

Research Article

Baoyan Ji, Xiongfei Cheng, Xiaojun Cai, Chuiyan Kong, Qingyan Yang, Ting Fu, Yahang Wang, Ying Song*

CK20 mRNA expression in serum as a biomarker for colorectal cancer diagnosis: A meta-analysis

<https://doi.org/10.1515/med-2017-0050>

received April 1, 2017; accepted April 25, 2017

Abstract: The aim of this study was to evaluate the diagnostic value of serum CK20 mRNA as a biomarker for colorectal cancer diagnosis by meta-analysis.

Clinical studies related to serum CK20 mRNA expression for colorectal cancer diagnosis were searched in the databases of Pubmed, Cochrane Library, Embase, ISI Web of Knowledge, CNKI and Wanfang. The number of true positive (tp), false positive (fp), false negative (fn) and true negative (tn) of the original included publications were extracted by two reviewers independently. The diagnostic sensitivity, specificity, positive likelihood ratio (+LR), negative likelihood ratio (-LR), diagnostic odds ratio (DOR) and area under the symmetric ROC curve (AUC) were pooled by random or fixed effect method according to the statistical heterogeneity among the studies.

After screening the databases, nineteen publications met the inclusion criteria and were finally included in this meta-analysis. The diagnostic sensitivity and specificity were pooled by random effect model ($I^2 > 50\%$). The pooled

diagnostic sensitivity and specificity of CK20 mRNA in serum as biomarker for colorectal cancer were 0.49 (95% CI: 0.46 to 0.51) and 0.94 (95% CI: 0.92-0.96) respectively. The pooled +LR and -LR were 10.90 (95% CI: 5.78 to 20.55) and 0.51 (95% CI: 0.45 to 0.57) respectively by random-effect method. The pooled DOR was 22.31 with the 95% CI of 11.65 to 42.71. The pooled area under the ROC curve (AUC) was 0.72 for CK20 mRNA in serum as a biomarker for colorectal cancer diagnosis. Conclusion Serum CK20 mRNA expression was significantly elevated in colorectal cancer patients which could be a promising serum biomarker for colorectal cancer diagnosis with high specificity.

Keywords: Colorectal cancer; CK20; Biomarker; Diagnosis; Meta-analysis.

1 Introduction

Colorectal cancer accounting for 1,000,000 new cases per year is one of the most diagnosed malignant carcinoma world-wide [1]. Its incidence ranks the third just behind lung cancer and breast cancer. In China, colorectal cancer is one of the leading causes of cancer related death with an incidence rate of 13.6/100,000 for males and 9.2/100,000 for females which is still on the rise [2]. The prognosis of colorectal cancer is relatively good for early stage patients. However, the long-term survival rate is very low for advanced metastatic disease. Therefore, early diagnosis is important for improving the survival of patients with colorectal cancer. Serum biomarkers for colorectal cancer screening or diagnosis is generally used clinically such as CEA, CA199 and *et al* [3, 4]. Cytokeratin 20 (CK20) protein is widely used as a serum biomarker for cancer screening or diagnosis. However, the efficacy of using the serum level of CK20mRNA as a biomarker for colorectal cancer diagnosis is not clear according to the previously published studies with small sample sizes. Therefore, we

*Corresponding author: Ying Song, Department of Oncology Center, The People's Hospital of Shiyan City Hubei Province 442000 China., E-mail: songying319@126.com

Baoyan Ji, Department of Oncology, Qinghai Province People's Hospital, Xining Qinghai 81000 China

Xiongfei Cheng, Xiaojun Cai, Ting Fu, Department of Oncology Center, The People's Hospital of Shiyan City Hubei Province 442000 China

Chuiyan Kong, Department of Oncology, The People's Hospital of Wuzhou Guangxi Province 543000, China

Qingyan Yang, Department of Oncology, The 188th Hospital of PLA, Chaozhou Guangdong Province 521000, China

Yahang Wang, Department of General surgery, People's Hospital Fenghua District Ningbo City Zhejiang Province, 315500 PR China

Baoyan Ji and Xiongfei Cheng contribute equally to this work.

performed this meta-analysis to further evaluate the diagnostic value of CK20mRNA level in serum for colorectal cancer.

