CASE REPORT



Novel Case Report: A Previously Reported, but Pathophysiologically Unexplained, Association Between Collagenous Colitis and Protein-Losing Enteropathy May Be Explained by an Undetected Link with Collagenous Duodenitis

Inayat Gill¹ · Aciel Ahmed Shaheen¹ · Ahmed Iqbal Edhi¹ · Mitual Amin^{2,3} · Ketan Rana¹ · Mitchell S. Cappell^{1,4}

Received: 19 November 2020 / Accepted: 17 December 2020 / Published online: 4 February 2021 © The Author(s), under exclusive licence to Springer Science+Business Media, LLC part of Springer Nature 2021

Abstract

Collagenous colitis (CC) is associated with non-bloody, watery diarrhea, which is pathophysiologically reasonable because normal colonic absorption (or excretion) of water and electrolytes can be blocked by the abnormally thick collagen layer in CC. However, CC has also been associated with six previous cases of protein-losing enteropathy (PLE), with no pathophysiologic explanation. The colon does not normally absorb (or excrete) amino acids/proteins, which is primarily the function of the small bowel. Collagenous duodenitis (CD) has not been associated with PLE. This work reports a novel case of CD (and CC) associated with PLE; a pathophysiologically reasonable mechanism for CD causing PLE (by the thick collagen layer of CD blocking normal intestinal amino acid absorption); and a novel association of PLE with severe COVID-19 infection (attributed to relative immunosuppression from hypoproteinemia, hypoalbuminemia, hypogammaglobulinemia, and malnutrition from PLE).

Keywords Collagenous colitis \cdot Collagenous duodenitis \cdot Protein-losing enteropathy \cdot Hypoalbuminemia \cdot Anasarca \cdot Malnutrition \cdot COVID-19 infection \cdot SARS-CoV-2 infection

Mitchell S. Cappell mitchell.cappell@beaumont.edu

Inayat Gill inayat.gill@beaumont.org

Aciel Ahmed Shaheen aciel.ahmedshaheen@beaumont.org

Ahmed Iqbal Edhi ahmed.edhi@beaumont.org

Mitual Amin mitual.amin@beaumont.edu

Ketan Rana ketan.rana@beaumont.edu

- Division of Gastroenterology, Department of Internal Medicine, William Beaumont Hospital at Royal Oak, 3535 W. Thirteen Mile Rd, Royal Oak, MI 48073, USA
- Department of Pathology, William Beaumont Hospital at Royal Oak, 3600 W. Thirteen Mile Rd, Royal Oak, MI 48073, USA
- Department of Pathology, Oakland University William Beaumont School of Medicine, 3600 W. Thirteen Mile Rd, Royal Oak, MI 48073, USA
- Division of Gastroenterology, Department of Internal Medicine, Oakland University William Beaumont School of Medicine, 3535 W. Thirteen Mile Rd, Royal Oak, MI 48073, USA



 Table 1
 Clinical presentation in seven reported cases of protein-losing enteropathy associated with collagenous colitis (suspected from undiagnosed collagenous duodenitis/enteritis)

