



Expanded human blood-derived $\gamma\delta$ T cells display potent antigen-presentation functions

Mohd Wajid A. Khan¹, Stuart M. Curbishley², Hung-Chang Chen¹, Andrew D. Thomas¹, Hanspeter Pircher³, Domenico Mavilio^{4,5}, Neil M. Steven⁶, Matthias Eberl¹ and Bernhard Moser^{1*}

¹ Institute of Infection and Immunity, Cardiff University School of Medicine, Cardiff, UK

² NIHR Biomedical Research Unit, Centre for Liver Research, University of Birmingham Medical School, Birmingham, UK

³ Department of Immunology, Institute of Medical Microbiology and Hygiene, University of Freiburg, Freiburg, Germany

⁴ Unit of Clinical and Experimental Immunology, Humanitas Clinical and Research Center, Rozzano, Milan, Italy

⁵ Department of Medical Biotechnologies and Translational Medicine, University of Milan, Milan, Italy

⁶ CR-UK Clinical Trials Unit, School of Cancer Sciences, University of Birmingham Medical School, Birmingham, UK

Edited by:

José Mordoh, Fundación Instituto Leloir, Argentina

Reviewed by:

Fabio Bagnoli, Novartis Vaccines, Italy
María Marcela Barrio, Fundación Cáncer FUCA, Argentina

*Correspondence:

Bernhard Moser, Institute of Infection and Immunity, Cardiff University School of Medicine, Henry Wellcome Building Heath Park, Cardiff CF14 4XN, Wales, UK
e-mail: moserb@cardiff.ac.uk

Cell-based immunotherapy strategies target tumors directly (via cytolytic effector cells) or aim at mobilizing endogenous anti-tumor immunity. The latter approach includes dendritic cells (DC) most frequently in the form of *in vitro* cultured peripheral blood monocytes-derived DC. Human blood $\gamma\delta$ T cells are selective for a single class of non-peptide agonists (“phosphoantigens”) and develop into potent antigen-presenting cells (APC), termed $\gamma\delta$ T-APC within 1–3 days of *in vitro* culture. Availability of large numbers of $\gamma\delta$ T-APC would be advantageous for use as a novel cellular vaccine. We here report optimal $\gamma\delta$ T cell expansion ($>10^7$ cells/ml blood) when peripheral blood mononuclear cells (PBMC) from healthy individuals and melanoma patients were stimulated with zoledronate and then cultured for 14 days in the presence of IL-2 and IL-15, yielding $\gamma\delta$ T cell cultures of variable purity (77 ± 21 and $56 \pm 26\%$, respectively). They resembled effector memory $\alpha\beta$ T (T_{EM}) cells and retained full functionality as assessed by *in vitro* tumor cell killing as well as secretion of pro-inflammatory cytokines (IFN γ , TNF α) and cell proliferation in response to stimulation with phosphoantigens. Importantly, day 14 $\gamma\delta$ T cells expressed numerous APC-related cell surface markers and, in agreement, displayed potent *in vitro* APC functions. Day 14 $\gamma\delta$ T cells from PBMC of patients with cancer were equally effective as their counterparts derived from blood of healthy individuals and triggered potent CD8⁺ $\alpha\beta$ T cell responses following processing and cross-presentation of simple (influenza M1) and complex (tuberculin purified protein derivative) protein antigens. Of note, and in clear contrast to peripheral blood $\gamma\delta$ T cells, the ability of day 14 $\gamma\delta$ T cells to trigger antigen-specific $\alpha\beta$ T cell responses did not depend on re-stimulation. We conclude that day 14 $\gamma\delta$ T cell cultures provide a convenient source of autologous APC for use in immunotherapy of patients with various cancers.

Keywords: $\gamma\delta$ T cells, antigen-presentation, vaccine, $\alpha\beta$ T cells

INTRODUCTION

Dendritic cells (DC) are master regulators of adaptive immunity and are long thought to be a prime target for the treatment of various diseases, including chronic infections, autoimmune diseases, and cancer (1, 2). In principle, DC-based immunotherapies involve two separate strategies: infusion of patients with *in vitro* generated, vaccine-loaded DC and injection of patients with biologicals targeting the patients' own DC (3). The former approach has the advantage of modifying *in vitro* cultured DC prior to their use as cellular vaccine. However, DC do not grow during *in vitro* culture and are scarce in peripheral blood. Therefore, a common strategy involves the *in vitro* generation of DC by culturing blood-derived monocytes for 6 days in the presence of IL-4 and GM-CSF [monocyte-derived DC (moDC)] (4). Again, this method does not yield unlimited numbers of moDC as the majority of cells die during the *in vitro* differentiation process. A hallmark of DC is their exquisite functional diversity underscored by the numerous

distinct DC subsets present in blood and peripheral tissue and their varied reactivity to maturation factors, including cytokines and microbial stimuli (5). These multiple factors may have limited the use of DC-based cellular vaccines in the clinic, explaining the paucity in approved cell products [except for Sipuleucel-T (6)], despite decades of fundamental and clinical research (3).

$\gamma\delta$ T-antigen-presenting cells (APC), activated $\gamma\delta$ T cells with antigen-presentation function, might be a valuable alternative to moDC for use as cellular vaccines in the treatment of patients with cancer (7). $\gamma\delta$ T-APC are generated during short-term activation of human peripheral blood $\gamma\delta$ T cells expressing V γ 9V δ 2-TCR. This particular $\gamma\delta$ T cell subset predominates in peripheral blood (1–5% of total T cells) and recognizes a class of non-peptide ligands, so-called “phosphoantigens.” The most potent phosphoantigen, (*E*)-4-hydroxy-3-methyl-but-2-enyl pyrophosphate (HMBPP), is of microbial origin and outperforms the structurally related phosphoantigen found also in higher organisms,

isopentenyl pyrophosphate (IPP), by $>10^3$ -fold (8). Recent evidence underscores the importance of butyrophilin 3A1/CD277 in presenting HMBPP or IPP to V γ 9V δ 2-TCR $^+$ $\gamma\delta$ T cells, and differences in binding kinetics may explain the striking difference in their potency to induce cell responses (9, 10). Activation of V γ 9V δ 2-TCR $^+$ $\gamma\delta$ T cells by HMBPP/IPP or aminobisphosphonates (nBP) that increase intracellular IPP levels due to inhibition of IPP-metabolizing enzymes, leads to multiple cell responses, including cytokine and chemokine secretion as well as tumor cell lysis (11–13). Indeed, the broad range of tumor cells that are being killed during *in vitro* culture with V γ 9V δ 2-TCR $^+$ $\gamma\delta$ T cells (abbreviated hereafter as $\gamma\delta$ T cells) provided the rationale for targeting these cells in current cancer immunotherapy trials (14, 15).

We here propose to explore the DC-like APC properties of $\gamma\delta$ T cells and to discuss the possibility of translating our findings into a novel type of cellular vaccine. The principles underlying the two $\gamma\delta$ T cell-based translational approaches, i.e., tumor cell-killing and $\alpha\beta$ T cell activation, differ fundamentally from each other. Most notably, tumor cell-killing requires that infused $\gamma\delta$ T cells reach the sites of tumors in order to kill tumor cells during cell-to-cell contact. By contrast, the APC properties of $\gamma\delta$ T cells target endogenous $\alpha\beta$ T cells and, in order to do so, tumor-antigen-presenting $\gamma\delta$ T cells need to interact with tumor-specific $\alpha\beta$ T cells within secondary lymphoid tissues (spleen, lymph nodes). We do not anticipate that the mobilization of tumor-specific $\alpha\beta$ T cells in spleen and lymph nodes is hindered by those $\gamma\delta$ T cells that home to the tumor tissue. In fact, it may well be that tumor cell-killing by itself leads to *in situ* prepared tumor-antigen-presenting $\gamma\delta$ T cells that may further enhance endogenous $\alpha\beta$ T cell responses.

What is the evidence for DC-like properties of activated $\gamma\delta$ T cells? Similar to tumor cell-killing, the APC functionality is the result of extensive *in vitro* studies facilitated by the fact that human peripheral blood $\gamma\delta$ T cells uniformly respond to HMBPP/IPP. Resting peripheral blood $\gamma\delta$ T cells express receptors for inflammatory chemokines and, similar to T_{EM} cells, are in pole position to be recruited to sites of inflammation (16–19). However, during short-term (1–2 days) activation with IPP, the inflammatory homing program in $\gamma\delta$ T cells is switched to a transient lymph node-homing program characterized by CCR7 expression, suggesting their contribution to lymph node activities (16). In addition to cytokine production, IPP stimulation results in surface expression of multiple receptors commonly associated with DC, including antigen-presentation molecules (MHC class II), co-stimulatory receptors (CD40, CD80, CD86), maturation markers (CD83), and adhesion receptors (CD11a, CD11b, CD11c, CD18, CD50, CD54) (20). Indeed, activated $\gamma\delta$ T cells are efficient antigen-processing and peptide–MHC-presenting cells shown to trigger primary (naïve) and memory responses in both CD4 $^+$ and CD8 $^+$ $\alpha\beta$ T cells (20). Activated $\gamma\delta$ T cells even trigger antibody production in B cells that may be facilitated by additional co-stimulatory molecules (ICOS, OX40, CD70) (16, 21). Relevant to tumor cell-killing by cytotoxic T cells, activated $\gamma\delta$ T cells are also able to trigger CD8 $^+$ $\alpha\beta$ T cells via antigen cross-presentation, a process involving the uptake and proteasomal processing of exogenous antigen and peptide loading onto intracellular MHC class I molecules (22, 23). Several laboratories have confirmed and extended these initial observations (24–28), corroborating the DC-like properties of short-term activated $\gamma\delta$ T cells, abbreviated as $\gamma\delta$ T-APC.

An obvious advantage over DC-based vaccines is the capacity of peripheral blood $\gamma\delta$ T cells to selectively expand to large numbers during *in vitro* culture in response to HMBPP/IPP or nBP. In fact, numerous studies have already demonstrated that large numbers (10^9 cells) of $\gamma\delta$ T cells can be grown from peripheral blood mononuclear cells (PBMC) of patients with cancer, although the level of expansion was shown to be influenced by disease stage and/or $\gamma\delta$ T cell frequency (29–40). Here, we report that expanded $\gamma\delta$ T cells maintain critical features of $\gamma\delta$ T-APC, including DC-like antigen-presenting and co-stimulatory molecules as well as the ability to induce antigen-specific CD4 $^+$ and CD8 $^+$ $\alpha\beta$ T cell responses. A protocol for the preparation of a $\gamma\delta$ T-APC-based autologous tumor vaccine is discussed.