2 Material and methods

2.1 Studies selection

Clinical studies related to serum CK20 mRNA for colorectal cancer diagnosis were searched in the databases of Pubmed, Cochrane Library, Embase, ISI Web of Knowledge, CNKI, and Wanfang. The “colorectal cancer/CRC”, “colon cancer”, “rectal cancer”, “colorectal carcinoma”, “colorectal neoplasm”, “colorectal tumor”, “colon carcinoma”, “rectal carcinoma”, “colon neoplasm”, “rectal neoplasm”, “colon tumor”, “rectal tumor”, “CK20”, “cytokertation 20” were used as the MeSH and free text words when searching the databases. The inclusion criteria were: colorectal cancer patients with histology or cytology conformation; the CK20 mRNA expression in serum was detected by real-time quantitative PCR (RT-qPCR) or other confirmed methods; the patients distribution of true positive(tp), false positive(fp), false negative(fn) and true negative(tn) in colorectal patients and controls could be drawn from the original included publications. All potential relevant studies were assessed in details, and all citations of the included articles were further evaluated in order to identify additional suitable studies. The publication searching was done by two reviewers independently and cross checked.

2.2 Data extraction from the original studies

The data and main information of each include publications were extracted by two authors independently and checked by the third reviewer. The sample size, first and corresponding authors, number of subjects distributed to tp, fp, fn and tn were carefully extracted from the included publications.

2.3 Statistical analysis

The statistical analysis was done by MetaDiSc1.4 (<http://www.biomedsearch.com>) software. Chi-square and I^2 test were used to calculate the statistical heterogeneity among

the included studies. Fixed or random effect method was used to pool the sensitivity, specificity, +LR, -LR, and DOR according to the statistical heterogeneity results. The publication bias was evaluated by funnel plot and Egger’s line regression test.

3 Results

3.1 General character of the included 19 publications

After searching the related electronic databases, 19 studies [5-23] met the inclusion criteria and were finally included in this meta-analysis. The publication searching procedure is demonstrated in Figure 1. The publication year ranges from 1999 to 2012. Serum CK20 mRNA was examined by RT-qPCR and FQ-PCR assay in 15 and 4 studies respectively. The general information of the included 19 publications is demonstrated in Table 1.

3.2 Pooled sensitivity

Due to significant statistical heterogeneity across the studies, the diagnostic sensitivity was pooled by random effect model ($I^2=85.6\%$). The pooled diagnostic sensitivity of CK20 mRNA in serum as a biomarker for colorectal cancer was 0.49 with the 95% CI of 0.46 to 0.51 through random effect method, Figure 2.

3.3 Pooled specificity

Statistical significance was also found in diagnostic specificity ($I^2=82.2\%$). The data was pooled by random effect method. The pooled specificity was 0.94 with the 95%CI of 0.92-0.96, Figure 3.

3.4 Pooled +LR and -LR

Significant heterogeneity was found in +LR and -LR ($I^2=74.9\%$ and 81.9%). The pooled +LR and -LR were 10.90 (95%CI:5.78 to 20.55) and 0.51 (95%CI:0.45 to 0.57) respectively for serum CK20 mRNA in the diagnosis of colorectal cancer, Figure 4.

