



# Review of Natural Resources With Vasodilation: Traditional Medicinal Plants, Natural Products, and Their Mechanism and Clinical Efficacy

## OPEN ACCESS

Fei Tang<sup>1†</sup>, Hong-Ling Yan<sup>1†</sup>, Li-Xia Wang<sup>1</sup>, Jin-Feng Xu<sup>1</sup>, Cheng Peng<sup>1\*</sup>, Hui Ao<sup>2\*</sup> and Yu-Zhu Tan<sup>1\*</sup>

### Edited by:

Yu-Yo Sun,  
University of Virginia,  
United States

### Reviewed by:

Simona Saponara,  
University of Siena, Italy  
Shaobo Ding,  
Southern Medical University, China

### \*Correspondence:

Cheng Peng  
pengchengchengdu@126.com  
Hui Ao  
aohui2005@126.com  
Yu-Zhu Tan  
tanyuzhu@cdutcm.edu.cn

<sup>†</sup>These authors have contributed  
equally to this work

### Specialty section:

This article was submitted to  
Cardiovascular and Smooth Muscle  
Pharmacology,  
a section of the journal  
Frontiers in Pharmacology

**Received:** 09 November 2020

**Accepted:** 29 January 2021

**Published:** 01 April 2021

### Citation:

Tang F, Yan H-L, Wang L-X, Xu J-F,  
Peng C, Ao H and Tan Y-Z (2021)  
Review of Natural Resources With  
Vasodilation: Traditional Medicinal  
Plants, Natural Products, and Their  
Mechanism and Clinical Efficacy.  
*Front. Pharmacol.* 12:627458.  
doi: 10.3389/fphar.2021.627458

<sup>1</sup>State Key Laboratory of Characteristic Chinese Medicine Resources in Southwest China, Pharmacy College, Chengdu University of Traditional Chinese Medicine, Chengdu, China, <sup>2</sup>Innovative Institute of Chinese Medicine and Pharmacy, Chengdu University of Traditional Chinese Medicine, Chengdu, China

For decades, chronic diseases including cardiovascular and cerebrovascular diseases (CCVDs) have plagued the world. Meanwhile, we have noticed a close association between CCVDs and vascular lesions, such as hypertension. More focus has been placed on TMPs and natural products with vasodilation and hypotension. TMPs with vasodilatory and hypotensive activities are mainly from *Compositae*, *Lamiaceae*, and *Orchidaceae* (such as *V. amygdalina Del.*, *T. prociuibens L.*, *M. glomerata Spreng.*, *K. galanga L.*, etc.) whereas natural products eliciting vasorelaxant potentials were primarily from flavonoids, phenolic acids and alkaloids (such as apigenin, puerarin, curcumin, sinomenine, etc.). Furthermore, the data analysis showed that the vasodilatory function of TMPs was mainly concerned with the activation of eNOS, while the natural products were primarily correlated with the blockage of calcium channel. Thus, TMPs will be used as alternative drugs and nutritional supplements, while natural products will be considered as potential therapies for CCVDs in the future. This study provides comprehensive and valuable references for the prevention and treatment of hypertension and CCVDs and sheds light on the further studies in this regard. However, since most studies are *in vitro* and preclinical, there is a need for more in-depth researches and clinical trials to understand the potential of these substances.

**Keywords:** traditional medicinal plants, natural products, vasodilation, mechanism, cardiovascular and cerebrovascular diseases

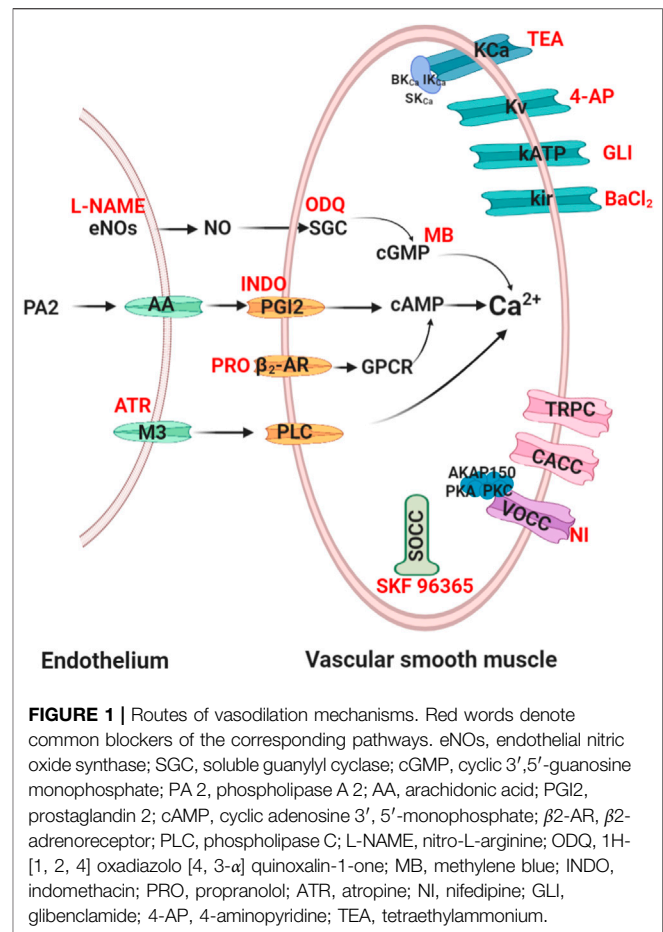
**Abbreviations:** SBP, systolic blood pressure; TC, total cholesterol; LDL-C, low-density lipoprotein cholesterol; HDL-C, high-density lipoprotein cholesterol; TG, triglycerides; non-HDL-C, non-high-density lipoprotein cholesterol; MS, metabolic syndrome; HAA, horizontal aspartate aminotransferase; ADAS-Cog, Alzheimer's disease assessment scale-cognitive subscale; GERRI, geriatric evaluation by relative's rating instrument; CGIC, clinical global impression of change; NIHSS, National Institutes of Health Stroke Scale.

## INTRODUCTION

It is universally acknowledged that the pathogenesis of cardiovascular and cerebrovascular diseases (CCVDs) is complex and long, which is an obstacle to the development of human health. In 2011, the United Nations recognised non-communicable diseases, including CCVDs, as the major health barriers for humans and developed plans to reduce the impact of these diseases (Mensah and Mayosi, 2013). According to WHO, 31% of the world's deaths in 2016 were from CCVDs. These causes included ischaemic heart disease (IHD), ischaemic stroke, haemorrhagic stroke, atrial fibrillation, peripheral arterial disease, aortic aneurysm, atherosclerotic, cardiomyopathy, and myocarditis, hypertensive heart disease, endocarditis, rheumatic heart disease, arrhythmias, etc (Kratz et al., 2017; Roth et al., 2017). Importantly, many common chronic CCVDs, such as heart failure, myocardial infarction, stroke, vascular dementia, and chronic kidney diseases, are closely related to hypertension (Sureda et al., 2017). However, hypertension is often caused by vascular lesions such as vessel wall thickening, vessel stenosis, vessel occlusion and endothelial cell injury (Mathieu et al., 2009; Farias, Pereira and Rosa 2010; Howlett 2014; Singh et al., 2017). For example, hypertension is more likely to cause IHD, stroke, cerebral haemorrhage and cerebral ischaemia in the elder population (Collaboration, 2002). Additionally, diabetic vasculopathy is concerned with vessel wall thickening and endothelial cell damage, which is the main cause of blindness, kidney failure, heart attacks, and stroke (Wimmer et al., 2019). Furthermore, hypertensive patients are also more likely to develop diabetes. (Sowers et al., 2001).

Remarkably, WHO encourages the use of traditional medicinal plants (TMPs) to improve various chronic diseases with increasing risk around the world. TMPs were shown to be beneficial for human health due to the richness of active compounds. Natural products, referring to small molecules derived from TMPs, also improve processes of biological metabolism through regulating the activity of enzymes in the body (Loai and Zohar, 2015; Briguglio et al., 2018). What's more, TMPs have been used as food supplements to treat chronic diseases without prescription. Currently, TMPs and natural products are playing an indispensable part for improving human health as a complementary alternative therapy, although some side effects have been found. In the past two decades, scientists have explored the vasodilation of TMPs and natural products, and roughly explained their mechanism. These natural resources have also been proved to be potentially effective for the treatment of CCVDs in clinic and drawn greater attention, even in developed countries including the United States and Australia. (Delfan et al., 2014; Liu and Huang, 2016; Shaito et al., 2020). However, the vasodilatory activities, the underlying mechanism and the clinical efficacy of TMPs and natural products, had not been comprehensively over-reviewed.

This article reviews the underlying mechanism and the clinical efficacy of TMPs and natural products with vasodilation, and puts forward a view that TMPs and natural products can be used as adjuvants in the prevention and treatment of CCVDs. We hope this review will lay the foundation for an in-depth investigation



**FIGURE 1 |** Routes of vasodilation mechanisms. Red words denote common blockers of the corresponding pathways. eNOS, endothelial nitric oxide synthase; SGC, soluble guanylyl cyclase; cGMP, cyclic 3',5'-guanosine monophosphate; PA 2, phospholipase A 2; AA, arachidonic acid; PGI<sub>2</sub>, prostaglandin 2; cAMP, cyclic adenosine 3', 5'-monophosphate; β<sub>2</sub>-AR, β<sub>2</sub>-adrenoreceptor; PLC, phospholipase C; L-NAME, nitro-L-arginine; ODQ, 1H-[1, 2, 4] oxadiazolo [4, 3-a] quinoxalin-1-one; MB, methylene blue; INDO, indomethacin; PRO, propranolol; ATR, atropine; NI, nifedipine; GLI, glibenclamide; 4-AP, 4-aminopyridine; TEA, tetraethylammonium.

on the development and utilization of nature resources in this field.

## SEARCH STRATEGY

This review aims to provide readers with a brief account of the main achievements in the field of vasodilation-based natural products and plant extracts from TMPs worldwide during the years of 1998–2020. To this end, the literatures from several databases including PubMed, Web of Science and Google Scholar were searched to obtain any studies evaluating TMPs and natural products with vasodilation. The data collected in this review were limited to English articles that primarily focused on the vasodilation and the underlying mechanisms of screening of TMPs and natural products.

## VASODILATION MECHANISM OF TMPs AND NATURAL PRODUCTS

The vascular tone changes of vascular smooth muscle cells (VSMCs) occurs through electromechanical and chemomechanical coupling and then acting on α-receptor, angiotensin-converting enzymes, angiotensin receptor or

potassium/calcium channels. The detailed mechanism of action is as follows (Hill et al., 2001; Cole and Welsh, 2011). **Figure 1** shows a brief description of the vascular tone changes of VSMs to support our understanding of vasodilatory mechanisms (Alexander et al., 2019a; Alexander et al., 2019b).

## Endothelium-Mediated Vasodilation Mechanism

Endothelial-derived relaxing factor (EDRF), including nitric oxide (NO) and prostacyclin from endothelial cells, plays an important role in vasodilation (Török, 2000; Tokoudagba et al., 2010).

### NO Signalling Cascade-Mediated Vasodilation

NO is the main EDRF that induces relaxation of VSMCs. In addition, NO also plays a crucial role in CCVDs such as atherosclerotic disease (Malekmohammad et al., 2020). Endothelial nitric oxide synthase (eNOS) catalyses the production of NO, which diffuses into VSMCs and then enhances cGMP synthesis. cGMP activates dependent protein kinase to cause vasodilation by reducing the intracellular  $\text{Ca}^{2+}$  concentration (Bian et al., 2008). Therefore, to validate the eNOS/NO/sGC/cGMP/signalling pathway, L-NAME (non-selective NO inhibitor) is used to inhibit NO synthesis. ODQ (sGC pathways inhibitor) and MB (cGMP pathways inhibitor) are usually used to inhibit the sGC/cGMP pathway (Bian et al., 2008).

### PGI2 Signalling Cascade-Mediated Vasodilation

Prostaglandin 2 (PGI<sub>2</sub>) is also a major EDRF, prostacyclin synthase-catalysed intermediate prostaglandin H<sub>2</sub>, and catalysed by cyclooxygenase (COX) for arachidonic acid synthesis. PGI<sub>2</sub> activates adenylate cyclase (AC) to produce cAMP and then regulates dependent protein kinase to reduce the intracellular  $\text{Ca}^{2+}$  concentration (Ameer O. Z. et al., 2010). Indomethacin is commonly used to inhibit COX and PGI<sub>2</sub> channels.

### Muscarinic Receptor Signalling Cascade-Mediated Vasodilation

M<sub>3</sub> receptor is a G $\alpha_q$ -protein-coupled receptor (GPCR) that presents in endothelium. (Walch et al., 2000). Atropine is commonly used to block phospholipase-C signalling pathways by inhibiting M<sub>3</sub>.

### VSMCs Mediated Vasodilation Mechanism

The contraction of VSMCs mainly rely on  $\text{Ca}^{2+}$  influx via regulating ion channels ( $\text{K}^+/\text{Ca}^{2+}$ ) or receptor channels.  $\text{Ca}^{2+}$  enter cells mainly through voltage dependent L-type  $\text{Ca}^{2+}$  channels (LTCC), store-operated calcium channels (SOCC) and transient receptor potential channels (TRPC). Of course,  $\text{Ca}^{2+}$  influx also relies on the regulation of  $\text{Ca}^{2+}$ -gated Cl channels (CACC) (Brozovich et al., 2016). Moreover,  $\text{K}^+$  channels also changes of vascular tone by regulating extracellular  $\text{Ca}^{2+}$  influx.

### $\beta_2$ -Adrenoreceptor-Mediated Vasodilation

$\beta_2$ -adrenoreceptor, a GPCR, only exists on the membrane of VSMCs. This receptor passes through the GPCR/AC pathway,

which catalyses the breakdown of ATP into cAMP and subsequently causes vasodilation (Ch'ng et al., 2017). Propranolol, a nonselective  $\beta_2$ -adrenergic receptor inhibitor, eventually causes vasoconstriction through inhibiting this channel.

### Potassium Ion Channels Mediated Vasodilation

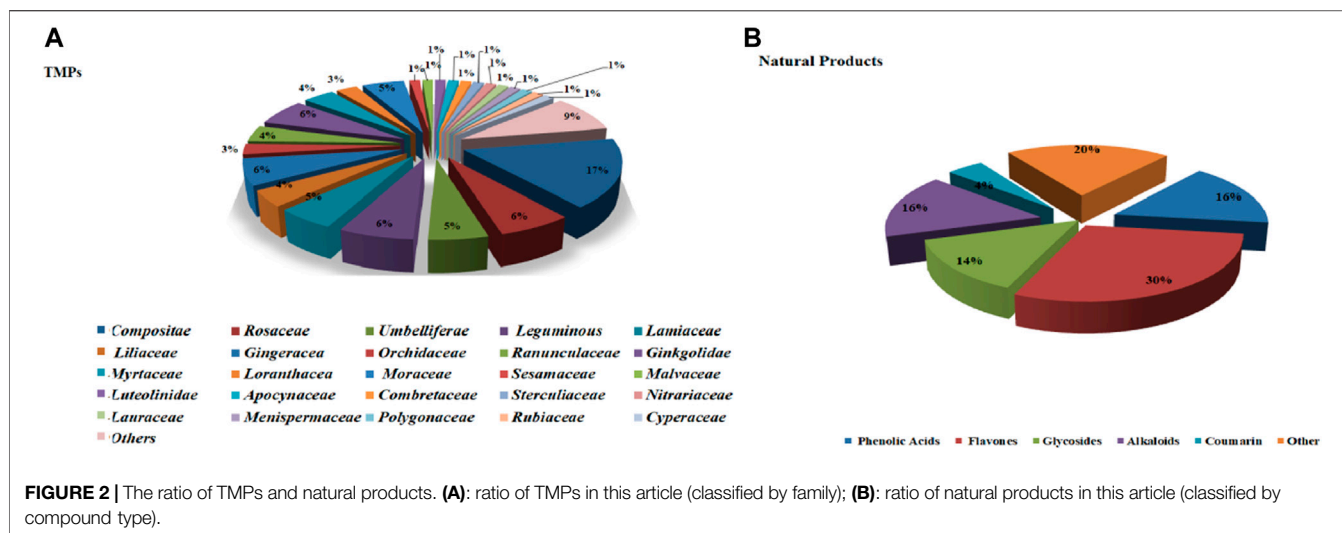
There are mainly four types of  $\text{K}^+$  channels in VSMCs: voltage-sensitive  $\text{K}^+$  channels (Kv), ATP-sensitive  $\text{K}^+$  channels (KATP), inward rectifier-type  $\text{K}^+$  channels (Kir) and  $\text{Ca}^{2+}$  activated  $\text{K}^+$  channels (KCa). KCa, widely appear in pulmonary artery smooth muscle cells, include large conductance  $\text{Ca}^{2+}$ -dependent K channels (BK channels), intermediate-conductance  $\text{Ca}^{2+}$  activated  $\text{K}^+$  channel (IKCa) and small-conductance  $\text{Ca}^{2+}$  activated  $\text{K}^+$  channel (SKCa), and the BK channel was associated with the  $\beta$ -1 subunit (Brenner et al., 2000). The four channels were blocked by 4-aminopyridine (4-AP), glibenclamide, BaCl<sub>2</sub>, and tetraethylammonium (TEA), respectively (Su et al., 2014). Furthermore, other channels such as KCNQ subfamily, EAG (ether-à-go-go or KCNH) subfamily,  $\text{Ca}^{2+}$ -activated Slo subfamily and  $\text{Ca}^{2+}$  and Na-activated SK subfamily were found in VSMCs (Brozovich et al., 2016; Barros et al., 2020).

### Calcium Ion Channels Mediated Vasodilation

There are mainly three types of  $\text{Ca}^{2+}$  channels in VSMCs membranes, including voltage-operated calcium channels (VOCC) and receptor-operated calcium channels (ROCC). VOCC were regulated by membrane potential-dependent voltage, and ROCC bind to GPCR, SOCC and so on. LTCC, as a predominant VOCC, has been shown to be concerned with the A-kinase anchor protein 150 (AKAP150) and protein kinase C/A (Navedo et al., 2008). SOCC is mediated by the sarcoplasmic reticulum (SR)  $\text{Ca}^{2+}$  sensor stromal interaction molecule (STIM) and orai channels (Trebak, 2012). TRPC, including TRPC, TRPM, TRPV, TRPA, TRPP, and TRPML, are nonselective cation channels that carry receptor-operated  $\text{Ca}^{2+}$  currents (ROCs) triggered by phospholipase C (PLC)-catalyzed hydrolysis of phosphatidylinositol 4,5-bisphosphate [PI(4, 5) P<sub>2</sub>] (Mori et al., 2015). These channels may be also stimulated in store-operated manner, via tyrosine kinases, or by lysophospholipids, hypoosmotic solutions, and mechanical stimuli (Chen et al., 2020). Several TRPC, such as TRPC1, TRPC3, TRPC6, and TRP M4 were found to be involved in vasoconstriction and more sensitive than LTCC (Beech, 2005). Additionally, it was indicated that both SOCC and ROCC are concerned with TRPC family (McFadzean and Gibson, 2002). The blockers of VOCC and SOCC are nifedipine, SKF 96365 or Gd<sup>3+</sup> respectively *in vitro* experiments (Xu et al., 2015; Alexander et al., 2019b).

## TRADITIONAL MEDICINAL PLANTS WITH VASODILATION *IN VITRO* AND ANTIHYPERTENSIVE ACTIVITIES *IN VIVO*

This section describes the vasodilatory and antihypertensive effects of plant extracts. Most extracts act on complex

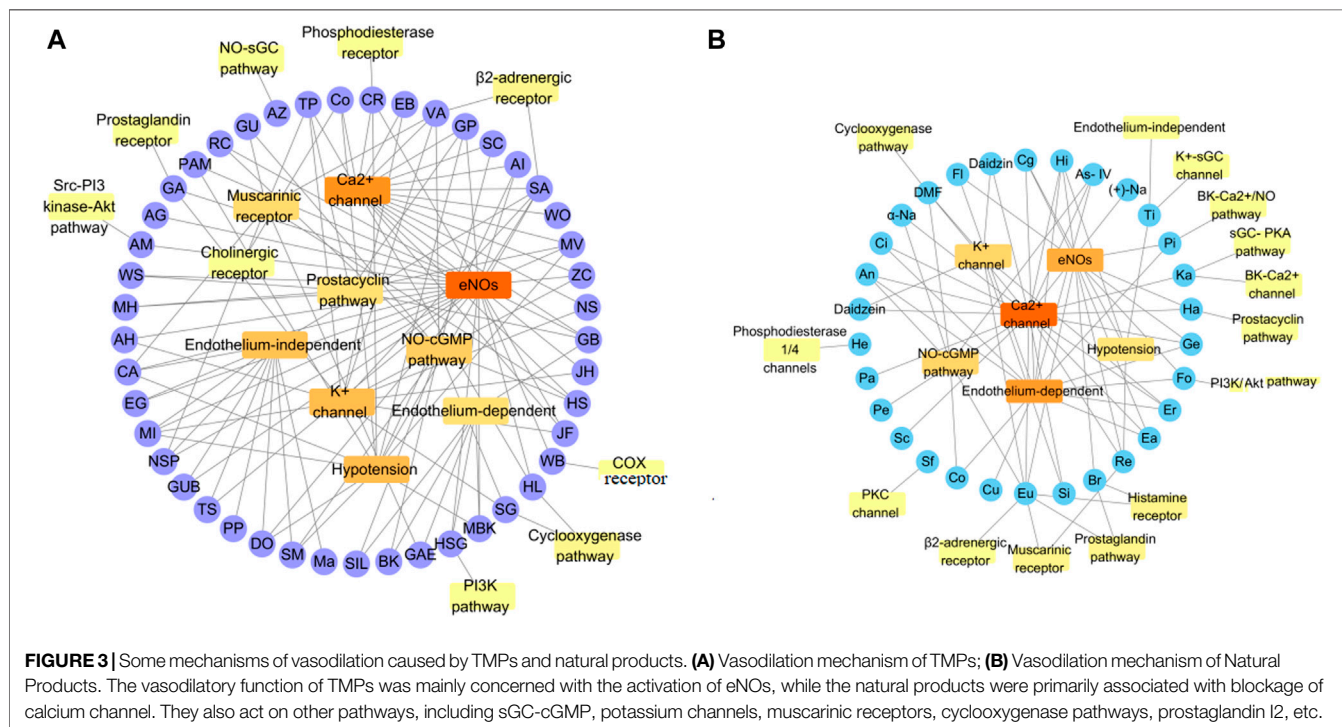


pathways for vasodilation or hypotension in rats. **Supplementary Table S1** shows the details of studies investigating vasodilation of TMPs. All the discussed TMPs, which were primarily from *Compositae* (17%), were shown in **Figure 2A**. TMPs mainly displayed vasodilation by acting on eNOS and Ca<sup>2+</sup> channels (**Figure 3A**). Most of TMPs, such as *V. amygdalina Del.*, *T. procuinbens L.*, *M. glomerata Spreng.*, *K. galanga L.*, etc., have significant vasodilatory bioactivities *in vitro*, but the hypotensive effect of only 40% of TMPs have been investigated *in vivo* at present, such as *G. procumbens*, *H. cere*, *Bidens pilosa Linn*, etc (**Figure 3A**). In addition, some TMPs

have significant vasodilation but the mechanistic exploration was insufficient, such as *G. procumbens*, *M glomerata Spreng.*

### Compositae *Erigeron breviscapus* (Vant.) Hand-Mazz.

*Erigeron breviscapus* (EB), as a traditional Chinese medicine, is commonly used for neuroprotection and vascular protection. EB induced relaxation on U44619 (Ca<sup>2+</sup> agonists, EC<sub>50</sub> 0.354 mg mL<sup>-1</sup>) pre-contracted aortic rings, which was abolished by glibenclamide (KATP inhibitor) or TEA (KCa channel inhibitor). However, this effect of EB was not affected by endothelial removal or ipilicoin





(BKCa inhibitor), BaCl<sub>2</sub> (Kir inhibitor) or 4-AP (Kv inhibitor). These results indicated that the vasodilatory activities of EB are mediated by Ca<sup>2+</sup> and KATP channels, rather than endothelial cells or K<sup>+</sup> channels (Pan et al., 2008).