Clinical presentation	Laboratory values	Endoscopy and histopathology	Clinical course	Reference (publication type)
hnitial hospitalization 65 y. o. F with history of tobacco abuse presented with nausea, vomiting, and abdominal pain. PE: BP = 103/65 mmHg, HR = 83 beats/min, RR = 22 breaths/min, T = 36.5 °C. Dry mucous membranes, absent axillary sweat, poor skin turgor, and diffusely mildly tender abdomen Later hospitalizations Presented with recurrent nausea, vomiting, and diarrhea. PE: new-onset anasarca (ascites and bilateral upper and lower extremity edema)	Initial hospitalization WBC=6900/mm³, Hb=13.9 g/dL, BUN=43 mg/dL, (GFR=14 mL/ min), K=2.7 mmol/L. Stool tests negative for Clostridioides difficile toxins A & B, and for ova and para- sites. Stool tests positive for lactofer- rin and calprotectin. Serologic panel for celiac disease: normal Later hospitalizations Albumin=1.4 g/dL, Prealbu- min=10 mg/dL, Normal liver function tests. 24-h urine total protein=276 mg, stool alpha-1 antit- rypsin=369 mg/dL (normal <54 mg/ dL)	Initial hospitalization Colonoscopy: abnormally thick (> 10 µm) subepithelial collagen band (diagnostic of collagenous colitis) and numerous intraepithelial lymphocytes in colonic mucosa Later hospitalizations EGD: diffuse nodular mucosa in first and second portion of duodenum. Duodenal biopsy: severe collagenous band (collagenous duodenitis)	Initial hospitalization Treated with budesonide and mesalamine, with improving in diarrhea and discharged Later hospitalizations Treated with budesonide, mesalamine and azathioprine, with improving diarrhea and discharged	Current report
15-month-old M with history of microcornea admitted for vomiting, diarrhea, and peripheral edema for 4 weeks. PE: ill-appearing, afebrile, and with upper and lower extremity edema	WBC=14,500/mm³, total protein=3.6 g/dL, albumin=2.4 mg/dL. Normal liver and renal function tests. Normal serologic tests for celiac disease. Stool tests: negative bacterial and viral cultures. Stool alpha-1 antitrypsin≥1.33 mg/g (normal<0.62 mg/g)	EGD: edematous antral and duodenal mucosa. Duodenal biopsy: mildly increased collagenous band, but not thick enough to diagnose collagenous duodenitis. Gastric biopsy: negative for collagen. Flexible sigmoidoscopy: normal-appearing colon and rectum. Biopsies: collagenous colitis	Therapy: budesonide and TPN. Methylprednisolone added due to failed improvement in diarrhea. Weaned off TPN and discharged to take budesonide and methylprednisolone as outpatient	Almadhoun et al. [2]
76 y. o. F admitted with recurrent diarrhea and edema for 5 months. PE: normal vital signs, tender lower abdomen, swollen face, and pretibial pitting edema	WBC=7960/mm³, total protein=4.8 g/dL, albumin=2.8 gm/dL, normal liver and renal function tests. No proteinuria	EGD: normal-appearing stomach and duodenum. Biopsies: normal villi, no lymphocytic infiltrate, no collagenous duodenitis. Colonoscopy: edematous mucosa. Biopsies consistent with collagenous colitis, with increased lymphocytes and plasma cells. ^{99m} Tc HSA scintigraphy: protein leakage in colon, but not in stomach or small bowel	Therapy: prednisolone 30 mg/day, with Sano et al. [3] improving diarrhea	Sano et al. [3]
82 y. o. F with CKD admitted for nausea, anorexia, and diarrhea for 1 week. PE: normal vital signs, bilateral pitting edema of lower extremities	Albumin = 1.2 g/dL, (3 months earlier albumin = 3.1 g/dL). Stool positive for occult blood. Stool culture negative for bacteria and viruses. No proteinuria. Normal liver function tests	CT scan: mural thickening of small bowel. ^{99m} Tc HSA scintigraphy: protein leakage from small bowel. EGD: normal-appearing stomach and duodenum. Colonoscopy: edematous mucosa. Biopsies: > 10 µm subepithelial collagenous band (diagnostic of collagenous colitis)	Therapy: loperamide, with improving diarrhea	Nakaya et al. [4]



Table 1 (continued)

iable i (continued)				
Clinical presentation	Laboratory values	Endoscopy and histopathology	Clinical course	Reference (publication type)
63 y. o. F chronically taking lansoprazole admitted with diarthea and generalized edema for 5 months. No abdominal pain. PE: BP= 128/90 mmHg, HR = 67 beats/min, T=36.5 °C, swollen face, bilateral pretibial and pedal edema	WBC=4800/mm³, Hb 14.3 g/dL, total protein=4.6 g/dL, albumin=2.8 g/dL, normal liver and renal function tests. U/A: no proteinuria	EGD: normal-appearing stomach and duodenum with normal biopsies. Colonoscopy: longitudinal lacerations in descending colon. Biopsies: thick collagen layer (> 10 µm thick), with subepithelial eosinophilic infiltrate. 9mrTc HSA scintigraphy: protein leakage in descending colon	Lansoprazole was discontinued and diarrhea improved. 3 weeks later albumin increased to 3.4 g/dL. No steroids administered	Ozeki et al. [5]
62 y. o. F presented with 6–8 watery stools/day, abdominal cramping, and 4 kg weight loss over 4 months. PE: normal vital signs, non-tender abdomen	WBC = $10,500/\text{nm}^3$, $K = 4.4 \text{ meq/L}$, total protein = 4.3 g/dL , albumin = 2.24 g/dL , normal liver and renal function tests. No proteinuria. IgA and IgG levels low. Stool for bacterial culture, ova and parasites, and fecal leukocytes were negative. Stool alpha-1 antitrypsin = $214 \text{ mL/24} \text{ h}$ (normal < $13 \text{ mL/24} \text{ h}$)	Abdominal ultrasound: normal-appearing liver, gallbladder, pancreas, and kidneys Small bowel barium series: normal EGD: normal-appearing duodenal mucosa with intact intestinal villi. Small bowel biopsy negative for celiac disease Colonoscopy: grossly normal. Biopsies consistent with collagenous colitis	Therapy: not discussed	Stark et al. [6]
64 y. o. F with breast cancer admitted for 8–10 watery, non-bloody bowel movements/day and 8 kg weight loss over 3 months. PE: normal vital signs, no abdominal pain	Bacterial stool cultures, stool for ova and parasites, fecal leukocytes, and celiac disease panel all negative. Elevated stool alpha-1 antitrypsin in 24 h collection. Low protein C, S, and antithrombin III levels	Abdominal CT scan: right lower lobe pulmonary embolus and renal vein thrombosis EGD: normal-appearing Colonoscopy: normal-appearing. Biopsy: collagenous colitis	Therapy: oral corticosteroids with resolution of diarrhea. Pulmonary embolus treated with IV heparin as bridge to Coumadin	Raimo et al. [7]