MATERIALS AND METHODS

MEDIA, REAGENTS, AND ANTIBODIES

The tissue culture medium used throughout was RPMI 1640 supplemented with 2 mM L-glutamine, 1% non-essential amino acids, 1% sodium pyruvate, 50 μ g/ml penicillin/streptomycin (GIBCO Life Technologies, Carlsbad, CA, USA), and combination of 5% FCS (Hyclone Laboratories, Logan, UT, USA) with 5% human serum (prepared in-house). Human recombinant IL-2 (Proleukin) and IL-15 were purchased from Novartis (Basel, Switzerland) and Miltenyi Biotec (Bergisch Gladbach, Germany), respectively. MicroBeads conjugated to mouse monoclonal anti-PE antibodies (Miltenyi Biotec) were used for magnetic cell separation. Mouse monoclonal antibodies (mAbs) specific for CD3 (UCHT1, SK7XX), CD4 (RPA-T4), CD8 (SK1, HIT8a, RPA-T8), CD11a (HI-111), CD14 (M5E2), CD25 (M-A251), CD27 (M-T271), CD45RA (HI100), CD45RO (UCHL-1), CD54 (HA58), CD56 (B159), CD57 (NK-1), CD62L (DREG-56), CD69 (FN50), HLA-DR (L243), CCR7 (3D12), V δ 2-TCR (B6.1), and IFN γ (β 27) were from BD Biosciences (San Jose, CA, USA); mouse mAbs specific for TCR-V γ 9 (Immu360) and CD40 (mAB89) were from Beckman Coulter (Brea, CA, USA); mouse mAbs specific for CD28 (28.2), CD86 (IT2.2), CD127 (A019D5), and HLA-ABC (w6/32) were from Biologend (San Diego, CA, USA); mouse mAb specific for CD19 (SJ25C1), CD25 (BC96), CD80 (2D10.4), NKG2D/CD314 (1D11), and PE-Cy7 conjugated streptavidin were from eBioscience (San Diego, CA, USA); mouse mAb anti TCR-V γ 9 (7A5) was from Thermo Scientific (Waltham, MA, USA); mouse IgG1 mAb (NCG01) was from Abcam (Cambridge, UK); mouse mAb specific for HLA-A2 (BB7.2) was from Serotec (Kidlington, UK). In addition, rat mAb specific for CCR7 (3D12) was from M. Lipp (Max Delbrück Center for Molecular Medicine, Berlin, Germany), and mouse mAb anti-CD18 (TS1-18) was from R. Pardi (Institute San Raffaele, Milano, Italy), and were used together with appropriate isotype controls during flow cytometric analyses. Biotin-SP-conjugated donkey anti-rat IgG was from Jackson ImmunoResearch Laboratories (West Grove, PA, USA). Carboxyfluorescein diacetate succinimidyl ester (CFSE), LIVE/DEAD® Fixable Aqua Dead Cell Stain Kit and Accucell counting beads were from Invitrogen-Life Technologies (Carlsbad, CA, USA). CellVue and PKH26 were from Sigma-Aldrich (St. Louis, MO, USA).

Peptides were purchased from Severn Biotech (Kidderminster, UK). Recombinant influenza PR8 M1 protein was generated in *E. coli*, isolated to $>95\%$ following standard chromatographic procedures and stored in aliquots at -80°C .

BLOOD DONORS

Donors comprised healthy donors and patients with advanced melanoma in the UK and patients with diverse advanced cancers in Italy. The study with venous blood from healthy human individuals was approved by the South East Wales Ethics Committee under reference number 08/WSE04/17. Samples from patients with melanoma, along with anonymized demographic and treatment data, were acquired from the Human Biomaterials Resource Centre (HBRC), University of Birmingham. The HBRC is a biorepository licensed by the Human Tissue Authority, UK and samples were taken with prospective written generic consent for research use. Samples of hematologic malignancies and metastatic colon cancer were obtained from patients enrolled in trials of translational and experimental medicine at Humanitas Research Hospital, Milan, Italy. These protocols have been approved by the Institutional Review Board (IRB) of the same institute (Approvals 1009 for hematologic malignancies and 1021 for metastatic colon cancer), and each patient had signed a consent form that has been approved by the IRB.

IN VITRO EXPANSION OF $\gamma\delta$ T CELLS

Whole blood samples from healthy donors and patients with cancer were taken into heparin/EDTA. PBMCs were isolated by Ficoll (Lymphoprep, Axis-Shield, Dundee, UK) gradient centrifugation from whole blood and frozen in liquid nitrogen until use. In order to deplete Treg cells, CD25⁺ cells in PBMC were stained with PE-conjugated anti-CD25 mAb in combination with anti-PE microbeads and then retained on magnetic columns according to the manufacturer's instructions (Miltenyi Biotec). The pass-through was evaluated for CD25⁺ cell depletion by flow cytometry and designated as CD25⁺ cell depleted PBMC preparations. For *in vitro* $\gamma\delta$ T cell expansion, PBMC or CD25⁺ cell depleted PBMC were stimulated with 1 μ M zoledronate (Zometa; Novartis, Basel, Switzerland) complete RPMI 1640 medium at a density of 10⁶ cells/ml in 24-well plates and cultured at 37°C in 5% CO₂. At day 5, 8, and 11, cell cultures were split in fresh medium and supplemented with 100 IU/ml IL-2 alone or with 100 IU/ml IL-2 and 20 ng/ml IL-15. At day 14, $\gamma\delta$ T cell cultures were examined by flow cytometry for $\gamma\delta$ T cell expansion, purity, and viability as well as expression of cell surface markers.

CYTOKINE PRODUCTION

Day 14 expanded $\gamma\delta$ T cells from PBMC of healthy individuals and melanoma patients were stimulated with 10 nM HMBPP at 1 \times 10⁶ cells/well in complete RPMI 1640 medium and cultured for 48 h. Cytokines and chemokines in cell-free culture medium were determined using ELISA kits for IL-1 β , IL-8, CCL-1, CCL-4, CXCL13 (R&D Systems, Minneapolis, MN, USA) and IFN γ , TNF α , IL-10, IL-17A, TGF β 1, CCL-2 (eBioscience) and a Dynex MRX II reader (DYNEX Technologies, Chantilly, VA, USA).

CYTOTOXICITY ASSAY

Day 14 $\gamma\delta$ T cell tumor cell-killing activity was tested on the chronic myeloid leukemia cell line KBM7 that was maintained in (1 \times) Iscove's Modified Dulbecco's Medium (GIBCO Life Technologies,

Carlsbad, CA, USA) supplemented with 10% fetal calf serum and penicillin/streptomycin. KBM7 cells were labeled with the membrane dye CellVue prior to co-culture with $\gamma\delta$ T cells. Alternatively, in order to determine the effect of sensitization of KBM7 cells with HMBPP on tumor cell-killing, KBM7 cells were cultured for 18 h in the presence of 1 μ M HMBPP and then labeled with the alternative cell membrane dye PKH26 and mixed with untreated CellVue labeled KBM7 cells at a 1:1 ratio. CellVue labeled KBM7 cells or a mixture of Pkh26 labeled, HMBPP-treated KBM7 cells and untreated CellVue labeled KBM7 cells were then incubated together with day 14 $\gamma\delta$ T cells at increasing effector/target cell ratios for 6 h at 37°C, 5% CO₂. KBM7 cell-killing was determined by flow cytometry by gating on CellVue⁺ and Aqua⁺ cells or PKH26⁺ and Aqua⁺ cells; Accucount counting beads were also used for dead cell count.

In order to examine the involvement of $\gamma\delta$ T cell receptors in KBM7 cell-killing, day 14 $\gamma\delta$ T cells pre-treated with antibodies to V γ 9-TCR (10 μ g/ml), to V δ 2-TCR (1:100 and 1:200 dilution), to CD18 (10 μ g/ml), and to NKG2D (10 μ g/ml) or control mAb (10 μ g/ml) before co-culture with KBM7 cells.

ANTIGEN-PRESENTATION ASSAY

Intracellular IFN γ assay

Day 14 $\gamma\delta$ T cells from HLA-A2⁺ blood donors were incubated for 16–24 h in the presence of 0.01–1 μ M influenza M1 protein and then washed and incubated with brefeldin-A pre-treated, HLA-A2⁺ restricted, M1 p58–66-specific CD8⁺ $\alpha\beta$ T (CD8⁺ responder) cells in 96-well round bottom plates at a ratio of 1:1 (1–0.5 \times 10⁶ cells/well) as described (23). Alternatively, day 14 $\gamma\delta$ T cells were pulsed with 0.1 μ M M1 p58–66 and then rigorously washed before use as APC. After 5–6 h, cells were stained with antibodies to CD3, V γ 9-TCR, and CD8, fixed, and then permeabilized in order to detect intracellular IFN γ by flow cytometry. Brefeldin-A treated CD8⁺ responder cells cultured alone or in the presence of PMA and ionomycin served as negative and positive control, respectively (22, 23).

Proliferation assays

Day 14 $\gamma\delta$ T cells from HLA-A2⁺ blood donors were treated with influenza M1 protein or pulsed with M1 p58–66 exactly as described in the intracellular IFN γ assay. Proliferation was measured in M1 p58–66-specific CD8⁺ $\alpha\beta$ T cells present in freshly prepared, autologous PBMC. Instead of preparing CD3⁺ or CD8⁺ $\alpha\beta$ T cells for use as responder cells (22, 23), PBMC were directly labeled with 1 μ M CFSE for 5 min at room temperature and then washed before use as responder cells. Influenza M1 antigen-treated $\gamma\delta$ T cells and CFSE-labeled PBMC were mixed at a ratio of 1:5 in 96-well round bottom plates. After 6, 8, and 10 days of culture at 37°C, 5% CO₂, responder cells were identified by staining with antibodies to CD3, CD8 in combination with M1 p58–66-tetramers; CFSE^{low}, M1 p58–66-tetramer⁺ cells were enumerated by flow cytometry. CFSE-labeled PBMC cultured alone or in the presence of day 14 $\gamma\delta$ T cells that were not pre-treated with influenza M1 antigens served as negative controls. In a second proliferation assay, day 14 $\gamma\delta$ T cells were treated with 0.01–10 μ g/ml *Mycobacterium tuberculosis* purified protein derivative (PPD) before co-culture with autologous, CFSE-labeled PBMC as

described for influenza M1 protein. Proliferating (CFSE^{low}) cells were detected after day 6, 8, and 10 of culture by staining for activated (CD25⁺) CD3⁺ T cells in combination with either CD4 or CD8 and enumerated by flow cytometry.

