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## Two Cases of Bacteremia Due to Roseomonas mucosa

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*Roseomonas* is a genus of pink-pigmented nonfermentative bacilli. These slow-growing, gram-negative cocobacilli form pink-colored colonies on sheep blood agar. They differ from other pink-pigmented nonfermenters, including *Methylobacterium*, in morphology, biochemical characteristics, and DNA sequence. *Roseomonas* strains are rarely isolated in clinical laboratories; therefore, we report two cases in order to improve our ability to identify these pathogens. We isolated two strains of *Roseomonas mucosa* from the venous blood cultures of two patients, an 84-yr-old woman with common bile duct obstruction and a 17-yr-old male with acute myeloid leukemia who had an indwelling central-venous catheter for chemotherapy. The isolated strains were confirmed as *R. mucosa* by 16S rRNA sequencing.

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Key Words: Pink-pigmented nonfermenters, Roseomonas mucosa, 16S rRNA sequencing

## INTRODUCTION

The genus *Roseomonas* was initially identified as 42 strains of pink-pigmented gram-negative bacteria. From these strains, six genomospecies, all of which were considered *Roseomonas* [1], were proposed. *Roseomonas* constituted three named genomospecies, i.e., *Roseomonas gilardii, R. cervicalis,* and *R. fauriae,* and three unnamed genomospecies [2]. From 1992 to 2002, 36 strains of *Roseomonas* were isolated from blood cultures at the MD Anderson Cancer Center, and a new species, *R. mucosa,* was proposed on the basis of 16S rRNA sequencing and phylogenetic analysis [3]. Currently, the genus *Roseomonas* constitutes 15 valid species, including *R. aquatica, R. aerilata, R. cervicalis, R. gilardii, R. lacus, R. mucosa, R. terrae, R. stagni, R. vinacea, R. fauriae,* and other unnamed genomospecies.

*Roseomonas* are slow-growing, gram-negative bacteria. The colonies they form on sheep blood agar (SBA) are slightly fastidious, very mucoid, and almost runny. Microscopically, *Roseomonas* are nonvacuolated, plump coccobacilli that form mostly pairs and short chains. Although *Roseomonas* show low human pathogenicity, they can cause systemic infection in patients with underlying diseases or those who are immunocompromised [1, 4-7]. We isolated *R. mucosa* from the venous blood of two patients. We present the findings of these two cases and review the literature of *R. mucosa* bacteremia.

## **CASE REPORT**

#### 1. Case 1

An 84-yr-old woman was admitted with jaundice and abdominal pain for three days. She had a history of cholecystitis and cholangitis and underwent cholecystectomy and percutaneous transhepatic biliary drainage. At admission, the values of AST, ALT, total bilirubin, and direct bilirubin were 73 IU/L, 44 IU/L, 14.93 mg/dL, and 13.5 mg/dL, respectively. Three sets of blood cultures were obtained before administration of antibiotics. Common bile duct obstruction was diagnosed by abdominal ultrasound and endoscopic retrograde cholangiopancreatography.

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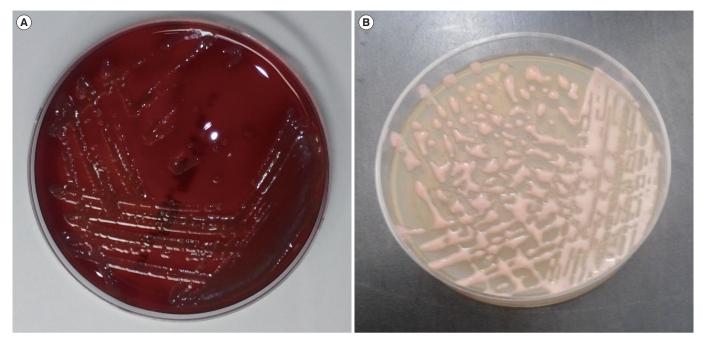


Fig. 1. (A) Roseomonas on blood agar showing nonhemolytic pale pink colonies after 72 hr incubation. (B) Roseomonas on potato cornmeal Tween 80 agar showing nonhemolytic, pink-colored, runny mucoid colonies after 24 hr incubation.

After three-day incubation of blood culture bottles, positive signals were observed, and light pink-colored colonies formed on blood agar that was subcultured from one of the bottles (Fig. 1A). The isolate was identified as *R. gilardii* by the Vitek2 system (bioMerieux, Marcy l'Etoile, France). Matrix assisted laser desorption ionization – time of flight mass spectrometry (MALDI-TOF) analysis by Vitek MS (bioMerieux) failed because of the incomplete database, but 16S rRNA gene sequencing confirmed *R. mucosa*. After three days of cefotaxime and metronidazole combination therapy, the patient showed no signs of infection, including fever and chilling sensation. She was eventually discharged with no complications.