### ***Vernonia amygdalina* Del.**

*Vernonia amygdalina* Del. (VA) is used to treat hypertension in Malaysia. The ethanol extract of VA leaves (VAE) caused the relaxation of Phenylephrine (PHE, 1 μM)-pre-contracted aortic rings (EC<sub>50</sub> 0.057 mg mL<sup>-1</sup>). However, its effects were significantly reduced by endothelial removal, TEA, 4-AP, BaCl<sub>2</sub>, glibenclamide, L-NAME (eNOS blocker), methylene blue (cGMP inhibitor), indomethacin (COX inhibitor), atropine (muscarinic receptor inhibitor) and propranolol (β<sub>2</sub>-adrenoreceptor inhibitor). Hence, the effect of VAE was related with the NO/cGMP/PGI<sub>2</sub> pathways, Ca<sup>2+</sup>/K<sup>+</sup> channels or β<sub>2</sub>-adrenergic receptor (Ch'ng et al., 2017). Additionally, VA (i.v. 10.0 mg kg<sup>-1</sup>) ultimately caused a reduction in blood pressure in SD rats (Taiwo et al., 2010).

### ***Tridax procuinbens* L.**

The water extract of the leaf from *Tridax procuinbens* (TP) has been shown to reduce blood pressure, but its mechanism remains unclear. The data showed that TP could significantly relieve the contraction caused by PHE (0.1 μM) and K<sup>+</sup> (60 mM). TP (10<sup>-9</sup>–10<sup>-5</sup> M) also antagonized the Ca<sup>2+</sup>-induced vasoconstriction in a Ca<sup>2+</sup>-free context by high K<sup>+</sup>. The activity of TP was repressed by BaCl<sub>2</sub> and apamin (K<sup>+</sup> channels blockers), L-NAME, indomethacin, atropine, propranolol, and methylene blue; however, it was not affected by glibenclamide, TEA, 4-AP or ODQ (guanylyl cyclase inhibitor). Therefore, this action is intervened by endothelial cells and partial ion channels (Salahdeen et al., 2015).

### ***Artemisia herba Alba* Asso.**

*Artemisia herba Alba* Asso (AH) is used to treat diabetes and hypertension in Morocco. AH relaxed the contraction elicited by noradrenalin (NE, 1 μM) in endothelium-containing aortas. However, its activity was significantly abolished by L-NAME (100 μM), endothelial removal, methylene blue (10 μM) and ODQ (50 μM) but not atropine (10 μM), TEA (5 mM), indomethacin (10 μM) or glibenclamide (10 μM). It was suggested that the vasodilation of AH occurs mainly through the activation of eNOS/SGC/cGMP pathways rather than K<sup>+</sup> channels (Skiker et al., 2010).

### ***Bidens pilosa* L.**

Previous studies have shown that the antihypertensive effect of *Bidens pilosa* (BP) extract is closely related to its vasodilatory activity. The extract of BP was shown to induce a concentration-dependent vasorelaxation of rat aortas pre-contracted by K<sup>+</sup> (60 mM, inhibition rate 90% at 1.5 mg mL<sup>-1</sup>) and NE (1 μM, inhibition rate 88% at 1.5 mg mL<sup>-1</sup>) which was significantly inhibited by indomethacin or pyrillamine maleate (histamine-1 inhibitor) (Nguelefack et al., 2005). Furthermore, the extract of BP leaves (10, 20, and 30 mg kg<sup>-1</sup>) decreased SBP by 18.26, 42.5

and 30% in normotensive rats and by 25.77, 38.96 and 28.64% in SHR, respectively. Moreover, it also induced hypotension by 27, 34.13 and 18.73% in salt-loaded hypertensive rats, respectively (Dimo et al., 2003). These results indicated that the effect is concerned with vasodilation.

### ***Gynura procumbens* L. Merr.**

*Gynura procumbens* (GP) has been shown to decrease blood pressure by inhibiting the angiotensin-converting enzyme. The present experiments showed that the aqueous extract of GP significantly reduced the contraction induced by angiotensin I and II in rat aortic rings, which was blocked by indomethacin (10 μM) or L-NAME (0.1 μM) (Poh et al., 2013). Moreover, the butanoic fraction of GP (2.5–5 mg mL<sup>-1</sup>) also released the PHE (1 μM)-/K<sup>+</sup> (80 mM)-induced contractions. GP (i.v., 10–20 mg kg<sup>-1</sup>) also reduced mean arterial pressure in anesthetized rats (Hoe et al., 2011). Therefore, it was suggested that GP causes vasodilation through upregulating eNOS levels.

### ***Helichrysum ceres* S. Moore.**

Ethnomedical evidences suggested that extracts from *Helichrysum* have anti-inflammatory and anti-allergic activities and are commonly used to treat renal and cardiopulmonary diseases. The researchers found that the ethanolic extract of *Helichrysum ceres* leaf (HCE) could relax atropine (1 μM)/NE (1 μM)/K<sup>+</sup> (20, 80 mM)-induced contractions which was weakened by L-NAME (100 μM) in rat aortic. Furthermore, HCE could cause hypotension in normotensive rats or Dahl salt sensitive hypertensive rats (Musabayane et al., 2008). It was shown that activity of HCE is related to the eNOS levels.

### ***Mikania glomerata* Spreng.**

*Mikania glomerata* (MG) is mainly used to treat respiratory diseases in Brazil. The aqueous extracts and hydroalcoholic extract (HAE) of MG leaves significantly inhibited the histamine contractions in isolated guinea pig tracheas. HAE also induced relaxation in guinea pig tracheas pre-contracted by histamine (IC<sub>50</sub> 0.34 mg mL<sup>-1</sup>), acetylcholine (IC<sub>50</sub> 0.72 mg mL<sup>-1</sup>) or K<sup>+</sup> (IC<sub>50</sub> 1.41 mg mL<sup>-1</sup>). Moreover, the dichloromethane fraction of MG could relax the isolated mesenteric vascular or aorta in rats (Soares de Moura et al., 2002). But the mechanism of MG needs more exploration in the future.

### ***Flos chrysanthemi* Indici**

The ethyl acetate extract from *Flos chrysanthemi* Indici (FCE, 9.4–150 mg/L) antagonized vasoconstriction induced by PHE (1 μM)/K<sup>+</sup> (60 mM), which was significantly inhibited by endothelium removal, L-NAME (10<sup>-4</sup> M), glibenclamide and methylene blue (10<sup>-5</sup> M). But this effect of FCE was not affected by TEA, BaCl<sub>2</sub>, 4-AP, 5-HD or propranolol. In addition, FCE attenuated PHE-induced contraction in calcium-free or potassium-free solutions by upregulated NO levels in rat aortic. Overall, it was showed that FCE is in association with regulation of the NO/cGMP pathway and inhibition of KATP (Jiang et al., 2005).

## Others

The hydroalcoholic extract of *Senecio nutans* sch. Bip and the aqueous extract of *Tanacetum vulgare* L were shown to relax the PHE/K<sup>+</sup>/NE-dependent contractions of rat aortic rings in an endothelium-dependent manner (Paredes et al., 2016), (Lahlou et al., 2008). Additionally, the dichloromethane extract of *Kaempferia galanga* L and the methanol extract from *Stevia rebaudiana* had shown antihypertensive activity in anaesthetised rats or SHR (Othman et al., 2006; Melis 1995). But the mechanism of these extracts is not sufficiently studied at present.

## Rosaceae

### *Crataegus gracilior* J. B. Phipps.

Hawthorn is used worldwide as traditional medicines to treat CCVDs, such as *Crataegus gracilior* J. B. Phipps (Mexican hawthorn, MH). The aqueous extracts of the leaves and fruits of MH elicited relaxation of aortic rings treated by PHE (1 μM), and its methanol extract (EC<sub>50</sub> 4.34 mg mL<sup>-1</sup>) had significant vasodilator effects (Hernández-Pérez et al., 2014). Additionally, the extract of *Crataegus* leaves and flowers caused relaxation in PHE (10 mM)-mediated rat aorta (IC<sub>50</sub> 15.1 μg mL<sup>-1</sup>) or human papillary artery (IC<sub>50</sub> 19.3 μg mL<sup>-1</sup>) from patients undergoing coronary artery bypass surgery. In short, the vasodilation of the extract is concerned with the eNOS pathway, but other pathways need to be further explored (Brixius et al., 2006).

### *Fragaria x Ananassa* Duch.

The vasodilation activity of the aqueous extract of *Fragaria x ananassa* Duch (wild strawberry, WS) leaves attenuated NE-induced (0.1 μM) vasoconstriction in rat aortas which was suppressed by L-NAME or indomethacin. Therefore, similar to the hawthorn aqueous extract, the aqueous extracts could cause endothelium-dependent vasodilation which were correlated with the NO/COX pathways (Hernández-Pérez et al., 2014).

### *Rubus chingii* Hu.

*Rubus chingii* (RC) is a commonly used traditional Chinese medicine that can improve renal function and treat polyuria. The ethanol extract of RC caused PHE (1 μM)-induced relaxation in rat aortas. However, this effect of RC was significantly attenuated by L-NAME (10 μM), ODQ (10 μM), diltiazem (10 μM, Ca<sup>2+</sup> antagonist), wortmannin (0.1 μM, PI3-kinase inhibitor), thapsigargin (1 μM, Ca<sup>2+</sup> modulator), Gd<sup>3+</sup> (10 μM, Ca<sup>2+</sup> modulator), 2-aminoethyl diphenylborinate (75 μM, Ca<sup>2+</sup> modulator) and 4-AP (1 mM), rather than TEA (1 mM), glibenclamide (10 μM), indomethacin, atropine, and propranolol. This study indicated that RC induces vasodilation in an endothelium-dependent manner, mainly by activating the Ca<sup>2+</sup>/eNOS and the NO/sGC/cGMP/KV channels (Su et al., 2014).

### *Aronia melanocarpa*

The high content of phenolic constituents of *Aronia melanocarpa* (AM) is characterized by a variety of biological activities.

Researchers have found that AM juice caused potent endothelium-dependent relaxations in porcine coronary artery rings, which was markedly abolished by L-NAME, PP2 (Src kinase inhibitor) and wortmannin. This result showed that AM juice promotes NO levels in the coronary artery endothelium by activating the Src/PI3 kinase/Akt pathway. Its main active components may be conjugated anthocyanins and chlorogenic acid (Kim et al., 2013).

### *Sorbus commixta* Hedl.

The cortex of this species has been used for antitussive purposes in oriental medicine. The methanol extract of *Sorbus commixta* cortex (SC) produced relaxation of the PHE-induced aorta which was attenuated by L-NAME, methylene blue, ODQ and endothelial removal except for indomethacin, glibenclamide, TEA, atropine or propranolol. Thus, its vasodilatory activity occurs through activation of the NO/cGMP pathway rather than blockage of KCa or KATP (Kang et al., 2005).

### *Carum roxburghianum* (DC.) Kurz.

*Carum roxburghianum* (CR) exhibit vasodilatory and cardiac modulatory actions. The crude extract of CR (10–100 mg kg<sup>-1</sup>) induced decrease arterial blood pressure of rats. Meanwhile, CR also inhibited high K<sup>+</sup> (80 mM)-/PHE (1 μM)-induced contractions in isolated rabbit aortas, which was not impaired by L-NAME. Therefore, CR extracts caused vasodilation might through antagonising Ca<sup>2+</sup> channels, regulating NO levels and inhibiting phosphodiesterase (Khan et al., 2015).

## Umbelliferae

### *Angelica dahurica* Bentham

*Angelica dahurica* Bentham (ADB) has been used for the treatment of CCVDs in Asia. The methanol extract of ADB (1 mg mL<sup>-1</sup>) resisted PHE (1 μM)-/K<sup>+</sup> (60 mM)-induced contractions in aortic rings but not caffeine (opener of ryanodine-sensitive receptors) in a Ca<sup>2+</sup>-free context (Lee et al., 2015). But the vasodilatory activity of ADB needs further investigation.

### *Ligusticum chuanxiong* Hort.

*Ligusticum chuanxiong* (Lc) have been widely used in the treatment of CCVDs in Asian countries. The CHCl<sub>3</sub> extract of Lc could inhibit contraction induced by NE in aortic strips and abolished Ca<sup>2+</sup>-independent contractions evoked by 12-deoxyphorbol 13-isobutyrate in Ca<sup>2+</sup>-free medium containing EGTA (1 mM). Furthermore, Lc significantly inhibited NE-mediated extracellular regulated protein kinases 1/2 (ERK1/2) activation but not P38 mitogen activated protein kinases (MAPK) (Kim B. et al. 2004).

### *Bupleurum fruticosum* L.

The studies have shown that *Bupleurum fruticosum* (BF) is beneficial to the heart and circulatory system. The chloroformic extract of the BF roots (0.1 mg mL<sup>-1</sup>) induced relaxation of rat thoracic aorta induced by NE or caffeine.

This activity was blocked by cyclopiazonic acid ( $\text{Ca}^{2+}$ -ATP blocker) (Testai et al., 2005).

### ***Coriandrum sativum* L.**

Coriander (Co) is traditionally used for gastrointestinal diseases and CCVDs. Researchers have found that Co could resist  $\text{K}^+$  (80 mM)-induced contractions in the rabbit jejunum and PHE (1  $\mu\text{M}$ )/ $\text{K}^+$  (80 mM)-induced contractions in rabbit aortas. Meanwhile, Co (1–30  $\text{mg mL}^{-1}$ ) caused hypotension in normal rats (Jabeen et al., 2009).

## **Leguminous**

### ***Glycyrrhiza uralensis* Fisch.**

*Glycyrrhiza uralensis* (GU) is used in traditional Chinese medicine that displays a variety of bioactivities. The GU caused relaxation of rat aortic rings treated by PHE (1  $\mu\text{M}$ )/ $\text{K}^+$  (80 mM) in endothelium-intact aortic rings. The vasodilatory activity of GU was weakened by L-NAME (10  $\mu\text{M}$ ), methylene blue (10  $\mu\text{M}$ ), indomethacin (10  $\mu\text{M}$ ), atropine (1  $\mu\text{M}$ ), propranolol (1  $\mu\text{M}$ ), glibenclamide (10  $\mu\text{M}$ ) and 4-AP (1  $\mu\text{M}$ ) except for TEA (1 mM) or  $\text{BaCl}_2$  (10  $\mu\text{M}$ ) (Tan et al., 2017). Thus, GU may relax blood vessels through activating the NO/sGC/cGMP pathway rather than  $\text{KCa}$  or Kir.

### ***Albizia inopinata* G. P. Lewis.**

The ethanol extract of *Albizia inopinata* G. P. Lewis (AI) could inhibit vasoconstriction induced by  $\text{K}^+$  (80 and 30 mM) or PHE (1  $\mu\text{M}$ ) ( $\text{IC}_{50}$  54, 52, and 65  $\mu\text{g mL}^{-1}$ , respectively). Additionally, AI antagonized contractions caused in  $\text{Ca}^{2+}$ -free medium induced by NE (1  $\mu\text{M}$ ) rather than caffeine (20 mM) in rat aortas (Pires et al., 2000). Furthermore, AI leaves abolished PHE (1  $\mu\text{M}$ )/ $\text{K}^+$  (80 mM)-induced vasoconstriction ( $\text{IC}_{50}$  65 and 54  $\mu\text{g mL}^{-1}$ , respectively) which was repressed by endothelial removal or L-NAME (10 and 100  $\mu\text{M}$ ) but not atropine (1  $\mu\text{M}$ ) or indomethacin (10  $\mu\text{M}$ ). Moreover, AI leaves (20  $\text{mg kg}^{-1}$ ) produced a significantly hypotensive effect and reduction in heart rate and cardiac output and total peripheral resistance in SHR (Maciel et al., 2004).

## **Others**

The total alkaloids of *Sophora alopecuroids* L. (SA, 40  $\text{mg L}^{-1}$ ) resisted concentrate induced by  $\text{K}^+$ / $\text{Ca}^{2+}$  in rabbit aortas, while was not significantly affected by removal of endothelium, L-NAME, indomethacin or propranolol (Zhang et al., 2009). Seeds of *Securigera securidaca* (SS) were used for the improvement of hyperlipidaemia in Iranian medicine. The hydroalcoholic extract of SS could improve vascular endothelium-dependent relaxation and decrease lipid levels and peroxidation in a rat model fed a high-fat diet (Garjani et al., 2009).

## **Lamiaceae**

### ***Mentha x villosa* Hubs.**

*Mentha x villosa* (MV) has anti-parasitic and tranquillising action and treatment of gastrointestinal diseases. Essential oil from MV induced significant and dose-dependent hypotensive and bradycardic responses, which were weakened by atropine

(2  $\text{mg kg}^{-1}$ ). Moreover, MV could attenuate PHE (1  $\mu\text{M}$ ) ( $\text{IC}_{50}$  255  $\mu\text{g mL}^{-1}$ ), prostaglandin F<sub>2</sub>- $\alpha$  (10  $\mu\text{M}$ ) ( $\text{IC}_{50}$  174  $\mu\text{g mL}^{-1}$ )/ $\text{K}^+$  (80 mM) ( $\text{IC}_{50}$  165  $\mu\text{g mL}^{-1}$ )-induced contractions in rat aortic rings. This effect was abolished by endothelial removal, L-NAME (100  $\mu\text{M}$ ) or indomethacin (10  $\mu\text{M}$ ) but not atropine (1  $\mu\text{M}$ ) (Guedes et al., 2004). The results suggested that activity of MV is primarily associated with endothelial cells, but the role of ion channels is not clear at present.

### ***Ziziphora clinopodioides* L.**

The  $\text{CHCl}_3$  extracts of *Ziziphora clinopodioides* L (ZC) inhibited PHE (1  $\mu\text{M}$ ,  $\text{EC}_{50}$  0.27  $\text{g L}^{-1}$ )/ $\text{K}^+$  (60 mM,  $\text{EC}_{50}$  0.34  $\text{g L}^{-1}$ )-induced contractions in rat aortic rings, which was significantly decreased by 4-AP (1 mM) or endothelial removal, rather than glibenclamide (100  $\mu\text{M}$ ), iberiotoxin (10 nm) or thapsigargin (100 nm). In  $\text{Ca}^{2+}$ -free solution, ZC also significantly inhibited vasoconstriction in  $\text{K}^+$ /PHE pre-contracted rings (Senejoux et al., 2010). It was shown that activity of ZC is mainly associated with endothelial cells and Kv.

### ***Salvia miltiorrhiza* Bqe.**

The roots of *Salvia miltiorrhiza* (SM) is used to treat CCVDs, such as angina pectoris and myocardial infarction in TCM. The aqueous extract of SM relaxed the NA-induced aorta which was abolished by L-NAME (100  $\mu\text{M}$ ), methylene blue (10  $\mu\text{M}$ ) and endothelium removal. Additionally, SM produced hypotensive response in normal rats through regulating release of angiotensin and bradykinin (Kamata et al., 1993). Moreover, SM caused hypotension in albino rats and rabbits, which was abolished by atropine and propranolol. Interestingly, low concentration of this extract, not higher concentration, induced vasodilation of the renal, mesenteric and femoral arteries (Lei and Chiou, 1986).

### ***Orthosiphon stamineus* Benth.**

*Orthosiphon stamineus* in folk medicine is used in the treatment of hypertension and kidney stones. The methanolic extract of *Orthosiphon stamineus* (CF) relieved PHE (1  $\mu\text{M}$ ) ( $\text{EC}_{50}$  2.21  $\mu\text{g mL}^{-1}$ )/ $\text{K}^+$  (60 mM) ( $\text{EC}_{50}$  3.32  $\mu\text{g mL}^{-1}$ )-induced constriction of rat thoracic aorta which was resisted by L-NAME (10  $\mu\text{M}$ ,  $\text{EC}_{50}$  17.8  $\mu\text{g mL}^{-1}$ ), methylene blue (10  $\mu\text{M}$ ,  $\text{EC}_{50}$  12.09  $\mu\text{g mL}^{-1}$ ), TEA (1 mM,  $\text{EC}_{50}$  24.35  $\mu\text{g mL}^{-1}$ ), 4-AP (1 mM,  $\text{EC}_{50}$  27.09  $\mu\text{g mL}^{-1}$ ),  $\text{BaCl}_2$  (10  $\mu\text{M}$ ,  $\text{EC}_{50}$  22.41  $\mu\text{g mL}^{-1}$ ), glibenclamide (10  $\mu\text{M}$ ,  $\text{EC}_{50}$  10.24  $\mu\text{g mL}^{-1}$ ) and propranolol (1  $\mu\text{M}$ ,  $\text{EC}_{50}$  7.76  $\mu\text{g mL}^{-1}$ ), rather than indomethacin (10  $\mu\text{M}$ ) ( $\text{EC}_{50}$  4.62  $\mu\text{g mL}^{-1}$ ) or ODQ (10  $\mu\text{M}$ ) ( $\text{EC}_{50}$  3.87  $\mu\text{g mL}^{-1}$ ). Thus, the vasodilatory activity of CF was in relationship with regulation of NO/cGMP pathway, muscarinic/ $\beta$ -adrenergic receptors and blockage of  $\text{Ca}^{2+}$  and  $\text{K}^+$  channels (Yam M. F. et al., 2016).

### ***Agastache mexicana* Ssp.**

The dichloromethane extract of *Agastache mexicana* (AM) significantly relaxed NA (0.1  $\mu\text{M}$ )-induced aortic contraction with endothelium ( $\text{EC}_{50}$  174  $\mu\text{g mL}^{-1}$ ) or without endothelium ( $\text{EC}_{50}$  293  $\mu\text{g mL}^{-1}$ ) which was significantly inhibited by TEA. In

conclusion, this activity is correlated with endothelial cells or K<sub>Ca</sub> (Flores-Flores et al., 2016).