STATISTICAL ANALYSIS

Statistical analysis was performed using Student's *t*-test and results were considered significant at *P*-values of <0.05.

RESULTS

OPTIMIZED $\gamma\delta$ T CELL EXPANSION DURING *IN VITRO* CULTURE

Previous $\gamma\delta$ T cell expansion studies commonly used IL-2 as T cell growth factor in combination with phosphoantigen or nBP as V γ 9V δ 2-TCR⁺ $\gamma\delta$ T cell-specific stimuli (29–40). In over 10 individual experiments, we have noticed substantial variations in the yield of $\gamma\delta$ T cells following the stimulation of PBMC from healthy individuals with zoledronate and their subsequent culture for 14 days in the presence of IL-2. The ability of $\gamma\delta$ T cells to expand to large numbers is an important aspect in considering their use in immunotherapy and represents a clear advantage over moDC that fail to grow during tissue culture. Therefore, we sought to optimize the culture conditions for reliable expansion of $\gamma\delta$ T cells by culturing zoledronate-stimulated PBMC from both healthy individuals and patients with cancer in the presence of IL-2 plus IL-15. In addition, due to their inhibitory properties, we examined the effect of removing Treg cells from PBMC on $\gamma\delta$ T cell proliferation during subsequent tissue culture. Accordingly, cultures were initiated by addition of zoledronate to PBMC either with or without prior depletion of CD25⁺ cells. After 5 days, IL-2 alone or in combination with IL-15 was added and cultures were maintained until day 14 when cells were harvested and analyzed for cell survival, $\gamma\delta$ T cell content, and purity. As summarized in **Table 1**, we demonstrate that the combination of IL-2 and IL-15 is approximately 10-fold better than IL-2 alone, yielding >20 × 10⁶ $\gamma\delta$ T cells per 10⁶ input PBMC in 8 out of 13 $\gamma\delta$ T cell cultures reaching >70% purity. Of note, consistently higher cell viability

was obtained in combination with IL-15 compared to IL-2 alone. Depletion of Treg cells (or CD4⁺ and CD8⁺ T cell subsets; not shown) in input PBMC preparations did not significantly improve the outcome of the IL-2 or IL-2/IL-15 cultures, both in terms of $\gamma\delta$ T cell purity and cell viability. In our hands, addition of IL-2 right from the start frequently led to substantial $\alpha\beta$ T cell growth and reduced $\gamma\delta$ T cell counts in day 14 cultures, which was prevented by delayed (day 5) IL-2 addition (alone or in combination with IL-15).

Next, we tested our $\gamma\delta$ T cell expansion protocol on PBMC from patients with cancer (**Table 2**). Out of 10 PBMC samples from melanoma patients with advanced disease, one sample with low $\gamma\delta$ T cell count (0.06% of PBMC) failed to respond during *in vitro* culture. However, the PBMC samples from the remaining nine patients responded well to zoledronate plus IL-2/IL-15 with a range of expansion factors that did not differ substantially from PBMC of healthy donors (**Table 3**). This finding was somewhat surprising considering the substantial variation in the cellular composition of PBMC from melanoma patients as compared to healthy individuals (**Figure 1**). For instance, the fraction of CD3⁺ T cells, and most notably CD8⁺ T cells, was consistently lower in PBMC from melanoma patients, whereas the opposite was found for CD14⁺ monocytes. Similarly, the V γ 9⁺ $\gamma\delta$ T cell count in PBMC from melanoma patients was approximately twofold lower than in PBMC from healthy individuals, which may have been related to the age gap between healthy and melanoma blood donors (41). Reduced $\gamma\delta$ T cell numbers in the original PBMC samples may have accounted for the twofold lower yields of $\gamma\delta$ T cells at day 14 of culture (**Tables 1 and 3**). Of interest, and in clear contrast to a recent report (35), initial depletion of Treg cells in PBMC did not further improve the outcome of $\gamma\delta$ T cell cultures; quite the contrary, the expansion fold and total yield of $\gamma\delta$ T cells was substantially and consistently reduced following removal of CD25⁺ cells, which was not observed in control experiments with PBMC from healthy individuals (**Figure 2**). We have not further examined this observation but suspect that removal of CD25⁺ cells may have

Table 1 | *In vitro* expansion of V γ 9V δ 2-TCR⁺ $\gamma\delta$ T cells from PBMC of healthy donors in response to zoledronate.

Treatment	PBMC		CD25 ⁺ depleted	
	IL-2	IL-2/IL-15	IL-2	IL-2/IL-15
Donors	6	13	6	12
Input of V γ 9 ⁺ T cells (×10 ⁶ /10 ⁶ PBMC) ^a	0.019 ± 0.010	0.021 ± 0.011	0.019 ± 0.010	0.022 ± 0.012
DAY 14 CULTURES				
Live cells (%) ^b	71 ± 11	85 ± 11	77 ± 9	87 ± 6
CD3 ⁺ cells (%) ^c	92 ± 10	96 ± 3	94 ± 4	95 ± 4
V γ 9 ⁺ T cells (%) ^c	56 ± 27	77 ± 21	62 ± 22	74 ± 18
Yield of V γ 9 ⁺ T cells (×10 ⁶ /10 ⁶ PBMC) ^d	2 ± 1.2	29 ± 10.4	2.9 ± 0.94	23 ± 8.7
Expansion fold ^e	104 ± 120	1370 ± 949	150 ± 94	1025 ± 721

^aInitiation of cultures at 10⁶ PBMC per well (24-well plate).

^bLive cells defined as Aqua⁻ cells.

^cPercentage of total live (Aqua⁻) cells.

^dNumber of V γ 9⁺ T cells per 10⁶ of input PBMC after 14 days of culture.

^eYield of V γ 9⁺ T cells divided by input V γ 9⁺ T cells.

Table 2 | Melanoma patients' characteristics.

Patient	Age (years)	Gender	Date of diagnosis	Disease	Surgery and radiotherapy	Therapy
1	68	F	2008	Lymph nodes and lung metastases	Regional nodal dissection	None
2	65	M	2011	Lung and brain metastases	Regional nodal dissection	None
3	49	M	2012	No current known metastases	Regional nodal dissection; excision distant cutaneous and brain metastases; whole brain radiotherapy	None
4	74	F	2010	Nodal, lung, and brain metastases	Regional nodal dissection	None
5	60	F	2011	Nodal and liver metastases	Regional nodal dissection	None
6	52	F	2011	Distant skin metastases	Regional nodal dissection	None
7	31	F	2007	Skin and lung metastases	Regional nodal dissection	None
8	33	F	2001	Lung metastases	Regional nodal dissection; excision gastro-intestinal metastases	None
9	33	M	2010	Skin, lung, and bone metastases	Regional nodal dissection	Dexame-thasone
10	83	M	2012	Nodal, lung, and liver metastases	Regional nodal dissection	Imatinib

F, female; M, male.

Table 3 | *In vitro* expansion of $V\gamma 9V\delta 2$ -TCR⁺ T cells from PBMC of melanoma patients in response to zoledronate.

Treatment of PBMC	IL-2/IL-15	CD25 ⁺ depleted IL-2/IL-15
Donors	9	9
Input of $V\gamma 9^+$ T cells ($\times 10^6/10^6$ PBMC) ^a	0.0095 \pm 0.01	0.0095 \pm 0.01
DAY 14 CULTURES		
Live cells (%) ^b	85 \pm 7	84 \pm 6
CD3 ⁺ cells (%) ^c	95 \pm 4	86 \pm 15
$V\gamma 9^+$ T cells (%) ^c	54 \pm 33	56 \pm 26
Yield of $V\gamma 9^+$ T cells ($\times 10^6/10^6$ PBMC) ^d	8.2 \pm 7.23	2.2 \pm 2.72
Expansion fold ^e	861 \pm 723	230 \pm 272

^aInitiation of cultures at 10^6 PBMC per well (24-well plate).

^bLive cells defined as Aqua⁻ cells.

^cPercentage of total live (Aqua⁻) cells.

^dNumber of $V\gamma 9^+$ T cells per 10^6 of input PBMC after 14 days of culture.

^eYield of $V\gamma 9^+$ T cells divided by input $V\gamma 9^+$ T cells.

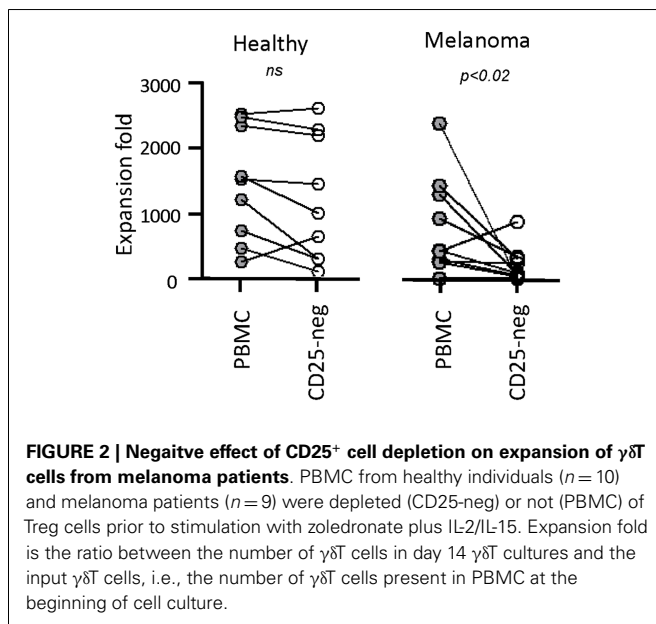
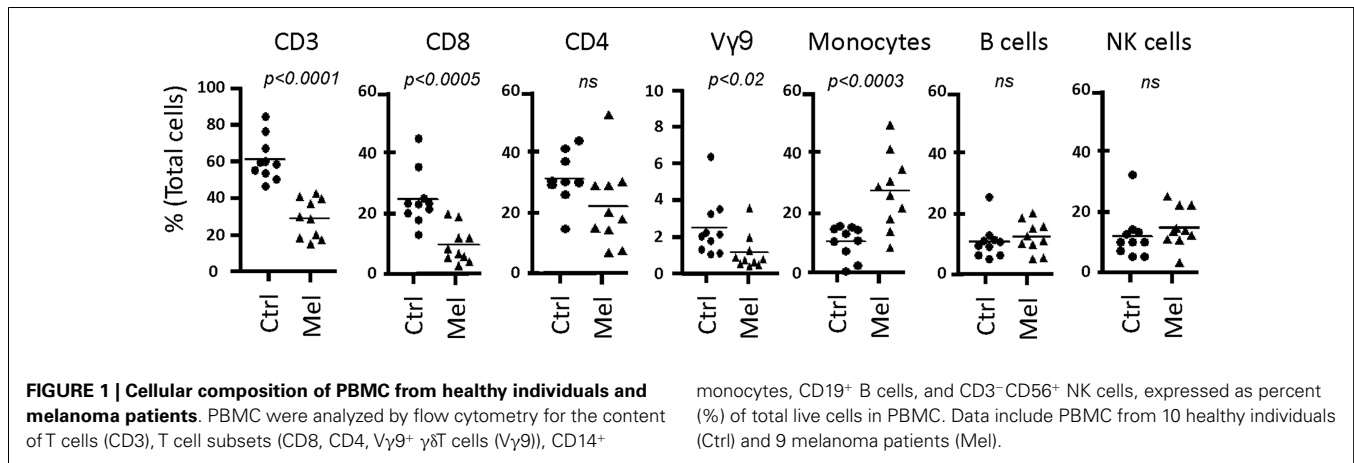
led to a reduction in zoledronate-reactive cells in PBMC samples of patients with cancer. Finally, we did not notice a significant difference in $\gamma\delta$ T cell yield and purity when zoledronate was substituted with IPP or HMBPP (not shown). Overall, our findings document that $\gamma\delta$ T cells in PBMC from melanoma patients responded well to zoledronate plus IL-2/IL-15. The observed donor-to-donor variations fit well with a recent study carried out with PBMC from 30 melanoma patients, although in the latter study PBMC cultures were terminated at day 7 (42).