y. o. years old, F female, M male, WBC white blood cell (count), Hb hemoglobin, BUN blood urea nitrogen, GFR glomerular filtration rate, K potassium, IgA immunoglobulin A, IgG immunoglobulin G, AP alkaline phosphatase, TPN total parenteral nutrition, U/A urinalysis, CT computed tomography, ⁹⁹ⁿTc HSA technetium-99m human serum albumin, PE physical exam, EGD esophagogastroduodenoscopy, HR heart rate, RR respiratory rate, BP blood pressure, T temperature



Introduction

Collagenous colitis (CC) is associated with non-bloody, watery diarrhea, which is pathophysiologically reasonable because normal colonic absorption (or excretion) of water and electrolytes can be blocked by the abnormally thick collagen layer [1]. However, CC has also been associated with six cases of protein-losing enteropathy (PLE) ([2–7]; Table 1), with no pathophysiologic explanation. The colon does not normally absorb (or excrete) amino acids/proteins, which is primarily the function of the small bowel [8].

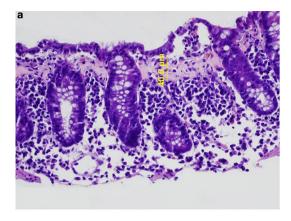
Collagenous duodenitis (CD)¹ (or enteritis) has not been associated with PLE [9–19].² This work reports a novel case of CD (and CC) *associated* with PLE; a pathophysiologically reasonable mechanism for CD causing PLE (the thick collagen layer of CD blocks normal intestinal amino acid absorption); and a novel association of PLE with severe COVID-19 infection (attributed to relative immunosuppression from hypoproteinemia, hypoalbuminemia, hypogammaglobulinemia, and malnutrition from PLE) [3, 6]. A potential association of CD with PLE is clinically important; PLE is a life-threatening, but potentially curable, syndrome.

Case Report

A 65-year-old African American woman with a 50-pack-year history of smoking tobacco, no prior renal insufficiency, and no other medical disorders, presented with nausea, vomiting, and generalized abdominal pain for 3 days; five non-bloody, watery stools/day; and 14 kg involuntary weight loss over the prior 3 months. Physical examination on admission revealed a minimally overweight female (BMI = 27.6 kg/m^2); normal vital signs; dry mucous membranes, absent axillary sweat, and poor skin turgor; a non-tender, non-distended abdomen without hepatosplenomegaly; and no fecal occult blood. The leukocyte count was 6900/mm³ (normal: 3500–10,100 leukocytes/mm³), and hemoglobin was 13.9 g/dL (normal: 13.5–17 g/dL). Serum creatinine was 3.61 mg/dL (normal: 0.6–1.3 mg/dL), blood urea nitrogen was 43 mg/dL (normal: 7–25 mg/dL), urine specific gravity was 1.023 (> 1.015 consistent with dehydration), and glomerular filtration rate was 14 mL/min (<15 mL/min indicates renal failure). Serum potassium was 2.7 mmol/L (normal: 3.5-5 mmol/L). Stool

² Excludes references with collagenous gastritis without collagenous duodenitis, because the stomach is not physiologically an important site of amino acid absorption, and therefore a thickened gastric collagen layer should not pathophysiologically cause protein-losing enteropathy [20–22].