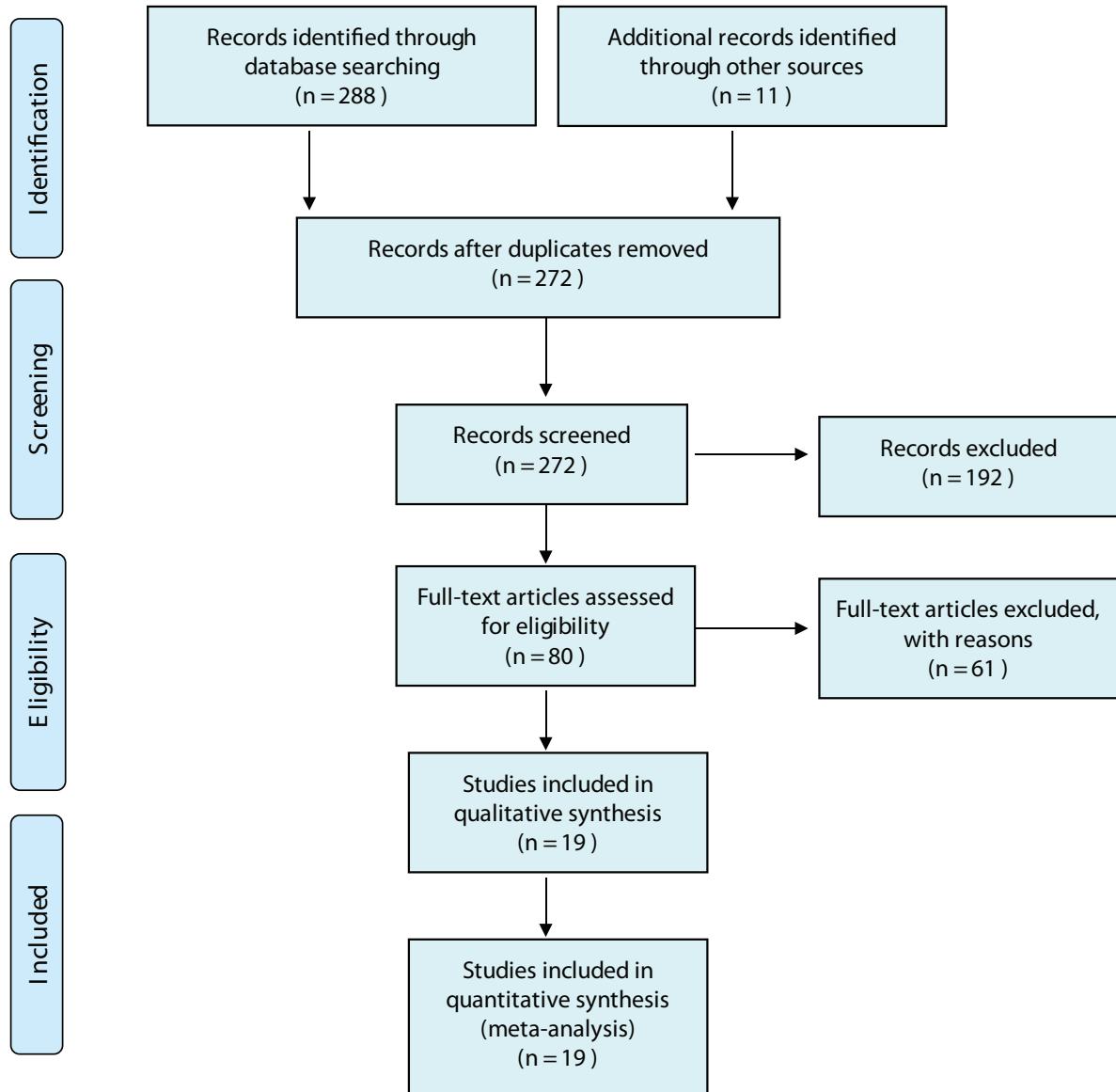


Figure 1: The publication searching flow chart

3.5 Pooled DOR

With statistical heterogeneity, the DOR was calculated by random effect method ($I^2=64.2\%$). The pooled DOR was 22.31 with the 95% CI of 11.65 to 42.71, Figure 5.

3.6 Pooled ROC

The pooled area under the ROC curve was 0.72 for CK20 mRNA in serum as a biomarker for colorectal cancer, Figure 6.

3.7 Subgroup analysis

We performed subgroup analysis according to control type and CK20 mRNA detection methods. The pooled sensitivity, specificity, +LR, -LR, DOR and AUC of the population based control group, hospital based control group, RT-qPCR group and FQ-PCR group are shown in Table 2. The diagnostic value changed a little for different subgroup analysis.