Day 14 $\gamma\delta$ T cells from melanoma patients were also similar to those derived from PBMC of healthy individuals in terms of their activation and differentiation markers (Figure 3). The degree of

differentiation was determined by correlating cell surface expression of CD27, CD28, CD45RA, and CD45RO, revealing that all day 14 $\gamma\delta$ T cells expanded in IL-2/IL-15 expressed the memory marker CD45RO. By contrast, expression of CD27 and CD28 was highly variable in both day 14 $\gamma\delta$ T cells derived from melanoma patients (3–19 and 2–76%, respectively; $n = 4$) and healthy donors (2–19 and 15–86%, respectively; $n = 5$), reminiscent of effector memory (T_{EM}) cells rather than terminally differentiated effector memory (T_{EMRA}) cells. CD127, a marker for homeostatic proliferation of $\alpha\beta$ T cells, was not detected. By contrast, killer cell lectin-like receptor G1 (KLRG1) was present on most expanded $\gamma\delta$ T cells (Figure 3). The inhibitory receptors PD-1 and CTLA-4 were not detected on day 14 $\gamma\delta$ T cells of patients or healthy blood donors, whereas the exhaustion marker CD57 was present at various levels (7–64%; $n = 8$). Finally, the combination of homing molecules suggests a preference for inflammatory tissues (CXCR3, CCR4, CCR5, CD11a, CD54, etc.) as opposed to secondary lymphoid organs (CXCR5, CCR7, CD62L) (not shown and see below). We conclude that the addition of IL-2 plus IL-15 to zoledronate-stimulated PBMC resulted in large numbers of *in vitro* expanded $\gamma\delta$ T cells with a T_{EM} cell-like phenotype while lacking markers commonly associated with inhibitory T cells.

RESPONSIVENESS OF EXPANDED $\gamma\delta$ T CELLS TO PHOSPHOANTIGENS

The use of expanded $\gamma\delta$ T cells in immunotherapy requires a detailed knowledge about their functionality pertaining to both APC-related and -unrelated cell responses. In agreement with phenotypic marker expression (see above), day 14 $\gamma\delta$ T cells retained responsiveness to HMBPP as evidenced by cytokine production (Table 4), most notably the pro-inflammatory cytokines IFN γ and TNF α . Secretion of TGF β and IL-10 was relatively low, yet highly variable and not affected by HMBPP treatment whereas IL-1 β ,



IL-12, and IL-17 were not detected. In addition, the chemokines CXCL8, CCL1, and CCL4 were found to be present. Again, cytokine secretion profiles did not differ substantially between day 14 $\gamma\delta$ T cells derived from healthy individuals as compared to melanoma patients. As expected (43), zoledronate alone was inactive and cytokine responses were only seen when zoledronate plus monocytes were added to day 14 $\gamma\delta$ T cells (not shown).

Functionality of day 14 $\gamma\delta$ T cells was also evidenced in their proliferation responses. In the presence of IL-2 and IL-15 but in the absence of HMBPP, day 14 $\gamma\delta$ T cells continued to proliferate at variable levels (zero to sixfold) during subsequent 7 days cultures, which may reflect the considerable variation in activation states, as evidenced by the expression of CD69 [75–86 and 24–96% in $\gamma\delta$ T cells from healthy individuals ($n = 5$) and melanoma patients ($n = 9$), respectively] and CD25 [2–36 and 2–76% in $\gamma\delta$ T cells from healthy individuals ($n = 5$) and melanoma patients ($n = 9$), respectively]. Addition of HMBPP killed > 50% of day 14 $\gamma\delta$ T cells within 24 h, but the residual cells resumed proliferation to levels

routinely exceeding the ones seen in cultures without addition of HMBPP.

We also examined *in vitro* tumor cell-killing activity by day 14 $\gamma\delta$ T cells. The underlying mechanisms are still debated but have been proposed to involve either $\gamma\delta$ TCR- or NK receptors-mediated processes or a combination of both pathways. We demonstrate that day 14 expanded $\gamma\delta$ T cells retained tumor cell-killing activity without the need for prior activation (Figure 4). However, the target tumor cells (myeloid KBM7 cells) were much more efficiently killed following overnight incubation with HMBPP. This effect was reduced to the level of untreated KBM7 cells when $\gamma\delta$ TCR-blocking Abs were included, demonstrating that the HMBPP pretreatment of tumor cells directly promoted the $\gamma\delta$ TCR-mediated KBM7 killing. $\gamma\delta$ TCR-blocking Abs did not affect the killing of untreated KBM7 cells whereas the addition of CD18- or NKG2D-blocking Abs reduced the killing of both untreated and HMBPP-pre-treated KBM7 cells (not shown). We conclude that day 14 $\gamma\delta$ T cells retained their ability to respond to $V\gamma 9V\delta 2$ -TCR-specific ligands and were able to generate cell responses that were typically seen with freshly isolated $\gamma\delta$ T cells.

ANTIGEN-PRESENTATION BY EXPANDED $\gamma\delta$ T CELLS ($\gamma\delta$ T-APCs)

The purpose of this study was to determine whether expanded $\gamma\delta$ T cells could be potentially used as a novel type of cellular vaccine for the treatment of patients with cancer. Therefore, we sought to establish whether day 14 $\gamma\delta$ T cells, as opposed to short-term activated $\gamma\delta$ T cells (20, 22, 23), expressed APC markers and were able to induce *in vitro* antigen-specific $\alpha\beta$ T cell responses. A hallmark of $\gamma\delta$ T-APC generated during short-term activation of peripheral blood $\gamma\delta$ T cells was their high-level expression of antigen-presentation (MHC I and II), co-stimulation (CD40, CD80, CD86, etc.), adhesion (CD11a, CD54, etc.), and lymph node homing (CD62L, CCR7) molecules (20). Of interest, and in clear contrast to $\gamma\delta$ T cells freshly isolated from peripheral blood, day 14 $\gamma\delta$ T cells retained many of these APC markers, most notably CD80, CD86, and HLA-DR (MHC II) (Figure 5). The adhesion molecules CD11a and CD54 were still uniformly expressed whereas CD40 and CCR7 (not shown) were low or absent. Maintenance of crucial APC markers may well reflect the presence (albeit at various levels) of activation (CD69) and/or proliferation (CD25)

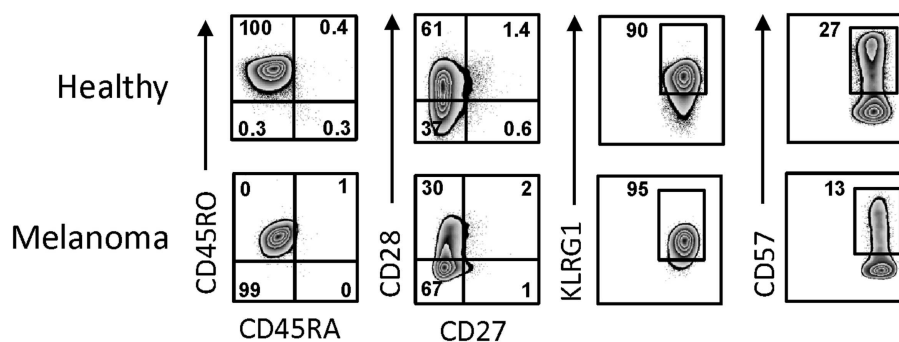


FIGURE 3 | Activation and differentiation marker expression on day 14 $\gamma\delta$ T cells from melanoma patients and healthy individuals. Day 14 $\gamma\delta$ T cells from PBMC of healthy individuals and melanoma patients were analyzed by flow cytometry for the expression of activation and

differentiation markers by gating on V γ 9⁺ cells in combination with the cell surface markers as indicated. One representative example for each of day 14 $\gamma\delta$ T cell cultures of healthy individuals ($n = 5$) and melanoma patients ($n = 4$) are shown.

Table 4 | Secretion of cytokines by day 14 $\gamma\delta$ T cells following stimulation with PA.

Stimulation ^b	Cytokines ^c (pg/ml)				
	IFN γ	TNF α	TGF β	IL-10	
Healthy ^a					
–	7900 \pm 7600	23 \pm 24	77 \pm 97	129 \pm 58	
HMBPP	32000 \pm 22000	550 \pm 427	32 \pm 55	95 \pm 57	
Melanoma ^a					
–	2800 \pm 1700	20 \pm 6	238 \pm 172	183 \pm 317 ^d	
HMBPP	26000 \pm 7000	130 \pm 64	247 \pm 45	205 \pm 353 ^d	

^aDay 14 $\gamma\delta$ T cells from PBMC of either three healthy individuals or three melanoma patients.

^bTreatment of day 14 $\gamma\delta$ T cells with or without 10 nM HMBPP.

^cCytokines determined by ELISA.

