## **Short Communication**

## Unexpected expression of the 250 kD melanoma-associated antigen in human sarcoma cells

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One of the melanoma-associated antigens most extensively studied is the core glycoprotein (Mol. wt = 250 kD) of a cell membrane chondroitin sulfate proteoglycan (Bumol & Reisfeld, 1982). Among several monoclonal antibodies that recognize this antigen (Kantor et al., 1982) the 9.2.27 antibody (Morgan et al., 1981), has been found to bind to tumour cells from more than 90% of human melanomas (Oldham et al., 1984). Although some early reports indicated that the 9.2.27 antibody may cross-react with some types of normal cells (Lloyd et al., 1982; Johnson & Riethmüller, 1982) and non-melanoma tumours (Carrel et al., 1982; Saxton et al., 1982), the antigen regonized by the 9.2.27 antibody has been considered to be rather specifically expressed in melanoma cells. Recently, the 9.2.27 antibody has been successfully used for radioimaging of human melanomas growing as xenografts in nude mice (Hwang et al., 1985), and it has been investigated for use as an in vivo carrier for toxins (Bumol et al., 1983). Clinically, the antibody has been employed for immunotherapeutic purposes (Oldham et al., 1984).

During testing of the binding specificity of two monoclonal antibodies (TP-1 and TP-3) developed in our laboratory against human sarcomaassociated antigens (Bruland *et al.*, 1986), we recently made the accidental observation that the 9.2.27 antibody, included as a supposedly negative control, showed significant binding to several human sarcoma cell lines, as judged by indirect immunofluoresence on unfixed cells.

To study the reactivity of the antibody with human sarcomas in more detail, the ability of two sarcoma cell lines to bind 9.2.27 antibody, labelled with <sup>125</sup>I by the Iodo-Gen method (Fraker & Speck, 1978), was measured and compared to that of FEMX melanoma cells. One non-small cell lung cancer and 4 primary cultures of skin fibroblasts were used as controls. It was found (Table I) that the sarcoma cell lines bound equal or higher

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amounts of the antibody than the FEMX melanoma line, which is known to have high levels of the 250 kD antigen (Godal *et al.*, 1986). Furthermore, appreciable amounts of labelled antibody were bound to cultured fibroblasts (Table I).

The binding of the 9.2.27 antibody to sarcomas and normal connective tissues obtained directly from patients was then examined. Table II demon-

 Table I Binding of <sup>125</sup>I-labelled 9.2.27 antibody

Cell-line	Bound c.p.m. (% of total)
Sarcoma 1 (OHS)	50
Sarcoma 2 (PE)	22
Melanoma (FEMX)	20
Fibroblast 1 (sarcoma patient)	24
Fibroblast 2 (sarcoma patient)	15
Fibroblast 3 (non-cancer patient)	16
Fibroblast 4 (non-cancer patient)	8
Lung cancer (SELS)	0.3ª

 $^{\rm a}$  <0.5% bound/total is considered as negative. One ml of a cell suspension containing 10<sup>5</sup> cells in PBS in the presence of 1 mg ml<sup>-1</sup> human serum albumin were incubated with 10 ng of labelled 9.2.27 antibody at 4°C for 2 h. After washing, the cell associated radioactivity was measured.

 Table II Immunostaining with the 9.2.27 antibody on frozen sections

No. pos./No. test.
7/9
5/7
2/3
2/2
0/3

Acetone-fixed cryostat sections were incubated with  $10 \,\mu g \,ml^{-1}$  of the 9.2.27 antibody for 1 h at room temperature. After washing, bound antibody was detected by the use of Vectastain peroxidase ABC kit, following the manufacturer's instructions.

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strates the results of immunohistochemical studies obtained with the antibody on acetone-fixed cryostat sections, using the Vectastain peroxydase ABC kit (Vector Lab. Inc., Burlingame, CA, USA). It is seen that 7 of 9 osteosarcomas were positively stained. Similarly, 2/3 malignant Schwannomas, 5/7 malignant fibrous histiocytomas and 2/2 synovial sarcomas were positive, indicating that the 250 kD antigen may be widely distributed in human sarcomas. The sections of fibrous connective tissues, however, were not stained with the antibody, in contrast to the finding (Table I) that cultured fibroblasts bound fairly high amounts of labelled antibody. Since it is known that the expression of cellular antigens may depend on the conditions under which the cells are growing (Hosoi et al., 1982; Zwadlo et al., 1985), it seems reasonable to assume that in cultured fibroblasts the expression of the 250 kD antigen may be induced during in vitro growth.

