural killer T cells in dendritic cell licensing, cross-priming, and memory CD8(+) T cell generation. Front Immunol (2015) 6:379. doi:10.3389/fimmu.2015.00379
- Burgdorf S, Lukacs-Kornek V, Kurts C. The mannose receptor mediates uptake of soluble but not of cell-associated antigen for cross-presentation. *J Immunol* (2006) 176:6770–6. doi:10.4049/jimmunol.176.11.6770
- Burgdorf S, Schuette V, Semmling V, Hochheiser K, Lukacs-Kornek V, Knolle PA, et al. Steady-state cross-presentation of OVA is mannose receptor-dependent but inhibitable by collagen fragments. *Proc Natl Acad Sci U S A* (2010) 107:E48–9. doi:10.1073/pnas.1000598107
- Bennett SR, Carbone FR, Karamalis F, Flavell RA, Miller JF, Heath WR. Help for cytotoxic-T-cell responses is mediated by CD40 signalling. *Nature* (1998) 393:478–80. doi:10.1038/30996

- Bennett SR, Carbone FR, Karamalis F, Miller JF, Heath WR. Induction of a CD8+ cytotoxic T lymphocyte response by cross-priming requires cognate CD4+ T cell help. J Exp Med (1997) 186:65–70. doi:10.1084/jem.186.1.65
- Chen T, Guo J, Han C, Yang M, Cao X. Heat shock protein 70, released from heat-stressed tumor cells, initiates antitumor immunity by inducing tumor cell chemokine production and activating dendritic cells via TLR4 pathway. J Immunol (2009) 182:1449–59. doi:10.4049/jimmunol.182.3.1449
- Bianchi ME. DAMPs, PAMPs and alarmins: all we need to know about danger. J Leukoc Biol (2007) 81:1–5. doi:10.1189/jlb.0306164
- Medzhitov R, Preston-Hurlburt P, Janeway CA Jr. A human homologue of the *Drosophila* toll protein signals activation of adaptive immunity. Nature (1997) 388:394–7. doi:10.1038/41131
- 63. Pandey S, Kawai T, Akira S. Microbial sensing by toll-like receptors and intracellular nucleic acid sensors. *Cold Spring Harb Perspect Biol* (2015) 7:a016246. doi:10.1101/cshperspect.a016246
- Murshid A, Gong J, Prince T, Borges TJ, Calderwood SK. Scavenger receptor SREC-I mediated entry of TLR4 into lipid microdomains and triggered inflammatory cytokine release in RAW 264.7 cells upon LPS activation. *PLoS One* (2015) 10:e0122529. doi:10.1371/journal.pone.0122529
- Zugel U, Kaufmann SH. Immune response against heat shock proteins in infectious diseases. *Immunobiology* (1999) 201:22–35. doi:10.1016/ S0171-2985(99)80044-8
- Kono H, Rock KL. How dying cells alert the immune system to danger. Nat Rev Immunol (2008) 8:279–89. doi:10.1038/nri2215
- Rock KL, Kataoka H, Lai JJ. Uric acid as a danger signal in gout and its comorbidities. Nat Rev Rheumatol (2013) 9:13–23. doi:10.1038/ nrrheum.2012.143
- Calderwood SK, Murshid A, Gong J. Heat shock proteins: conditional mediators of inflammation in tumor immunity. Front Immunol (2012) 3:75. doi:10.3389/fimmu.2012.00075
- van Eden W, Spiering R, Broere F, van der Zee R. A case of mistaken identity: Hsps are no DAMPs but DAMPERs. *Cell Stress Chaperones* (2012) 17:281–92. doi:10.1007/s12192-011-0311-5
- Moudgil KD, Chang TT, Eradat H, Chen AM, Gupta RS, Brahn E, et al. Diversification of T cell responses to carboxy-terminal determinants within the 65-kD heat-shock protein is involved in regulation of autoimmune arthritis. J Exp Med (1997) 185:1307–16. doi:10.1084/jem.185.7.1307
- van Eden W, van der Zee R, Prakken B. Heat-shock proteins induce T-cell regulation of chronic inflammation. Nat Rev Immunol (2005) 5:318–30. doi:10.1038/nri1593
- Motta A, Schmitz C, Rodrigues L, Ribeiro F, Teixeira C, Detanico T, et al. *Mycobacterium tuberculosis* heat-shock protein 70 impairs maturation of dendritic cells from bone marrow precursors, induces interleukin-10 production and inhibits T-cell proliferation in vitro. Immunology (2007) 121:462–72. doi:10.1111/j.1365-2567.2007.02564.x
- Theriault JR, Adachi H, Calderwood SK. Role of scavenger receptors in the binding and internalization of heat shock protein 70. *J Immunol* (2006) 177:8604–11. doi:10.4049/jimmunol.177.12.8604
- 74. de Wolf C, van der Zee R, den Braber I, Glant T, Maillere B, Favry E, et al. An arthritis suppressive and regulatory T cell-inducing CD4 T cell epitope is functional in the context of HLA-restricted T cell responses. *Arthritis Rheumatol* (2016). 68: 639–47. doi: 10.1002/art.39444
- Delneste Y, Magistrelli G, Gauchat J, Haeuw J, Aubry J, Nakamura K, et al. Involvement of LOX-1 in dendritic cell-mediated antigen cross-presentation. Immunity (2002) 17:353–62. doi:10.1016/S1074-7613(02)00388-6
- 76. Cohen IR. Biomarkers, self-antigens and the immunological homunculus. *J Autoimmun* (2007) **29**:246–9. doi:10.1016/j.jaut.2007.07.016
- Udono H, Srivastava PK. Heat shock protein 70-associated peptides elicit specific cancer immunity. JExp Med (1993) 178:1391–6. doi:10.1084/ jem.178.4.1391
- Liu Z, Li X, Qiu L, Zhang X, Chen L, Cao S, et al. Treg suppress CTL responses upon immunization with Hsp gp96. Eur J Immunol (2009) 39:3110–20. doi:10.1002/eji.200939593
- Wallin RP, Lundqvist A, Moré SH, von Bonin A, Kiessling R, Ljunggren HG. Heat-shock proteins as activators of the innate immune system. *Trends Immunol* (2002) 23:130–5. doi:10.1016/S1471-4906(01)02168-8
- Suzue K, Young RA. Adjuvant-free Hsp70 fusion protein system elicits humoral and cellular immune responses to HIV-1 p24. *J Immunol* (1996) 156:873–9.