Table 1: The general characters of the included 19 publications

Author	Year	Sample size	Distribution				Control type	Country	Methods
			TP	FP	FN	TN			
Richard Q[5]	1999	170	35	1	65	69	Population based	Britain	RT-qPCR
Chausovsky G[6]	1999	57	15	0	20	22	Population based	Israel	RT-qPCR
Lin GL[7]	2002	67	23	0	24	20	Mixed	China	RT-qPCR
Sun JW[8]	2004	62	19	0	23	20	Population based	China	RT-qPCR
Cui M[9]	2004	122	62	0	40	20	Population based	China	RT-qPCR
Zeng QG[10]	2004	89	41	6	12	30	Hospital based	China	RT-qPCR
Yin HZ[11]	2005	57	34	0	13	10	Population based	China	FQ-PCR
Yin HZ[11]	2005	57	34	0	13	10	Hospital based	China	FQ-PCR
Dandachi N[17]	2005	134	46	17	36	35	Mixed	Austria	RT-qPCR
Guo J[18]	2005	50	31	0	9	10	Hospital based	China	FQ-PCR
Xu D[19]	2006	198	46	2	122	28	Population based	China	RT-qPCR
Katsumata K[20]	2006	54	18	0	22	14	Population based	Japan	RT-qPCR
Wang ZC[12]	2007	211	35	0	101	73	Mixed	China	FQ-PCR
Shen C[21]	2008	281	74	21	82	104	Mixed	China	RT-qPCR
Lagoudianakis EE[22]	2009	58	28	0	14	14	Population based	Greece	RT-qPCR
Wong SC[23]	2009	342	62	3	70	207	Mixed	China	FQ-PCR
Wu F[13]	2009	142	44	0	48	50	Population based	China	RT-qPCR
Chen P[14]	2009	56	18	0	18	20	Population based	China	RT-qPCR
Chen P[14]	2009	57	18	1	18	10	Hospital based	China	RT-qPCR
He Q[15]	2010	151	49	0	52	50	Population based	China	RT-qPCR
Li LH[16]	2012	110	48	3	32	27	Hospital based	China	RT-qPCR
Li LH[16]	2012	110	48	0	32	30	Population based	China	RT-qPCR

3.8 Publication bias

The Deeks' funnel plot showed no significant publication bias in the aspect of serum CK20 mRNA as a biomarker for colorectal cancer diagnosis ($t=0.56$, $P=0.58$), Figure 7.

4 Discussion

Colorectal cancer, also known as bowel cancer and colon cancer, is the development of cancer from the colon or

rectum. It is one of the leading causes of cancer related death world-wide [24, 25]. Population based colorectal cancer screening for high risk patients is a key method for identifying early stage patients and improving the prognosis. The National Comprehensive Cancer Network (NCCN) colorectal cancer guideline recommends fecal occult blood tests plus colonoscopy for colorectal cancer screening [26, 27]. Colonoscopy examination for colorectal cancer diagnosis has high specificity and sensitivity. However, colonoscopy examination is mildly invasive, expensive, and a complex operation procedure, which

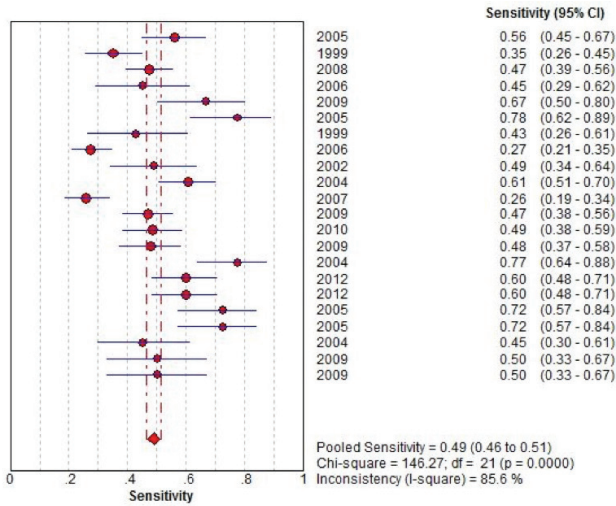


Figure 2: The forest plot of diagnostic sensitivity for serum CK20 mRNA in diagnosis of colorectal cancer