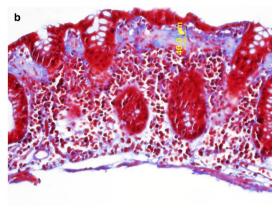


Fig. 1 a High-power photomicrograph of hematoxylin and eosin stained section of a random colonic biopsy from endoscopically normal-appearing colon shows a markedly thickened subepithelial collagen (pink) band measuring approximately 40 μ m (diagnostic of collagenous colitis). b High-power photomicrograph of the same colonic biopsy stained with Masson's trichrome, which highlights the collagenous band in blue and confirms the thickened (approximately 40 μ m) collagen layer

tests for *Clostridioides difficile* toxins A and B and for ova and parasites were negative. Stool culture for bacterial pathogens was sterile. Stool calprotectin level was 225 mcg/g (normal: 0–50 mcg/g). Abdominopelvic computed tomography (CT) on the initial admission was within normal limits, with no intra-abdominal lymphadenopathy, mural bowel thickening, dilated bowel loops, streaky mesentery, or organomegaly.

The patient received lactated Ringer's solution intravenously at 150 mL/h to reverse dehydration and acute renal failure. Tests for celiac disease revealed tissue transglutaminase immunoglobulin (Ig) A of 8.4 units/mL (normal: 0–19.9 units/mL), and total IgA of 247 mg/dL (normal: 70–365 mg/dL). Colonoscopy, performed for chronic diarrhea, revealed an endoscopically normal colon, but histopathology of biopsies taken throughout the colon showed an abnormally thick (> 10 μ m) subepithelial collagen layer diagnostic of CC (Fig. 1a, b). The patient was initially

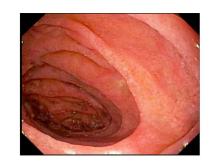
¹ CD herein denotes collagenous duodenitis and not Crohn's disease.

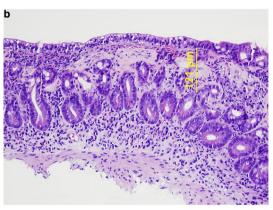
treated with oral budesonide 9 mg/day, with gradually decreasing diarrhea, but mesalamine 800 mg twice daily was added as therapy because of persisting non-bloody diarrhea, with subsequent improvement. The patient was advised to cease smoking, as recommended for patients with collagenous colitis [23], but refused.

She was readmitted 1 month later for refractory nausea and vomiting. Physical exam revealed anasarca, manifested by ascites and 3+ pitting edema of all four extremities. Serum albumin was 1.4 g/dL (normal: 3.5-4.9 g/dL), and prealbumin was 10 mg/dL (normal: 18-44 mg/dL). Hypoalbuminemia was not from liver disease: all liver function tests were within normal limits. Hypoalbuminemia was not from protein-losing nephropathy: urine collection revealed only 276 mg of protein/24 h (nephrotic syndrome: > 3.5 g/24 h). Ascites was not from congestive heart failure: electrocardiogram was within normal limits, and chest X-ray did not reveal cardiomegaly. Stool alpha-1 antitrypsin was 369 mg/ dL (normal: < 54 mg/dL), a finding diagnostic of PLE [24]; anasarca and hypoalbuminemia were therefore due to PLE. Iron studies were within normal limits. Vitamin B12, folate, and vitamin K were within normal limits, as were cholesterol and triglyceride levels. Vitamin D level was 14 ng/mL (normal 30-100 ng/mL).

Esophagogastroduodenoscopy (EGD) revealed minimally nodular mucosa in the first and second portions of the duodenum (Fig. 2a). Histopathology of duodenal biopsies revealed severe CD [Fig. 2b, c; collagen layer in (c) measuring approximately 332 μ m, > 10 μ m characteristic of CD)], and of gastric biopsies revealed no *Helicobacter pylori*. Azathioprine was added to the regimen of mesalamine and budesonide. A dietician recommended a protein-rich diet supplemented with medium-chain triglycerides, but patient refused this diet as unpalatable, and was discharged on total parenteral nutrition (TPN).