#### 2. Case 2

A 17-yr-old man was admitted to our hospital for consolidation chemotherapy. He had been diagnosed as having acute myeloid leukemia and received induction chemotherapy with no complications. During the consolidation chemotherapy, the patient exhibited fever and elevated C-reactive protein level with no focal infection. Ceftazidime and aminoglycoside combination as empirical antibiotics were initiated. Despite antibiotic therapy, fever persisted, and the C-reactive protein level further increased. Central catheter-related infection was suspected, and vancomycin was added to the antibiotic therapy after collection of two sets of central blood samples and three sets of venous blood samples. We found an isolate that was later identified as *R. gilardii* by the Vitek2 system but *R. mucosa* by 16S rRNA sequencing. Therefore, we reported this isolate as *R. mucosa*. Vancomycin and carbapenem combination therapy was continued, even though the patient did not exhibit significant clinical signs. The patient was discharged with no complications caused by the infection.

## DISCUSSION

*Roseomonas* is found easily in the environment, including the soil, water, and air. The mechanism of infection caused by *Roseomonas* and its clinical significance are not well understood. Studies have proposed that *Roseomonas* can be found in turbid water from dirty or contaminated faucets [8, 9]. However, although *Roseomonas* species have been isolated from specimens, only approximately 60% of isolates are clinically significant because of their low pathogenicity [4]. After their initial identification, several case reports have described clinically significant infections due to *Roseomonas* spp. Because of its low pathogenicity, *Roseomonas* can cause infection in patients with underlying disease or immunosuppression. Several cases have described catheter-related bacteremia with immunosuppression [5, 10-12], bacteremia during chemotherapy for leukemia [13, 14], and peritonitis during continuous ambulatory peritoneal di-

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**Table 1.** Biochemical profiles of the isolated *R. mucosa* strains and the number of positive biochemical reactions of the reference strain

|                             | Patient 1 | Patient 2 | MDA5527 group<br>(n=22) [3] |
|-----------------------------|-----------|-----------|-----------------------------|
| Enzyme activities           |           |           |                             |
| Arginine decarboxylase      | -         | -         | 0 (0.0%)                    |
| Catalase                    | +         | +         | 22 (100.0%)                 |
| Lysine decarboxylase        | -         | -         | n/a                         |
| Ornithine decarboxylase     | -         | -         | n/a                         |
| Oxidase                     | +         | +         | 6 (27.3%)                   |
| Other activities            |           |           |                             |
| Citrate                     | -         | +         | 20 (90.9%)                  |
| Esculin hydrolysis          | -         | -         | 0 (0.0%)                    |
| Hydrogen sulfide production | -         | -         | n/a                         |
| Urea utilization            | +         | +         | 21 (95.5%)                  |
| Acidification of sugars     |           |           |                             |
| Arabinose                   | -         | -         | 22 (100.0%)                 |
| Glucose                     | -         | -         | 15 (68.2%)                  |
| Inositol                    | -         | -         | n/a                         |
| Melibiose                   | -         | -         | n/a                         |
| Raffinose                   | -         | -         | n/a                         |
| Rhamnose                    | -         | -         | n/a                         |
| Sorbitol                    | -         | -         | n/a                         |
| Sucrose                     | +         | -         | n/a                         |

Abbreviations: MDA, MD Anderson Cancer Center; n/a, not available.

alysis with end-stage renal disease [8, 15]. However, *R. mucosa* infection in immunocompetent individuals has also been reported [6, 16-18].

In our cases, *R. mucosa* was isolated by using conventional, routine methods. We used BACTEC Plus Aerobic/F and BACTEC Lytic/10 Anaerobic/F culture bottles and the BD Diagnostic System for blood culture. If a bottle showed a positive signal, subcultures were placed on SBA and MacConkey agar plates, which were incubated at 35°C in a 5% CO<sub>2</sub> incubator. Finally, the colonies were identified by using the Vitek2 system. Biochemical characterization of the isolated strains from our two patients showed positivity for catalase and urease, negativity for methanol oxidation, and a lack of UV light absorption [3, 8], as summarized in Table 1. In order to identify prominent colonies of *Rosemonas* species, we attempted to subculture the isolates on potato cornmeal Tween 80 agar, which our institution uses for fungal subculture [19]; we found noticeable pink-colored runny mucoid colonies (Fig. 1B).

For molecular confirmation of the isolated strains, we per-

formed 16S rRNA gene sequence analysis. The 16S rRNA gene was amplified in order to obtain a complete sequence, and the obtained sequence was then compared with the consensus sequence provided by GenBank (National Center for Biotechnology Information [NCBI]). The sequences were compared by using the Basic Local Alignment Search Tool provided by NCBI, and the result showed that the 16S rRNA gene sequence of *R. mucosa* with the highest similarity to the GenBank source matched with GenBank accession number NR\_028857.1. The isolated *Roseomonas* species were susceptible to aminoglycosides (e.g., amikacin), carbapenems (e.g., imipenem), and fluoroquinolones (e.g., ciprofloxacin). The isolates had intermediate susceptibility to cefotaxime but were resistant to ampicillin and ceftazidime. This overall susceptibility pattern was consistent with the previous findings [2, 3, 7].

In summary, we report two cases of bacteremia due to *R. mucosa*, which is not exactly identified by conventional method of bacterial identification but confirmed by 16S rRNA gene sequence analysis.

# Authors' Disclosures of Potential Conflicts of Interest

No potential conflicts of interest relevant to this article were reported.

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