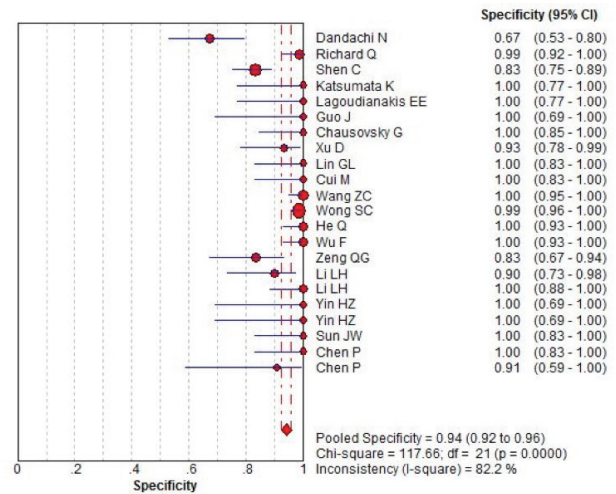


Figure 3: The forest plot of diagnostic specificity for serum CK20 mRNA in diagnosis of colorectal cancer

is not suitable for population based screening. It was believed that the ideal colorectal cancer screening or early diagnosis method should be of high sensitivity, high specificity, non-invasive, and easy to perform. Serum biomarkers are easy to access, non-invasive, less expensive and easy to test, which makes them suitable for colorectal cancer screening or diagnosis [28].

CK20 is a member of the intermediate filament protein family and a prominent component of the intestinal epithelium. CK20 expression is confined to gastrointestinal epithelium, urothelium, and Merkel cells of the epidermis, as well as malignancies that originate from the aforementioned sites [29, 30]. CK20 mRNA expression in serum detected by RT-qPCR was widely investigated for colorectal cancer screening. For most studies published in the previ-

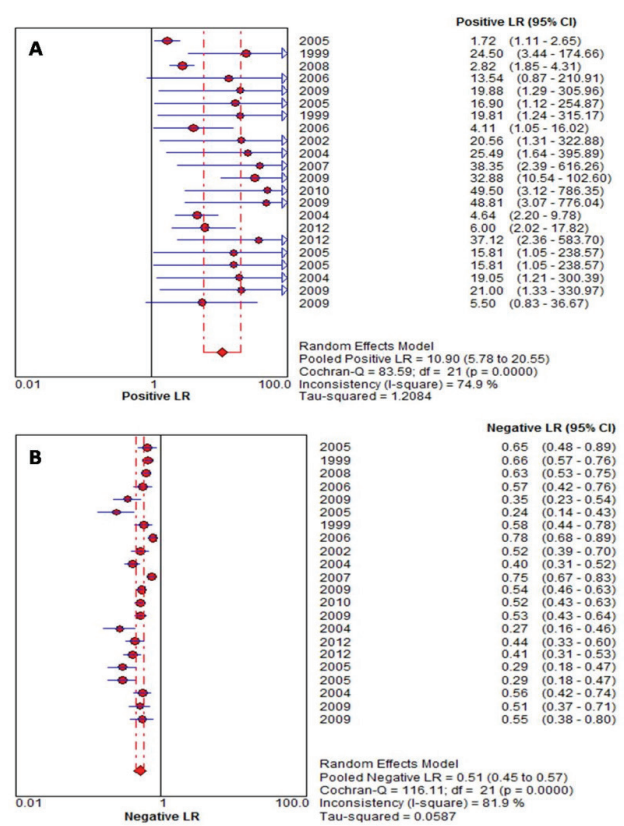


Figure 4: The forest plot of diagnostic +LR and -LR for serum CK20 mRNA in diagnosis of colorectal cancer

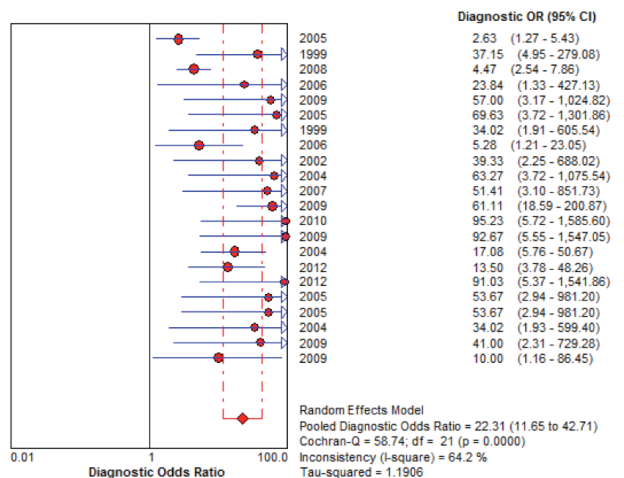


Figure 5: The forest plot of DOR for serum CK20 mRNA in diagnosis of colorectal cancer.