- Daniels GA, Sanchez-Perez L, Diaz RM, Kottke T, Thompson J, Lai M, et al. A simple method to cure established tumors by inflammatory killing of normal cells. *Nat Biotechnol* (2004) 22:1125–32. doi:10.1038/ nbt1007
- Kottke T, Sanchez-Perez L, Diaz RM, Thompson J, Chong H, Harrington K, et al. Induction of Hsp70-mediated Th17 autoimmunity can be exploited as immunotherapy for metastatic prostate cancer. *Cancer Res* (2007) 67:11970–9. doi:10.1158/0008-5472.CAN-07-2259
- 83. Melcher A, Gough M, Todryk S, Vile R. Apoptosis or necrosis for tumor immunotherapy: what's in a name? *J Mol Med (Berl)* (1999) 77:824–33. doi:10.1007/s001099900066
- Marino G, Kroemer G. Mechanisms of apoptotic phosphatidylserine exposure. Cell Res (2013) 23:1247–8. doi:10.1038/cr.2013.115
- Poon IK, Hulett MD, Parish CR. Molecular mechanisms of late apoptotic/ necrotic cell clearance. Cell Death Differ (2010) 17:381–97. doi:10.1038/ cdd.2009.195
- Yamaguchi H, Maruyama T, Urade Y, Nagata S. Immunosuppression via adenosine receptor activation by adenosine monophosphate released from apoptotic cells. Elife (2014) 3:e02172. doi:10.7554/eLife.02172
- 87. Gaipl US, Kuenkele S, Voll RE, Beyer TD, Kolowos W, Heyder P, et al. Complement binding is an early feature of necrotic and a rather late event during apoptotic cell death. Cell Death Differ (2001) 8:327–34. doi:10.1038/ si.cdd.4400826
- Massa C, Guiducci C, Arioli I, Parenza M, Colombo MP, Melani C. Enhanced efficacy of tumor cell vaccines transfected with secretable Hsp70. Cancer Res (2004) 64:1502–8. doi:10.1158/0008-5472.CAN-03-2936
- Massa C, Melani C, Colombo MP. Chaperon and adjuvant activity of Hsp70: different natural killer requirement for cross-priming of chaperoned and bystander antigens. *Cancer Res* (2005) 65:7942–9.
- Wang R, Town T, Gokarn V, Flavell RA, Chandawarkar RY. Hsp70 enhances macrophage phagocytosis by interaction with lipid raft-associated TLR-7 and upregulating p38 MAPK and PI3K pathways. *J Surg Res* (2006) 136:58–69. doi:10.1016/j.jss.2006.06.003
- Todryk S, Melcher AA, Hardwick N, Linardakis E, Bateman A, Colombo MP, et al. Heat shock protein 70 induced during tumor cell killing induces Th1 cytokines and targets immature dendritic cell precursors to enhance antigen uptake. *J Immunol* (1999) 163:1398–408.
- Zhou Z, Hartwieg E, Horvitz HR. CED-1 is a transmembrane receptor that mediates cell corpse engulfment in C. elegans. Cell (2001) 104:43–56. doi:10.1016/S0092-8674(01)00190-8
- Murshid A, Borges TJ, Calderwood SK. Emerging roles for scavenger receptor SREC-I in immunity. Cytokine (2015) 75(2):256–60. doi:10.1016/j. cvto.2015.02.009
- Calderwood SK, Theriault J, Gray PJ, Gong J. Cell surface receptors for molecular chaperones. *Methods* (2007) 43:199–206. doi:10.1016/j. ymeth.2007.06.008
- Berwin B, Delneste Y, Lovingood RV, Post SR, Pizzo SV. SREC-I, a type F scavenger receptor, is an endocytic receptor for calreticulin. *J Biol Chem* (2004) 279:51250–7. doi:10.1074/jbc.M406202200
- Moriwaki H, Kume N, Sawamura T, Aoyama T, Hoshikawa H, Ochi H, et al. Ligand specificity of LOX-1, a novel endothelial receptor for oxidized low density lipoprotein. *Arterioscler Thromb Vasc Biol* (1998) 18:1541–7. doi:10.1161/01.ATV.18.10.1541
- Pluddemann A, Neyen C, Gordon S. Macrophage scavenger receptors and host-derived ligands. *Methods* (2007) 43:207–17. doi:10.1016/j. vmeth.2007.06.004
- Murshid A, Gong J, Ahmad R, Borges TJ, Calderwood SK. Scavenger receptor SREC-I promotes double stranded RNA-mediated TLR3 activation in human monocytes. *Immunobiology* (2015) 220:823–32. doi:10.1016/j. imbio.2014.12.011
- Kzhyshkowska J, Gratchev A, Martens JH, Pervushina O, Mamidi S, Johansson S, et al. Stabilin-1 localizes to endosomes and the trans-Golgi network in human macrophages and interacts with GGA adaptors. *J Leukoc Biol* (2004) 76:1151–61. doi:10.1189/jlb.0504300
- 100. Ali SR, Fong JJ, Carlin AF, Busch TD, Linden R, Angata T, et al. Siglec-5 and Siglec-14 are polymorphic paired receptors that modulate neutrophil and amnion signaling responses to group B Streptococcus. J Exp Med (2014) 211:1231–42. doi:10.1084/jem.20131853