### ***Marrubium vulgare* L.**

The water extract of *Marrubium* (Ma) showed a potent inhibition on K<sup>+</sup>-induced rat aorta contractions and decreased SBP by improving endothelial function in SHR (El Bardai et al., 2003). But mechanisms of Ma needs more exploration *in vivo* or *in vitro* in the future.

## **Liliaceae**

### ***Allium fistulosum* L.**

Welsh onion (WO, *Allium fistulosum* L.) has been consumed to prevent CCVDs. The WO extracts, particularly green-leaf extracts (RG, 30–40 mg mL<sup>-1</sup>), could resist NE (30 nM)-induced contractions in aortic ring. The activity was abolished by L-NAME (100 μM), TEA (1 mM) and SQ29548 (TXA<sub>2</sub>-receptor antagonist, 10 μM) (Chen et al., 1999). The hydroalcoholic extract of onion peel relaxed vasoconstriction induced by K<sup>+</sup>/PHE which was not attenuated by endothelial removal, L-NAME (100 μM), methylene blue (10 μM) and indomethacin (10 μM) in rat thoracic aortas (Naseri et al., 2008). Thus, the activity of WO is independent of the eNOS/cGMP pathway and whether it acts on ion channels needs further investigation.

### ***Allium sativum* L.**

The extracts of *Allium sativum* (AS) induce hypotension in hypertensive patients. AS could relieve NE (3 μM)-induced contractions in the aortic ring which was resisted by L-NAME (100 μM) and indomethacin (5 μM) (Takashima et al., 2017). Thus, the application about AS also needs to be supported by more research in CCVDs.

## **Gingeraceae**

### ***Alpinia zerumbet* (Pers.) Burt. et Smith.**

The hydroalcoholic extract of *Alpinia zerumbet* leaves (AZ) could resist NE-induced contractions which was not suppressed by indomethacin, 4-AP or glibenclamide except for L-NAME and ODQ (de Moura et al., 2005). The essential oil of AZ (0.01–3,000 mg mL<sup>-1</sup>) also relieved PHE-induced contractions which was inhibited by L-NAME and endothelial removal but not TEA (500 mM) or indomethacin (10 mM) (Pinto et al., 2009). Thus, the activity of AZ is mainly concerned with endothelial cells.

### ***Curcuma comosa* Roxb.**

Researchers have found that *Curcuma comosa* Roxb (CC) prevented the impairment of vascular relaxation by regulating the eNOS and ER-α protein levels in aorta of ovariectomised rats (Intapad et al., 2012). Additionally, CC promoted the phosphorylation of serine 1,177 in eNOS and serine 473 in Akt protein (Intapad et al., 2009). CC also improved the diastolic function of the aorta in hypercholesterolemic rats by activation of HSP70 and Bcl-2 levels, improving activity of antioxidant enzymes (Kam et al., 2012).

### ***Curcuma longa* L.**

The methanolic extract of *Curcuma longa* (i. v, CL, 10, 20 and 30 mg kg<sup>-1</sup>) could reduce blood pressure (2.0, 27.1, and 26.7%) and heart rate (5.8, 19.3, and 22.9%) in normal rats. CL (1–1,000 μg mL<sup>-1</sup>) also relieved PHE (10 μM)/K<sup>+</sup> (80 mM)-induced aortic contraction, which was attenuated by glibenclamide, BaCl<sub>2</sub>, TEA or 4-AP. In addition, CL inhibited CaCl<sub>2</sub> (1–30 mM) induced contraction in Ca<sup>2+</sup> free medium. This suggested that CL relaxes blood vessels by blockage of K<sup>+</sup> channels (Adaramoye et al., 2009).

## **Others**

The methanol and water extracts of curcuma herbs such as *C. kwangsiensis* (1 mg mL<sup>-1</sup>), *C. phaeocaulis* (1 mg mL<sup>-1</sup>), *C. wenyujin* (1 mg mL<sup>-1</sup>), and *C. zedoaria* (1 mg mL<sup>-1</sup>) could relieve prostaglandin F-2α (6 μM)-induced contractions in aortic ring. Additionally, curcuma herbs such as *C. zedoaria* (3% wt/wt) could lower blood pressure and protect endothelial cells in SHR (Goto et al., 2005). Therefore, curcuma herbs, with activities of invigorating blood circulation and eliminating stasis according to Chinese Medicine, may have significant potential for the prevention and treatment of CCVDs.

## **Orchidaceae**

### ***Orchis mascula*. ex L.**

*Orchis mascula* (OM) is used to treat CCVDs in Pakistan and India. The OM extract inhibited PHE (1 μM)-/K<sup>+</sup> (80 mM)-induced contractions in isolated rabbit aortas. OM (10 and 30 mg kg<sup>-1</sup>) also reduced SBP and improved endothelial function in SHR. Moreover, OM decreased TG, LDL-C levels in tyloxapol and high-fat diet-induced hyperlipidemia (Aziz et al., 2009).

### ***Dendrobium officinale* Kimura. et Migo.**

*Dendrobium officinale* (DO) has been found to improve metabolic diseases including hypertension and diabetes mellitus. In addition, the extract of DO (3.1 g kg<sup>-1</sup>) significantly reduced SBP and mean arterial pressure in the hypertensive rats. Moreover, it reversed thoracic aortic thickening and endothelial cell apoptosis, decreased plasma ET-1/TXB<sub>2</sub> levels and upregulated PGI<sub>2</sub>/NO levels (Liang et al., 2018).

## **Ranunculaceae**

### ***Paeonia suffruticosa* Andrew.**

The methanolic extract of the root bark of *Paeonia suffruticosa* (PS) showed a vasodilation in rat aortas pre-contracted by PHE (0.3 μM, IC<sub>50</sub> 16.8 μg mL<sup>-1</sup>). PS increased the endothelium and SOD function in rats fed a high-fat diet. Therefore, PM elicited vasorelaxant activity by protecting endothelial cells (Yoo et al., 2006).

### ***Nigella sativa* L.**

*Nigella sativa* (NS, 2–14 mg mL<sup>-1</sup>) extract induced a dose-dependent relaxation in aortic rings treated by PHE (1 μM)/K<sup>+</sup> (60 mM) which was abolished by diltiazem, TEA and



glibenclamide except for L-NAME, indomethacin or ruthenium red (LTCC inhibitor). This finding was suggested that effect of NS is concerned with activating on  $K^+$  channels (Niazmand et al., 2014).

### **Ginkgo biloba L.**

*Ginkgo biloba* leaf extract (GB) has been clinically used to improve peripheral vascular disease in France and Germany. Researchers have found that GB produced a dose-dependent relaxation in aortic rings treated by NE, which was alleviated by L-NAME (100  $\mu$ M), TEA (100  $\mu$ M) and indomethacin (100  $\mu$ M). In contrast, the effects of GB (3 mg mL<sup>-1</sup>) was strongly attenuated to 53% in Ca<sup>2+</sup>-free medium (Kubota et al., 2001), (Nishida and Satoh, 2003). Additionally, GB significantly reduced SBP in rats fed with 8.0% NaCl or SHR and potentiated the relaxation in response to acetylcholine in aortic (Kubota et al., 2006). However, GB was significantly increased serum alanine aminotransferase and hepatic CYP2B protein levels in aged SHR (Tada et al., 2008). It was suggested that the effect of GB was caused by Ca<sup>2+</sup> channels inhibition and NO levels promotion, endothelial cells protection in SHR. Notably, terpenoids and flavonoids may be the main active components of GB (Nishida and Satoh, 2003; Seiichiro and Hiroyasu, 2004).

### **Myrtaceae**

#### ***Syzygium guineense***

The hydroalcohol extract of the leaves of *Syzygium guineense* (SG) (50, 100, and 150 mg kg<sup>-1</sup>, orally) reduced systolic/diastolic blood pressure (6.9, 34.0, and 40.8 mmHg)/(5.0, 18.3, and 25.9 mmHg) in 1-kidney-1-clip hypertensive rat model. Moreover, the extract (70 mg kg<sup>-1</sup>) caused relaxation in aortas pre-contracted by K<sup>+</sup> (80 mM), with a maximum relaxation of 56.22%. The relaxation mechanism was concerned with muscarinic or histamine receptors, KATP and COX/NO/cGMP pathway, independent of the endothelium system (Ayele et al., 2010).

#### ***Myrciaria cauliflora***

Jabuticaba (*Myrciaria cauliflora*) hydroalcoholic extract (JH, 0.38–1.92 mg mL<sup>-1</sup>) resisted K<sup>+</sup>-PHE-induced aortic ring contractions, which was weakened by TEA, glibenclamide and 4-AP. Therefore, JH might activates the K<sup>+</sup>/Ca<sup>2+</sup> channels to cause vasodilation rather than SR Ca<sup>2+</sup>-ATPase or endothelium (Andrade et al., 2016).

#### ***Pimenta dioica* L.**

The aqueous extract of *Pimenta dioica* (EC<sub>50</sub> 45 mg kg<sup>-1</sup>) decreased the blood pressure in SHR which was not related to ion concentration (K<sup>+</sup>, Na<sup>+</sup>, Ca<sup>2+</sup>, and Mg<sup>2+</sup>),  $\alpha/\beta$ -adrenoceptor and cholinergic receptor (Suárez et al., 2000).

### **Loranthaceae**

#### ***Loranthus ferrugineus* Roxb.**

*Loranthus ferrugineus* Roxb could be successively fractionated by chloroform, ethyl acetate and n-butanol. n-butanol fraction of LFME (NBF-LFME) produced a significant inhibition of PHE-/

K<sup>+</sup>-induced aortic ring contractions. Moreover, NBF-LFME lowered blood pressure more compared with the other fractions in normal rats. Thus, the effects of LFME were attributed to content of terpenoids in the n-butanol fraction (Ameer O. et al., 2010).

#### ***Agelanthus dodoneifolius***

The ethanolic extract of *Agelanthus dodoneifolius* (AD, 0.01–10 mg kg<sup>-1</sup>, i.v.) could decrease the systolic and diastolic blood pressure in normotensive rats but not heart rate. Further, the extract was divided into 14 fractions (F1-F14). F4, contained most of the dihydropyranone dodoneine, produced the most effective activation (ED<sub>50</sub> 160  $\mu$ g mL<sup>-1</sup>) in NE-induced aortic rings contractions and reduced systolic and diastolic blood pressure by 56.9 and 81.6%, respectively in SHR (Ouedraogo et al., 2011). But the relationship between hypotensive activity and vasodilation of AD needs further investigation in the future.

### **Moraceae**

#### ***Morus bombycis* Koidzumi**

The ethanol extract of *Morus bombycis* koidzumi (MBK) exhibited vasodilatory effect (IC<sub>50</sub> 3.9  $\mu$ g mL<sup>-1</sup>) which was abolished by L-NAME or endothelial removal in rat aortas. Moreover, MBK extract (10, 30 and 100 mg kg<sup>-1</sup>) dose-dependently reduced SBP and attenuated liver lipid peroxidation and DNA-damage in SHR (Oh et al., 2007a). Therefore, the hypotensive activity of MBK is closely related to vasodilation, but the mechanism of vasodilation needs more exploration.

#### ***Morus alba* L.**

The major components of Mulberry leaves (ML) are polyphenols, flavonoids, carbohydrates, proteins and lipids. Researchers have found that ML could produce vasorelaxation of aortas treated by high K<sup>+</sup> (60 mM) or PHE (1  $\mu$ M) in arteries, which was abolished by ruthenium red (Xia et al., 2007). But mechanistic studies are notably imperfect such as endothelial cells and ion channels.

#### ***Ficus sycomorus* L.**

The extracts of *Ficus sycomorus* leaves (decoction, macerated and ethanol extract) exhibited a significant vasodilation in rat aortas (IC<sub>50</sub> 1.27, 0.38, and 0.13 mg mL<sup>-1</sup>, respectively). But the effect of extract was inhibited by L-NAME except for ethanol extract (Ramdé-Tiendrébéogo et al., 2014). Thus, the vasodilatory potential of the extracts needs further exploration in the future.

#### ***Humulus lupulus* L.**

The extracts of *Humulus lupulus* (HL, 10<sup>-9</sup>–10<sup>-2</sup> g L<sup>-1</sup>) inhibited NE (0.1  $\mu$ M)-induced arterial ring contractions in sham-ovariectomised female rats. The vasorelaxation was strongly abolished by L-NAME (100  $\mu$ M), indomethacin (10  $\mu$ M) and thapsigargin (100  $\mu$ M). The data suggested that vasodilation of HL is concerned with regulation of eNOS/COX levels and inhibition of Ca<sup>2+</sup> channel (Figard et al., 2008).

### **Other Sources**

#### ***Sesamum indicum* L.**

The petroleum ether soluble fraction of the root extract of *Sesamum indicum* L (SIL, 180  $\mu$ g mL<sup>-1</sup>) significantly inhibited

PHE (2  $\mu\text{M}$ )-/ $\text{K}^+$  (80 mM)-induced contractions (98.13 and 70.19%, respectively) in rat aortas. Additionally, which was abolished by L-NAME (300  $\mu\text{M}$ ) or methylene blue (10  $\mu\text{M}$ ) rather than propranolol (10  $\mu\text{M}$ ), atropine (1  $\mu\text{M}$ ), indomethacin (10  $\mu\text{M}$ ) or glibenclamide (10  $\mu\text{M}$ ). These results revealed that the vasodilation of SIL is chiefly mediated by endothelium-dependent pathway (Suresh Kumar et al., 2008).

#### ***Hibiscus sabdariffa* L.**

The methanolic extract of the calyces of *Hibiscus sabdariffa* L (HS) inhibited high  $\text{K}^+$  (80 mM)-/PHE (1  $\mu\text{M}$ )-induced vasoconstriction in SHR. Meanwhile, the activity of HS was eliminated by atropine (1  $\mu\text{M}$ ), L-NAME (10  $\mu\text{M}$ ) or methylene blue (10  $\mu\text{M}$ ) except for indomethacin (10  $\mu\text{M}$ ). These results indicated that the effects of HS are likely mediated by the NO/cGMP pathway or  $\text{Ca}^{2+}$  channels (Ajay et al., 2007).

#### ***Jasminum* Spp.**

The ethanol extract of Jasmine flower (JF) caused a concentration-dependent relaxation in endothelium-intact rings treated by PHE (10  $\mu\text{M}$ )/ $\text{K}^+$  (60 mM), which was decreased by L-NAME (3 mM),  $\text{BaCl}_2$  (1 mM), 4-AP (5 mM) and TEA (1 mM), expect for indomethacin (10  $\mu\text{M}$ ), and glibenclamide (10  $\mu\text{M}$ ). Additionally, JF inhibited the contraction of PHE after endothelium removal in  $\text{Ca}^{2+}$ -free medium. The activity of JF was concerned with the NO levels or  $\text{K}^+$  channels (Yin et al., 2014).

#### ***Hancornia speciosa* Gomes.**

The extract of leaves from *Hancornia speciosa* Gomes (HSG) produced vasodilatation ( $\text{pIC}_{50}$  5.6). This effect was suppressed by endothelial removal, L-NAME (100  $\mu\text{M}$ ), indomethacin (10  $\mu\text{M}$ ) or wortmannin (0.3  $\mu\text{M}$ ). In conclusion, HSG induced vasodilation in rat aortas possibly through PI3K activation (Ferreira et al., 2007).

#### ***Pseuderanthemum palatiferum***

Water extracts from fresh leaves of *Pseuderanthemum palatiferum* (PP) caused the relaxation of NE-contracted endothelium-intact aorta rings ( $\text{EC}_{50}$  81.0  $\mu\text{g mL}^{-1}$ ), removal endothelium ( $\text{EC}_{50}$  136.4  $\mu\text{g mL}^{-1}$ ). Neither L-NAME nor atropine altered the vasodilation effect of the extract. The vasodilatory effect was partially dependent on endothelium rather than muscarinic receptor (Khonsung et al., 2011).

#### ***Terminalia superba* Engl.et Diels**

The aqueous, the methanolic and the methylene chloride extracts from the stem bark of *Terminalia superba* (TS) induced vasodilation on  $\text{K}^+$ -/PHE-induced contractions in rat aortic rings. In contrast, the effect of TS was endothelium-dependent which was decreased by L-NAME (Tom et al., 2010).

#### ***Guazuma ulmifolia* Lam.**

The procyanidin fraction of *Guazuma ulmifolia* bark (GUB, 10  $\text{mg kg}^{-1}$ ) decreased systolic arterial pressure and heart rate in hypertensive rats which was attenuated by L-NAME

(31  $\text{mg kg}^{-1}$ ). GUB reduced the contractions induced by NE (0.1  $\mu\text{M}$ ) in the aortic rings of normotensive ( $\text{IC}_{50}$  35.3  $\text{ng mL}^{-1}$ ) or hypertensive rats ( $\text{IC}_{50}$  101  $\text{ng mL}^{-1}$ ). This activity was attenuated by endothelial removal or L-NAME (30  $\mu\text{M}$ ) but not indomethacin (10  $\mu\text{M}$ ) or atropine (10  $\mu\text{M}$ ) (Magos et al., 2008). Thus, the potential of GUB is mainly associated with the endothelial rather than the PGI2 pathway.

#### ***Nitraria sibirica* Pall.**

The extract from the fruits of *Nitraria sibirica* Pall (NSP, 0.1–10  $\text{g L}^{-1}$ ) produced vasodilation in PHE (1  $\mu\text{M}$ )-/ $\text{K}^+$  (60 mM)-induced pre-contracted aortic rings which was significantly suppressed by endothelial removal, L-NAME (100  $\mu\text{M}$ ), atropine (1  $\mu\text{M}$ ) and charybdotoxin (30 nM,  $\text{K}^+$  channel blocker) plus apamin (30 nM). Furthermore, NSP (i.v. 1, 5, 10, and 20  $\text{mg kg}^{-1}$ ) induced hypotensive effect in SHR and Wistar rats (Senejoux et al., 2012). But the relationship between the hypotensive activity and the vasodilation needs further exploration.

#### ***Persea americana* Mill.**

The aqueous extract of *Persea americana* Mill (PAM) abolished the positive inotropic and chronotropic responses induced by NE ( $10^{-10}$ – $10^{-5}$  M)/ $\text{Ca}^{2+}$  (5–40 mM) in guinea pig atrial muscle. However, this effect was markedly inhibited by L-NAME (10  $\mu\text{M}$ ). Furthermore, PAM (i.v. 25–400  $\text{mg kg}^{-1}$ ) significant reduced blood pressure and heart rates in normotensive and hypertensive rats (Ojewole et al., 2007).

#### ***Stephania abyssinica* Walp.**

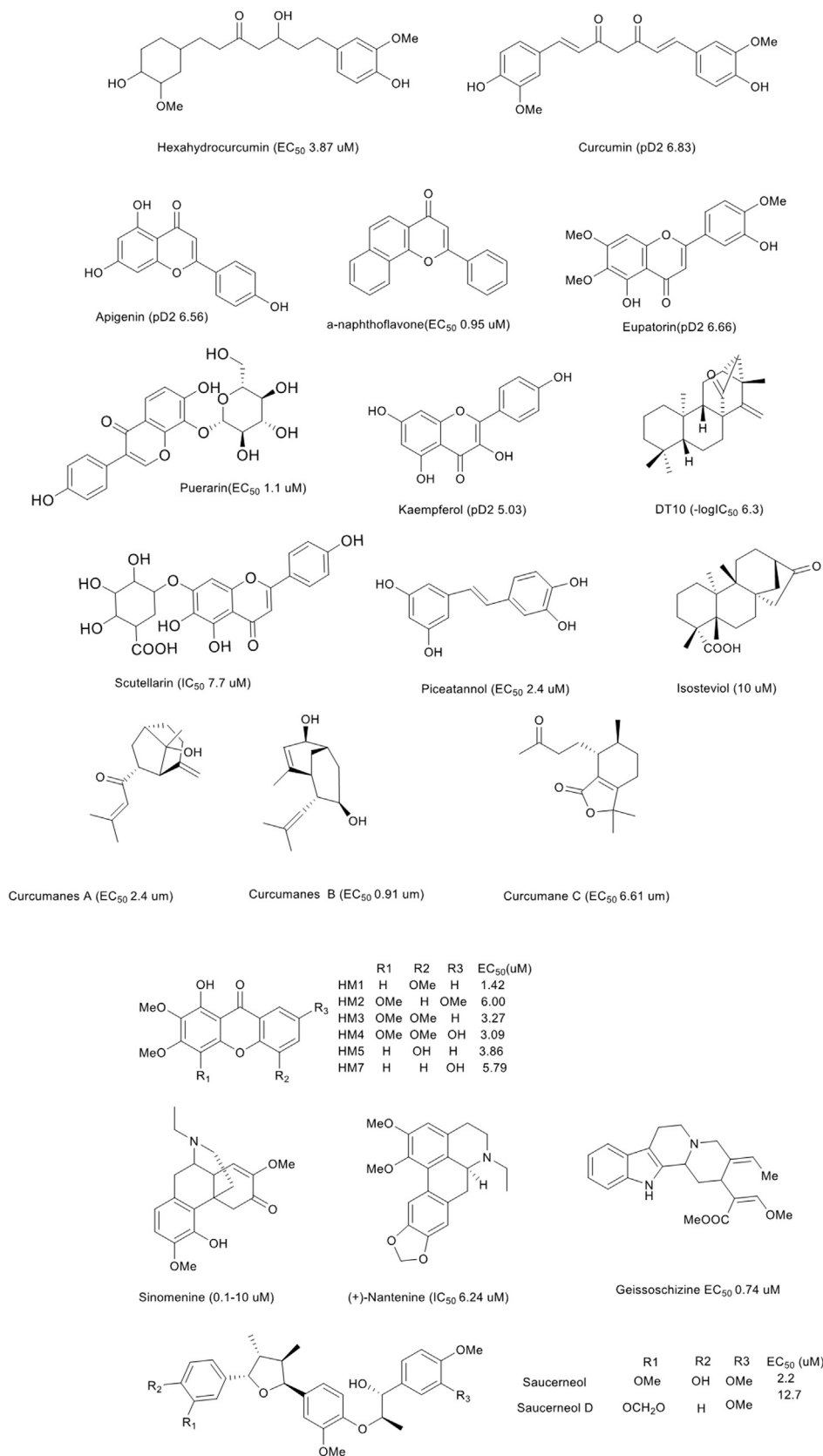
*Stephania abyssinica* Walp (SA) is used to treat arterial hypertension in west region of Cameroon. A previous study indicated that aqueous (ASAW) and methanol (MSAW) extracts from fresh leaves of *S. abyssinica* exhibited vasorelaxation by  $\text{K}^+$ -induced contractions ( $\text{EC}_{50}$  0.16, 0.35  $\text{mg kg}^{-1}$ ) in aortic rings. ASAW ( $\text{EC}_{50}$ , 0.18  $\text{mg kg}^{-1}$ ) also resisted contracted by PHE in aortic rings which was affected by TEA, glibenclamide, and propranolol but not L-NAME. These results indicated that the vasodilation of ASAW is mediated by  $\text{Ca}^{2+}$ /KATP channels (Nguelefack et al., 2015).