The ability of the 9.2.27 antibody to bind to sarcomas *in vivo* was studied by immunoscintigraphy. One  $\mu g$  (40  $\mu$ Ci) of  $^{131}$ I-labelled F(ab')<sub>2</sub>fragments of the antibody was injected into nude mice carrying a human osteogenic sarcoma in their left flank and a human malignant melanoma in their right. Scintigrams obtained 20 h after injection of the labelled antibody, showed that the radioactivity localized even better in the sarcoma than in the melanoma (Figure 1). In contrast, uptake of labelled TP-1 anti-sarcoma antibody was restricted to the sarcoma xenograft (not shown).

In attempts to confirm that the antigen in sarcoma cells and cultured fibroblasts, to which the 9.2.27 antibody binds, actually is the 250 kD core



**Figure 1** Immunoscintigram of a nude mouse bearing two human osteogenic sarcoma xenografts in its left flank and a human malignant melanoma xenograft in its right flank. Picture taken 20 h after i.v. injection of  $1 \mu g (40 \mu Ci)^{131}$ -labelled 9.2.27 antibody. (Posterior view).

protein, immunoprecipitation was carried out as described by Bumol & Reisfeld (1982) on extracts from cells labelled for 20 h with <sup>35</sup>S-methionine  $(0.1 \text{ mCi ml}^{-1})$ . The immunoprecipitate was analyzed on a 5% polyacrylamide gel in the presence of SDS (Laemmli, 1970). It is seen (Figure 2) that the 9.2.27 antibody precipitates the 250 kD core protein from extracts of sarcoma cells and fibroblasts as well as from extracts of melanoma cells. In addition, higher mol. wt products were precipitated from all extracts. These precipitates showed, however, different migration patterns depending on the cell type from which they originated. This finding probably indicates that varying amounts of chondroitin sulfate side-chains may be associated with the protein core in the different cells examined. A protein of mol. wt  $\sim 210 \text{ kD}$  was precipitated from the extract of fibroblasts, presumably representing a precursor for the 250 kD protein similar to that described of Bumol et al. (1984). None of these high mol. wt proteins was detected in the extract from the SELS lung carcinoma cells, which had failed to bind (Table I) labelled 9.2.27 antibody.

Interestingly, when immunoprecipitation was carried out on spent medium obtained after incubation of labelled sarcoma and melanoma cells for 24 h in normal medium, appreciable amounts of the high mol. wt (HMW) proteoglycan as well as

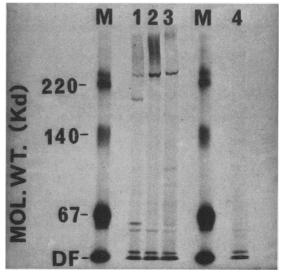


Figure 2 Autoradiograph of immunoprecipitates obtained with 9.2.27 antibody on extracts from  $^{35}$ Smethionine-labelled cells analyzed on a 5% polyacrylamide gel in the presence of SDS. 1: cultured fibroblasts; 2: osteogenic sarcoma; 3: malignant melanoma; 4: lung carcinoma; M:  $^{125}$ I-labelled mol. wt marker (HMW, Pharmacia Fine Chemicals, Uppsala, Sweden); DF: dye front.

the 250 kD antigen were detected (not shown), showing that the antigen is shed by both sarcoma and melanoma cells. This observation is in contradiction to the findings of Bumol *et al.* (1984), who in spent medium from melanoma cells were able to detect only the HMW proteoglycan. The discrepancy may possibly be ascribed to differences in the experimental procedures followed, as in their case the tumour cells were labelled with <sup>35</sup>Smethionine for a very short period of time (10 min).

In conclusion, the present data show that the 250 kD core protein is present on the cell surface of human sarcoma cells and cultured human fibroblasts as well as of malignant melanoma cells. The demonstration here of a wider distribution of this

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antigen than previously recognized raises interesting questions as to its biological function. Moreover, it is clear from the present findings that the 9.2.27 antibody cannot be used for diagnostic purposes to distinguish between melanomas and sarcomas. That the 250 kD antigen was found to be expressed in sarcomas *in vivo*, however, opens the possibility of using the 9.2.27 antibody in combination with antisarcoma antibodies in immunoscintigraphy and therapy of human sarcomas.

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