ous years, CK20 mRNA was elevated in colorectal cancer patients compared to healthy controls or benign colorectal disease patients. However, its clinical value as a biomarker for colorectal cancer diagnosis is not clear for different study design. And, because of small sample size for

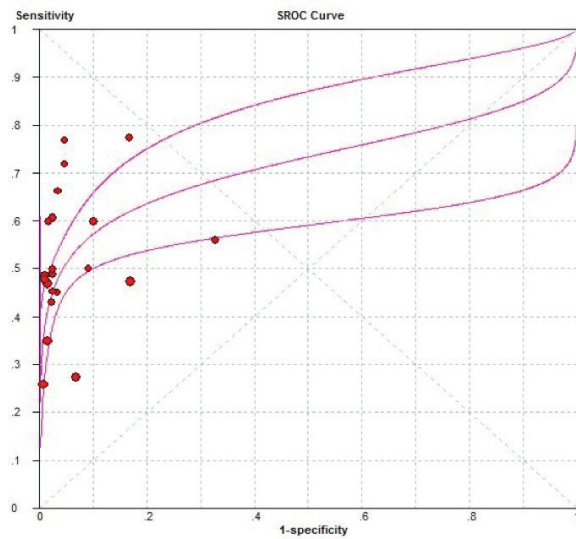


Figure 6: ROC curve of CK20 mRNA in serum as biomarker for colorectal cancer diagnosis.

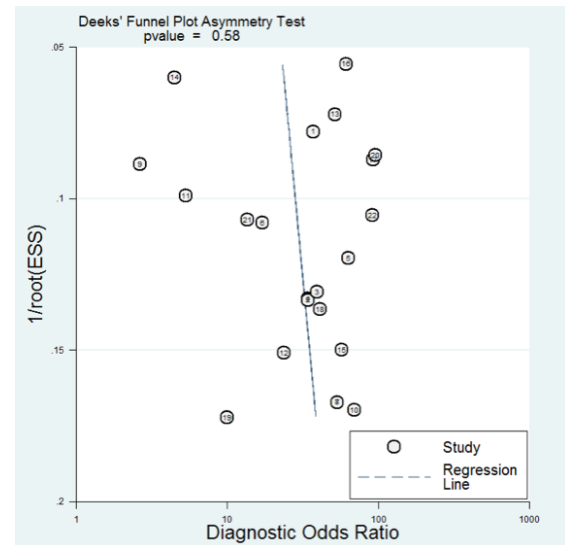


Figure 7: Publication bias evaluated by Deeks' funnel plot

Table 2: Subgroup analysis according to the control type and CK20 mRNA detection methods

Factors	Sen(95%CI)	Spe(95%CI)	+LR(95%CI)	-LR(95%CI)	DOR(95%CI)	AUC
Control type						
Population based	0.47(0.44-0.50)	0.99(0.98-1.00)	15.51(7.81-30.83)	0.51(0.44-0.60)	29.37(14.30-60.34)	0.86
Hospital based	0.67(0.61-0.73)	0.90(0.82-0.95)	5.61(3.21-9.82)	0.36(0.26-0.49)	17.44(8.46-35.94)	0.88
Methods						
RT-qPCR	0.49(0.46-0.52)	0.92(0.89-0.94)	8.30(4.37-15.60)	0.53(0.47-0.60)	16.60(8.42-32.75)	0.71
FQ-PCR	0.49(0.44-0.54)	0.99(0.77-1.00)	26.77(11.17-64.15)	0.41(0.27-0.61)	59.21(23.64-148.29)	0.95

each individual study, the results were inconclusive and clinical value was limited. To further assess the diagnostic value of serum CK20 mRNA as a biomarker for colorectal cancer diagnosis or screening, we searched the open published studies related to CK20 mRNA and colorectal cancer diagnosis and performed this meta-analysis. We found that CK20 mRNA almost cannot be detected in the serum of healthy people or benign colorectal disease patients. This made the false positive rate very low. However, the positive rate of serum CK20 mRNA in colorectal patients was also relative low. This made the serum CK20 mRNA a high specificity and low sensitivity biomarker for colorectal cancer diagnosis. Due to the low sensitivity (0.49, 95%CI:0.46-0.51), serum CK20 mRNA is not suitable for colorectal screening especially for the population based trials. However, a very high specificity (0.94, 95%CI: of 0.92-0.96), means that serum CK20 mRNA can be used as a confirmation test for colorectal cancer diagnosis.