#### ***Fagopyrum esculentum* Moench.**

The hot-water extract of *Fagopyrum esculentum* (FE) evoked a significant vasodilation in rat aortic rings contracted by PHE (1  $\mu\text{M}$ ,  $\text{EC}_{50}$  2.2  $\text{mg mL}^{-1}$ ) or  $\text{K}^+$  (50  $\mu\text{M}$ ,  $\text{EC}_{50}$  1.9  $\text{mg mL}^{-1}$ ). Moreover, the acidic fraction of FE ( $\text{EC}_{50}$  0.25  $\text{mg mL}^{-1}$ ) had markedly stronger effect which was weakened by endothelial removal or L-NAME (100  $\mu\text{M}$ ). These results suggested that acidic partial vasodilation is mediated by the NO/cGMP pathway (Ushida et al., 2008).

#### ***Uncariae ramulus et Uncus.***

The alcohol extract of *Uncariae ramulus et Uncus* displayed a vasodilatory effect. The underlying mechanism consists of  $\text{Ca}^{2+}$  channel blockage and endothelium-dependent. The alkaloids and tannins might be the main active components. Additionally, *Uncaria sinensis* hexane extracts (HEUS), ethyl acetate extracts (EAEUS) and methanol extracts (MEUS) had a protective effect



**FIGURE 4 |** Natural products with significant vasodilation effect.

against photothrombotic ischaemic injury in mice. HEUS (10, 50, and 100 mg kg<sup>-1</sup>) also reduced infarct volume and edema compared with EAEUS and MEUS. However, HEUS did not reduce eNOS levels of brain infarction in mice, suggesting that the protective effect of HEUS is primarily concerned with endothelium (Yuzurihara et al., 2002), (Goto et al., 2000).

### Capparis aphylla

The extract of *Capparis aphylla* (CA) could inhibit PHE (1 μM) (EC<sub>50</sub> 0.1 mg mL<sup>-1</sup>)/K<sup>+</sup> (80 mM) (EC<sub>50</sub> 1.22 mg mL<sup>-1</sup>)-induced contractions in rabbit aortic rings, which was abolished by endothelium removal, L-NAME and atropine. Moreover, CA (3–100 mg kg<sup>-1</sup>, i.v.) reduced mean arterial pressure in normotensive rats, which was partially blocked by atropine (2 mg kg<sup>-1</sup>) (Shah and Gilani, 2011). Thus, the activity of CA may be associated with activation of muscarinic receptors or eNOS.

### Others

The aqueous extract of *Rheum undulatum* L. (Ru) could alleviate PHE-induced constriction in rat aortic, which was attenuated by endothelial removal, L-NAME, methylene blue and ODQ. These results suggested that activity is mediated by NO/cGMP pathways (Kim H. H. et al., 2004). The aqueous extract of *Echinodorus grandiflorus* (EG, 0.1–10 mg) significantly induced renal vasodilation in rabbits treated by NE-constriction which was attenuated by L-NAME (100 μM) and methylene blue (20 μM). However, its activity was not affected by charybdotoxin (100 nM, Ca<sup>2+</sup> blocker) or glibenclamide (3 μM) (Tibirić et al., 2007). The extracts of *Maytenus ilicifolia* (MI) leaves reduced the mean arterial pressure and heart rate in anaesthetised rats which was significantly reduced by L-NAME, methylene blue, ODQ, TEA, 4-AP and glibenclamide except for atropine, propranolol (Crestani et al., 2009). The extract of *Globularia alypum* (GA) also relieved diastolic PHE (2–4 ng mL<sup>-1</sup>)-induced mesenteric artery contractions in rats, which was inhibited by endothelial removal or atropine, but not indomethacin (10 μM) or L-NAME (100 μM) (Chokri et al., 2012). The extract of *Gmelina arborea* hexane leaves (GAE) produced vasodilatory effects in PHE (1 μM)-induced contractions which was reduced by L-NAME (2 μM), indomethacin (2 μM) and glibenclamide (2 μM) (Wansi et al., 2012).

## NATURAL PRODUCTS WITH VASODILATION IN VITRO AND ANTIHYPERTENSIVE ACTIVITIES IN VIVO

This section summarized the vasodilation of natural compounds. Some compounds have significant vasodilation (EC<sub>50</sub>/IC<sub>50</sub> < 10 μM), whose mechanism are related to multi-pathways (Supplementary Table S2). The natural products, mainly flavones (30%), were shown in Figure 2B. Importantly, they showed vasodilation mainly by acting on Ca<sup>2+</sup> channels and eNOS (Figure 3B).

Additionally, we briefly analysed the structure-activity relationship of the compounds with remarkable functions, as

shown in Figure 4. Among the phenolic acids, hexahydrocurcumin and curcumin had the prominent vasodilatory effects. The difference in bioactivity may be related to the number of double bonds. All compounds, specifically flavones, displayed the most excellent potential. To summarise, the vasodilation of these compounds was related to the number and position of double bonds, carbonyls, phenolic hydroxyl groups and methoxy groups. Furthermore, some compounds, such as chrysin glucoside, tilianin, reticuline and hirsutine also exhibited hypotension *in vivo* (Figure 3B).

## Flavones

### Apigenin

Apigenin (Ap) was a flavonoid in the Chinese herbal medicine Flos Chrysanthemi that displays anti-hypertensive and anti-inflammatory activities. Ap markedly reduced rat thoracic aorta contractions induced by pyrogallol or acetylcholine (pD2 6.56, 5.31), which was weakened by L-NAME rather than aminoguanidine or indomethacin. Additionally, Ap significantly reduced blood pressure of in hypertension rat by ameliorating NO levels and nitrite urinary excretion (Jin et al., 2009).

### α-Naphthoflavone

α-Naphthoflavone (α-NF) induced relaxation in PHE-induced aortas (EC<sub>50</sub> 0.95 μM), which was significantly inhibited by endothelial removal, L-NAME or methylene blue. In HUVECs, α-NF also activated NO channels which was blocked by SKF 96365 (Ca<sup>2+</sup> channel blockers) and Ni<sup>2+</sup> (Ca<sup>2+</sup> channel blockers). Thus, the vasodilatory activities of α-NF might be mediated by the Ca<sup>2+</sup> channels and the NO/cGMP pathway (Cheng et al., 2003).

### Formononetin

Formononetin (Fo), as a methoxylated isoflavone, enhanced NO levels or endothelial cell function and also induced vasodilation by K<sup>+</sup> pre-contractions in rat aortas (EC<sub>50</sub> 107.2 μM). This action was reduced by endothelial removal and L-NAME. Moreover, the vasodilatory activity was concerned with the phosphatidylinositol 3-kinase/protein kinase B (PI3K/Akt) pathway (Li et al., 2018).

### Puerarin

Puerarin (Pu) is the main isoflavone found in *Pueraria lobata*, which has been used for CCVDs. Pu (10<sup>-10</sup>–10<sup>-8</sup> M) induced relaxation in PHE-induced aortas through promoting HO function in endothelial removal. Additionally, Pu caused relaxation of aortic rings contracted with NE (1 μM) (EC<sub>50</sub> 1.1 μM), which was significantly repressed by iberiotoxin (50 nM, BKCa blocker). Pu also alleviated high glucose-induced endothelium-dependent vascular dysfunction in rat aortic rings (Meng et al., 2009), (Sun et al., 2007). Thus, the activities of Pu are concerned with endothelial cells or BKCa.

### Hesperetin

Hesperetin (He) was shown to relieve NE (1 μM)-/K<sup>+</sup> (60 mM)-induced contractions (IC<sub>50</sub> 62.8, 62.2 μM). The effects were not inhibited by endothelial removal, glibenclamide (10 mM), TEA (2 mM) or nifedipine (0.1 μM). Moreover, He inhibited



calmodulin-activated PDE1 and PDE4 isolated from bovine aortas ( $IC_{50}$  74, 70  $\mu$ M) (Orallo et al., 2004).

### 5, 7-Dimethoxyflavone

In Thai traditional medicine, *Kaempferia parviflora* is used to treat CCVDs, including hypertension, asthma or diarrhoea. 5, 7-dimethoxyflavone (DMF, 1–100  $\mu$ M), which was the primary component of *Kaempferia parviflora*, caused relaxations in aortic rings pre-contracted by methoxamine. This effect was abolished by endothelial removal, L-NAME (300  $\mu$ M), indomethacin (10  $\mu$ M), TEA (5 mM), glibenclamide (10  $\mu$ M), 4-AP (1 mM),  $BaCl_2$  (10  $\mu$ M) or ODQ (10  $\mu$ M). Therefore, DMF-induced relaxation was mediated through the activation of the NO/cGMP/COX pathways and the inhibition of  $Ca^{2+}$  channels (Tep-Areenan et al., 2010).

### Eupatorin

Eupatorin (Eu) is the primary component in *Orthosiphon stamineus*, used in Malaysia to treat hypertension. Eu produced relaxation of aortic rings pre-contracted by PHE in intact endothelium. This effect was reduced by L-NAME (pD2 4.60), methylene blue (pD2 6.05), ODQ (pD2 5.84), indomethacin (pD2 6.27), TEA (pD2 6.09), 4-AP (pD2 6.34),  $BaCl_2$  (pD2 6.47), atropine (pD2 6.36) and propranolol (pD2 6.49). Overall, the effect of Eu was related to activation of the NO/SGC/cGMP pathway, regulation of muscarinic/ $\beta$ -adrenergic receptor and inhibition of  $K^+$  channels (Yam M. et al., 2016).

### Scutellarin

Scutellarin (Sc), obtained from *Erigeron breviscapus* Hand. Mazz, has been used to treat CCVDs such as hypertension. Sc aroused relaxation in rat aortic rings pre-contracted by NE (1  $\mu$ M,  $IC_{50}$  7.7  $\mu$ M), which was not influenced by TEA (10 mM), glibenclamide (10  $\mu$ M), atropine (100 nM), propranolol (1  $\mu$ M), indomethacin (10  $\mu$ M) or L-NAME (100  $\mu$ M). Additionally, Sc inhibited the increase of intracellular  $Ca^{2+}$  induced by NE and had no effect on phorbol-12, 13-diacetate-induced contractions in  $Ca^{2+}$ -free solution (Pan et al., 2008). It was shown that the activity of Sc may be through acting on ion channels rather than endothelial cells.

### Daidzein/Daidzin

Daidzein and daidzin were shown to produce relaxation in aortas pre-contracted by U46619 (100 nM) ( $IC_{50}$  20 and 140  $\mu$ M, respectively), which was weakened by glibenclamide (1  $\mu$ M), TEA (100 mM), iberiotoxin (100 nM), 4-AP (1 mM) or  $BaCl_2$  (100  $\mu$ M). Daidzein-induced vasodilation of rat cerebral basilar arteries was partially dependent on BKCa channels in VSMCs (Zhang H.-T. et al., 2010). Therefore, the effects of daidzein and daidzin were concerned with inhibition of  $K^+$  channels (Sun et al., 2007).

### Kaempferol

Kaempferol (Ka) produced relaxation in pulmonary arterial rings pre-contracted by PHE (1  $\mu$ M) (pD2 5.03), which was not affected by glibenclamide,  $BaCl_2$ , 4-AP (1 mM), L-NAME or indomethacin. However, this activity was repressed by TEA (10 mM) and ODQ. Therefore, Ka caused vasodilation through

blocking BKCa/LTCC or regulating the sGC/PKA pathway (Mahobiya et al., 2018).

### Baicalein/Luteolin

Baicalein and luteolin were shown to significantly alleviate insulin resistance (IR)-induced SBP elevation which were inhibited by bisphenol A diglycidyl ether in rats fed fructose for 12 weeks. Meanwhile, they also reduced excessive vasoconstriction of PHE/ $K^+$  in IR animals through elevating NO/ROS (reactive oxygen species) levels (Huang et al., 2004).

### Genistein

Genistein (4',5,7-trihydroxyisoflavone) is naturally occurring flavonoid found in the Leguminosae plant. Studies had shown that genistein elicited vasodilatory, anti-thrombotic and anti-atherosclerotic. This vasodilation was closely related to regulation of eNOS/angiotensin levels or inhibition of  $K^+$ / $Ca^{2+}$  channels (Sureda et al., 2017).

### Others

Hydroxy-2,3,5-trimethoxy-xanthone (HM-1,  $EC_{50}$  1.42  $\mu$ M), 1-hydroxy-2,3,4,7-tetramethoxy-xanthone (HM-2,  $EC_{50}$  6.00  $\mu$ M), 1-hydroxy-2,3,4,5-tetramethoxy-xanthone (HM-3,  $EC_{50}$  5.27  $\mu$ M), 1,7-dihydroxy-2,3,4,5-tetramethoxy-xanthone (HM-4,  $EC_{50}$  3.09  $\mu$ M), 1,5-dihydroxy-2,3-dimethoxy-xanthone (HM-5,  $EC_{50}$  3.86  $\mu$ M) and 1,7-dihydroxy-2,3-dimethoxy-xanthone (HM-7,  $EC_{50}$  5.76  $\mu$ M) caused vasodilation in coronary arteries pre-contracted by 5-hydroxytryptamine (5-HT, 1  $\mu$ M). Furthermore, removal of the endothelium decreased the vasodilation of HM-1 and HM-7 but did not affect HM-2, HM-3, HM-4 or HM-5 (Wang et al., 2009). The number of methoxy groups (HM 1, HM 2) and the substitution position of hydroxyl groups (HM 5, HM 7) had significant effects. In addition, Floranol (Fo) was shown to induce vasodilation pre-contracted by PHE (0.1  $\mu$ M) ( $IC_{50}$  19.9  $\mu$ M), which was not affected by L-NAME or endothelium removal (Lemos et al., 2006).

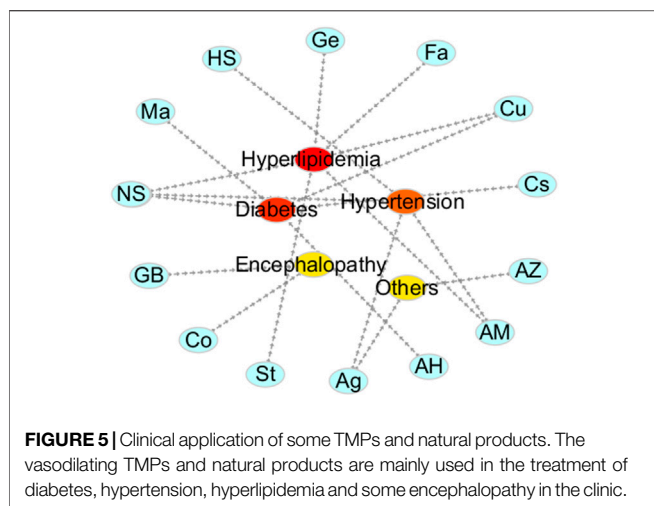
### Alkaloids

#### Harmaline

Harmaline (Ha) has shown hypotensive activity *in vivo*, but the mechanism is no clear at present. Researchers had found that Ha (3, 10, and 30  $\mu$ M) induce relaxation in aortas pre-contracted by NE/ $K^+$ , which was significantly suppressed by L-NAME, indomethacin, prazosin ( $\alpha$ -adrenoreceptors blocker) or diltiazem. Thus, the effect of Ha was related with activation of the prostacyclin or eNOS pathway (Berrougui et al., 2006).

#### Reticuline

Reticuline (Re) resisted contractions induced by PHE (1  $\mu$ M) and  $K^+$  (80 mM) ( $IC_{50}$  40 and 240  $\mu$ M, respectively). Additionally, Re (i.v. 5, 10 and 20 mg  $kg^{-1}$ ) produced hypotensive effect in normotensive rats, which was attenuated by L-NAME (20 mg  $kg^{-1}$ ) or atropine (2 mg  $kg^{-1}$ ). These results suggested that Re reduced blood pressure by activating the muscarinic or eNOS receptors, and blocking  $Ca^{2+}$  channels (Dias et al., 2004).



### Sinomenine

Sinomenine acutum Rehder has been used in the treatment of rheumatoid arthritis in China and its extract is also found to have vasodilator activity. Sinomenine (Si, 0.1–10  $\mu\text{M}$ ), as the mainly compound from Sinomenine acutum Rehder, produced relaxation in aortic rings pre-contracted by PHE (10 nM) or  $\text{K}^+$  (40 mM) which was attenuated by glibenclamide. Similarly, Si (1–100  $\mu\text{M}$ ) also reduced  $\text{Ca}^{2+}$  concentration in aortic smooth muscle (A7r5) cells induced by PHE (1  $\mu\text{M}$ ) or  $\text{K}^+$  (40 mM), which was eliminated by glibenclamide. Moreover, Si (i.v. 2.5–10  $\text{mg kg}^{-1}$ ) decreased SBP in SHR (Lee et al., 2007). Thus, the activity of Si was concerned with activation of KATP.

### (+)-Nantenine

(+)-Nantenine (Na), isolated from *Nandina domestica*, was widely used for the treatment of asthma, uterine bleeding and diabetes. Na could relieve contractions induced by NE ( $\text{IC}_{50}$  6.24  $\mu\text{M}$ ) or high  $\text{K}^+$  (60 mM,  $\text{IC}_{50}$  5.23  $\mu\text{M}$ ) in rat aortic rings, which was not modified by endothelial removal, glibenclamide (10  $\mu\text{M}$ ) or TEA (2 mM) (Orallo and Alzueta, 2001). The activity of Si was concerned with acting on KATP or endothelial cells.

### Cassiarin A

The leaves of *Cassia siamea* were used for the treatment of hypertension in traditional folk medicine. Cassiarin A (Ca), isolated from *Cassia siamea*, was shown to induce relaxation in mesenteric arteries pre-contracted with PHE (1  $\mu\text{M}$ ,  $\text{EC}_{50}$  6.4  $\mu\text{M}$ ), which was significantly reduced by endothelial removal, L-NAME (100  $\mu\text{M}$ ), ODQ (10  $\mu\text{M}$ ), TEA (1 mM) and iberiotoxin (100 nM) rather than indomethacin (10 mM), glibenclamide (10 mM) and 4-AP (1 mM). These results suggested that Ca activity is mediated by NO and BKCa channels (Matsumoto et al., 2010).

### 2-Benzyl-5-Hydroxy-6-Methoxy-3,4-Dihydroisoquinolin-1-One

Isoquinolinone alkaloids have antihypertensive, antiarrhythmic and other activities. Researchers have found that 2-Benzyl-5-hydroxy-6-methoxy-3, 4-dihydroisoquinolin-1-one (ZC2) relax

mesenteric artery segments pre-contracted by KCl ( $\text{pEC}_{50}$  4.56), PHE ( $\text{pEC}_{50}$  5.39) or U46619 ( $\text{pEC}_{50}$  4.67), which was not modified by endothelial removal or glibenclamide (10  $\mu\text{M}$ ). Thus, ZC2 induced vasodilation through inhibiting VOCC and ROCC (Xu et al., 2012).

### Geissoschizine Methyl Ether/Hirsutine

Geissoschizine methyl ether (Ge) and hirsutine resisted NE (10 nM)-induced contractions ( $\text{EC}_{50}$  0.74, 10.6  $\mu\text{M}$ ) which were abolished by endothelial removal or L-NAME (100  $\mu\text{M}$ ). Furthermore, hirsutine reduced the SBP and heart rate of SHR (Yuzurihara et al., 2002). These results were suggested that the vasodilatory activity of NE is closely related to endothelial cells.

### Piperine

Piperine (Pi), as main compound in Sahatsatara formula, had been demonstrated to have hypotensive effects in L-NAME-induced endothelial dysfunction rats. Moreover, it relaxed the thoracic aorta and has vascular protection effects against hypertension in rats (Booranasubkajorn et al., 2017).

### Uncarialin A

*Uncaria rhynchophylla* Miq. ex Havil has been used to treat hypertension and epilepsy in clinics. Uncarialin A, separated from the hooks of *U. rhynchophylla*, resisted PHE-induced contractions ( $\text{IC}_{50}$  0.18 M) independent of endothelial cells. Additionally, Uncarialin A reduced concentration of  $\text{Ca}^{2+}$  in VSMCs by inhibiting the LTCC subunit  $\alpha\text{-1c}$  (Cav1.2) (Yun et al., 2020).

### 8-Oxo-9-Dihydromakomakine

8-Oxo-9-dihydromakomakine (Di), separated from the leaves of *Aristolochia chilensis*, produced dose-dependent relaxation of aortic rings pre-contracted by PHE (1  $\mu\text{M}$ )/KCl (60 mM), which was inhibited by TEA and  $\text{BaCl}_2$ . It was suggested that the activity of Di is related to reduction intracellular  $\text{Ca}^{2+}$  concentration by inhibiting KCa or Kir (Cifuentes et al., 2018).

### Curine

Curine could inhibit contractions induced by KCl and Bay K8644 in rat aortas and decreased intracellular  $\text{Ca}^{2+}$  concentration in VSMCs. The activity was not abolished by 3-isobutyl-1-methylxanthine (phosphodiesterase inhibitor), dibutyryl cyclic AMP (protein kinase A activator) or 8-br-cyclic GMP (protein activator kinase G) (Medeiros et al., 2011). It was indicated that potential of curine is associated with endothelial cells and ion channels.

### Phenolic Acids

#### Ethyl Rosmarinate

Ethyl rosmarinate (Er), as a major component of *Rosmarinus officinalis* and *Hyptis suaveolens*, has cardioprotective activities. Er produced relaxation pre-contracted by PHE (10  $\mu\text{M}$ ) ( $\text{pD}_2$  4.42) or  $\text{K}^+$  (80 mM) ( $\text{pD}_2$  4.56) in endothelium-denuded rings. However, the effect was not abolished by TEA (5 mM), glibenclamide (10  $\mu\text{M}$ ),  $\text{BaCl}_2$  (1 mM) and ODQ (1  $\mu\text{M}$ ) except

for 4-AP (1 mM). Er also reduced the contractions induced by deoxyepinephrine (10  $\mu$ M) and caffeine (20 mM) in a  $\text{Ca}^{2+}$ -free solution and inhibited extracellular  $\text{Ca}^{2+}$  influx. Therefore, the vasodilation of Er was mediated by endothelial cells or Kv (Wicha et al., 2015).

