

Electronic Supplementary Material

Hydroxyapatite nanoparticles drive the potency of Toll-like receptor 9 agonist for amplified innate and adaptive immune response

Qin Zeng^{1,2,3} (✉), Ruiqi Wang⁴, Yuchen Hua^{1,3}, Hongfeng Wu^{1,3}, Xuening Chen^{1,3}, You-cai Xiao⁴, Qiang Ao^{2,3}, Xiangdong Zhu^{1,3} (✉), and Xingdong Zhang^{1,2,3}

¹ National Engineering Research Center for Biomaterials, Sichuan University, Chengdu 610064, China

² NMPA Key Laboratory for Quality Research and Control of Tissue Regenerative Biomaterials & Institute of Regulatory Science for Medical Devices & NMPA Research Base of Regulatory Science for Medical Devices, Sichuan University, Chengdu 610064, China

³ College of Biomedical Engineering, Sichuan University, Chengdu 610064, China

⁴ Key Laboratory of Drug-Targeting and Drug Delivery System of the Education Ministry and Sichuan Province, Sichuan University, Chengdu 610041, China

Supporting information to <https://doi.org/10.1007/s12274-022-4683-x>

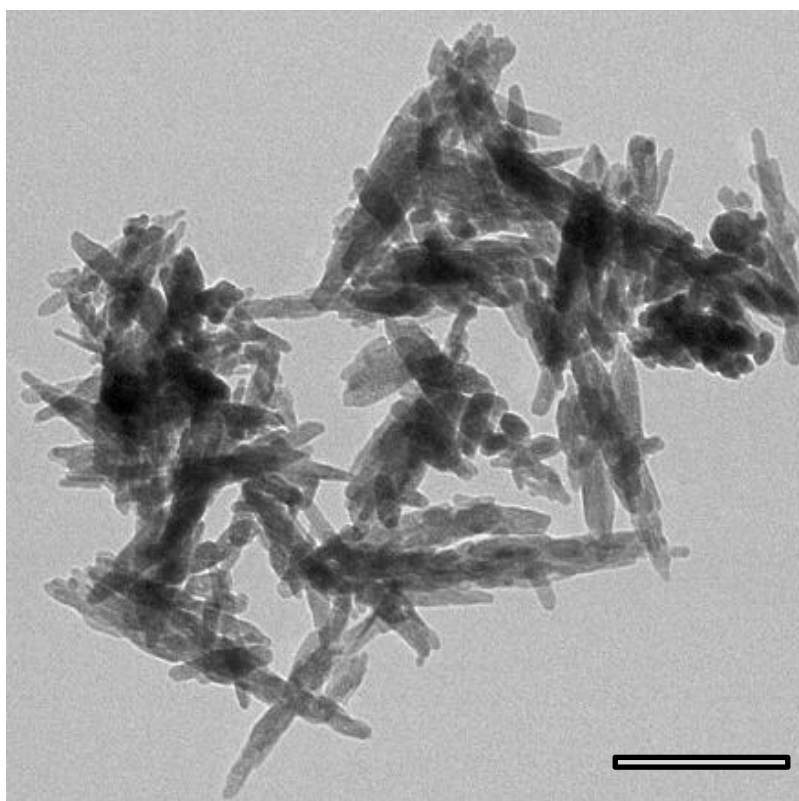


Figure S1 The TEM (HT7800, HITACHI) image of HANPs after sterilization at 200 °C for 2 h. scan bar = 200 nm.