Conflicts of interest: The authors declare no conflicts of interest regarding the publication of this manuscript.

Reference

- [1] Siegel RL., Miller KD., Jemal A. Cancer statistics, 2016.,CA. Cancer. J. Clin., 2016, 66, 7-30
- [2] Chen W., Zheng R., Baade PD., Zhang S., Zeng H., Bray F., et al., Cancer statistics in China, 2015, CA. Cancer. J. Clin., 2016, 66, 115-132
- [3] Nakayama T., Watanabe M., Teramoto T., Kitajima M., Prognostic values of serum CA19-9 and CEA levels for colorectal cancer, Oncol. Rep. 1997, 4, 819-822
- [4] Vukobrat-Bijedic Z., Husic-Selimovic A., Bijedic N., Mujkic A., Sofic A., Gogov B., Mehmedovic A, Bjelogriic I, Glavas S and Djuran A. Sensitivity of Symptomatology Versus Diagnostic Procedures and Concentration of CEA and CA19-9 in the Early Detection of Colorectal Cancer, Acta Inform. Med., 2014, 22, 89-93

- [5] Wharton RQ., Jonas SK., Glover C., Khan ZA., Klokouzas A., Quinn H., et al., Increased detection of circulating tumor cells in the blood of colorectal carcinoma patients using two reverse transcription-PCR assays and multiple blood samples, *Clin. Cancer. Res.*, 1999, 5, 4158-4163
- [6] Chausovsky G., Luchansky M., Figer A., Shapira J., Gottfried M., Novis B., et al. Expression of cytokeratin 20 in the blood of patients with disseminated carcinoma of the pancreas, colon, stomach, and lung, *Cancer.*, 1999, 86, 2398-2405
- [7] Lin GL., Qiu HZ., Xu T., Qian JM., Detection of micrometastasis in peripheral blood of patients with colorectal carcinoma before and during operative procedure, *Chinese. Journal of General. Surgery.*, 2002, 17, 605-607
- [8] Sun JW., Hua YK., Jia L., Lin RL., Yang KX., Chen JM., Expression of CK20 mRNA in Peripheral Blood of Patients with Colorectal Cancer and Its Clinical Significance, *Chin. J. Bases. Clin. General. Surg.*, 2004, 11, 119-121
- [9] Cui M., Liu WH., Yang Y., Xu H. Prognostic value of the combined detection of CK20 mRNA and CEA mRNA in peripheral blood of patients with colorectal cancer, *Chin. J. Exp. Surg.*, 2004, 21, 615-616
- [10] Zeng QG., Wang Can., Wang ZC. Research on diagnosing colorectal carcinoma by CK20 with the RT-PCR. *Chongqing. medical. journal.*, 2004, 33, 568-569
- [11] Hong-Zhuan Y., Xia H., Qi S., Shuo-Dong W., En-Qing L., Detection of CK20 in peripheral blood of rectal cancer patients with fluorescence quantitative polymerase chain reaction and its significance. *World Chinese Journal of Digestology.*, 2005, 13, 2892-2895
- [12] Zhong-cheng W., Can W., Qing-gui Z., Detection of cytokeratin20 mRNA in peripheral blood from colonic cancer patients by fluorescence quantitative polymerase chain reaction, *Chin. J. Oncol.*, 2007, 29, 600-603
- [13] Fang W., Xu-qian W., Jing S., Gu-yin L., Xue-feng W., Jie-ru J., Clinical research on detecting multimarker genes in peripheral blood of patients with colorectal carcinoma and its clinical significance, *China. oncology.*, 2009, 19, 353-357
- [14] Chen P., Jin XM., Zhang D., Yin QY., Liu XL., Study of CK20mRNA Expression in Peripheral Blood of Colorectal Carcinoma Patients and Tumor Micrometastasis, *Ningxia. Med. J.*, 2009, 31, 499-502
- [15] He Q., Jiao JR., Wei XQ., Zhao YJ., Wu F., Clinical Research on Detecting of Tumour Marker in Peripheral Blood of Patients with Gastric and Colorectal Carcinoma, *Chin. J. of clin. gastroenterol.*, 2012, 22, 67-75