### Paeonol

Paeonol (Pa) is the main component of the Chinese herbs *Paeonia suffruticosa* Andr. and *Cynanchum paniculatum* (Bunge) Kitagawa. Pa displayed anti-ischemia reperfusion injury, antihypertensive, anti-platelet aggregation, scavenges oxygen free radicals, anti-atherosclerosis and anti-VSMCs proliferation activities. Researchers have found that Pa relaxed PHE-induced isolated rat aortic rings ( $\text{EC}_{50}$  290  $\mu$ M). Additionally, Pa significantly inhibited vasoconstriction induced by angiotensin II, prostaglandin F-2 $\alpha$ , 5-HT, dopamine, vasopressin and endothelin-1. Moreover, the activity of Pa was not affected by L-NAME, ODQ, propranolol, glibenclamide, TEA and  $\text{BaCl}_2$  in the rings. Therefore, the vasodilatory effect of Pa was in relationship with regulation of  $\text{Ca}^{2+}$  channels (Li et al., 2010).

### Hexahydrocurcumin/Curcumin

Curcumin (Cu) was the main active component of the roots of *Curcuma longa* L. Hexahydrocurcumin (HCC) was a reduction product of Cu and metabolites in mice or humans. HCC has a wide variety of pharmacological effects, including anti-inflammatory, anti-viral, anti-fungal, anti-oxidant and anti-cancer effects. Moreover, HHC relaxed endothelium-intact aortic rings pre-contracted by PHE (10  $\mu$ M,  $\text{EC}_{50}$  3.87  $\mu$ M) or  $\text{K}^+$  (80 mM,  $\text{EC}_{50}$  95.12  $\mu$ M). This agent relieved the  $\text{CaCl}_2$ -induced contractions in  $\text{K}^+$  solutions and also suppressed the contractions induced by PHE/caffeine in  $\text{Ca}^{2+}$ -free solutions. Additionally, HHC relieved phobal-12-myristate-13-acetate (PMA, activator of PKC) pre-contracted aortic rings ( $\text{EC}_{50}$  93.36  $\mu$ M). Collectively, it was suggested that the effect of HHC is related to endothelium cells, VOCC, ROCC and PKC-mediated  $\text{Ca}^{2+}$  channels (Moohammadaree et al., 2015). Cu (pD2 6.83) also relaxed the aortic rings pre-contracted by PHE (1  $\mu$ M). Further, Cu reversed the vasodilatory dysfunction induced by high glucose (44 mmol/L) by improving HO-1 function in aortic rings. However, the activities were resisted by protoporphyrin IX zinc (1  $\mu$ M, inhibitor of HO-1) or methylene blue (1  $\mu$ M) (Sasaki et al., 2003).

### Sodium Ferulate/Ferulic Acid

Ferulic acid (Fa) was the main component of *Radix Angelicae Sinensis* and *Rhizoma Chuanxiong*. Studies have shown that sodium ferulate (Sf) or Fa displayed anti-platelet aggregative, anti-inflammatory, antioxidative activities. Sf relaxed the aortic rings pre-contracted with PHE/ $\text{K}^+$  (pD2 2.7 and 2.6, respectively), which was not affected by TEA, glibenclamide, 4-AP and  $\text{BaCl}_2$ . Sf also inhibited contraction in  $\text{K}^+$ /PE/PMA pre-contracted rings in  $\text{Ca}^{2+}$ -free solution (Chen G. et al., 2009). Moreover, Fa decreased superoxide anion levels in SHR aortas and improved acetylcholine-induced vasodilation in SHR but not in WKY rats (Suzuki et al., 2007). This activity was related to the methoxy modified 3-position in the benzene ring and 2-propylene (Fukuda et al., 2015).

### Ellagic Acid

Ellagic acid (Ea), a polyphenolic compound, has anti-hypertensive, anti-diabetic, anti-oxidative, anti-inflammatory and anti-hyperlipidaemia effects. Ea relaxed the aortic pre-contracted by PHE (pD2 5.60), which was partially abolished by endothelium removal and L-NAME rather than indomethacin. Therefore, the activity of Ea was related to endothelium cells and  $\text{Ca}^{2+}$  channels (Yilmaz and Usta, 2013).

### Glycosides

#### Astragaloside IV

*Astragalus membranaceus* has been used to treat and prevent CCVDs such as haemorrhagic stroke and viral myocarditis. Astragaloside IV (As-IV), the main component of *A. membranaceus*, has anti-cardiac hypertrophy, anti-inflammatory and anti-oxidant activities. As-IV could antagonise contractions induced by PHE/ $\text{K}^+$  in aortic rings from normal rats and SHR. Similarly, As-IV attenuated the vasoconstriction induced by angiotensin II or PHE in presence of perivascular adipose tissue (Zhang et al., 2006). This activity of As-IV was abolished by L-NAME or ODQ rather than 7-nitroindazole (neuronal eNOS inhibitor). Therefore, this vasodilatory was associated with blockage of calcium channel, and activation of NO/cGMP pathway (Zhang et al., 2007). Additionally, As-IV (orally, 40–80 mg  $\text{kg d}^{-1}$ ) improved the expression of eNOS, SOD and GSH-Px in the thoracic aorta and decreased ROS levels in a streptozotocin-induced diabetic rat model. It was suggested that As-IV ameliorates endothelial damages in the thoracic aorta of diabetic rats via regulating levels of ROS and calpain-1 (Nie et al., 2019).

#### Cornuside

Cornuside (Co, 100  $\mu$ M) relaxed PHE (3  $\mu$ M) pre-contracted rat aortas, which was inhibited by endothelial removal, L-NAME and ODQ rather than diltiazem (10  $\mu$ M), TEA (100  $\mu$ M), glibenclamide (10  $\mu$ M), indomethacin (1  $\mu$ M), atropine (1  $\mu$ M) or propranolol (1  $\mu$ M). Co also increased cGMP levels in HUVECs, which was inhibited by L-NAME (10  $\mu$ M) or ODQ (1  $\mu$ M). Therefore, Co displayed vasodilatory activity through activating the NO/cGMP pathway (Kang et al., 2007).

#### Chrysin Glucoside

Chrysin glucoside (Cg) could inhibit NE (1  $\mu$ M)-induced contractions ( $\text{IC}_{50}$  52  $\mu$ M) in rat aortas, which was inhibited by L-NAME and endothelial removal. Cg (2.5 mg  $\text{kg}^{-1}$ ) significantly increased urine flow, glomerular filtration and electrolyte excretion ( $\text{Na}^+/\text{K}^+$ ) in rats. Additionally, Cg (i.v. 10 mg  $\text{kg}^{-1}$ ) caused an immediate decrease in the mean arterial blood pressure of the anaesthetised rats, which was inhibited by L-NAME (Kang et al., 2007).

#### Tilianin

*Agastache mexicana* is used as an antihypertensive drug in Mexico. Tilianin (Ti, 0.002–933  $\mu$ M), derived from *A. mexicana*, produced significant relaxation in aortic rings pre-contracted by NE (0.1  $\mu$ M) or serotonin (5-HT, 100  $\mu$ M). This relaxation was markedly abolished by endothelium removal, L-NAME (10  $\mu$ M), TEA

(5 mM), 4-AP (0.1  $\mu\text{M}$ ) or ODQ (1  $\mu\text{M}$ ) but not by indomethacin (10  $\mu\text{M}$ ) or atropine (1  $\mu\text{M}$ ). Additionally, Ti (50 mg  $\text{kg}^{-1}$ , orally) significantly reduced systolic and diastolic blood pressure in an SHR (Kang et al., 2007). The vasodilatory activity of Ti was concerned with endothelial cells or  $\text{K}^+$  channels.

### Jujuboside B

*Zizyphi Spinosi* Semen (ZSS) has protective effects against myocardial ischemic injury and hypertension. Jujuboside B (Ju), obtained from ZSS, reduced the tension of rat aorta with intact endothelium. However, the vasodilatory activity of Ju was attenuated by L-NAME, KN93, EGTA, SKF96365, iberiotoxin and glibenclamide rather than indometacin. Therefore, Ju displays its vasodilation through promoting eNOS levels and inhibiting  $\text{K}^+/\text{Ca}^{2+}$  channels (Zhao et al., 2016).

### Glucosyl hesperidin

Researchers have found that hesperidin possessed anti-oxidant, anti-hypertensive. Moreover, glucosyl hesperidin (Gh) has antihypertensive effects and improves lipid metabolism. Gh (50 mg  $\text{kg}^{-1}$ , orally) could reduce blood pressure in SHR and enhance endothelium-dependent vasodilation induced by acetylcholine. Additionally, Gh improved endothelial function by inhibiting the expression of nicotinamide adenine dinucleotide phosphate oxidase in SHR aorta (Yamamoto et al., 2008).

## Terpenoids

### Ent-Trachyloban-14, 15-Dione

*Croton zambesicus* is used to treat hypertension in Benin. Ent-trachyloban-14, 15-dione (DT10), separated from *Croton zambesicus*, relieved  $\text{K}^+$ -induced (100 mM) contractions in rat aortas ( $-\log\text{IC}_{50}$  6.3), which was significantly decreased by L-NAME ( $-\log\text{IC}_{50}$  5.7). Furthermore, DT10 inhibited  $\text{K}^+$ -evoked contractions in aorta rings and SH-SY5Y (human neuroblastoma cells) (Martinsen et al., 2010).

### Sesquiterpenoids

Zerumbone resisted aortas contracted by  $\text{K}^+$  (60 mM,  $\text{IC}_{50}$  16  $\mu\text{M}$ ) (Fusi et al., 2015). Sesquiterpenoids [curcumanes A, B, C, and ( $\pm$ ) D], which were isolated from *Curcuma longa*, also have significant vasodilation. They could significant resist KCl-induced aortic contractions in rats ( $\text{EC}_{50}$  2.40, 0.91, 6.61, 14.56, and 16.03  $\mu\text{M}$ , respectively) and curcumanes A, B, and C also inhibited PHE-induced aortic ring contractions in rats ( $\text{EC}_{50}$  3.37, 0.83, and 4.26  $\mu\text{M}$ , respectively). Additionally, curcumane C promoted the growth of human umbilical vein endothelial cells, which was inhibited by L-NAME. Curcumanes A and B produced vasodilation through regulation of VDCC and ROCC (Qiao et al., 2019; Liu et al., 2019). Therefore, sesquiterpenoids with significant activities should be paid more attention in the prevention and treatment of CCVDs.

## Coumarin

### Imperatorin/Isoimperatorin

Imperatorin and isoimperatorin were shown to relax rat aorta contractions by PHE. The effect of imperatorin was significantly

stronger than that of isoimperatorin. However, this activity was inhibited by endothelial removal and L-NAME (Nie et al., 2009).

### (+)-Cis-4'-O-Acetyl-3'-O-Angeloylkhellactone

(+)-cis-4'-O-acetyl-3'-O-angeloylkhellactone (Al) relaxed rat aortas pre-contracted by PHE (1  $\mu\text{M}$ ,  $\text{EC}_{50}$  17.8  $\mu\text{M}$ ) which was diminished by endothelial removal, L-NAME (100  $\mu\text{M}$ ) or methylene blue (30  $\mu\text{M}$ ). However, this function was not eliminated by indomethacin (30  $\mu\text{M}$ ), atropine (0.1  $\mu\text{M}$ ), triprolidine (10  $\mu\text{M}$ ), TEA (10 mM) and propranolol (3  $\mu\text{M}$ ). These results suggested that activity is mediated by  $\text{Ca}^{2+}$  channels and the NO/cGMP pathways (Lee et al., 2002).

## Others

### Perillaldehyde

Perillaldehyde (Pe), is major compound from aqueous extract of *Perilla* leaves, improves NO levels in VSMCs. Pe (0.01–1 mM) also resisted aorta contraction by prostaglandin F-2 $\alpha$  or NE, which was weakened by L-NAME, endothelial removal, propranolol, theophylline, TEA or glibenclamide. Therefore, the vasodilatory effect of Pe was concerned with blockage of  $\text{Ca}^{2+}$  channel (Takagi et al., 2005). Further, Pe (150 mg  $\text{kg}^{-1}$ ) reduced aortic atherosclerotic plaques, improved endothelial function, increased tetrahydrobiopterin and NO levels in carotid arteries of mice and rats fed high-fat diet (Yu and Liu 2018).

### Cinnamaldehyde

Cinnamaldehyde (Ci), separated from *Cinnamomi Cortex*, has anti-platelet aggregation by regulating arachidonic acid. Ci (1–1,000  $\mu\text{M}$ ) also resisted rat aortas contracted by prostaglandin F-2 $\alpha$  (5  $\mu\text{M}$ ), NE (0.1  $\mu\text{M}$ ) or  $\text{K}^+$  (60 mM). The activity was not inhibited by indomethacin, propranolol (10  $\mu\text{M}$ ), theophylline (100 mM, phosphodiesterase inhibitor), TEA (1 mM) or glibenclamide, instead of L-NAME (100 mM) (Yanaga et al., 2006). Further, Ci inhibited LTCC in mice ventricular myocytes and mesenteric artery SMCs (Buglak et al., 2018), (Alvarez-Collazo et al., 2014). The vasodilatory effect was related to endothelium cells and the  $\text{Ca}^{2+}$  channels.

### Phthalides

*Ligusticum chuanxiong* is used for CCVDs to promote the circulation of blood and removed stasis in TCM. Ligustilide (Li), main phthalides of *L. chuanxiong*, was used to treat CCVDs. Li was shown to relax rat mesenteric arteries pre-treated by KCl,  $\text{CaCl}_2$ , NA or 5-hydroxytryptamine (5-HT). But the activates was not affect by propranolol, glibenclamide, TEA and  $\text{BaCl}_2$ . It was indicated that Li induces vasodilation through regulating VOCC and ROCC rather than endothelial cells (Cao et al., 2006). Further, the recent research found that the activity of phthalide dimers was generally superior to that of monomeric phthalides. Phthalides dimers, such as Chuanxiongdiolides R4 and R6, inhibited KCl-induced (60 mM) vasoconstriction. Moreover, Chuanxiongdiolides R4 and R6 also significantly inhibited the LTCC subunit  $\alpha$ -1c (Cav1.2) (Tang et al., 2020).

### Isosteviol

Isosteviol (Is, 10  $\mu\text{M}$ ) significantly relaxed the vasopressin ( $10^{-8}$  M)-/ $\text{K}^+$  (100 mM)-induced vasoconstriction in aortic rings,



which was resisted by apamin and glibenclamide rather than  $K^+$  (30 mM). Therefore, this effect might be attributive to inhibition of KATP or SKCa (Wong et al., 2004).

### Piceatannol

Piceatannol (Pi) caused relaxation in aortas pre-contracted by PHE ( $EC_{50}$  2.4  $\mu$ M). This effect was reduced by endothelial removal, L-NAME, methylene blue, ODQ, 4-AP and TEA rather than indomethacin, atropine, propranolol, nifedipine,  $BaCl_2$  or glibenclamide. Moreover, charybdotoxin and iberia toxin (BKCa channel blockers) could also reduce the activity of Pi. Therefore, this vasodilation of Pi may be related to the activation of BKCa and NO pathway (Oh et al., 2007b).

### Eudesmin

Eudesmin (Eu) induced relaxation of rat aortic pre-contracted by PHE ( $IC_{50}$  10.69  $\mu$ g  $ml^{-1}$ ), which was blocked by endothelial removal, L-NAME, ODQ, indomethacin and diphenhydramine (type I histamine receptor), instead of atropine, propranolol and glibenclamide. This effect was mediated by regulation of histamine receptors and NO/prostaglandin pathways (Raimundo et al., 2009).

### Brazilin

Studies have found that brazilin (Br) has anti-diabetic, anti-inflammatory, anti-asthmatic, anti-platelet aggregation, anti-tumour and anti-oxidant. Further, Br resisted the  $NE/K^+$ -induced contraction of aortic rings ( $EC_{50}$  83.51 and 79.79  $\mu$ M, respectively), which was significantly attenuated by endothelium removal, L-NAME, methylene blue or indomethacin. Thus, the activity of Br was related to inhibition of ERK1/2 phosphorylation or blockage of  $Ca^{2+}$  channels (Yan et al., 2015).

### Tanshinone IIA

Tanshinone IIA (Tan IIA) could reduce infarct area and blood pressure in hamsters. Moreover, Tan IIA caused endothelium-dependent relaxation, which was blocked by oestrogen receptor antagonist ICI 182 and 780. Therefore, the activity of Tan IIA was related to oestrogen receptor, NO channels and ERK1/2 phosphorylation (Fan et al., 2011).

### Sodium Danshensu

Sodium danshensu (So) has anticoagulation and arrhythmia resistance on myocardial ischaemia-reperfusion injury. Moreover, So (1–3  $g L^{-1}$ ) inhibited the PHE- $K^+$  induced contraction of rat aortic, which was partially antagonised by TEA and apamin (SKCa blocker). However, the vasodilatory effect was not abolished by iberiotoxin (BKCa blocker),  $BaCl_2$  and glibenclamide (Zhang N. et al., 2010). Thus, the vasodilation of So was concerned with SKCa rather than BKCa, Kir and KATP.

### Caracasamide

Caracasamide (i.v.) was shown to reduce blood pressure and increase cardiac muscle strength, respiratory rate and tidal volume in rats. Additionally, it also induced vasodilation in rats through acting on cardiac  $\beta_1$ -adrenergic receptors (Delle Monache et al., 1993).

### Others

Other lignans (saucerneol, saucerneol D and machilin D) exhibited vasodilation in rat aortic treated by PHE (10  $\mu$ M,  $EC_{50}$  2.2, 12.7 and 17.8  $\mu$ M, respectively). The activities were significantly inhibited by L-NAME or endothelial removal. Additionally, saucerneol and saucerneol D were shown to significantly reduce left ventricular pressure (Oh et al., 2008).

### Clinical Application

We also summarized the clinical applications of TMPs and natural products with vasodilatory activities (Figure 5). TMPs and natural products, listed in Figure 5, were mainly used in the treatment of diabetes, hypertension, hyperlipidaemia, and some encephalopathy, which demonstrated the relationship between the vasodilatory activity and the treatment of CCVDs. Some TMPs, such as *M. Vulgare* and *Nigella sativa*, etc, combined with conventional therapeutics improved the efficacy and tolerability of the drugs, and reduced their adverse reactions and side effects. Other TMPs, such as *Hibiscus sabdariffa* L, showed obviously antihypertensive effect. However, the detailed analysis of the drug's underlying mechanism was not performed. In this present study, the role of TMPs and natural products in the prevention and treatment of CCVDs was positive and encouraging, but there were serious limitations in the druggability studies of this field due to lack of researches on toxicology and pharmacokinetics of TMPs and natural products with vasodilatory activities.

### Diabetes

In addition to the medical treatment with glibenclamide, the patients suffering from II diabetes were treated with *M. vulgare* (Ma, 3 weeks). The results showed that Ma could reduce levels of blood glucose (0.64%), cholesterol (4.16%) and TG (5.78%) in patients. *C. obtusifolia* lowered blood glucose (15.25%), cholesterol (14.62%) and TG (42.0%), respectively (Herrera-Arellano et al., 2004a). Therefore, it showed significant differences between the hypoglycemic effect produced by *C. obtusifolia* and *M. vulgare*. The type II diabetes treatment with curcumin (Cu, 1,000  $mg d^{-1}$ ) plus piperine (absorption enhancer, 10  $mg d^{-1}$ ) or placebo plus standard care for 12 weeks, significantly reduced the levels of TC (−21.86 vs. −17.06), non-HDL-C (−23.42 vs. −16.84) and lipoprotein (1.50 vs. −0.34), and increased the levels of HDL-C (1.56 vs. −0.22) as compared with the placebo group (Panahi et al., 2017). *Nigella sativa* (NS) (Kaatabi et al., 2015), *Artemisia herba alba asso* (AH) (Al-Waili 1986) and *Coriandrum sativum* (Cs) (Waheed et al., 2006) can significantly reduce fasting blood glucose in type II diabetes. Furthermore, NS could significantly reduce fasting blood glucose, glycated hemoglobin and glutathione, but also improve the levels of total antioxidant capacity, SOD, catalase and glutathione.

### Hypertension

NS oil (5  $mL d^{-1}$ , 8 weeks) significantly reduced diastolic and SBP in healthy volunteers without adverse effects. But there was no effect on levels of HAA, alanine aminotransferase, alkaline phosphatase, and creatinine (Fallah Huseini et al., 2013). *Hibiscus sabdariffa* L (HS, 9.6  $mg d^{-1}$ ) also significantly

reduced SBP in 30–80 year old hypertension patients, and there was no significant difference with captopril (50 mg d<sup>-1</sup>) (Herrera-Arellano et al., 2004b). Furthermore, aged garlic (Ag, 480/960 mg d<sup>-1</sup>, 12 weeks) significantly reduced SBP in hypertension patients (Ried et al., 2013).

### Hyperlipidemia

NS (Sabzghabae et al., 2012) (2 g d<sup>-1</sup>, 4 weeks) and Strawberries (Basu et al., 2014) (St, 5 or 50 g d<sup>-1</sup>, 12 weeks) significantly reduced serum malondialdehyde, TC, TG, LDL-C or HDL-C in adults. Ferulic acid (Fa, 1 g d<sup>-1</sup>, 6 weeks) could reduce levels of TC, TG, LDL-C or HDL-C in hyperlipidemia patients (Bumrungpert et al., 2018). AM (300 mg d<sup>-1</sup>, 2 months) and Cu (1 g d<sup>-1</sup>, 8 weeks) reduced levels of TC, LDL-C, HDL-C, TG and non-HDL-C in metabolic syndrome (MS) patients (Sikora et al., 2012). Additionally, Genistein (Ge, 54 mg d<sup>-1</sup>, 6 months) improved brachial artery vasodilation in postmenopausal women with MS, while lowering levels of TC, TG, and homocysteine (Irace et al., 2013). Furthermore, Cu (500 mg d<sup>-1</sup>, 10 weeks) reduced body mass index, waist circumference, hip circumference and HDL-C levels in obese girls, but there was no significant difference with control groups (Panahi et al., 2014).

### Encephalopathy

*Ginkgo biloba* extract (GB, 120 mg d<sup>-1</sup>, 52 weeks) improved cognitive performance in dementia patients and improved ADAS-Cog, GERR and CGIC scores in patients. In addition, GB leaves also improved the prognosis with acute ischemic stroke and increased the NIHSS score in patients. GB leaves (240 mg d<sup>-1</sup>, 3 months) could improve memory and attention in senile Alzheimer's disease (Bars et al., 1997; Maurer et al., 1997; Oskouei et al., 2013). Coriander (Co, 4 weeks) also relieved migraine compared with control group by the Akaike criteria (Mansouri et al., 2015).

## CONCLUSION

This review discussed TMPs and natural products with vasodilation *in vitro*. Their possible mechanisms and clinical application were also summarised. Notably, TMPs with vasodilation are mainly from Compositae while natural products are flavonoids. The vasodilatory function of TMPs and natural products is mainly attributed to regulation of eNOS or Ca<sup>2+</sup> channels. Further, we analysed briefly the structure-activity relationship of the compounds with significant vasodilatory effects. The vasodilation of the natural compounds was related to the number and position of double bonds, carbonyls, phenolic hydroxyl groups, and methoxy groups. Overall, these evidences suggested that TMPs and natural products are emerging alternatives for the prevention or treatment of CCVDs such as hypertension.