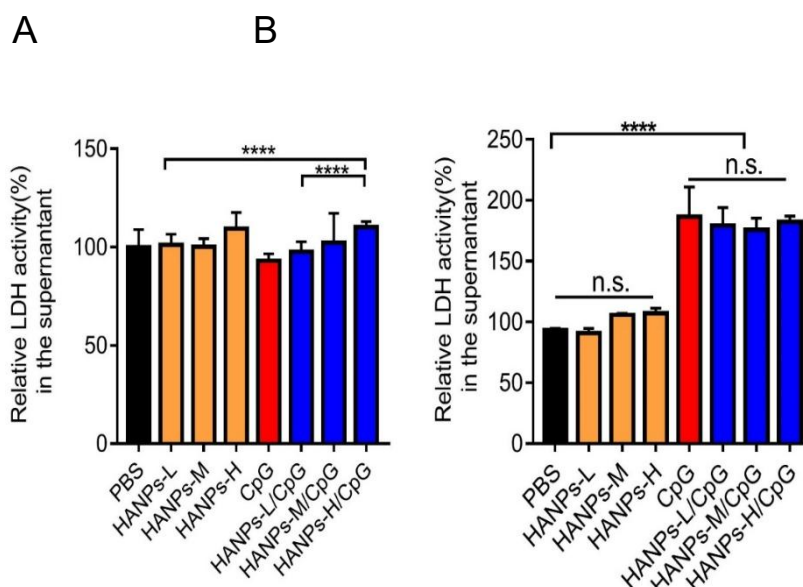


Figure S2 Relative LDH activity in the cell culture supernatant after macrophages were treated with PBS, HANPs at varying concentrations (50, 250 and 1000 $\mu\text{g/mL}$), CpG alone (0.5 $\mu\text{g/mL}$) or HANPs and CpG co-stimulation for 24 h (A) or 72 h (B). One-way ANOVA with Tukey's multiple comparisons test was performed for statistical analysis with ** ($p < 0.01$), *** ($p < 0.001$), **** ($p < 0.0001$). n.s. indicated no significance.

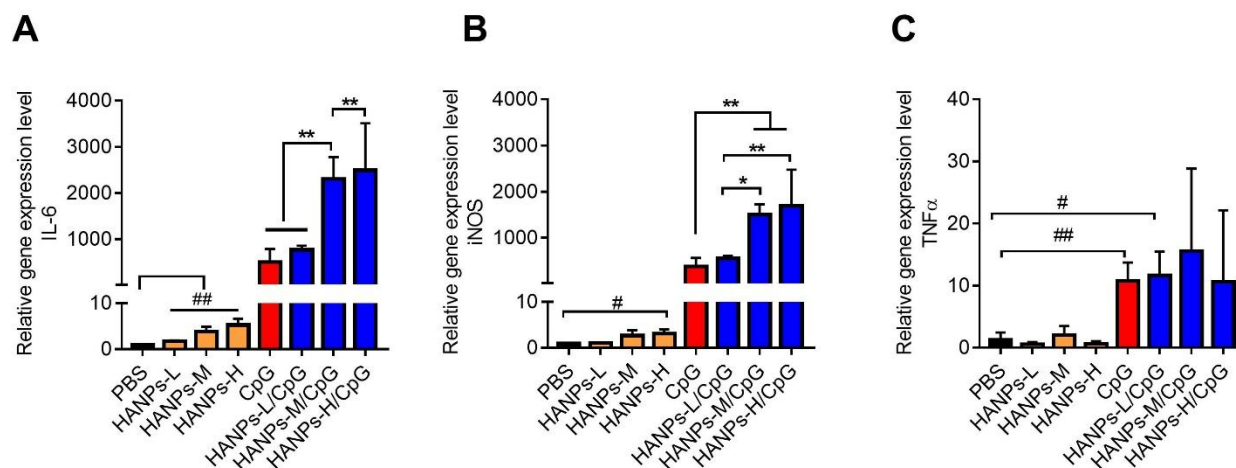


Figure S3 qRT-PCR analysis for mRNA expressing TNF- α (A), IL-6 (B) and INOS (C) by macrophage after the cells were treated with PBS, CpG ODN alone (0.5 $\mu\text{g/mL}$), HANPs at varying concentration (50, 250, and 1000 $\mu\text{g/mL}$), or co-stimulators of CpG ODN and HANPs for 24 h. One-way ANOVA with Tukey's multiple comparisons test was performed for statistical analysis with * ($p < 0.05$), ** ($p < 0.01$). Student's t-test was used to test compare two groups with # ($p < 0.05$), ## ($p < 0.01$).