- [16] Linhai L., Hui L., Mingwan J., Tianwei D., The relationship of VEGF-C and CK20 mRNA expression in peripheral blood and lymphatic metastasis in patients with colon carcinoma, *Chin. J. Gastroenterol. Hepatol.*, 2012, 21, 126-128
- [17] Dandachi N., Balic M., Stanzer S., Halm M., Resel M., Hinterleitner TA., et al., Critical evaluation of real-time reverse transcriptase-polymerase chain reaction for the quantitative detection of cytokeratin 20 mRNA in colorectal cancer patients, *J. Mol. Diagn.*, 2005, 7, 631-637
- [18] Guo J., Xiao B., Jin Z., Qin L., Chen J., Chen H., et al., Detection of cytokeratin 20 mRNA in the peripheral blood of patients with colorectal cancer by immunomagnetic bead enrichment and real-time reverse transcriptase-polymerase chain reaction, *J. Gastroenterol. Hepatol.*, 2005, 20, 1279-1284
- [19] Xu D., Li XF., Zheng S., Jiang WZ., Quantitative real-time RT-PCR detection for CEA, CK20 and CK19 mRNA in peripheral blood of colorectal cancer patients, *J. Zhejiang. Univ. Sci. B.*, 2006, 7, 445-451
- [20] Katsumata K., Sumi T., Mori Y., Hisada M., Tsuchida A., Aoki T., Detection and evaluation of epithelial cells in the blood of colon cancer patients using RT-PCR, *Int. J. Clin. Oncol.*, 2006, 11, 385-389
- [21] Shen C., Hu L., Xia L., Li Y., Quantitative real-time RT-PCR detection for survivin, CK20 and CEA in peripheral blood of colorectal cancer patients, *Jpn. J. Clin. Oncol.*, 2008, 38, 770-776
- [22] Lagoudianakis EE., Katakaki A., Manouras A., Memos N., Papadima A., Derventzi A., et al., Detection of epithelial cells by RT-PCR targeting CEA, CK20, and TEM-8 in colorectal carcinoma patients using OncoQuick density gradient centrifugation system, *J. Surg. Res.* 2009, 155, 183-190
- [23] Wong SC., Chan CM., Ma BB., Hui EP., Ng SS., Lai PB., et al., Clinical significance of cytokeratin 20-positive circulating tumor cells detected by a refined immunomagnetic enrichment assay in colorectal cancer patients, *Clin. Cancer. Res.*, 2009, 15, 1005-1012
- [24] Jemal A., Bray F., Center MM., Ferlay J., Ward E., Forman D., Global cancer statistics, *CA. Cancer. J. Clin.*, 2011, 61, 69-90
- [25] Torre LA., Bray F., Siegel RL., Ferlay J., Lortet-Tieulent J., Jemal A. Global cancer statistics, 2012, *CA. Cancer. J. Clin.*, 2015, 65, 87-108
- [26] Burt RW., Barthel JS., Dunn KB., David DS., Drelichman E., Ford JM., et al., NCCN clinical practice guidelines in oncology. Colorectal cancer screening, *J. Natl. Compr. Canc. Netw.*, 2010, 8, 8-61
- [27] Williams CD., Grady WM., Zullig LL., Use of NCCN Guidelines, Other Guidelines, and Biomarkers for Colorectal Cancer Screening, *J. Natl. Compr. Canc. Netw.* 2016, 14, 1479-1485
- [28] Corbo C., Cevenini A., Salvatore F., Biomarker discovery by proteomics-based approaches for early detection and personalized medicine in colorectal cancer, *Proteomics. Clin. Appl.*, 2017, 11, 5-6
- [29] Moll R., Lowe A., Laufer J., Franke WW., Cytokeratin 20 in human carcinomas. A new histodiagnostic marker detected by monoclonal antibodies, *Am. J. Pathol.*, 1992, 140, 427-447
- [30] Moll R., Zimbelmann R., Goldschmidt MD., Keith M., Laufer J., Kasper M., et al., The human gene encoding cytokeratin 20 and its expression during fetal development and in gastrointestinal carcinomas, *Differentiation.*, 1993, 53, 75-93