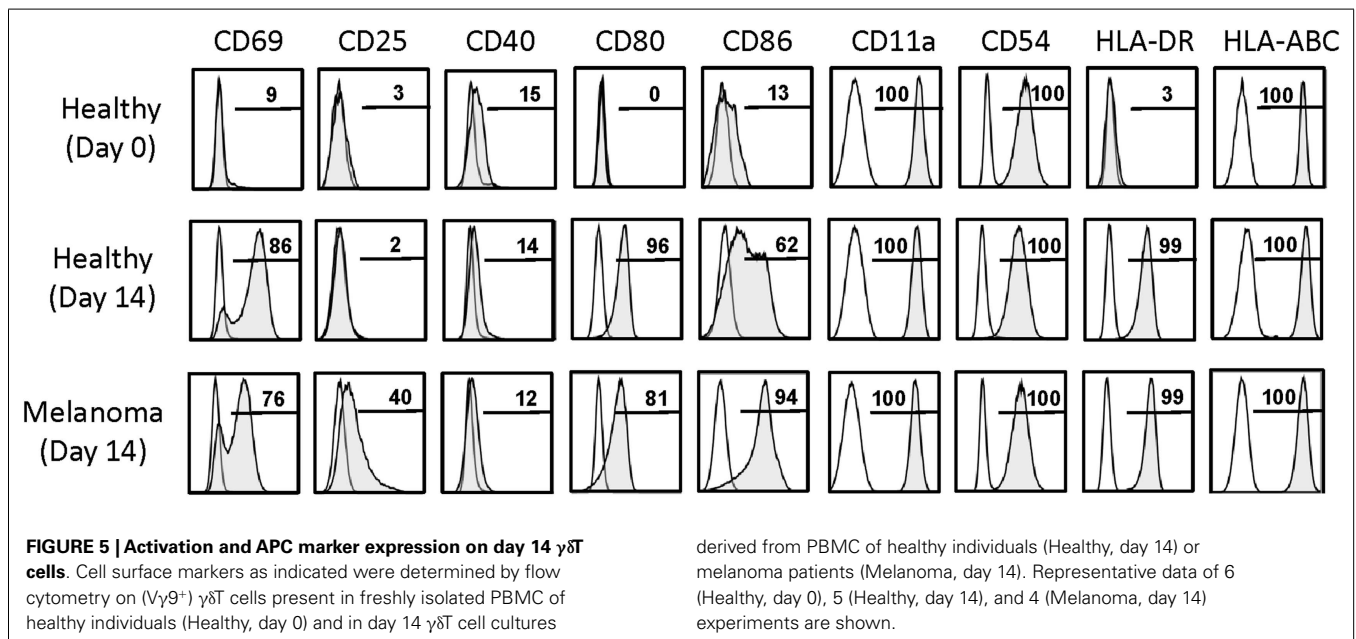
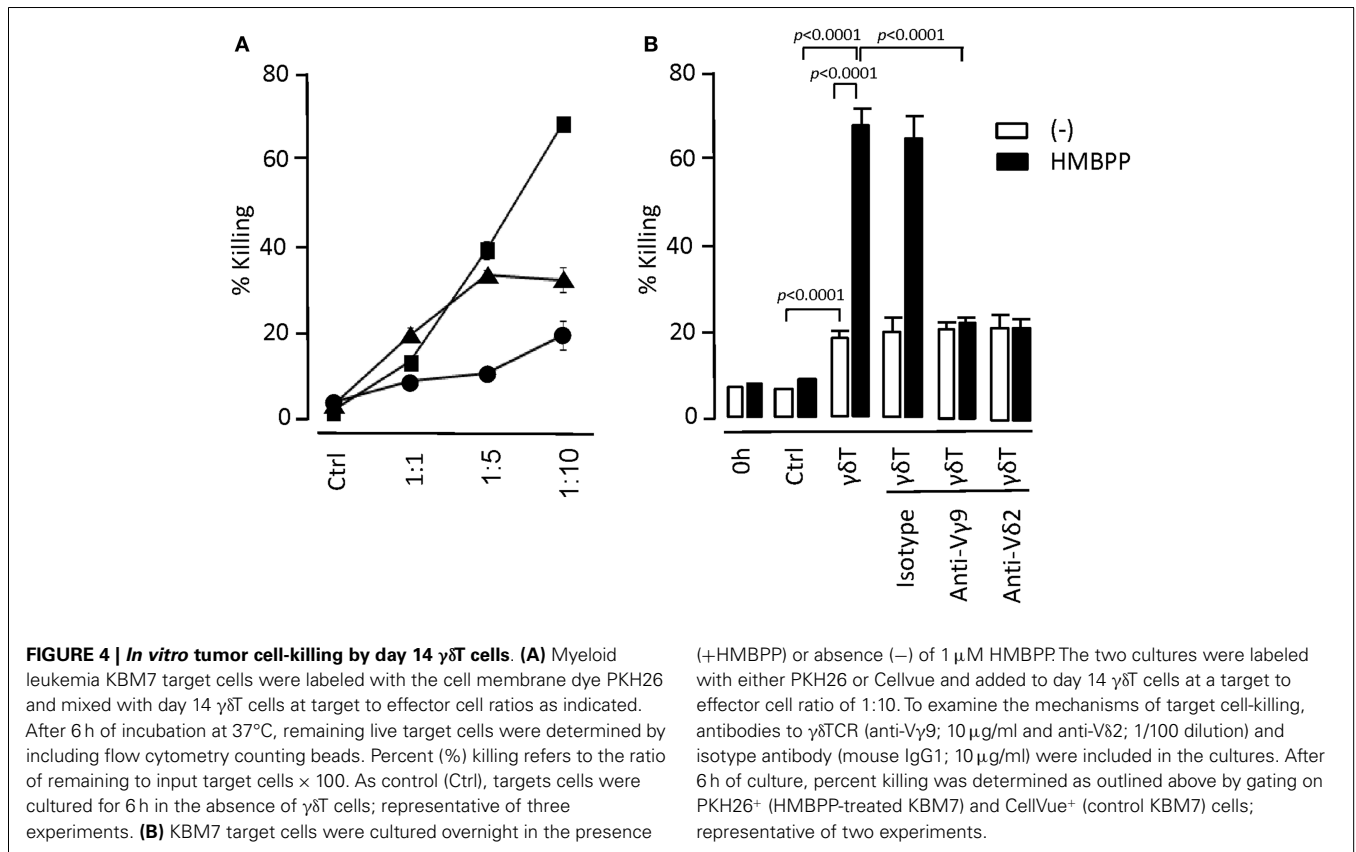
^dTwo out of three melanoma $\gamma\delta$ T cell cultures has no (0 pg/ml) IL-10, irrespective of HMBPP treatment.

markers, which were low/absent on resting (day 0) $\gamma\delta$ T cells. Of interest, re-stimulation of day 14 $\gamma\delta$ T cells with HMBPP for 3 days resulted in moderate CCR7 and CD62L expression (4–16% of cells), indicating that extensively expanded $\gamma\delta$ T cells retained in part the ability to home to secondary lymphoid tissues. Together, these data demonstrate that day 14 $\gamma\delta$ T cells retained many APC markers that are routinely induced in short-term activated $\gamma\delta$ T cells and, importantly, that APC marker expression did not differ between day 14 $\gamma\delta$ T cells derived from healthy individuals and melanoma patients. In fact, preliminary data with PBMC from melanoma-unrelated patients with cancer, including patients with Hodgkin's lymphoma (HL), non-Hodgkin's lymphoma (NHL), and colorectal carcinoma (CRC), revealed similar APC marker expression on expanded $\gamma\delta$ T cells (not shown), indicating that the APC phenotype is a general characteristic of expanded $\gamma\delta$ T cells.

Antigen-presenting cell marker expression suggested that day 14 $\gamma\delta$ T cells would function as APC without the need for re-stimulation, i.e., treatment with phosphoantigens, which is in clear contrast to $\gamma\delta$ T cells freshly isolated from peripheral blood that required stimulation (20). First, we tested the ability of day 14 $\gamma\delta$ T cells to induce $\alpha\beta$ T cell proliferation in response to *M. tuberculosis* PPD as carried out previously (20). To mimic the situation of a first-in-man clinical trial, we used unfractionated PBMC (as opposed to purified $\alpha\beta$ T cells) as responder cells. Following

treatment with PPD, day 14 $\gamma\delta$ T cells were mixed with 5-(and 6-)carboxyfluorescein diacetate succinimidyl ester (CFSE)-labeled, autologous PBMC and proliferating CD4⁺ and CD8⁺ $\alpha\beta$ T cells were identified as CD25⁺ CFSE^{low} cells at d4, d6, and d8 of culture. The representative example in **Figure 6** clearly demonstrates that day 14 $\gamma\delta$ T cells were able to process PPD and present PPD-derived peptides on MHC class I and II to CD8⁺ and CD4⁺ $\alpha\beta$ T cells, respectively, at PPD concentrations as low as 0.1 μ g/ml. Furthermore, CD4⁺ $\alpha\beta$ T cells proliferated much better than CD8⁺ $\alpha\beta$ T cells as evidenced by their higher background (absence of $\gamma\delta$ T cells) and maximal responses ($\gamma\delta$ T cells treated with 10 μ g/ml PPD). Although clearly evident, CD8⁺ $\alpha\beta$ T cell responses were smaller, which may reflect the inefficiency of antigen cross-presentation, a lower frequency of PPD-specific CD8⁺ memory T cells among PBMC, lack of endogenous IL-2 production required for proliferation or a combination of these factors.