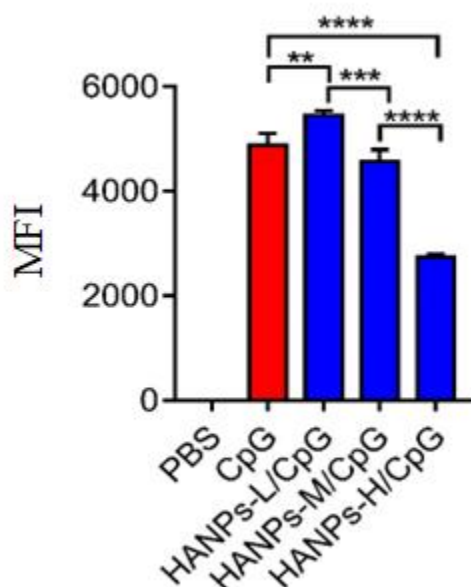


Figure S4 Mean fluorescence intensity (MFI) of Cy5-CpG in macrophage following the cells received treatments of PBS, CpG (0.5 $\mu\text{g/mL}$) in the presence or absence of HANPs at varying concentrations (50, 250, 1000 $\mu\text{g/mL}$). One-way ANOVA with Tukey's multiple comparisons test was performed for statistical analysis with ** ($p<0.01$), *** ($p<0.001$), **** ($p<0.0001$).

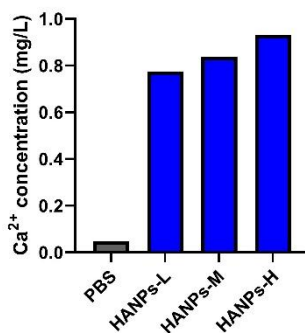


Figure S5 The release of Ca^{2+} after HANPs with varying concentration were incubated in PBS (pH 7.4) for 24 hours.

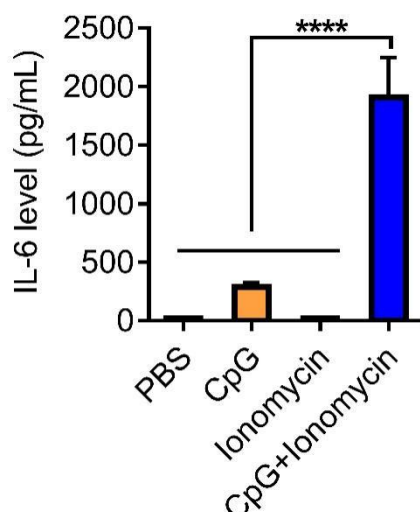


Figure S6 The level of IL-6 secreted by RAW 264.7 exposed to PBS, CpG (0.5 $\mu\text{g/mL}$), Ionomycin (1 μM) and co-stimulation of CpG and Ionomycin for 48 h. One-way ANOVA with Tukey's multiple comparisons test was performed for statistical analysis with **** ($p < 0.0001$).

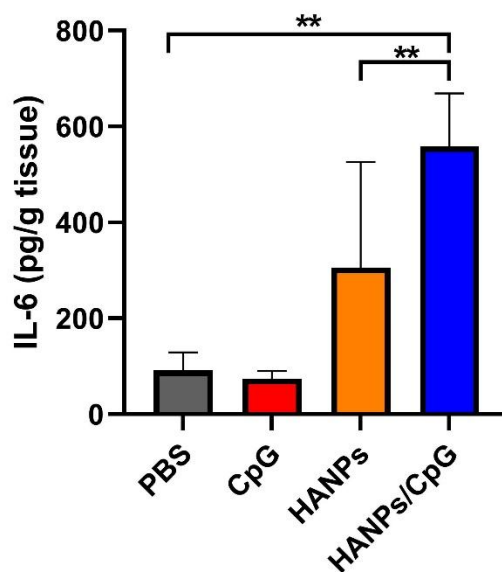


Figure S7 The level of IL-6 in the injection tissue. C57BL/6 mice were injected subcutaneously with PBS, CpG (5 μg), HANPs (5 mg), or a mixture of HANPs and CpG. 7 days later, the injection site was excised and homogenized, and IL-6 was determined by ELISA.