- 101. Chen GY, Brown NK, Wu W, Khedri Z, Yu H, Chen X, et al. Broad and direct interaction between TLR and Siglec families of pattern recognition receptors and its regulation by Neu1. *Elife* (2014) 3:e04066. doi:10.7554/ eLife.04066
- 102. Fong JJ, Sreedhara K, Deng L, Varki NM, Angata T, Liu Q, et al. Immunomodulatory activity of extracellular Hsp70 mediated via paired receptors Siglec-5 and Siglec-14. EMBO J (2015) 34(22):2775–88. doi:10.15252/embj.201591407
- 103. Calderwood SK, Murshid A. Siglecs take a TOLL on inflammation: deciphering the Hsp70 riddle. EMBO J (2015) 34:2733–4. doi:10.15252/ embj.201593172
- Robert J. Evolution of heat shock protein and immunity. *Dev Comp Immunol* (2003) 27:449–64. doi:10.1016/S0145-305X(02)00160-X
- 105. Okuya K, Tamura Y, Saito K, Kutomi G, Torigoe T, Hirata K, et al. Spatiotemporal regulation of heat shock protein 90-chaperoned self-DNA and CpG-oligodeoxynucleotide for type I IFN induction via targeting

- to static early endosome. JImmunol (2010) **184**:7092–9. doi:10.4049/jimmunol.1000490
- Callahan MK, Wolchok JD, Allison JP. Anti-CTLA-4 antibody therapy: immune monitoring during clinical development of a novel immunotherapy. Semin Oncol (2010) 37:473–84. doi:10.1053/j.seminoncol.2010.09.001

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2016 Calderwood, Gong and Murshid. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.