It was foreseeable that they will receive more attention in the future, although there were still some limitations based on literatures. Firstly, the current thoracic aortic models were usually used as the main mode to investigate the vasodilation,

whereas different vessels, such as cerebral artery, abdominal aorta and mesenteric artery, have not sufficiently explored (Pfaltzgraff and Bader 2015). Secondly, the various channel blockers, such as TEA, BaCl<sub>2</sub>, and 4-AP had been applied to block K<sup>+</sup> channels. However, the relationship between the vasodilation of TMPs and natural products and other, such as KCNQ, TRPC, and TACC, had not been adequately investigated. Therefore, the underlying mechanisms, especially in relationship with various types of ion channels such as KCa, should be further explored. Thirdly, the clinical researches on the vasodilatory activity of TMPs and natural products is severely insufficient in this regard, although they exhibited remarkable potential in animal models. In addition, current the clinical studies on TMPs and natural products also have some deficiencies. For example, mechanisms of action usually remain unknown and only a small number of patients had been reported in almost all the related literatures. Finally, the application of TMPs is troublesome owing to difficulties in source identification, active ingredients, quality standard and mechanism study. Generally, natural products always maintain unsatisfactory druggability owing to their poor oral bioavailability, low plasma concentrations and so on. The ingested natural products was either excreted unabsorbably or metabolized rapidly after absorption, such as apigenin (Tang et al., 2017), sinomenine (Chen W. et al., 2009) and Kaempferol (Calderón-Montaño et al., 2011), etc.

However, developments in technologies such as metabolomics, proteomics and genomics will facilitate the application of TMPs and natural products in the treatment of CCVDs (Harvey et al., 2015). The bioavailability of natural products will be greatly improved by amelioration of hydrophilicity or alteration of administration modes. For instance, the curcumin nanoparticles with higher hydrophilicity achieved by loading into sophorolipid micelles had an appreciably higher bioavailability than that of free curcumin crystals (Peng et al., 2018). In addition, compared to oral administration, nasal administered paeonol was absorbed rapidly in rats (Adki and Kulkarni, 2020). The bioavailability of intravenous injection of cinnamaldehyde was also superior to that of oral administration (Zhu et al., 2017), etc.

In brief, we are still optimistic about the prospect of TMPs and natural products. TMPs will be used as alternative drugs and nutritional supplements, while natural products will be considered as potential therapies for CCVDs in the future. This study provides comprehensive and valuable references for the prevention and treatment of hypertension and CCVDs and sheds light on the further studies in this regard. In the next few years, it is necessary to investigate absorption, distribution, metabolism, and excretion of TMPs and natural products with vasodilation *in vivo*. Also, the activities of the major metabolites of these natural resources should be concerned.

## AUTHOR CONTRIBUTIONS

FT contributed to the drafting of the manuscript. Others were involved in searching, screening the search results, translation, and

data collection. Y-ZT, HA, and CP obtained funding, designed, conceived and supervised process, and revised the manuscript. All the authors have read and approved the final manuscript.

## FUNDING

This work was financially supported by the Program for the National Natural Science Foundation of China (Nos. 81703693 and U19A2010), Sichuan Science and Technology Program (No. 2021JDJQ0040), China Postdoctoral Science

Foundation (No. 2019M653363), and Xinglin Scholar Research Promotion Project of Chengdu University of TCM (No. QNXZ2018009).

## SUPPLEMENTARY MATERIAL

The Supplementary Material for this article can be found online at: <https://www.frontiersin.org/articles/10.3389/fphar.2021.627458/full#supplementary-material>.

## REFERENCES

- Adaramoye, O. A., Anjos, R. M., Almeida, M. M., Veras, R. C., Silvia, D. F., Oliveira, F. A., et al. (2009). Hypotensive and endothelium-independent vasorelaxant effects of methanolic extract from *Curcuma longa* L. in rats. *J. Ethnopharmacol.* 124, 457–462. doi:10.1016/j.jep.2009.05.021
- Adki, K. M., and Kulkarni, Y. A. (2020). Chemistry, pharmacokinetics, pharmacology and recent novel drug delivery systems of paeonol. *Life Sci.* 250, 117544. doi:10.1016/j.lfs.2020.117544
- Ajay, M., Chai, H. J., Mustafa, A., Gilani, A., and Mustafa, M. (2007). Mechanisms of the anti-hypertensive effect of *Hibiscus sabdariffa* L. calyces. *J. Ethnopharmacol.* 109, 388–393. doi:10.1016/j.jep.2006.08.005
- Alexander, S. P. H., and Kelly, E., Mathie, A., Peters, J. A., Veale, E. L., Armstrong, J. F., et al. (2019a) THE CONCISE GUIDE TO PHARMACOLOGY 2019/20: introduction and other protein targets. *J. pharmacol.* 176(Suppl. 1), S1–S20. doi:10.1111/bph.14747
- Alexander, S. P. H., Mathie, A., Peters, J. A., Veale, E. L., Striessnig, J., Kelly, et al. (2019b) The concise guide to pharmacology 2019/20: ion channels. *J. Pharmacol.* 176(Suppl. 1), S142–S228. doi:10.1111/bph.14749
- Alvarez-Collazo, J., Alonso-Carabajo, L., López-Medina, A. I., Alpizar, Y. A., Tajada, S., Nilius, B., et al. (2014). Cinnamaldehyde inhibits L-type calcium channels in mouse ventricular cardiomyocytes and vascular smooth muscle cells. *Pflugers Arch. Eur. J. Physiol.* 466, 2089–2099. doi:10.1007/s00424-014-1472-8
- Al-Waili, N. (1986). Treatment of diabetes mellitus by *Artemisia herba-alba* extract: preliminary study. *Clin. Exp. Pharmacol. Physiol.* 13, 569–574. doi:10.1111/j.1440-1681.1986.tb00940.x
- Ameer, O., Salman, I., Siddiqui, M., Yam, M., Sriramaneni, R., Sadikun, A., et al. (2010). Cardiovascular activity of the n-butanol fraction of the methanol extract of *Loranthus ferrugineus* Roxb. *Braz. J. Med. Biol. Res.* 43, 186–194. doi:10.1590/s0100-879x2010005000002
- Ameer, O. Z., Salman, I. M., Siddiqui, M. J. A., Yam, M. F., Sriramaneni, R. N., Mohamed, A. J., et al. (2010). Pharmacological mechanisms underlying the vascular activities of *Loranthus serrugineus* Roxb. in rat thoracic aorta. *J. Ethnopharmacol.* 127, 19–25. doi:10.1016/j.jep.2009.09.057
- Andrade, D., Borges, L., Torres, I., Conceição, E., and Rocha, M. (2016). Jabuticaba-induced endothelium-independent vasodilating effect on isolated arteries. *Arq. Bras. Cardiol.* 107, 223–229. doi:10.5935/abc.20160118
- Ayele, Y., Urga, K., and Engidawork, E. (2010). Evaluation of *in vivo* antihypertensive and *in vitro* vasodepressor activities of the leaf extract of *Syzygium guineense* (Willd) D.C. *Phytother. Res.* 24, 1457–1462. doi:10.1002/ptr.3141
- Aziz, N., Mehmood, M., Siddiqi, H., Mandukhail, S., Sadiq, F., Maan, W., et al. (2009). Antihypertensive, antidyslipidemic and endothelial modulating activities of *Orchis mascula*. *Hypertens. Res.* 32, 997–1003. doi:10.1038/hr.2009.148
- Barros, F., de la Peña, P., Domínguez, P., Sierra, M. L., and Pardo, L. A. (2020). The EAG voltage-dependent K(+) channel subfamily: similarities and differences in structural organization and gating. *Front. Pharmacol.* 11, 411. doi:10.3389/fphar.2020.00411
- Bars, L. P. L., Katz, M. M., Berman, N., Itil, T. M., Freedman, A. M., et al. (1997). A placebo-controlled, double-blind, randomized trial of an extract of *Ginkgo biloba* for dementia. *JAMA* 278, 1327–1332. doi:10.1001/jama.278.16.1327
- Basu, A., Betts, N. M., Nguyen, A., Newman, E. D., Fu, D., and Lyons, T. J. (2014). Freeze-dried strawberries lower serum cholesterol and lipid peroxidation in adults with abdominal adiposity and elevated serum lipids. *J. Nutr.* 144, 830–837. doi:10.3945/jn.113.188169
- Beech, D. J. (2005). Emerging functions of 10 types of TRP cationic channel in vascular smooth muscle. *Clin. Exp. Pharmacol. Physiol.* 32, 597–603. doi:10.1111/j.1440-1681.2005.04251.x
- Berrougui, H., Martín-Cordero, C., Khalil, A., Hmamouchi, M., Ettaib, A., Marhuenda, E., et al. (2006). Vasorelaxant effects of harmine and harmaline extracted from *Peganum harmala* L. seeds in isolated rat aorta. *Pharmacol. Res.* 54, 150–157. doi:10.1016/j.phrs.2006.04.001
- Bian, K., Doursout, M. F., and Murad, F. (2008). Vascular system: role of nitric oxide in cardiovascular diseases. *J. Clin. Hypertens.* 10, 304–310. doi:10.1111/j.1751-7176.2008.06632.x
- Booranasubkajorn, S., Huabprasert, S., Wattanarangsarn, J., Chotitham, P., Jutasompakorn, P., Laohapand, T., et al. (2017). Vasculoprotective and vasodilatation effects of herbal formula (Sahatsatara) and piperine in spontaneously hypertensive rats. *Phytomedicine* 24, 148–156. doi:10.1016/j.phymed.2016.11.013
- Brenner, R., Peréz, G. J., Bonev, A. D., Eckman, D. M., Kosek, J. C., Wiler, S. W., et al. (2000). Vasoregulation by the beta1 subunit of the calcium-activated potassium channel. *Nature* 407, 870–876. doi:10.1038/35038011
- Briguglio, M., Hrelia, S., Malaguti, M., Serpe, L., Canaparo, R., Dell'Osso, B., et al. (2018). Food bioactive compounds and their interference in drug pharmacokinetic/pharmacodynamic profiles. *Pharmaceutics* 10, 277. doi:10.3390/pharmaceutics10040277
- Brixius, K., Willms, S., Napp, A., Tossios, P., Ladage, D., Bloch, W., et al. (2006). Crataegus special extract WS 1442 induces an endothelium-dependent, NO-mediated vasorelaxation via eNOS-phosphorylation at serine 1177. *Cardiovasc. Drugs Ther.* 20, 177–184. doi:10.1007/s10557-006-8723-7
- Brozovich, F. V., Nicholson, C. J., Degen, C. V., Gao, Y. Z., Aggarwal, M., and Morgan, K. G. (2016). Mechanisms of vascular smooth muscle contraction and the basis for pharmacologic treatment of smooth muscle disorders. *Pharmacol. Rev.* 68, 476–532. doi:10.1124/pr.115.010652
- Buglak, N. E., Jiang, W., and Bahnsen, E. S. (2018). Cinnamic aldehyde inhibits vascular smoothmuscle cell proliferation and neointimal hyperplasia in Zucker diabetic fatty rats. *Redox Biol.* 19, 166–178. doi:10.1016/j.redox.2018.08.013
- Bumrungpert, A., Lilitchan, S., Tuntipopipat, S., Tirawanchai, N., and Komindr, S. (2018). Ferulic acid supplementation improves lipid profiles, oxidative stress, and inflammatory status in hyperlipidemic subjects: a randomized, double-blind, placebo-controlled clinical trial. *Nutrients* 10, 713. doi:10.3390/nu10060713
- Calderón-Montaño, J. M., Burgos-Morón, E., Pérez-Guerrero, C., and López-Lázaro, M. (2011). A review on the dietary flavonoid kaempferol. *Mini Rev. Med. Chem.* 11, 298–344. doi:10.2174/138955711795305335
- Cao, Y. X., Zhang, W., He, J. Y., He, L. C., and Xu, C. B. (2006). Ligustilide induces vasodilatation via inhibiting voltage dependent calcium channel and receptor-mediated Ca<sup>2+</sup> influx and release. *Vasc. Pharmacol.* 45, 171–176. doi:10.1016/j.vph.2006.05.004
- Chen, J., Tsai, S., and Chen, H. (1999). Welsh onion (*Allium fistulosum* L.) extracts alter vascular responses in rat aortae. *J. Cardiovasc. Pharmacol.* 33, 515–520. doi:10.1097/00005344-199904000-00001

- Chen, G., Ye, Y., Li, L., Yang, Y., Qian, A., and Hu, S. (2009). Endothelium-independent vasorelaxant effect of sodium ferulate on rat thoracic aorta. *Life Sci.* 84, 81–88. doi:10.1016/j.lfs.2008.11.003
- Chen, W., Zhou, Y., Kang, J., Li, Q., and Feng, X. (2009). Study on pharmacokinetics and absolute bioavailability of sinomenine in beagle dogs. *Zhongguo Zhong Yao Za Zhi* 34, 468–471.
- Chen, X., Souch, G., Demaree, I. S., White, F. A., and Obukhov, A. G. (2020). Transient receptor potential canonical (TRPC) channels: then and now. *Cells* 9, 1983. doi:10.3390/cells9091983
- Cheng, Y., Li, C., Lee, C., and Kang, J. (2003). Alpha-naphthoflavone induces vasorelaxation through the induction of extracellular calcium influx and NO formation in endothelium. *Naunyn. Schmiedeberg. Arch. Pharmacol.* 368, 377–385. doi:10.1007/s00210-003-0820-6
- Ch'ng, Y., Loh, Y., Tan, C., Ahmad, M., Asmawi, M., Wan Omar, W., et al. (2017). Vasorelaxant properties of Vernonia amygdalina ethanol extract and its possible mechanism. *Pharm. Biol.* 55, 2083–2094. doi:10.1080/13880209.2017.1357735
- Chokri, A., El Abida, K., Zegzouti, Y., and Ben Cheikh, R. (2012). Endothelium-dependent vascular relaxation induced by Globularia alypum extract is mediated by EDHF in perfused rat mesenteric arterial bed. *Can. J. Physiol. Pharmacol.* 90, 607–616. doi:10.1139/y2012-035
- Cifuentes, F., Palacios, J., Paredes, A., Nwokocho, C. R., and Paz, C. (2018). 8-oxo-9-dihydromakomakine isolated from *Aristolelia chilensis* induces vasodilation in rat aorta: role of the extracellular calcium influx. *Molecules* 23, 3050. doi:10.3390/molecules23113050
- Cole, W. C., and Welsh, D. G. (2011). Role of myosin light chain kinase and myosin light chain phosphatase in the resistance arterial myogenic response to intravascular pressure. *Arch. Biochem. Biophys.* 510, 160–173. doi:10.1016/j.abb.2011.02.024
- Collaboration, P. S. (2002). Age-specific relevance of usual blood pressure to vascular mortality: a meta-analysis of individual data for one million adults in 61 prospective studies. *Lancet* 360, 1903–1913. doi:10.1016/s0140-6736(02)11911-8
- Crestani, S., Rattmann, Y., Cipriani, T., de Souza, L., Iacomini, M., Kassuya, C., et al. (2009). A potent and nitric oxide-dependent hypotensive effect induced in rats by semi-purified fractions from *Maytenus ilicifolia*. *Vascul. Pharmacol.* 51, 57–63. doi:10.1016/j.vph.2009.02.005
- de Moura, R., Emiliano, A., de Carvalho, L., Souza, M., Guedes, D., Tano, T., et al. (2005). Antihypertensive and endothelium-dependent vasodilator effects of *Alpinia zerumbet*, a medicinal plant. *J. Cardiovasc. Pharmacol.* 46, 288–294. doi:10.1097/01.fjc.0000175239.26326.47
- Delfan, B., Saki, K., Bahmani, M., Rangsz, N., Delfan, M., Mohseni, N., et al. (2014). A study on anti-diabetic and anti-hypertension herbs used in Lorestan province, Iran. *J. HerbMed Pharmacol.* 3, 71–76.
- Delle Monache, G., Botta, B., Delle Monache, F., Espinal, R., De Bonnevaux, S., De Luca, C., et al. (1993). Novel hypotensive agents from *Verbesina caracasana*. 2. Synthesis and pharmacology of caracasamide. *J. Med. Chem.* 36, 2956–2963. doi:10.1021/jm00072a016
- Dias, K., Da Silva Dias, C., Barbosa-Filho, J., Almeida, R., De Azevedo Correia, N., and Medeiros, I. (2004). Cardiovascular effects induced by reticuline in normotensive rats. *Planta Med.* 70, 328–333. doi:10.1055/s-2004-818944
- Dimo, T., Nguetefack, T., Tan, P., Yewah, M., Dongo, E., Rakotonirina, S., et al. (2003). Possible mechanisms of action of the neutral extract from *Bidens pilosa* L. leaves on the cardiovascular system of anaesthetized rats. *Phytother. Res.* 17, 1135–1139. doi:10.1002/ptr.1132
- El Bardai, S., Morel, N., Wibo, M., Fabre, N., Llabres, G., Lyoussi, B., et al. (2003). The vasorelaxant activity of marrubenol and marrubiin from *Marrubium vulgare*. *Planta Med.* 69, 75–77. doi:10.1055/s-2003-37042
- Fallah Huseini, H., Amini, M., Mohtashami, R., Ghamarchehre, M., Sadeqhi, Z., Kianbakht, S., et al. (2013). Blood pressure lowering effect of *Nigella sativa* L. seed oil in healthy volunteers: a randomized, double-blind, placebo-controlled clinical trial. *Phytother. Res.* 27, 1849–1853. doi:10.1002/ptr.4944
- Fan, G., Zhu, Y., Guo, H., Wang, X., Wang, H., and Gao, X. (2011). Direct vasorelaxation by a novel phytoestrogen tanshinone IIA is mediated by nongenomic action of estrogen receptor through endothelial nitric oxide synthase activation and calcium mobilization. *J. Cardiovasc. Pharmacol.* 57, 340–347. doi:10.1097/fjc.0b013e31820a0da1
- Farias, D., Pereira, A. F., and Rosa, G. (2010). Metabolic syndrome in coronary artery and occlusive vascular diseases: a systematic review. *Arq. Bras. Cardiol.* 94, 150–78. doi:10.1590/s0066-782x2010000600024
- Ferreira, H., Serra, C., Lemos, V., Braga, F., and Cortes, S. (2007). Nitric oxide-dependent vasodilatation by ethanolic extract of *Hancornia speciosa* via phosphatidylinositol 3-kinase. *J. Ethnopharmacol.* 109, 161–164. doi:10.1016/j.jep.2006.06.009
- Figard, H., Girard, C., Mougou, F., Demougeot, C., and Berthelot, A. (2008). Effects of aqueous hop (*Humulus lupulus* L.) extract on vascular reactivity in rats: mechanisms and influence of gender and hormonal status. *Phytomedicine* 15, 185–193. doi:10.1016/j.phymed.2007.09.016
- Flores-Flores, A., Hernández-Abreu, O., Rios, M. Y., León-Rivera, I., Aguilar-Guadarrama, B., Castillo-España, P., et al. (2016). Vasorelaxant mode of action of dichloromethane-soluble extract from *Agastache mexicana* and its main bioactive compounds. *Pharm. Biol.* 54, 2807–2813. doi:10.1080/13880209.2016.1184690
- Fukuda, T., Kuroda, T., Kono, M., Hyoguchi, M., Tanaka, M., and Matsui, T. (2015). Augmentation of ferulic acid-induced vasorelaxation with aging and its structure importance in thoracic aorta of spontaneously hypertensive rats. *Naunyn. Schmiedeberg. Arch. Pharmacol.* 388, 1113–1117. doi:10.1007/s00210-015-1171-9
- Fusi, F., Durante, M., Sgaragli, G., Khanh, P., Son, N., Huong, T., et al. (2015). *In vitro* vasoactivity of zerumbone from *Zingiber zerumbet*. *Planta Med.* 81, 298–304. doi:10.1055/s-0034-1396307
- Garjani, A., Fathiazad, F., Zakheri, A., Akbari, N., Azarmie, Y., Fakhrajoo, A., et al. (2009). The effect of total extract of *Securigergera securidaca* L. seeds on serum lipid profiles, antioxidant status, and vascular function in hypercholesterolemic rats. *J. Ethnopharmacol.* 126, 525–532. doi:10.1016/j.jep.2009.09.003
- Goto, H., Sakakibara, I., Shimada, Y., Kasahara, Y., and Terasawa, K. (2000). Vasodilator effect of extract prepared from *Uncariae ramulus* on isolated rat aorta. *Am. J. Chin. Med.* 28, 197–203. doi:10.1142/s0192415x00000246
- Goto, H., Sasaki, Y., Fushimi, H., Shibahara, N., Shimada, Y., and Komatsu, K. (2005). Effect of curcuma herbs on vasomotion and hemorheology in spontaneously hypertensive rat. *Am. J. Chin. Med.* 33, 449–457. doi:10.1142/s0192415x05003053
- Guedes, D., Silva, D., Barbosa-Filho, J., and de Medeiros, I. (2004). Endothelium-dependent hypotensive and vasorelaxant effects of the essential oil from aerial parts of *Mentha x villosa* in rats. *Phytomedicine* 11, 490–497. doi:10.1016/j.phymed.2004.04.002
- Harvey, A. L., Edrada-Ebel, R., and Quinn, R. J. (2015). The re-emergence of natural products for drug discovery in the genomics era. *Nat. Rev. Drug Discov.* 14, 111–129. doi:10.1038/nrd4510
- Hernández-Pérez, A., Bah, M., Ibarra-Alvarado, C., Rivero-Cruz, J., Rojas-Molina, A., Rojas-Molina, J., et al. (2014). Aortic relaxant activity of *Crataegus Gracilior* Phipps and identification of some of its chemical constituents. *Molecules* 19, 20962–20974. doi:10.3390/molecules191220962
- Herrera-Arellano, A., Aguilar-Santamaria, L., Garcia-Hernandez, B., Nicasio-Torres, P., and Tortoriello, J. (2004a). Clinical trial of *Cecropia obtusifolia* and *Marrubium vulgare* leaf extracts on blood glucose and serum lipids in type 2 diabetics. *Phytomedicine* 11, 561–566. doi:10.1016/j.phymed.2004.01.006
- Herrera-Arellano, A., Flores-Romero, S., Chavez-Soto, M., and Tortoriello, J. (2004b). Effectiveness and tolerability of a standardized extract from *Hibiscus sabdariffa* in patients with mild to moderate hypertension: a controlled and randomized clinical trial. *Phytomedicine* 11, 375–382. doi:10.1016/j.phymed.2004.04.001
- Hill, M. A., Zou, H., Potocnik, S. J., Meininger, G. A., and Davis, M. J. (2001). Invited review: arteriolar smooth muscle mechanotransduction: Ca<sup>2+</sup> signaling pathways underlying myogenic reactivity. *J. Appl. Physiol.* 91, 973–983. doi:10.1152/jappl.2001.91.2.973
- Hoe, S., Lee, C., Mok, S., Kamaruddin, M., and Lam, S. (2011). Gynura procumbens Merr. decreases blood pressure in rats by vasodilatation via inhibition of calcium channels. *Clinics (Sao Paulo)* 66, 143–150. doi:10.1590/s1807-59322011000100025
- Howlett, J. G. (2014). Nebivolol: vasodilator properties and evidence for relevance in treatment of cardiovascular disease. *Can. J. Cardiol.* 30, S29–S37. doi:10.1016/j.cjca.2014.03.003
- Huang, Y., Wong, C., Lau, C., Yao, X., Tsang, S., Su, Y., et al. (2004). Inhibition of nitric oxide/cyclic GMP-mediated relaxation by purified flavonoids, baicalin