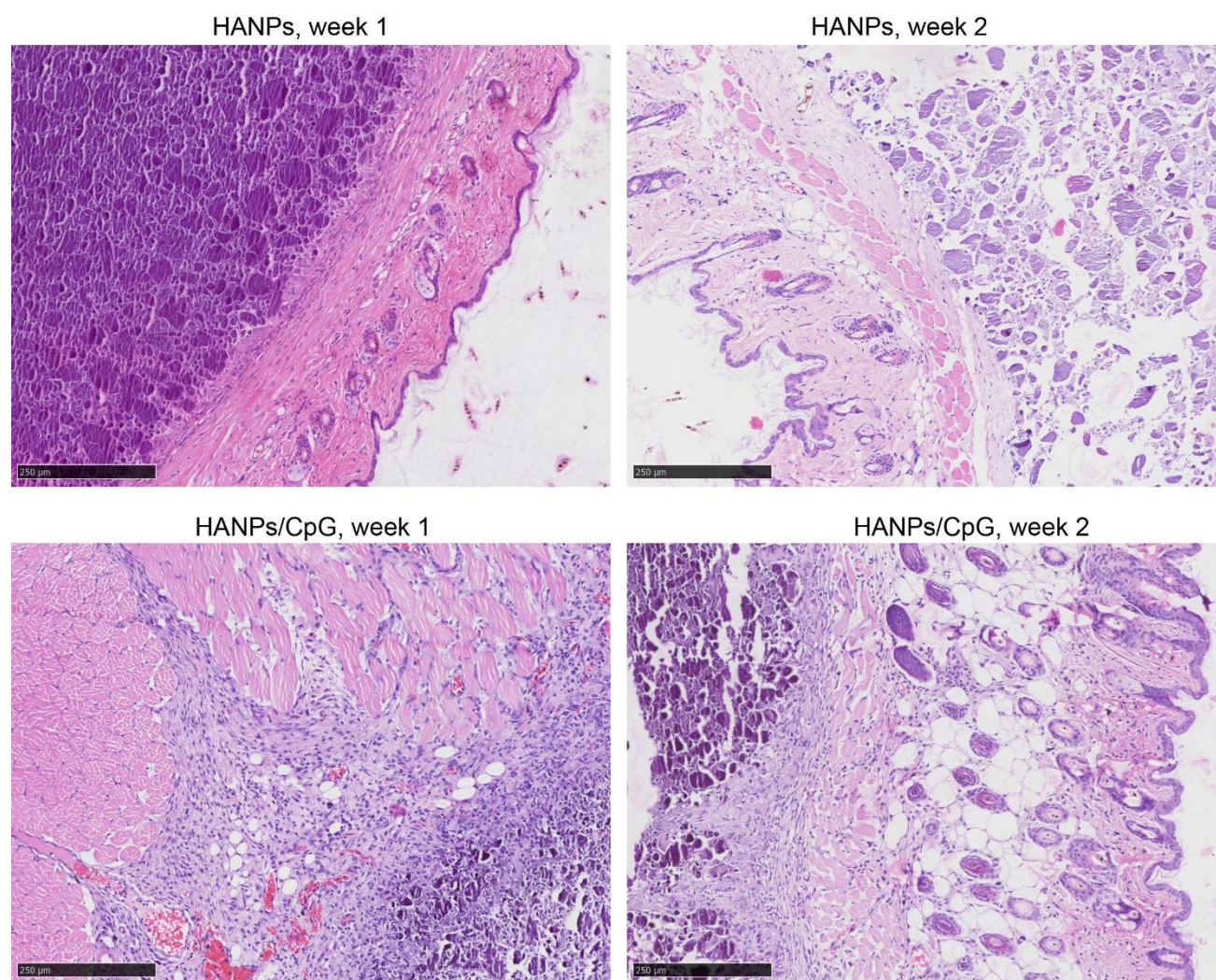


Figure S8 H&E staining of the surrounding tissues together with the biomaterials after HANPs or HANPs/CpG were subcutaneously injected into the mice for 1 and 2 weeks. Scale bar = 250 μm.