The patient presented 2 months later with nausea, vomiting, worsening diarrhea, cough, and dyspnea for 7 days. She was taking budesonide, but was noncompliant with azathioprine and mesalamine. Physical examination on admission revealed a mildly thin woman (BMI = 23.47 kg/m^2); pulse of 100 beats/min, temperature of 36.6 °C, 18 breaths/ min, and O2 saturation of 99% on room air; dry mucous membranes; moderate ascites; and soft, non-distended, and non-tender abdomen. Laboratory tests revealed 7900 leukocytes/mm³ (normal: 3500–10,100 leukocytes/mm³), creatinine of 0.87 mg/dL (normal: 0.6–1.3 mg/dL), potassium of 2.4 mmol/L (normal: 3.5-5 mmol/L), albumin of 2.3 g/ dL (normal: 3.5-4.9 g/dL), prealbumin of 8 mg/dL (normal: 18–44 mg/dL), and serum IgG of 491 mg/dL (normal: 550–1650 mg/dL), findings consistent with malnutrition from PLE. Nasopharyngeal swab was positive for COVID-19 infection by nucleic acid amplification, as confirmed by polymerase chain reaction. C-reactive protein was 48.4 mg/L





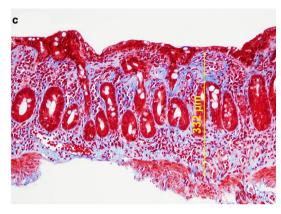


Fig. 2 a Esophagogastroduodenoscopy (EGD) revealed endoscopically essentially normal (minimally nodular) mucosa in first and second portions of duodenum, as illustrated for the second portion of the duodenum. **b** Medium-power photomicrograph of hematoxylin and eosin-stained section of a random biopsy of endoscopically relatively normal-appearing second portion of duodenum shows severe to total blunting of the villi, with no appreciable villous height. The collagen band measures approximately 121 microns (> 10 microns diagnostic of collagenous duodenitis). This collagen band is thicker than that observed in the colon (Fig. 1a). **c** Medium-power photomicrograph of the same duodenal biopsy stained with Masson's trichrome, which highlights collagen in blue, shows extensive collagenization of duodenal mucosa, with more discernable collagenization visualized in this stain (measuring approximately 332 μm of entire thickness of the lamina propria) than demonstrated in the hematoxylin and eosin stain

(normal: 0–7.9 mg/L), attributed to active COVID-19 infection. Stool studies for *Clostridioides difficile* toxins A and B and for ova and parasites were negative. Stool culture for bacterial pathogens was sterile. Repeat EGD with duodenal



biopsies revealed persistent CD. She received intravenous hydration, TPN, hydroxychloroquine for COVID-19 infection, and budesonide for CD. She did well without developing respiratory distress and was discharged 11 days after admission.

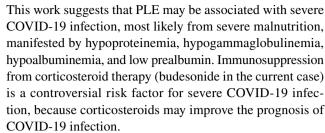
Discussion

Etiologies of PLE include erosive GI mucosal diseases such as Crohn's disease or GI malignancies; diseases causing intestinal lymphatic fluid loss from lymphatic obstruction, such as primary intestinal lymphangiectasia, or lymphoma; and diseases increasing intestinal mucosal permeability such as Ménétrier's disease [25, 26]. CC is a reported cause of PLE but lacks a pathophysiologic basis. The current work suggests a novel association of CD (and CC) with PLE. Linkage of CC with CD can explain the pathophysiology of PLE by disruption of normal amino acid absorption by the thick intestinal collagen layer in CD (12× upper limit of normal in the current case). Patients with CC and PLE may be reasonably tested to exclude CD by intestinal biopsies. Diagnosing PLE etiology is important to initiate specific therapy.

The current diagnosis of CD was challenging because of negative serology for celiac disease and absence of iron deficiency anemia. CD often occurs with celiac disease [27], which by itself has a mildly increased subepithelial collagen layer [28]. Initiation of a gluten-free diet causes regression of collagen in celiac disease, but not in CD. Collagenous celiac disease (collagenous sprue) has been rarely reported but has not been associated with PLE, even though it has been associated with hypoalbuminemia [29]. Although this patient had severe to total blunting of the intestinal villi (Fig. 2b), the patient did not have collagenous sprue, as shown by negative serology for celiac disease.