- and baicalein, in rat aortic rings. *Biochem. Pharmacol.* 67, 787–794. doi:10.1016/j.bcp.2003.10.002
- Intapad, S., Saengsirisuwan, V., Prasannarong, M., Chuncharunee, A., Suvitayawat, W., Chokchaisiri, R., et al. (2012). Long-term effect of phytoestrogens from *Curcuma comosa* Roxb. on vascular relaxation in ovariectomized rats. *J. Agric. Food Chem.* 60, 758–764. doi:10.1021/jf203173b
- Intapad, S., Suksamrarn, A., and Piyachaturawat, P. (2009). Enhancement of vascular relaxation in rat aorta by phytoestrogens from *Curcuma comosa* Roxb. *Vascul. Pharmacol.* 51, 284–290. doi:10.1016/j.vph.2009.07.005
- Trace, C., Marini, H., Bitto, A., Altavilla, D., Polito, F., Adamo, E. B., et al. (2013). Genistein and endothelial function in postmenopausal women with metabolic syndrome. *Eur. J. Clin. Invest.* 43, 1025–1031. doi:10.1111/eci.12139
- Jabeen, Q., Bashir, S., Lyoussi, B., and Gilani, A. (2009). Coriander fruit exhibits gut modulatory, blood pressure lowering and diuretic activities. *J. Ethnopharmacol.* 122, 123–130. doi:10.1016/j.jep.2008.12.016
- Jiang, H.-D., Cai, J., Xu, J.-H., Zhou, X.-M., and Xia, Q. (2005). Endothelium-dependent and direct relaxation induced by ethyl acetate extract from *Flos chrysanthemi* in rat thoracic aorta. *J. Ethnopharmacol.* 101, 221–226. doi:10.1016/j.jep.2005.04.018
- Jin, B., Qian, L., Chen, S., Li, J., Wang, H., Bruce, I., et al. (2009). Apigenin protects endothelium-dependent relaxation of rat aorta against oxidative stress. *Eur. J. Pharmacol.* 616, 200–205. doi:10.1016/j.ejphar.2009.06.020
- Kaatabi, H., Bamosa, A. O., Badar, A., Al-Elq, A., Abou-Hozaifa, B., Lebda, F., et al. (2015). Nigella sativa improves glycemic control and ameliorates oxidative stress in patients with type 2 diabetes mellitus: placebo controlled participant blinded clinical trial. *PLoS One* 10, e0113486. doi:10.1371/journal.pone.0113486
- Kam, T., Wong, C., Kwan, P., Fat-Yiu, W., Chiu, S., Chan, S., et al. (2012). Effects and mechanism of turmeric vasorelaxation of the thoracic aorta in hypercholesterolemic rats. *J. Med. Food* 15, 190–199. doi:10.1089/jmf.2011.1625
- Kamata, K., Iizuka, T., Nagai, M., and Kasuya, Y. (1993). Endothelium-dependent vasodilator effects of the extract from *Salvia miltiorrhizae* radix. A study on the identification of lithospermic acid B in the extracts. *Gen. Pharmacol.* 24, 977–981. doi:10.1016/0306-3623(93)90176-x
- Kang, D., Lee, J., Choi, D., Sohn, E., Moon, M., and Lee, H. (2005). Vascular relaxation by the methanol extract of Sorbus cortex via NO-cGMP pathway. *Biol. Pharm. Bull.* 28, 860–864. doi:10.1248/bpb.28.860
- Kang, D., Choi, D., Lee, J., Lee, Y., Moon, M., Yang, S., et al. (2007). Endothelial NO/cGMP-dependent vascular relaxation of cornuside isolated from the fruit of *Cornus officinalis*. *Planta Med.* 73, 1436–1440. doi:10.1055/s-2007-990243
- Khan, M., Khan, A., Najeeb-ur-Rehman and Gilani, A. (2015). Blood pressure lowering, vasodilator and cardiac-modulatory potential of *Carum roxburghianum* seed extract. *Clin. Exp. Hypertens.* 37, 102–107. doi:10.3109/10641963.2014.913602
- Khonsung, P., Panthong, A., Chiranthanut, N., and Intahphuak, S. (2011). Hypotensive effect of the water extract of the leaves of *Pseuderanthemum palatiferum*. *J. Nat. Med.* 65, 551–558. doi:10.1007/s11418-011-0540-z
- Kim, B., Kim, J., Kim, A., Kim, Y., Lee, Y., Bae, Y., et al. (2004). Ligusticum wallichii-induced vasorelaxation mediated by mitogen-activated protein kinase in rat aortic smooth muscle. *J. Ethnopharmacol.* 90, 397–401. doi:10.1016/j.jep.2003.11.003
- Kim, H. H., Park, S. Y., Ahn, D. K., and Park, S. K. (2004). Vasodilation effects between the water extract of *Rheum palmatum* L. and *Rheum nudatum* I. In rat thoracic aorta. *Korea. J. Herbol.* 19, 99. doi:10.1002/ptr.2042
- Kim, J., Auger, C., Kurita, I., Anselm, E., Rivoarilala, L., Lee, H., et al. (2013). Aronia melanocarpa juice, a rich source of polyphenols, induces endothelium-dependent relaxations in porcine coronary arteries via the redox-sensitive activation of endothelial nitric oxide synthase. *Nitric Oxide* 35, 54–64. doi:10.1016/j.niox.2013.08.002
- Kratz, J. M., Grienke, U., Scheel, O., Mann, S. A., and Rollinger, J. M. (2017). Natural products modulating the hERG channel: heartaches and hope. *Natural Prod. Rep.* 34, 957–980. doi:10.1039/c7np00014f
- Kubota, Y., Tanaka, N., Umegaki, K., Takenaka, H., Mizuno, H., Nakamura, K., et al. (2001). *Ginkgo biloba* extract-induced relaxation of rat aorta is associated with increase in endothelial intracellular calcium level. *Life Sci.* 69, 2327–2336. doi:10.1016/s0024-3205(01)01303-0
- Kubota, Y., Tanaka, N., Kagota, S., Nakamura, K., Kunitomo, M., Umegaki, K., et al. (2006). Effects of *Ginkgo biloba* extract feeding on salt-induced hypertensive Dahl rats. *Biol. Pharm. Bull.* 29, 266–269. doi:10.1248/bpb.29.266
- Lahlou, S., Tangi, K., Lyoussi, B., and Morel, N. (2008). Vascular effects of *Tanacetum vulgare* L. leaf extract: *in vitro* pharmacological study. *J. Ethnopharmacol.* 120, 98–102. doi:10.1016/j.jep.2008.07.041
- Lee, J., Roh, T., Rho, M., Kim, Y., and Lee, H. (2002). Mechanisms of relaxant action of a pyranocoumarin from *Peucedanum japonicum* in isolated rat thoracic aorta. *Planta Med.* 68, 891–895. doi:10.1055/s-2002-34934
- Lee, P., Chen, W., Liu, L., and Cheng, J. (2007). Vasodilatation induced by sinomenine lowers blood pressure in spontaneously hypertensive rats. *Clin. Exp. Pharmacol. Physiol.* 34, 979–984. doi:10.1111/j.1440-1681.2007.04668.x
- Lee, K., Shin, M., Ham, I., and Choi, H. (2015). Investigation of the mechanisms of *Angelica dahurica* root extract-induced vasorelaxation in isolated rat aortic rings. *BMC. Comple. Altern. Med.* 15, 395. doi:10.1186/s12906-015-0889-8
- Lei, X., and Chiou, G. (1986). Cardiovascular pharmacology of panax notoginseng (burk) F. H. Chen and *Salvia miltiorrhiza*. *Am. J. Chin. Med.* 14, 145–152. doi:10.1142/s0192415x86000235
- Lemos, V., Côrtes, S., dos Santos, M., Ellena, J., Moreira, M., and Doriguetto, A. (2006). Structure and vasorelaxant activity of floranol, a flavonoid isolated from the roots of *Dioclea grandiflora*. *Chem. Biodivers.* 3, 635–645. doi:10.1002/cbdv.200690066
- Li, T., Zhong, Y., Tang, T., Luo, J., Cui, H., Fan, R., et al. (2018). Formononetin induces vasorelaxation in rat thoracic aorta via regulation of the PI3K/PTEEN/Akt signaling pathway. *Drug Des. Devel Ther.* 12, 3675–3684. doi:10.2147/dddt.s180837
- Li, Y., Bao, J., Xu, J., Murad, F., and Bian, K. (2010). Vascular dilation by paeonol—a mechanism study. *Vascul. Pharmacol.* 53, 169–176. doi:10.1016/j.vph.2010.07.001
- Liang, K., Fang, P., Shi, Q., Su, J., Li, B., Chen, S., et al. (2018). [Antihypertensive effect and mechanism of *Dendrobium officinale* flos on high-blood pressure rats induced by high glucose and high fat compound alcohol]. *Zhongguo Zhong Yao Za Zhi* 43, 147–153. doi:10.19540/j.cnki.cjcm.20171027.020
- Liu, C., and Huang, Y. (2016). Chinese herbal medicine on cardiovascular diseases and the mechanisms of action. *Front. Pharmacol.* 7, 469. doi:10.3389/fphar.2016.00469
- Liu, Y., Liu, F., Qiao, M.-M., Guo, L., Chen, M.-H., Peng, C., et al. (2019). Curcumanes A and B, two bicyclic sesquiterpenoids with significant vasorelaxant activity from curcuma longa. *Org. Lett.* 21, 1197–1201. doi:10.1021/acs.orglett.9b00149
- Loai, B., and Zohar, K. (2015). Interactions between CYP3A4 and dietary polyphenols. *Oxid. Med. Cell Longev.* 2015, 854015. doi:10.1155/2015/854015
- Maciel, S., Dias, K., and Medeiros, I. (2004). Calcium mobilization as the endothelium-independent mechanism of action involved in the vasorelaxant response induced by the aqueous fraction of the ethanol extract of *Albizia inopinata* G. P. Lewis (AFL) in the rat aorta. *Phytomedicine* 11, 130–134. doi:10.1078/0944-7113-00361
- Magos, G., Mateos, J., Páez, E., Fernández, G., Lobato, C., Márquez, C., et al. (2008). Hypotensive and vasorelaxant effects of the procyanidin fraction from *Guazuma ulmifolia* bark in normotensive and hypertensive rats. *J. Ethnopharmacol.* 117, 58–68. doi:10.1016/j.jep.2008.01.015
- Mahobiya, A., Singh, T., Rungsung, S., Kumar, T., Chandrasekaran, G., Parida, S., et al. (2018). Kaempferol-induced vasorelaxation via endothelium-independent pathways in rat isolated pulmonary artery. *Pharmacol. Rep.* 70, 863–874. doi:10.1016/j.pharep.2018.03.006
- Malek mohammad, K., Sewell, R. D., and Rafieian-Kopaei, M. (2020). Mechanisms of medicinal plant activity on nitric oxide (NO) bioavailability as prospective treatments for atherosclerosis. *Current Phar. Design.* 9, 301. doi:10.3390/biom9080301
- Mansouri, A., Zayeri, F., Baghestani, A. R., Ghorbanifar, Z., Delavar Kasmaei, H., and Sheidaei, A. (2015). Effect of coriander fruit on clinical course of migraine patients: a comparison between random effect and transition models. *Horizon Med. Sci.* 21, 129–134. doi:10.18869/acadpub.hms.21.2.129
- Martinsen, A., Baccelli, C., Navarro, I., Abad, A., Quetin-Leclercq, J., and Morel, N. (2010). Vascular activity of a natural diterpene isolated from *Croton zambesicus* and of a structurally similar synthetic trachylobane. *Vascul. Pharmacol.* 52, 63–69. doi:10.1016/j.vph.2009.11.002

- Mathieu, P., Poirier, P., Pibarot, P., Lemieux, I., and Després, J. P. (2009). Visceral obesity: the link among inflammation, hypertension, and cardiovascular disease. *Hypertension* 53, 577–584. doi:10.1161/hypertensionaha.108.110320
- Matsumoto, T., Kobayashi, T., Ishida, K., Hirasawa, Y., Morita, H., Honda, T., et al. (2010). Vasodilator effect of cassiarin A, a novel antiplasmodial alkaloid from *Cassia siamea*, in rat isolated mesenteric artery. *Biol. Pharm. Bull.* 33, 844–848. doi:10.1248/bpb.33.844
- Maurer, K., Ihl, R., Dierks, T., and Frölich, L. (1997). Clinical efficacy of *Ginkgo biloba* special extract EGB 761 in dementia of the Alzheimer type. *J. Psychiatr. Res.* 31, 645–655. doi:10.1016/s0022-3956(97)00022-8
- McFadzean, I., and Gibson, A. (2002). The developing relationship between receptor-operated and store-operated calcium channels in smooth muscle. *Br. J. Pharmacol.* 135, 1–13. doi:10.1038/sj.bjp.0704468
- Medeiros, M. A., Pinho, J. F., De-Lira, D. P., Barbosa-Filho, J. M., Araújo, D. A., Cortes, S. F., et al. (2011). Curine, a bisbenzylisoquinoline alkaloid, blocks L-type Ca<sup>2+</sup> channels and decreases intracellular Ca<sup>2+</sup> transients in A7r5 cells. *Eur. J. Pharmacol.* 669, 100–107. doi:10.1016/j.ejphar.2011.07.044
- Melis, M. (1995). Chronic administration of aqueous extract of *Stevia rebaudiana* in rats: renal effects. *J. Ethnopharmacol.* 47, 129–134. doi:10.1016/0378-8741(95)01271-e
- Meng, X., Ni, C., Zhu, L., Shen, Y., Wang, L., and Chen, Y. (2009). Puerarin protects against high glucose-induced acute vascular dysfunction: role of heme oxygenase-1 in rat thoracic aorta. *Vascul. Pharmacol.* 50, 110–115. doi:10.1016/j.vph.2008.11.003
- Mensah, G. A., and Mayosi, B. M. (2013). The 2011 United Nations high-level meeting on non-communicable diseases: the Africa agenda calls for a 5-by-5 approach. *South Afr. Med. J.* 103, 77–79. doi:10.7196/samj.6347
- Mohammadare, A., Changtam, C., Wicha, P., Suksamrarn, A., Tocharus, J., and Tocharus, C. (2015). Mechanisms of vasorelaxation induced by hexahydrocurcuminin isolated rat thoracic aorta. *Phytother. Res.* 29, 1806–1813. doi:10.1002/ptr.5448
- Mori, M. X., Itsuki, K., Hase, H., Sawamura, S., Kurokawa, T., Mori, Y., et al. (2015). Dynamics of receptor-operated Ca<sup>2+</sup> currents through TRPC channels controlled via the PI(4,5)P<sub>2</sub>-PLC signaling pathway. *Front. Pharmacol.* 6, 22. doi:10.3389/fphar.2015.00022
- Musabayane, C., Kamadyaapa, D., Gondwe, M., Moodley, K., and Ojewole, J. (2008). Cardiovascular effects of *Helichrysum ceres* S Moore [Asteraceae] ethanolic leaf extract in some experimental animal paradigms. *Cardiovasc. J. Afr.* 19, 246–253.
- Nasari, M., Arabian, M., Badavi, M., and Ahangarpour, A. (2008). Vasorelaxant and hypotensive effects of *Allium cepa* peel hydroalcoholic extract in rat. *Pak. J. Biol. Sci.* 11, 1569–1575. doi:10.3923/pjbs.2008.1569.1575
- Navedo, M. F., Nieves-Cintrón, M., Amberg, G. C., Yuan, C., Votaw, V. S., Lederer, W. J., et al. (2008). AKAP150 is required for stuttering persistent Ca<sup>2+</sup> sparklets and angiotensin II-induced hypertension. *Circ. Res.* 102, e1–e11. doi:10.1161/circresaha.107.167809
- Nguelefack, T., Dimo, T., Mbuyo, E., Tan, P., Rakotonirina, S., and Kamanyi, A. (2005). Relaxant effects of the neutral extract of the leaves of *Bidens pilosa* Linn on isolated rat vascular smooth muscle. *Phytother. Res.* 19, 207–210. doi:10.1002/ptr.1646
- Nguelefack, T., Fodem, C., Nguelefack-Mbuyo, E., Nyadjeu, P., Wansi, S., Watcho, P., et al. (2015). Endothelium nitric oxide-independent vasorelaxant effects of the aqueous extract from *Stephania abyssinica* on the isolated rat thoracic aorta. *J. Comple. Integr. Med.* 12, 15–21. doi:10.1515/jcim-2014-0022
- Niazmand, S., Fereidouni, E., Mahmoudabady, M., and Mousavi, S. (2014). Endothelium-independent vasorelaxant effects of hydroalcoholic extract from *Nigella sativa* seed in rat aorta: the roles of Ca<sup>2+</sup> and K<sup>+</sup> channels. *Biomed. Res. Int.* 2014, 247054. doi:10.1155/2014/247054
- Nie, H., Meng, L., Zhou, J., Fan, X., Luo, Y., and Zhang, G. (2009). Imperatorin is responsible for the vasodilatation activity of *Angelica dahurica* var. *Formosana* regulated by nitric oxide in an endothelium-dependent manner. *Chin. J. Integr. Med.* 15, 442–447. doi:10.1007/s11655-009-0442-z
- Nie, Q., Zhu, L., Zhang, L., Leng, B., and Wang, H. (2019). Astragaloside IV protects against hyperglycemia-induced vascular endothelial dysfunction by inhibiting oxidative stress and calpain-1 activation. *Life. Sci.* 232, 116662. doi:10.1016/j.lfs.2019.116662
- Nishida, S., and Satoh, H. (2003). Mechanisms for the vasodilations induced by *Ginkgo biloba* extract and its main constituent, bilobalide, in rat aorta. *Life Sci.* 72, 2659–2667. doi:10.1016/s0024-3205(03)00177-2
- Oh, K., Han, W., Wang, M., and Lee, B. (2007a). The effects of chronic treatment with *Morus bombycis* KOIDZUMI in spontaneously hypertensive rats. *Biol. Pharm. Bull.* 30, 1278–1283. doi:10.1248/bpb.30.1278
- Oh, K., Ryu, S., Kim, Y., and Lee, B. (2007b). Large conductance Ca<sup>2+</sup>-activated K<sup>+</sup> (BKCa) channels are involved in the vascular relaxations elicited by piceatannol isolated from *Rheum undulatum* rhizome. *Planta Med.* 73, 1441–1446. doi:10.1055/s-2007-990246
- Oh, K. S., Choi, Y. H., Ryu, S. Y., Oh, B. K., Seo, H. W., Yon, G. H., et al. (2008). Cardiovascular effects of lignans isolated from *Saururus chinensis*. *Planta Med.* 74, 233–238. doi:10.1055/s-2008-1034310
- Ojewole, J., Kamadyaapa, D., Gondwe, M., Moodley, K., and Musabayane, C. (2007). Cardiovascular effects of *Persea americana* Mill (Lauraceae) (avocado) aqueous leaf extract in experimental animals. *Cardiovasc. J. Afr.* 18, 69–76. doi:10.1111/j.1600-0765.1986.tb01485.x
- Orallo, F., and Alzueta, A. (2001). Preliminary study of the vasorelaxant effects of (+)-nantenine, an alkaloid isolated from *Platycapnos spicata*, in rat aorta. *Planta Med.* 67, 800–806. doi:10.1055/s-2001-18848
- Orallo, F., Alvarez, E., Basaran, H., and Lugnier, C. (2004). Comparative study of the vasorelaxant activity, superoxide-scavenging ability and cyclic nucleotide phosphodiesterase-inhibitory effects of hesperetin and hesperidin. *Naunyn. Schmiedebergs Arch. Pharmacol.* 370, 452–463. doi:10.1007/s00210-004-0994-6
- Oskouei, D. S., Rikhtegar, R., Hashemilar, M., Sadeghi-Bazargani, H., Sharifi-Bonab, M., Sadeghi-Hokmabadi, E., et al. (2013). The effect of *Ginkgo biloba* on functional outcome of patients with acute ischemic stroke: a double-blind, placebo-controlled, randomized clinical trial. *J. Stroke Cerebrovasc. Dis.* 22, e557–e563. doi:10.1016/j.jstrokecerebrovasdis.2013.06.010
- Othman, R., Ibrahim, H., Mohd, M., Mustafa, M., and Awang, K. (2006). Bioassay-guided isolation of a vasorelaxant active compound from *Kaempferia Galanga* L. *Phytomedicine* 13, 61–66. doi:10.1016/j.phymed.2004.07.004
- Ouedraogo, M., Ruiz, M., Vardelle, E., Carreyre, H., Coustard, J., Potreau, D., et al. (2011). From the vasodilator and hypotensive effects of an extract fraction from *Agelanthus dononeifolius* (DC) *Danser* (Loranthaceae) to the active compound dononeine. *J. Ethnopharmacol.* 133, 345–352. doi:10.1016/j.jep.2010.10.002
- Pan, Z., Feng, T., Shan, L., Cai, B., Chu, W., Niu, H., et al. (2008). Scutellarin-induced endothelium-independent relaxation in rat aorta. *Phytother. Res.* 22, 1428–1433. doi:10.1002/ptr.2364
- Panahi, Y., Khalili, N., Hosseini, M. S., Abbasnazar, M., and Sahebkar, A. (2014). Lipid-modifying effects of adjunctive therapy with curcuminoids–piperine combination in patients with metabolic syndrome: results of a randomized controlled trial. *Comple. Ther. Med.* 22, 851–857. doi:10.1016/j.ctim.2014.07.006
- Panahi, Y., Khalili, N., Sahebi, E., Namazi, S., Reiner, Ž., Majeed, M., et al. (2017). Curcuminoids modify lipid profile in type 2 diabetes mellitus: a randomized controlled trial. *Comple. Ther. Med.* 33, 1–5. doi:10.1016/j.ctim.2017.05.006
- Paredes, A., Palacios, J., Quispe, C., Nwokocho, C., Morales, G., Kuzmich, J., et al. (2016). Hydroalcoholic extract and pure compounds from *Senecio nutans* Sch. Bip (Compositae) induce vasodilation in rat aorta through endothelium-dependent and independent mechanisms. *J. Ethnopharmacol.* 192, 99–107. doi:10.1016/j.jep.2016.07.008
- Peng, S., Li, Z., Zou, L., Liu, W., Liu, C., and McClements, D. J. (2018). Enhancement of curcumin bioavailability by encapsulation in sophorolipid-coated nanoparticles: an *in vitro* and *in vivo* study. *J. Agric. Food Chem.* 66, 1488–1497. doi:10.1021/acs.jafc.7b05478
- Pfaltzgraff, E. R., and Bader, D. M. (2015). Heterogeneity in vascular smooth muscle cell embryonic origin in relation to adult structure, physiology, and disease. *Dev. Dyn.* 244, 410–416. doi:10.1002/dvdy.24247
- Pinto, N. V., Assreuy, A. M. S., Coelho-de-Souza, A. N., Ceccatto, V. M., Magalhães, P. J. C., Lahlou, S., et al. (2009). Endothelium-dependent vasorelaxant effects of the essential oil from aerial parts of *Alpinia zerumbet* and its main constituent 1, 8-cineole in rats. *Phytomedicine* 16, 1151–1155. doi:10.1016/j.phymed.2009.04.007
- Pires, S., de Assis, T., de Almeida, R., Filho, J., Julien, C., and de Medeiros, I. (2000). Endothelium-derived nitric oxide is involved in the hypotensive and vasorelaxant responses induced by the aqueous fraction of the ethanolic extract of the leaves of *Albizia inopinata* (Harms) G. P. Lewis in rats. *Phytomedicine* 7, 91–98. doi:10.1016/s0944-7113(00)80079-3
- Poh, T., Ng, H., Hoe, S., and Lam, S. (2013). *Gynura procumbens* causes vasodilation by inhibiting angiotensin II and enhancing bradykinin actions. *J. Cardiovasc. Pharmacol.* 61, 378–384. doi:10.1097/fjc.0b013e31828685b3