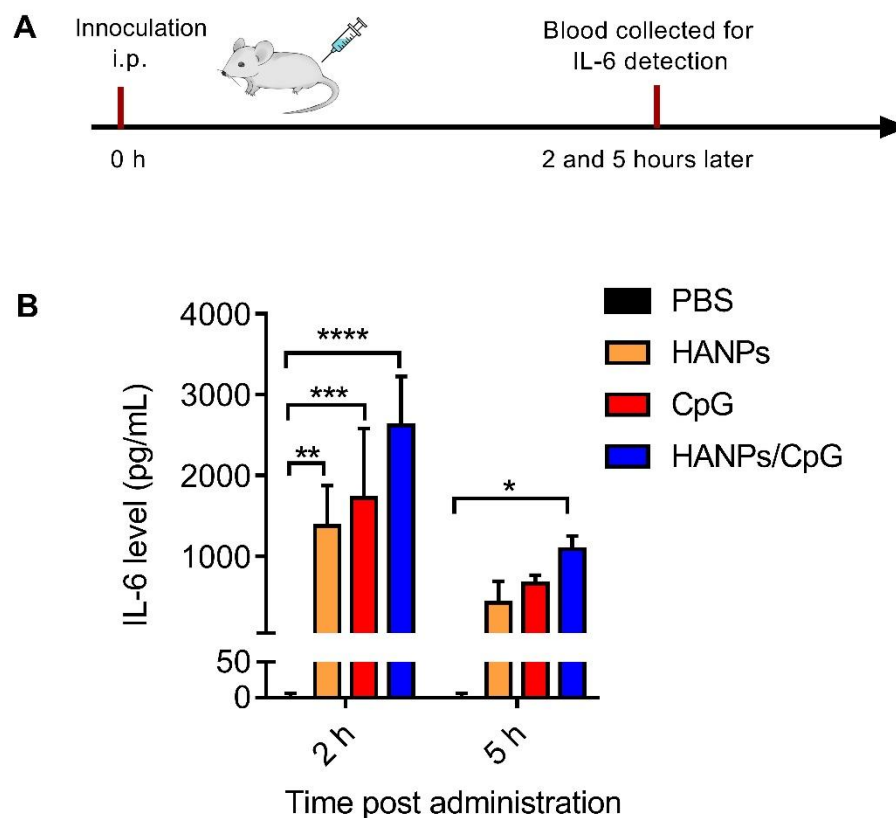


Figure S9 The schematic of administration and serum collection. Serum IL-6 level of mice following intraperitoneal (i.p.) treatment with PBS, HANPs, CpG alone or HANPs and CpG co-stimulation for 2 and 5 h (B). One-way ANOVA with Tukey's multiple comparisons test was performed for statistical analysis with ** ($p < 0.01$), *** ($p < 0.001$), **** ($p < 0.0001$).

Table 1 qPCR primer sequences for mRNA encoding INOS, TNF α and IL-6

Gene	Primer sequences
TNF α	F: 5'-TGGGAGTAGACAAGGTACAACCC-3'
	R: 5'-CATCTTCTCAAAATTCGAGTGACAA-3'
IL-6	F: 5'-GAGGATACCACTCCCAACAGACC-3'
	R: 5'-AAGTGCATCATCGTTGTTCATACA-3'
INOS	F: 5'-GTGACGGCAAACATGACTT-3'
	R: 5'-TCGATGCACAACCTGGGT-3'