Collagen deposition in CC is believed to be immune-mediated; autoimmune diseases, medications, viruses, and bacteria are speculated triggers [19, 28, 30], including one case of *Yersinia enterocolitica* [14]. Collagen deposition and inflammation in CC may extend from the colonic to intestinal mucosa and cause CD by dysfunction of subepithelial myofibroblasts [10, 17]. CC is characterized by an abnormally thick (> 10 μ m diameter) subepithelial hyalinized collagen band [19, 21, 27, 30]. Although CC is frequently reported, CD is rarely reported [13]. Because of its rarity, CD is not usually suspected in patients with CC.

Immunosuppression from human immunodeficiency virus infection, cancer, solid organ transplantation, immunomodulatory therapy for autoimmune diseases, and malnutrition can cause severe COVID-19 outcomes, including prolonged hospitalization, intensive care unit admission, renal failure, intubation, mechanical ventilation, and mortality [31–33].



COVID-19 can affect the digestive system [34–36]. The receptor-binding domain of COVID-19 binds strongly to human angiotensin converting enzyme-2 (ACE-2), which is abundantly expressed in the intestines [37]. Studies have identified COVID-19 RNA in stool specimens of infected patients, suggesting the virus may infect the digestive system [34]. It is interesting to speculate whether COVID-19 infection may be more severe in patients with intestinal disease, such as CD.

This report is limited by its single-case retrospective nature. However, an association of PLE with CD has a solid pathophysiologic basis because the small intestine is the primary site of amino acid (protein) absorption, which should be severely compromised by a thick intestinal collagen layer. In contrast, the colon does not play a physiologic role in amino acid absorption and CC should not affect amino acid absorption. A minor criticism is that the patient's collagenous colitis was treated about 1 year ago with budesonide and mesalamine. Mesalamine has recently been delisted as recommended therapy for collagenous colitis [38]. Strengths of this work include a strongly positive diagnosis of PLE by alpha-1 antitrypsin levels, a strongly positive pathologic diagnosis of CD, and exclusion of other major causes of hypoalbuminemia and anasarca aside from PLE. This work requires confirmation in a large prospective trial, but such a trial may be problematic because CD is relatively rare.

Conclusion

This case demonstrates that presentation of CD (with CC) can result in PLE, attributed to a thick collagen layer preventing absorption of amino acids/proteins in small intestine. Immunosuppression from hypoproteinemia, hypoalbuminemia, hypogammaglobulinemia, and malnutrition from PLE likely constitutes a high risk factor for severe COVID-19 infection.

Author's contribution The initial draft of the manuscript was written by Drs. IG, AAS, AIE, KR, and MSC. The manuscript was thoroughly revised, and major additions were made by Dr. MSC, who served as the mentor. Drs. KR and MSC provided clinical gastroenterologic care to the patient. Dr. MA provided the histopathology images and description. Drs. IG and MSC are the primary authors.



Compliance with Ethical Standards

Conflict of interest None for all authors. None of the authors received any financial support for writing this case report. In particular, Dr. Cappell, as a consultant of the United States Food and Drug Administration (FDA) Advisory Committee for Gastrointestinal Drugs, affirms that this paper does not discuss any proprietary, confidential, pharmaceutical data submitted to the FDA. Dr. Cappell was also a member of the speaker's bureau for AstraZeneca and Daiichi Sankyo, co-marketers of Movantik until > 2 years ago. Dr. Cappell received one-time honoraria from Shire and Mallinckrodt > 2 years ago. This work does not discuss any drug manufactured or marketed by AstraZeneca, Daiichi Sankyo, Shire, or Mallinckrodt.

Ethical approval Exemption/approval was obtained from the Beaumont Health System Institutional Review Board on June 1, 2020.