In order to examine antigen cross-presentation in more detail, we incubated day 14 $\gamma\delta$ T cells with the influenza M1 protein, which requires proteasomal processing and loading of the M1 immunodominant peptide p58–66 onto intracellular HMC class I (HLA-A2) molecules (22, 23). Following 24 h incubation with graded concentrations of M1 protein, day 14 $\gamma\delta$ T cells were washed and incubated with M1 responder T cells (HLA-A2⁺, M1p58-66-specific CD8⁺ $\alpha\beta$ T cells). After 6 h, intracellular IFN γ production



in responder cells was measured by flow cytometry (Figure 7A). Processing of 1 and 0.1 μ M M1 protein by day 14 $\gamma\delta$ T cells led to robust IFN γ responses, similar to what we have published with short-term activated $\gamma\delta$ T cells (22, 23). The nine amino acid M1p58–66 peptide can be pulsed directly (without the need

of prior uptake and intracellular processing) onto cell surface HLA-A2 on APCs such as DC or, as shown here, $\gamma\delta$ T cells. Thus, M1p58–66-pulsed, day 14 HLA-A2⁺ $\gamma\delta$ T cells initiated maximal intracellular IFN γ responses. Day 14 $\gamma\delta$ T cells from HLA-A2- individuals were unable to present the M1p58–66 peptide and,

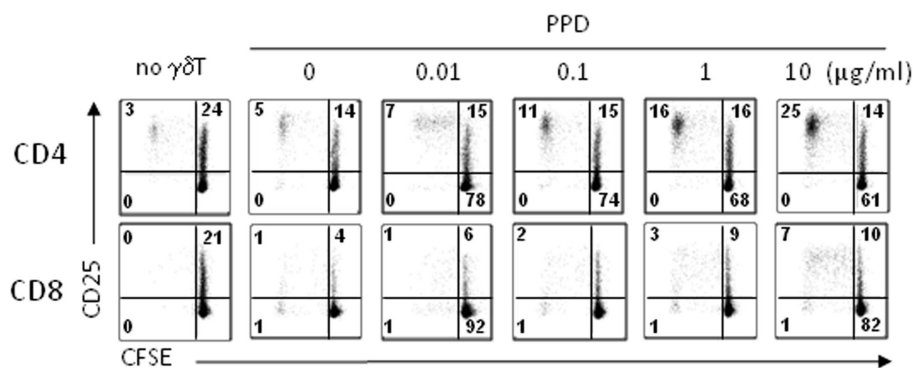


FIGURE 6 | Proliferation responses to PPD-treated day 14 $\gamma\delta$ T cells. Day 14 $\gamma\delta$ T cells from healthy individuals were treated overnight with PPD at concentrations as indicated, washed, and mixed with freshly isolated, CFSE-labeled PBMC from donor-matched blood samples at a $\gamma\delta$ T cell to PBMC ratio of 1:5. At day 6 (and days 8 and

10; not shown), proliferating CD4⁺ and CD8⁺ $\alpha\beta$ T cells were identified by flow cytometry in the V γ 9⁻ CD3⁺ T cell gate as activated (CD25⁺) CFSE^{low} cells. Control cultures (no $\gamma\delta$ T) contained CFSE-labeled PBMC but not $\gamma\delta$ T cells. One representative of four independent experiments is shown.

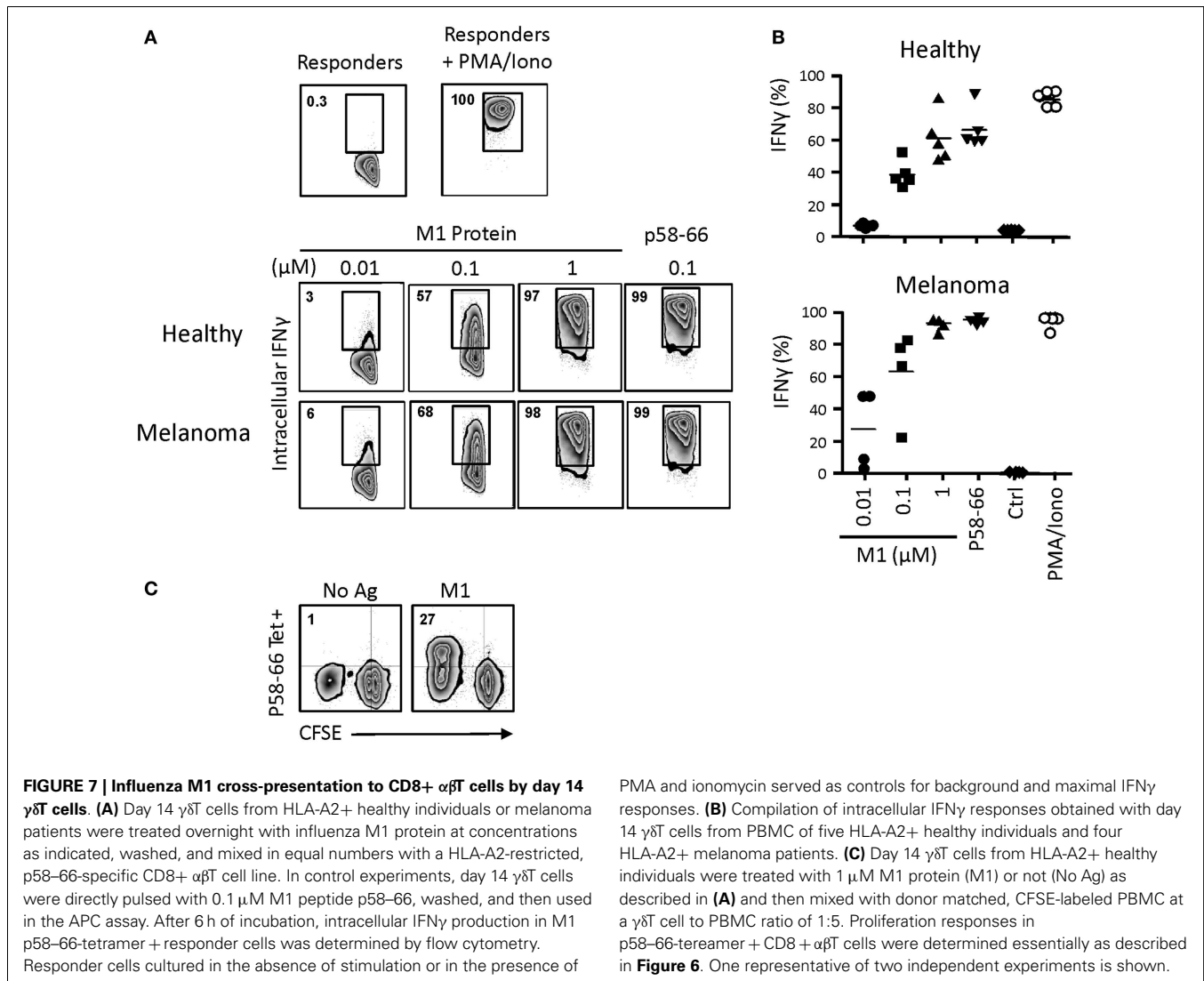
thus, served as negative control (not shown). Of note, we did not detect substantial differences with day 14 $\gamma\delta$ T cells derived from either healthy individuals or melanoma patients in their potencies to induce intracellular IFN γ responses (Figure 7B). Also, donor-matched monocyte-derived DC from PBMC of healthy individuals gave similar results, although these experiments were carried out on separate days (7 days after PBMC isolation; data not shown). The influenza results were replicated with CMV UL83 peptides and UL83 p495–503-specific CD8⁺ $\alpha\beta$ T responder cells (not shown). Moreover, day 14 $\gamma\delta$ T cells from melanoma-unrelated patients (HL, NHL and CRC; see above) were also capable of cross-presenting influenza M1 p58–66 to CD8⁺ $\alpha\beta$ T responder cells (not shown). Finally, we investigated the ability of day 14 $\gamma\delta$ T cells to induce influenza-specific proliferation responses in primary CD8⁺ $\alpha\beta$ T cells (as opposed to cultured cell lines) in unfractionated PBMC as described above. Data in Figure 7C show that influenza M1 treated, day 14 $\gamma\delta$ T cells but not untreated day 14 $\gamma\delta$ T cells induced robust proliferation responses in p58–66-specific CD8⁺ $\alpha\beta$ T cells. In fact, half of all proliferating (CFSE^{low}) cells were p58–66-specific whereas these cells did not expand at all in response to untreated day 14 $\gamma\delta$ T cells, underscoring the M1p58–66 specificity of this response. We conclude that day 14 $\gamma\delta$ T cells behaved similarly to short-term activated $\gamma\delta$ T cells (20) in their ability to process simple (influenza M1) and complex (PPD) antigens and to induce antigen-specific CD8⁺ and CD4⁺ $\alpha\beta$ T cell responses. Equally important, day 14 $\gamma\delta$ T cells from patients with cancer maintained their APC functionality, suggesting that patients-derived $\gamma\delta$ T cells may provide a valuable source for the generation of an autologous cellular vaccine.

DISCUSSION

The principal aim of the present study was the detailed characterization of the APC properties of $\gamma\delta$ T cells following their expansion during *in vitro* culture. It is already well known that V γ 9V δ 2-TCR⁺ $\gamma\delta$ T cells in PBMC respond vigorously to phosphoantigens or nBP leading to extremely large numbers during *in vitro* culture in the presence of T cell growth factors (29–40). Of note, treatment of

PBMC with phosphoantigen or nBP is highly selective for blood $\gamma\delta$ T cells such that unrelated lymphocytes, including $\alpha\beta$ T cells and NK cells, do not respond and are “left behind” during *in vitro* culture. This simple technique represents a considerable advantage over $\alpha\beta$ T cells when large numbers are needed for autologous cell therapy. Since $\gamma\delta$ T cells themselves do not produce sufficient IL-2, this or other T cell growth factors need to be included during *in vitro* $\gamma\delta$ T cell expansion. In our hands, delayed addition of IL-2 prevented early expansion of unrelated lymphocytes and, consequently, prevented outgrowth of $\alpha\beta$ T cells or NK cells. Since our previous work with IL-2 provided inconsistent results in terms of yield and purity of expanded $\gamma\delta$ T cells, we decided to modify the *in vitro* culture procedure by including either IL-15 or by depleting Treg cells prior to cell culture. In short, Treg cell depletion, i.e., removal of CD25⁺ cells by MACS beads isolation columns, did not affect $\gamma\delta$ T cell cultures from PBMC of healthy individuals whereas the addition of IL-15 was clearly beneficial (see below). Interestingly, we observed that removal of CD25⁺ cells from PBMC of melanoma patients was inhibitory, supporting the view that this treatment removed zoledronate-responsive cells that were required for optimal $\gamma\delta$ T cell activation. In fact, PBMC from patients with cancer contained numerous CD25⁺ monocytes, which were rare in PBMC from healthy individuals, suggesting that their removal might have contributed to the reduced responsiveness to zoledronate.

IL-2 is known to induce terminal T cell differentiation and, eventually, activation-induced cell death, which may limit the expansion and/or viability of cultured T cells (44). By contrast, and in addition to its T cell growth factor properties, IL-15 induces T cell survival (45, 46) and, in CD8⁺ $\alpha\beta$ T cells, enhances cytotoxic functions (47, 48). IL-15 alone or in combination with IL-2 has been used successfully in cultures of tumor-infiltrating $\alpha\beta$ T cells isolated from melanoma (49) and Merkel cell carcinoma patients (50). The action of IL-15 on human blood $\gamma\delta$ T cells is not well understood. In agreement with its involvement in homeostatic proliferation of $\gamma\delta$ T cells in mice (51, 52), persistence of $\gamma\delta$ T cells in patients with cancer has been explained by the presence of



endogenous IL-15 (53). Here, we report that the combination of IL-2 and IL-15 made a substantial difference as compared to IL-2 alone during culturing of $\gamma\delta$ T cells derived from both patients and healthy blood donors.