- Qiao, M.-M., Liu, F., Liu, Y., Guo, L., Zhou, Q.-M., Peng, C., et al. (2019). Curcumane C and (±)-curcumane D, an unusual seco-cadinane sesquiterpenoid and a pair of unusual nor-bisabolane enantiomers with significant vasorelaxant activity from *Curcuma longa*. *Bioorg. Chem.* 92, 103275. doi:10.1016/j.bioorg.2019.103275
- Raimundo, J., Trindade, A., Velozo, L., Kaplan, M., Sudo, R., and Zapata-Sudo, G. (2009). The lignan eudesmin extracted from *Piper truncatum* induced vascular relaxation via activation of endothelial histamine H1 receptors. *Eur. J. Pharmacol.* 606, 150–154. doi:10.1016/j.ejphar.2009.01.038
- Ramdé-Tiendrébéogo, A., Tibiri, A., Ouedraogo, M., Ouedraogo, S., Nacoulma, O., and Guissou, I. (2014). Study of antisickling and vasorelaxant activities and acute toxicity assessment of crude extracts of leaves of *Ficus sycamorua* L. (Moraceae). *Pak. J. Biol. Sci.* 17, 829–835. doi:10.3923/pjbs.2014.829.835
- Ried, K., Frank, O., and Stocks, N. (2013). Aged garlic extract reduces blood pressure in hypertensives: a dose–response trial. *Eur. J. Clin. Nutr.* 67, 64–70. doi:10.1038/ejcn.2012.178
- Roth, G. A., Johnson, C., Abajobir, A., Abd-Allah, F., Abera, S. F., Abyu, G., et al. (2017). Global, regional, and national burden of cardiovascular diseases for 10 causes, 1990 to 2015. *J. Am. Coll. Cardiol.* 70, 1–25. doi:10.1016/j.jacc.2017.04.052
- Sabzghabae, A. M., Dianatkah, M., Sarrafzadegan, N., Asgary, S., and Ghannadi, A. (2012). Clinical evaluation of *Nigella sativa* seeds for the treatment of hyperlipidemia: a randomized, placebo controlled clinical trial. *Med. Arch.* 66, 198–200. doi:10.5455/medarh.2012.66.198-200
- Salahdeen, H., Adebare, A., Murtala, B., and Alada, A. (2015). Potassium channels and prostacyclin contribute to vasorelaxant activities of *Tridax procumbens* crude aqueous leaf extract in rat superior mesenteric arteries. *Afr. J. Med. Med. Sci.* 44, 5–19.
- Sasaki, Y., Goto, H., Tohda, C., Hatanaka, F., Shibahara, N., Shimada, Y., et al. (2003). Effects of curcuma drugs on vasomotion in isolated rat aorta. *Biol. Pharm. Bull.* 26, 1135–1143. doi:10.1248/bpb.26.1135
- Senejoux, F., Girard, C., Kerram, P., Aisa, H., Berthelot, A., Bévalot, F., et al. (2010). Mechanisms of vasorelaxation induced by *Ziziphora clinopodioides* Lam. (Lamiaceae) extract in rat thoracic aorta. *J. Ethnopharmacol.* 132, 268–273. doi:10.1016/j.jep.2010.08.028
- Senejoux, F., Girard, C., Aisa, H., Bakri, M., Kerram, P., Berthelot, A., et al. (2012). Vasorelaxant and hypotensive effects of a hydroalcoholic extract from the fruits of *Nitraria sibirica* Pall. (Nitrariaceae). *J. Ethnopharmacol.* 141, 629–634. doi:10.1016/j.jep.2011.08.012
- Seiichiro, N., and Hiroyasu, S. (2004). Comparative vasodilating actions among terpenoids and flavonoids contained in *Ginkgo biloba* extract. *Clin. Chim. Acta* 339, 855. doi:10.1016/j.cccn.2003.10.004
- Shah, A., and Gilani, A. (2011). Blood pressure lowering effect of the extract of aerial parts of *Capparis aphylla* is mediated through endothelium-dependent and independent mechanisms. *Clin. Exp. Hypertens.* 33, 470–477. doi:10.3109/10641963.2010.549273
- Shaito, A., Thuan, D. T. B., Phu, H. T., Nguyen, T. H. D., Hasan, H., Halabi, S., et al. (2020). Herbal medicine for cardiovascular diseases: efficacy, mechanisms, and safety. *Front. Pharmacol.* 11, 422. doi:10.3389/fphar.2020.00422
- Sikora, J., Broncel, M., Markowicz, M., Chałubiński, M., Wojdan, K., and Mikiciuk-Olasik, E. (2012). Short-term supplementation with *Aronia melanocarpa* extract improves platelet aggregation, clotting, and fibrinolysis in patients with metabolic syndrome. *Eur. J. Nutr.* 51, 549–556. doi:10.1007/s00394-011-0238-8
- Singh, A., Laribi, S., Teerlink, J. R., and Mebazaa, A. (2017). Agents with vasodilator properties in acute heart failure. *Eur. Heart J.* 38, 317–325. doi:10.1093/eurheartj/ehv755
- Skiker, M., Mekhfi, H., Aziz, M., Haloui, B., Lahlou, S., Legssyer, A., et al. (2010). *Artemisia herba-alba* Asso relaxes the rat aorta through activation of NO/cGMP pathway and K(ATP) channels. *J. Smooth Muscle Res.* 46, 165–174. doi:10.1540/jsmr.46.165
- Soares de Moura, R., Costa, S., Jansen, J., Silva, C., Lopes, C., Bernardo-Filho, M., et al. (2002). Bronchodilator activity of *Mikania glomerata* Sprengel on human bronchi and Guinea-pig trachea. *J. Pharm. Pharmacol.* 54, 249–256. doi:10.1211/0022357021778277
- Sowers, J. R., Epstein, M., and Frohlich, E. D. (2001). Diabetes, hypertension, and cardiovascular disease: an update. *Hypertension* 37, 1053–1059. doi:10.1161/01.hyp.37.4.1053
- Su, X., Duan, R., Sun, Y., Wen, J., Kang, D., Lee, H., et al. (2014). Cardiovascular effects of ethanol extract of *Rubus chingii* Hu (Rosaceae) in rats: an *in vivo* and *in vitro* approach. *J. Physiol. Pharmacol.* 65, 417–424.
- Suárez, A., Ulate, G., and Ciccio, J. (2000). Hypotensive action of an aqueous extract of *Pimenta dioica* (Myrtaceae) in rats. *Rev. Biol. Trop.* 48, 53–58.
- Sun, X., Ding, J., Li, H., Pan, N., Gan, L., Yang, X., et al. (2007). Activation of large-conductance calcium-activated potassium channels by puerarin: the underlying mechanism of *Puerarin-mediated* vasodilation. *J. Pharmacol. Exp. Ther.* 323, 391–397. doi:10.1124/jpet.107.125567
- Sureda, A., Silva, A. S., Sánchez-Machado, D. I., López-Cervantes, J., Daglia, M., Nabavi, S. F., et al. (2017). Hypotensive effects of genistein: from chemistry to medicine. *Chem. Biol. Inter.* 268, 37–46. doi:10.1016/j.cbi.2017.02.012
- Suresh Kumar, P., Patel, J., and Saraf, M. (2008). Mechanism of vasorelaxant activity of a fraction of root extract of *Sesamum indicum* Linn. *Indian. J. Exp. Biol.* 46, 457–464. doi:10.1158/1535-7163.MCT-08-0363
- Suzuki, A., Yamamoto, M., Jokura, H., Fujii, A., Tokimitsu, I., Hase, T., et al. (2007). Ferulic acid restores endothelium-dependent vasodilation in aortas of spontaneously hypertensive rats. *Am. J. Hypertens.* 20, 508–513. doi:10.1016/j.amjhyper.2006.11.008
- Tada, Y., Kagota, S., Kubota, Y., Nejime, N., Nakamura, K., Kunitomo, M., et al. (2008). Long-term feeding of *Ginkgo biloba* extract impairs peripheral circulation and hepatic function in aged spontaneously hypertensive rats. *Biol. Pharm. Bull.* 31, 68–72. doi:10.1248/bpb.31.68
- Taiwo, I. A., Odeigah, P. G. C., Jaja, S., and Mojiminiyi, F. (2010). Cardiovascular effects of *Vernonia amygdalina* in rats and the implications for treatment of hypertension in diabetes. *Researcher* 2, 76–79.
- Takagi, S., Goto, H., Shimada, Y., Nakagomi, K., Sadakane, Y., Hatanaka, Y., et al. (2005). Vasodilative effect of perillaldehyde on isolated rat aorta. *Phytomedicine* 12, 333–337. doi:10.1016/j.phymed.2003.08.004
- Takashima, M., Kanamori, Y., Kodera, Y., Morihara, N., and Tamura, K. (2017). Aged garlic extract exerts endothelium-dependent vasorelaxant effect on rat aorta by increasing nitric oxide production. *Phytomedicine* 24, 56–61. doi:10.1016/j.phymed.2016.11.016
- Tan, C., Ch'ng, Y., Loh, Y., Zaini Asmawi, M., Ahmad, M., and Yam, M. (2017). Vasorelaxation effect of *Glycyrrhizae uralensis* through the endothelium-dependent pathway. *J. Ethnopharmacol.* 199, 149–160. doi:10.1016/j.jep.2017.02.001
- Tang, D., Chen, K., Huang, L., and Li, J. (2017). Pharmacokinetic properties and drug interactions of apigenin, a natural flavone. *Expert Opin. Drug Metab. Toxicol.* 13, 323–330. doi:10.1080/17425255.2017.1251903
- Tang, F., Yan, Y. M., Yan, H. L., Wang, L. X., Hu, C. J., Wang, H. L., et al. (2020). Chuanxiongdiolides R4 and R5, phthalide dimers with a complex polycyclic skeleton from the aerial parts of *Ligusticum chuanxiong* and their vasodilator activity. *Bioorg. Chem.* 107, 104523. doi:10.1016/j.bioorg.2020.104523
- Tep-Areenan, P., Sawasdee, P., and Randall, M. (2010). Possible mechanisms of vasorelaxation for 5,7-dimethoxyflavone from *Kaempferia parviflora* in the rat aorta. *Phytother. Res.* 24, 1520–1525. doi:10.1002/ptr.3164
- Testai, L., Silvio, C., Ammar, B., Luisa, P., Vincenzo, C., and Martinotti, E. (2005). Vasorelaxant effects of the chloroformic crude extract of *Bupleurum fruticosum* L. (Umbelliferae) roots on rat thoracic aorta. *J. Ethnopharmacol.* 96, 93–97. doi:10.1016/j.jep.2004.08.024
- Tibiriçá, E., Almeida, A., Cailleaux, S., Pimenta, D., Kaplan, M., Lessa, M., et al. (2007). Pharmacological mechanisms involved in the vasodilator effects of extracts from *Echinodorus Grandiflorus*. *J. Ethnopharmacol.* 111, 50–55. doi:10.1016/j.jep.2006.10.030
- Tokoudagba, J., Auger, C., Bréant, L., N'Gom, S., Chabert, P., Idris-Khodja, N., et al. (2010). Procyranidin-rich fractions from *Parkia Biglobosa* (Mimosaceae) leaves cause redox-sensitive endothelium-dependent relaxation involving NO and EDHF in porcine coronary artery. *J. Ethnopharmacol.* 132, 246–250. doi:10.1016/j.jep.2010.08.031
- Tom, E., Girard, C., Dimo, T., Mbafor, J., Berthelot, A., and Demougeot, C. (2010). Vasorelaxant effects of extracts of the stem bark of *Terminalia superba* Engler and Diels (Combretaceae). *J. Ethnopharmacol.* 127, 335–340. doi:10.1016/j.jep.2009.10.036
- Török, J. (2000). Histamine-induced relaxation in pulmonary artery of normotensive and hypertensive rats: relative contribution of prostanoids,



- nitric oxide and hyperpolarization. *Physiol. Res.* 49, 107–114. doi:10.1086/316715
- Trebak, M. (2012). STIM/Orai signalling complexes in vascular smooth muscle. *J. Physiol.* 590, 4201–4208. doi:10.1113/jphysiol.2012.233353
- Ushida, Y., Matsui, T., Tanaka, M., Matsumoto, K., Hosoyama, H., Mitomi, A., et al. (2008). Endothelium-dependent vasorelaxation effect of rutin-free tartary buckwheat extract in isolated rat thoracic aorta. *J. Nutr. Biochem.* 19, 700–707. doi:10.1016/j.jnutbio.2007.09.005
- Waheed, A., Miana, G., Ahmad, S., and Khan, M. A. (2006). Clinical investigation of hypoglycemic effect of *Coriandrum sativum* in type-2 (NIDDM) diabetic patients. *Pakistan J. Pharmacol.* 23, 7–11.
- Walch, L., Gascard, J. P., Dulmet, E., Brink, C., and Norel, X. (2000). Evidence for a M1 muscarinic receptor 1756 on the endothelium of human pulmonary veins. *Br. J. Pharmacol.* 130, 73–78. doi:10.1038/sj.bjp.0703301
- Wang, Y., Shi, J., Wang, M., Che, C., and Yeung, J. (2009). Vasodilatory actions of xanthenes isolated from a Tibetan herb, *Halenia elliptica*. *Phytomedicine* 16, 1144–1150. doi:10.1016/j.phymed.2009.03.015
- Wansi, S., Nyadjeu, P., Nguetefack, T., Fodouop, S., Donatien, A., and Kamanyi, A. (2012). *In vivo* antioxidant and vasodilating activities of *Gmelina arborea* (Verberaceae) leaves hexane extract. *J. Comple. Integr. Med* 9, doi:10.1515/1553-3840.1623
- Wicha, P., Tocharus, J., Nakaew, A., Pantan, R., Suksamrarn, A., and Tocharus, C. (2015). Ethyl rosmarinate relaxes rat aorta by an endothelium-independent pathway. *Eur. J. Pharmacol.* 766, 9–15. doi:10.1016/j.ejphar.2015.09.003
- Wimmer, R. A., Leopoldi, A., Aichinger, M., Wick, N., Hantusch, B., Novatchkova, M., et al. (2019). Human blood vessel organoids as a model of diabetic vasculopathy. *Nature* 565, 505–510. doi:10.1038/s41586-018-0858-8
- Wong, K., Chan, P., Yang, H., Hsu, F., Liu, L., Cheng, Y., et al. (2004). Isosteviol acts on potassium channels to relax isolated aortic strips of Wistar rat. *Life Sci.* 74, 2379–2387. doi:10.1016/j.lfs.2003.09.065
- Xia, M., Gao, Q., Zhou, X., Qian, L., Shen, Z., Jiang, H., et al. (2007). Vascular effect of extract from mulberry leaves and underlying mechanism. *Zhejiang Da Xue Xue Bao Yi Xue Ban* 36, 48–53.
- Xu, W.-Q., Xiong, Z. Z., Chen, T. T., Gao, X. Y., Yu, H., Zhang, S. Q., et al. (2012). Vasodilation effect of 2-benzyl-5-hydroxy-6-methoxy-3, 4-dihydroisoquinolin-1-one. *Arch. Pharmacol. Res.* 35, 1471–1477. doi:10.1007/s12272-012-0818-z
- Xu, Y. J., Elimban, V., and Dhalla, N. S. (2015). Reduction of blood pressure by store-operated calcium channel blockers. *J. Cell Mol. Med.* 19, 2763–2770. doi:10.1111/jcmm.12684
- Yam, M., Tan, C., Ahmad, M., and Shibao, R. (2016). Mechanism of vasorelaxation induced by eupatorin in the rat aortic ring. *Eur. J. Pharmacol.* 789, 27–36. doi:10.1016/j.ejphar.2016.06.047
- Yam, M. F., Tan, C. S., Ahmad, M., and Ruan, S. (2016). Vasorelaxant action of the chloroform fraction of *Orthosiphon stamineus* via NO/cGMP pathway, potassium and calcium channels. *Am. J. Chin. Med.* 44, 1413–1439. doi:10.1142/s0192415x16500798
- Yamamoto, M., Suzuki, A., Jokura, H., Yamamoto, N., and Hase, T. (2008). Glucosyl hesperidin prevents endothelial dysfunction and oxidative stress in spontaneously hypertensive rats. *Nutrition* 24, 470–476. doi:10.1016/j.nut.2008.01.010
- Yan, Y., Chen, Y., Lin, Y., Guo, J., Niu, Z., Li, L., et al. (2015). Brazilin isolated from the heartwood of *Caesalpinia sappan* L induces endothelium-dependent and -independent relaxation of rat aortic rings. *Acta Pharmacol. Sin.* 36, 1318–1326. doi:10.1038/aps.2015.113
- Yanaga, A., Goto, H., Nakagawa, T., Hikiami, H., Shibahara, N., and Shimada, Y. (2006). Cinnamaldehyde induces endothelium-dependent and -independent vasorelaxant action on isolated rat aorta. *Biol. Pharm. Bull.* 29, 2415–2418. doi:10.1248/bpb.29.2415
- Yilmaz, B., and Usta, C. (2013). Ellagic acid-induced endothelium-dependent and endothelium-independent vasorelaxation in rat thoracic aortic rings and the underlying mechanism. *Phytother. Res.* 27, 285–289. doi:10.1002/ptr.4716
- Yin, Y., Ying, X., Luan, H., Zhao, Z., Lou, J., Wang, D., et al. (2014). UPLC-DAD/Q-TOF-MS based ingredients identification and vasorelaxant effect of ethanol extract of jasmine flower. *Evid. Based Compl. Alternat. Med.* 2014, 707908. doi:10.1155/2014/707908
- Yoo, M., Lee, B., Choi, Y., Lee, J., Seo, J., Oh, K., et al. (2006). Vasorelaxant effect of the rootbark extract of *Paeonia moutan* on isolated rat thoracic aorta. *Planta Med.* 72, 1338–1341. doi:10.1055/s-2006-951678
- Yu, L., and Liu, H. (2018). Perillaldehyde prevents the formations of atherosclerotic plaques through recoupling endothelial nitric oxide synthase. *J. Cell Biochem.* 119, 10204–10215. doi:10.1002/jcb.27362
- Yun, W. J., Zhang, X. Y., Liu, T. T., Liang, J. H., Sun, C. P., Yan, J. K., et al. (2020). The inhibition effect of uncarialin A on voltage-dependent L-type calcium channel subunit alpha-1C: inhibition potential and molecular stimulation. *Inter. J. Biol. Macromol.* 159, 1022–1030. doi:10.1016/j.ijbiomac.2020.05.100
- Yuzurihara, M., Ikarashi, Y., Goto, K., Sakakibara, I., Hayakawa, T., and Sasaki, H. (2002). Geissoschizine methyl ether, an indole alkaloid extracted from *Uncaria ramulus* et *Uncus*, is a potent vasorelaxant of isolated rat aorta. *Eur. J. Pharmacol.* 444, 183–189. doi:10.1016/s0014-2999(02)01623-0
- Zhang, W., Zhang, C., Wang, X., Gao, P., Zhu, D., Chen, H., et al. (2006). Astragaloside IV dilates aortic vessels from normal and spontaneously hypertensive rats through endothelium-dependent and endothelium-independent ways. *Planta Med.* 72, 621–626. doi:10.1055/s-2006-931572
- Zhang, C., Wang, X., Zhong, M., Liu, R., Li, H., Zhang, W., et al. (2007). Mechanisms underlying vasorelaxant action of astragaloside IV in isolated rat aortic rings. *Clin. Exp. Pharmacol. Physiol.* 34, 387–392. doi:10.1111/j.1440-1681.2007.04564.x
- Zhang, T., Niu, C., Mai, W., Jing, H., and Liu, H. (2009). Vasorelaxational effects of total alkali *Sophora Alopecuroids* on rabbit aorta *in vitro*. *Zhongguo Zhong Yao Za Zhi* 34, 2379–2382.
- Zhang, H. T., Wang, Y., Deng, X. L., Dong, M. Q., Zhao, L. M., and Wang, Y. W. (2010). Daidzein relaxes rat cerebral basilar artery via activation of large-conductance Ca<sup>2+</sup>-activated K<sup>+</sup> channels in vascular smooth muscle cells. *Eur. J. Pharmacol.* 630, 100–106. doi:10.1016/j.ejphar.2009.12.032
- Zhang, N., Zou, H., Jin, L., Wang, J., Zhong, M., Huang, P., et al. (2010). Biphasic effects of sodium danshensu on vessel function in isolated rat aorta. *Acta Pharmacol. Sin.* 31, 421–428. doi:10.1038/aps.2010.24
- Zhao, Y., Zhang, X., Li, J., Bian, Y., Sheng, M., Liu, B., et al. (2016). Jujuboside B reduces vascular tension by increasing Ca<sup>2+</sup> influx and activating endothelial nitric oxide synthase. *PLoS One* 11, e0149386. doi:10.1371/journal.pone.0149386
- Zhu, R., Liu, H., Liu, C., Wang, L., Ma, R., Chen, B., et al. (2017). Cinnamaldehyde in diabetes: a review of pharmacology, pharmacokinetics and safety. *Pharmacol. Res.* 122, 78–89. doi:10.1016/j.phrs.2017.05.019

**Conflict of Interest:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Copyright © 2021 Tang, Yan, Wang, Xu, Peng, Ao and Tan. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) and the copyright owner(s) are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.