References

- Boland K, Nguyen GC. Microscopic colitis: a review of collagenous and lymphocytic colitis. Gastroenterol Hepatol (N Y) 2017:13:671–677
- Almadhoun OF, Katzman PJ, Rossi T. Collagenous colitis associated with protein losing enteropathy in a toddler. Case Rep Gastrointest Med. 2014. https://doi.org/10.1155/2014/209624.
- Sano S, Yamagami K, Tanaka A, Nishio M, Nakamura T, Kubo Y et al. A unique case of collagenous colitis presenting as protein-losing enteropathy successfully treated with prednisolone. World J Gastroenterol 2008;14:6083–6086. https://doi. org/10.3748/wjg.14.6083.
- Nakaya Y, Hosokawa SK, Kataoka Y, Hirabayashi M, Yamamoto S, Takasu K, Kitamura S, Omae T, Yoshimatsu Y. Acute onset collagenous colitis associated with protein-losing enteropathy. *J Gen Fam Med* 2017;18:135–137. https://doi.org/10.1002/jgf2.13.
- Ozeki T, Ogasawara N, Izawa S, Mizuno M, Yanamoto K, Noda H, Okaniwa N, Tanabe A, Sasaki M, Kasugai K. Protein-losing enteropathy associated with collagenous colitis cured by withdrawal of a proton pump inhibitor. *Intern Med* 2013;52:1183– 1187. https://doi.org/10.2169/internalmedicine.52.0232.
- Stark ME, Batts KP, Alexander GL. Protein-losing enteropathy with collagenous colitis. Am J Gastroenterol 1992;87:780–783
- Raimo J, Coronel M, Criss A, Storch I. Collagenous colitis associated with protein losing enteropathy presenting with multiple venous thromboses. Am J Gastroenterol 2010;105:S342
- Patrick F, Hagihara PF, Griffen WO Jr. Physiology of the colon and rectum. Surg Clin North Am 1972;52:797–805. https://doi. org/10.1016/s0039-6109(16)39779-1.
- Kim YJ, Satapathy S, Iqbal S, Cerulli M. Collagenous duodenitis in patient with collagenous colitis. Am J Gastroenterol 2010;105:S183-S184
- Meier PN, Otto P, Ritter M, Stolte M. Collagenous duodenitis and ileitis in a patient with collagenous colitis. *Leber Magen Darm* 1991;21:231–232
- Soeda A, Mamiya T, Hiroshima Y, Sugiyama H, Shidara S, Dai Y, Nakahara A, Ikezawa K. Collagenous gastroduodenitis coexisting repeated Dieulafoy ulcer: a case report and review of collagenous gastritis and gastroduodenitis without colonic involvement. *Clin J Gastroenterol* 2014;7:402–409. https://doi.org/10.1007/s1232 8-014-0526-y.
- Koide T, Mochizuki T, Kawai N, Yashiro K, Inoue T, Tsujimoto M, Nishigaki T. Collagenous gastroduodenitis with recurrent

- gastric ulcer in 12-year-old-girl. *Pediatr Int* 2015;57:754–757. https://doi.org/10.1111/ped.12615.
- Rustagi T, Rai M, Scholes JV. Collagenous gastroduodenitis. J Clin Gastroenterol 2011;45:794–799. https://doi.org/10.1097/ MCG.0b013e31820c6018.
- Navarro-Llavat M, Domenech E, Masnou H, Ojanguren I, Manosa M, Lorenzo-Zuniga V et al. Collagenous duodeno-ileo-colitis with transient IgG deficiency preceded by *Yersinia enterocolitica* intestinal infection: case report and review of literature. *Gastroenterol Hepatol* 2007;30:219–221. https://doi.org/10.1157/13100588.
- Cocq P, Scrumeda D, Baron P, Duthoit D, Korgami B, Colombel JK. Collagenous gastritis, duodenitis, ileitis and colitis. *Gastro-enterol Clin Biol* 2011;25:1027–1029
- Schreiber FS, Eidt S, Hidding M, Schmidt-Walczuch J, Werning C. Collagenous duodenitis and collagenous colitis: a short clinical course as evidenced by sequential endoscopic and histologic findings. *Endoscopy* 2001;33:555. https://doi.org/10.1055/s-2001-14968.
- Stolte M, Ritter M, Borchard F, Koch-Scherrer G. Collagenous gastroduodenitis on collagenous colitis. *Endoscopy* 1990;22:186– 187. https://doi.org/10.1055/s-2007-1012837.
- Borchard F, Niederau C. Collagenous gastroduodenitis. Dtsch Med Wochenschr 1989;114:1345
- Eckstein RP, Dowsett JF, Riley JW. Collagenous enterocolitis: a case of collagenous colitis with involvement of the small intestine. Am J Gastroenterol 1988;83:767–771
- Gopal P, McKenna BJ. The collagenous gastroenteritides: similarities and differences. *Arch Pathol Lab Med* 2010;134:1485–1489. https://doi.org/10.1043/2010-0295-CR.1.
- Nielsen OH, Riis LB, Danese S, Bojesen RD, Soendergaard C. Proximal collagenous gastroenteritides: clinical management. A systematic review. *Ann Med* 2014;46:311–317. https://doi. org/10.3109/07853890.2014.899102.
- Chen B, Popescu O, Barker C. A226 pediatric collagenous gastritis: endoscopic and histologic evolution. *J Can Assoc Gastroenterol* 2018;1:394–396. https://doi.org/10.1093/jcag/gwy008.227.
- Jaruvongvanich V, Poonsombudlert K, Ungprasert P. Smoking and risk of microscopic colitis: a systematic review and meta-analysis. *Inflamm Bowel Dis* 2019;25:672–678. https://doi.org/10.1093/ibd/ izy296.
- Perrault J, Markowitz H. Protein-losing gastroenteropathy and the intestinal clearance of serum-alhpa-1-antitrypsin. *Mayo Clin Proc* 1984;59:278–279. https://doi.org/10.1016/s0025-6196(12)61263-4.
- Cappell MS, Edhi A, Amin M. Case report of primary intestinal lymphangiectasia diagnosed in an octogenarian by ileal intubation and by push enteroscopy after missed diagnosis by standard colonoscopy and EGD. *Medicine* 2018;97:e9649. https://doi. org/10.1097/MD.00000000000009649.
- Levitt DG, Levitt MD. Protein losing enteropathy: comprehensive review of the mechanistic association with clinical and subclinical disease states. Clin Exp Gastroenterol 2017;10:147–168. https:// doi.org/10.2147/CEG.5136803.
- Shor J, Churrango G, Hosseini N, Marshall C. Management of microscopic colitis: challenges and solutions. *Clin Exp Gaster-oenterol* 2019;12:111–120. https://doi.org/10.2147/CEG.S1650 47.
- Freeman HJ. Collagenous sprue. Can J Gasteroenterol 2011;25:189–192. https://doi.org/10.1155/2011/821976.
- Soendergaard C, Buhl Riis L, Haagen Nielsen O. Collagenous sprue: a coeliac disease look-alike with different treatment strategy. BMJ Case Rep 2014;2014:bcr2014203721. https://doi.org/10.1136/bcr-2014-203721.
- Castellano VM, Muñoz MT, Colina F, Nevado M, Casis B, Solís-Herruzo JA. Collagenous gastrobulbitis and collagenous colitis:



- case report and review of the literature. *Scand J Gastroenterol* 1990;34:632–638. https://doi.org/10.1080/003655299750026128.
- Fung M, Babik JM. COVID-19 in immunocompromised hosts: what we know so far. *Clin Infect Dis*. 2020. https://doi. org/10.1093/cid/ciaa863.
- CDC (Centers for Disease Control and Prevention). Your health: evidence used to update the list of underlying medical conditions that increase a person's risk of severe illness from COVID-19. www.cdc.gov/coronavirus/2019-ncov/need-extra-precautions/ evidence-table.html
- Payette C, Terry AT. ACEP (American College of Emergency Physicians). COVID-19 field guide: immunocompromised patients-special populations. https://www.acep.org/corona/covid -19-field-guide/special-populations/immunosuppressed-patients/
- 34. Wong SH, Lui RN, Sung JJ. COVID-19 and the digestive system. *J Gastroenterol Hepatol* 2020;35:744–748. https://doi.org/10.1111/jgh.15047.
- Pan L, Mu M, Yang P, Sun Y, Wang R, Yan J et al. Clinical characteristics of COVID-19 patients with digestive symptoms in Hubei, China: a descriptive, cross sectional, multicenter study.

- Am J Gastroenterol. 2020;115:766–773. https://doi.org/10.14309/ajg.00000000000000020.
- Cappell MS. Moderately severe diarrhea and impaired renal function with COVID-19 infection. *Am J Gastroenterol* 2020;115:947–948. https://doi.org/10.14309/ajg.0000000000000881.
- Walls AC, Park YJ, Tortorici A, Wall A, McGuire AT, Veesler D. Structure, function and antigenicity of the SARS-CoV-2 spike glycoprotein. *Cell* 2020;181:281-292.e6. https://doi.org/10.1016/j.cell.2020.02.058.
- Miehlke S, Guagnozzi D, Zabana Y, Tontini GE, Fiehn AK, Wildt S et al. European guidelines on microscopic colitis: United European Gastroenterology (UEG) and European Microscopic Colitis Group (EMCG) statements and recommendations. *United Eur Gastroenterol J.* 2020. https://doi.org/10.1177/205064062095190

Publisher's Note Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