A high yield of expanded, autologous $\gamma\delta$ T cells is certainly desirable for their further use as cellular vaccine. However, the quality of expanded $\gamma\delta$ T cells in terms of their state of differentiation and responsiveness to re-stimulation are equally important parameters to consider. Day 14 $\gamma\delta$ T cells bear surface markers reminiscent of T_{EM} cells, although the validity of adopting this terminology for $\gamma\delta$ T cells is questionable. T_{EM} cells and central-memory T (T_{CM}) cells describe subsets of memory $\alpha\beta$ T cells present in peripheral blood and were originally defined on the basis of the expression of lymph node-homing markers (CD62L, CCR7) (54). Memory subsets, most notably CD8+ $\alpha\beta$ T cell subsets, were further defined by the expression of CD45RA and CD45RO in combination with co-stimulatory receptors (CD27, CD28) (55). Except for CCR7/CD27/CD28 triple-negative CD8+ $\alpha\beta$ T cells, all T cell subsets purely reflect a hallmark of $\alpha\beta$ T cell biology, namely

their need to interact with DC in lymph nodes or peripheral tissues in order to receive appropriate activation/differentiation signals. $\gamma\delta$ T cells, including the dominant subset of V γ 9V δ 2-TCR+ $\gamma\delta$ T cells in peripheral blood, are not controlled by DC in secondary lymphoid tissues, suggesting fundamental mechanistic differences in controlling responsiveness between these two classes of T cells. Nevertheless, day 14 $\gamma\delta$ T cells share hallmarks of late-differentiated T_{EM} cells with intact replication capacity because they lack CD45RA, CCR7, and for the most part CD27 but largely maintain CD28 expression. Uniform expression of KLRG1, otherwise a marker for replication-defective CD8+ $\alpha\beta$ T cells (56), may signify a role for these cells in epithelial tissues where the KLRG1 ligand, E-cadherin, is ubiquitously expressed (57). In agreement, day 14 $\gamma\delta$ T cells responded well to re-stimulation with HMBPP or zoledronate plus PBMC as assessed by cytokine production and $\gamma\delta$ T cell proliferation. Of note, day 14 $\gamma\delta$ T cells primarily produced pro-inflammatory cytokines (IFN γ , TNF α), much like activated peripheral blood $\gamma\delta$ T cells (58) but failed to secrete regulatory cytokines TGF β and IL-10 in response to HMBPP or to

express the inhibitory receptors PD-1 and CTLA-4. Day 14 $\gamma\delta$ T cells also maintained tumor cell-killing activity, most notably when HMBPP-pre-treated tumor cells were used as target cells. Collectively, stimulation of PBMC with zoledronate and culturing in the presence of IL-2 plus IL-15 resulted in large numbers of functionally competent $\gamma\delta$ T cells featuring immunostimulatory and cytotoxic as opposed to inhibitory properties.

Most relevant for the potential use of expanded $\gamma\delta$ T cell as a novel cellular vaccine were the abundant expression of APC-relevant cell surface markers and the capacity of day 14 $\gamma\delta$ T cells to trigger antigen-specific $\alpha\beta$ T cell responses *in vitro*. Professional APC, such as DC, have to be able to initiate adhesive interactions with $\alpha\beta$ T cells in order to allow $\alpha\beta$ T cells to screen the APC surface for the presence of their cognate peptide–MHC complexes (59). Engagement of $\alpha\beta$ TCR on CD8⁺ and CD4⁺ $\alpha\beta$ T cells with peptide–MHC class I and class II complexes on DC, respectively, leads to $\alpha\beta$ T cells priming and subsequent engagement of co-stimulatory receptors initiates $\alpha\beta$ T cell proliferation and differentiation. Interference in any of these early steps has profound effects on $\alpha\beta$ T cell responses. We here show that day 14 $\gamma\delta$ T cells abundantly express CD11a, CD54 (adhesion) and HLA-DR and HLA-ABC (antigen-presentation) molecules as well as the CD28 ligands CD80 and CD86, which trigger essential CD28-mediated co-stimulatory signals in $\alpha\beta$ T cells (60). Indeed, this APC phenotype is mirrored by the APC function, as demonstrated by the induction of antigen-specific CD4⁺ and CD8⁺ $\alpha\beta$ T cell proliferation in response to expanded $\gamma\delta$ T cells that have processed either the complex microbial antigen PPD or influenza M1 protein. We did not notice significant differences between day 14 $\gamma\delta$ T cells derived from PBMC of healthy individuals as compared to patients with cancer with regard to their ability to cross-present M1 protein. Of note, the cellular composition of PBMC from melanoma patients differed substantially from PBMC of healthy individuals, possibly reflecting the history of cancer treatment regimens and potential immunosuppressive conditions (61). However, day 14 $\gamma\delta$ T cells have undergone many rounds of cell division and, thus, appear “far removed” from the inhibitory conditions present in the original PBMC samples, which may explain their undiminished functionality. In this respect, our data confirm previous findings about $\gamma\delta$ T cell-induced CD8⁺ $\alpha\beta$ T cell responses although patients-derived $\gamma\delta$ T cells were examined at their peak-time (day 7/8) of proliferation (25, 27).

In summary, we demonstrated that expanded $\gamma\delta$ T cells from PBMC of both healthy individuals and patients with cancer displayed potent APC functions and responsiveness to $\gamma\delta$ T cell antigens. Our data support the view that expanded $\gamma\delta$ T cells may be exploited as a novel cellular vaccine for the treatment of patients with cancer. Relatively small (\approx 100 ml) blood samples of patients suffice to generate large numbers ($>10^9$) of expanded $\gamma\delta$ T cells within 2 weeks of *in vitro* culture in response of zoledronate and the T cell growth and survival factors IL-2 and IL-15. Cultured $\gamma\delta$ T cells efficiently take up, process, and cross-present complex antigens (20, 22), including proteins present in cell extracts (23). Therefore, we propose to treat expanded $\gamma\delta$ T cells with tumor antigens, either in the form of cellular extracts derived from patients' own tumor cells or from heterologous tumor cell lines. The latter approach would also provide highly stimulatory alloantigens

that may boost the adaptive immune response. Tumor-antigen-loaded $\gamma\delta$ T cells are then infused into patients. Since expanded $\gamma\delta$ T cells retain the capacity to secrete pro-inflammatory cytokines (IFN γ , TNF α) and to express, albeit at reduced levels, lymph node-homing markers (CCR7, CD62L), the infusion of tumor-antigen-loaded $\gamma\delta$ T cells may be followed by a single treatment with zoledronate. Thus, a fraction of activated $\gamma\delta$ T cells may acquire the ability to home to secondary lymphoid tissues (spleen, lymph nodes), where mobilization of endogenous tumor-specific $\alpha\beta$ T cells takes place. The anti-tumor activity mediated by this new wave of endogenous effector T cells may be further supported by the infused $\gamma\delta$ T cells that retained direct tumor cell-killing activity. A substantial body of evidence by numerous laboratories has already documented the safety of large (and repetitive) infusions of $\gamma\delta$ T cells (33, 37, 38, 62–67). Now, we need to test whether the clinical outcome is improved by treating patients with tumor-antigen-presenting $\gamma\delta$ T cells targeting the patients' own tumor-specific T cells as opposed to tumor cells. It is possible that the immune stimulatory effects of $\gamma\delta$ T cell-based vaccines are further enhanced when applied in combination with promising immune checkpoint inhibitors (anti-PD-1 and anti-CTLA-4 antibodies) (68).

ACKNOWLEDGMENTS

The authors thank A. Sewell and G. Dolton (Cardiff, UK) for the generous provision of influenza M1-specific tetramers and initial influenza M1-specific CD8⁺ responder cell lines, M. Luo (Birmingham, AL, USA) for the influenza M1 expressing DNA plasmid, H. Jomaa (Giessen, Germany) for HMBPP, P. Lehner (Cambridge, UK) for KBM7 cells, and R. Pardi (Milano, Italy) for the anti-CD18 antibody. This study has been supported by the Birmingham Experimental Cancer Medicine Centre, the Early Drug Development team in the Cancer Research UK Clinical Trials Unit, Birmingham and the Human Biomaterials Resource Centre, University of Birmingham and University Hospital Birmingham NHS Foundation Trust. This work was funded by a grant from the CR-UK to Bernhard Moser, and a Tenovus Ph.D. studentship to Hung-Chang Chen; Bernhard Moser is the recipient of a Royal Society Wolfson Research Merit Award.

REFERENCES

- Banchereau J, Palucka AK. Dendritic cells as therapeutic vaccines against cancer. *Nat Rev Immunol* (2005) 5:296–306. doi:10.1038/nri1592
- Steinman RM, Banchereau J. Taking dendritic cells into medicine. *Nature* (2007) 449:419–26. doi:10.1038/nature06175
- Palucka K, Banchereau J. Dendritic-cell-based therapeutic cancer vaccines. *Immunity* (2013) 39:38–48. doi:10.1016/j.immuni.2013.07.004
- Sallusto F, Lanzavecchia A. Efficient presentation of soluble antigen by cultured human dendritic cells is maintained by granulocyte/macrophage colony-stimulating factor plus interleukin 4 and downregulated by tumor necrosis factor alpha. *J Exp Med* (1994) 179:1109–18. doi:10.1084/jem.179.4.1109
- Banchereau J, Steinman RM. Dendritic cells and the control of immunity. *Nature* (1998) 392:245–52. doi:10.1038/32588
- Kantoff PW, Higano CS, Shore ND, Berger ER, Small EJ, Penson DE, et al. Sipuleucel-T immunotherapy for castration-resistant prostate cancer. *N Engl J Med* (2010) 363:411–22. doi:10.1056/NEJMoa1001294
- Moser B, Eberl M. gammadelta T-APCs: a novel tool for immunotherapy? *Cell Mol Life Sci* (2011) 68:2443–52. doi:10.1007/s00018-011-0706-6
- Eberl M, Moser B. Monocytes and gammadelta T cells: close encounters in microbial infection. *Trends Immunol* (2009) 30:562–8. doi:10.1016/j.it.2009.09.001

9. Harly C, Guillaume Y, Nedellec S, Peigné CM, Mönkkönen H, Mönkkönen J, et al. Key implication of CD277/butyrophilin-3 (BTN3A) in cellular stress sensing by a major human gammadelta T-cell subset. *Blood* (2012) **120**:2269–79. doi:10.1182/blood-2012-05-430470
10. Vavassori S, Kumar A, Wan GS, Ramanjaneyulu GS, Cavallari M, El Daker S, et al. Butyrophilin 3A1 binds phosphorylated antigens and stimulates human gammadelta T cells. *Nat Immunol* (2013) **14**:908–16. doi:10.1038/ni.2665
11. Hayday AC. Gammadelta T cells and the lymphoid stress-surveillance response. *Immunity* (2009) **31**:184–96. doi:10.1016/j.immuni.2009.08.006
12. Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* (2010) **10**:467–78. doi:10.1038/nri2781
13. Vantourout P, Hayday A. Six-of-the-best: unique contributions of gammadelta T cells to immunology. *Nat Rev Immunol* (2013) **13**:88–100. doi:10.1038/nri3384
14. Caccamo N, Meraviglia S, Scarpa F, La Mendola C, Santini D, Bonanno CT, et al. Aminobisphosphonate-activated gammadelta T cells in immunotherapy of cancer: doubts no more. *Expert Opin Biol Ther* (2008) **8**:875–83. doi:10.1517/14712598.8.7.875
15. Gomes AQ, Martins DS, Silva-Santos B. Targeting gammadelta T lymphocytes for cancer immunotherapy: from novel mechanistic insight to clinical application. *Cancer Res* (2010) **70**:10024–7. doi:10.1158/0008-5472.CAN-10-3236
16. Brandes M, Willmann K, Lang AB, Nam KH, Jin C, Brenner MB, et al. Flexible migration program regulates gamma delta T-cell involvement in humoral immunity. *Blood* (2003) **102**:3693–701. doi:10.1182/blood-2003-04-1016
17. Cipriani B, Borsellino G, Poccia F, Placido R, Tramonti D, Bach S, et al. Activation of C-C β -chemokines in human peripheral blood gamma δ T cells by isopentenyl pyrophosphate and regulation by cytokines. *Blood* (2000) **95**:39–47.
18. Glatzel A, Wesch D, Schiemann F, Brandt E, Janssen O, Kabelitz D. Patterns of chemokine receptor expression on peripheral blood gamma delta T lymphocytes: strong expression of CCR5 is a selective feature of V delta 2/V gamma 9 gamma delta T cells. *J Immunol* (2002) **168**:4920–9. doi:10.4049/jimmunol.168.10.4920
19. Dieli F, Poccia F, Lipp M, Sireci G, Caccamo N, Di Sano C, et al. Differentiation of effector/memory Vdelta2 T cells and migratory routes in lymph nodes or inflammatory sites. *J Exp Med* (2003) **198**:391–7. doi:10.1084/jem.20030235
20. Brandes M, Willmann K, Moser B. Professional antigen-presentation function by human gammadelta T Cells. *Science* (2005) **309**:264–8. doi:10.1126/science.1110267
21. Bansal RR, Mackay CR, Moser B, Eberl M. IL-21 enhances the potential of human gammadelta T cells to provide B-cell help. *Eur J Immunol* (2012) **42**:110–9. doi:10.1002/eji.201142017
22. Brandes M, Willmann K, Bioley G, Lévy N, Eberl M, Luo M, et al. Cross-presenting human gammadelta T cells induce robust CD8+ α beta T cell responses. *Proc Natl Acad Sci U S A* (2009) **106**:2307–12. doi:10.1073/pnas.0810059106
23. Meuter S, Eberl M, Moser B. Prolonged antigen survival and cytosolic export in cross-presenting human gammadelta T cells. *Proc Natl Acad Sci U S A* (2010) **107**:8730–5. doi:10.1073/pnas.1002769107
24. Wu Y, Wu W, Wong WM, Ward E, Thrasher AJ, Goldblatt D, et al. Human gamma delta T cells: a lymphoid lineage cell capable of professional phagocytosis. *J Immunol* (2009) **183**:5622–9. doi:10.4049/jimmunol.0901772
25. Landmeier S, Altvater B, Pscherer S, Juergens H, Varnholt L, Hansmeier A, et al. Activated human gammadelta T cells as stimulators of specific CD8+ T-cell responses to subdominant Epstein Barr virus epitopes: potential for immunotherapy of cancer. *J Immunother* (2009) **32**:310–21. doi:10.1097/CJI.0b013e31819b7c30
26. Hu C, Qian L, Miao Y, Huang Q, Miao P, Wang P, et al. Antigen-presenting effects of effector memory Vgamma9Vdelta2 T cells in rheumatoid arthritis. *Cell Mol Immunol* (2012) **9**:245–54. doi:10.1038/cmi.2011.50
27. Altvater B, Pscherer S, Landmeier S, Kailayangiri S, Savoldo B, Juergens H, et al. Activated human gammadelta T cells induce peptide-specific CD8+ T-cell responses to tumor-associated self-antigens. *Cancer Immunol Immunother* (2012) **61**:385–96. doi:10.1007/s00262-011-1111-6
28. Himoudi N, Morgenstern DA, Yan M, Vernay B, Saraiva L, Wu Y, et al. Human gammadelta T lymphocytes are licensed for professional antigen presentation by interaction with opsonized target cells. *J Immunol* (2012) **188**:1708–16. doi:10.4049/jimmunol.1102654
29. Kobayashi H, Tanaka Y, Yagi J, Osaka Y, Nakazawa H, Uchiyama T, et al. Safety profile and anti-tumor effects of adoptive immunotherapy using gamma-delta T cells against advanced renal cell carcinoma: a pilot study. *Cancer Immunol Immunother* (2007) **56**:469–76. doi:10.1007/s00262-006-0199-6
30. Bouet-Toussaint F, Cabillic F, Toutirais O, Le Gallo M, Thomas de la Pintière C, Daniel P, et al. Vgamma9Vdelta2 T cell-mediated recognition of human solid tumors. Potential for immunotherapy of hepatocellular and colorectal carcinomas. *Cancer Immunol Immunother* (2008) **57**:531–9. doi:10.1007/s00262-007-0391-3
31. Burjanadzé M, Condomines M, Reme T, Quittet P, Latry P, Lugagne C, et al. In vitro expansion of gamma delta T cells with anti-myeloma cell activity by Phosphostim and IL-2 in patients with multiple myeloma. *Br J Haematol* (2007) **139**:206–16. doi:10.1111/j.1365-2141.2007.06754.x
32. Salot S, Laplace C, Saiagh S, Bercegeay S, Tenaud I, Cassidani A, et al. Large scale expansion of gamma 9 delta 2 T lymphocytes: innacell gamma delta cell therapy product. *J Immunol Methods* (2007) **326**:63–75. doi:10.1016/j.jim.2007.07.010
33. Bennouna J, Bompas E, Neidhardt EM, Rolland F, Philip I, Galéa C, et al. Phase-I study of Innacell gammadelta, an autologous cell-therapy product highly enriched in gamma9delta2 T lymphocytes, in combination with IL-2, in patients with metastatic renal cell carcinoma. *Cancer Immunol Immunother* (2008) **57**:1599–609. doi:10.1007/s00262-008-0491-8
34. Kondo M, Sakuta K, Noguchi A, Ariyoshi N, Sato K, Sato S, et al. Zoledronate facilitates large-scale ex vivo expansion of functional gammadelta T cells from cancer patients for use in adoptive immunotherapy. *Cytotherapy* (2008) **10**:842–56. doi:10.1080/14653240802419328
35. Kunzmann V, Kimmel B, Herrmann T, Einsele H, Wilhelm M. Inhibition of phosphoantigen-mediated gammadelta T-cell proliferation by CD4+ CD25+ FoxP3+ regulatory T cells. *Immunology* (2009) **126**:256–67. doi:10.1111/j.1365-2567.2008.02894.x
36. Cabillic F, Toutirais O, Lavoué V, de LaPintière CT, Daniel P, Rioux-Leclerc N, et al. Aminobisphosphonate-pretreated dendritic cells trigger successful Vgamma9Vdelta2 T cell amplification for immunotherapy in advanced cancer patients. *Cancer Immunol Immunother* (2010) **59**:1611–9. doi:10.1007/s00262-010-0887-0
37. Kobayashi H, Tanaka Y, Yagi J, Minato N, Tanabe K. Phase I/II study of adoptive transfer of gammadelta T cells in combination with zoledronic acid and IL-2 to patients with advanced renal cell carcinoma. *Cancer Immunol Immunother* (2011) **60**:1075–84. doi:10.1007/s00262-011-1021-7
38. Nicol AJ, Tokuyama H, Mattarollo SR, Hagi T, Suzuki K, Yokokawa K, et al. Clinical evaluation of autologous gamma delta T cell-based immunotherapy for metastatic solid tumours. *Br J Cancer* (2011) **105**:778–86. doi:10.1038/bjc.2011.293
39. Yamasaki A, Onishi H, Morisaki T, Katano M. Induction of cytotoxic T lymphocytes by CEA peptide-pulsed gammadelta T-cells isolated from patients with advanced cancer. *Anticancer Res* (2011) **31**:2419–24.
40. Sugie T, Murata-Hirai K, Iwasaki M, Morita CT, Li W, Okamura H, et al. Zoledronic acid-induced expansion of gammadelta T cells from early-stage breast cancer patients: effect of IL-18 on helper NK cells. *Cancer Immunol Immunother* (2013) **62**:677–87. doi:10.1007/s00262-012-1368-4
41. Caccamo N, Dieli F, Wesch D, Jomaa H, Eberl M. Sex-specific phenotypical and functional differences in peripheral human Vgamma9/Vdelta2 T cells. *J Leukoc Biol* (2006) **79**:663–6. doi:10.1189/jlb.1105640
42. Petrini I, Pacini S, Galimberti S, Taddei MR, Romanini A, Petrini M. Impaired function of gamma-delta lymphocytes in melanoma patients. *Eur J Clin Invest* (2011) **41**:1186–94. doi:10.1111/j.1365-2362.2011.02524.x
43. Roelofs AJ, Jauhainen M, Monkkinen H, Rogers MJ, Monkkinen J, Thompson K. Peripheral blood monocytes are responsible for gammadelta T cell activation induced by zoledronic acid through accumulation of IPP/DMAPP. *Br J Haematol* (2009) **144**:245–50. doi:10.1111/j.1365-2141.2008.07435.x
44. Liao W, Lin JX, Leonard WJ. Interleukin-2 at the crossroads of effector responses, tolerance, and immunotherapy. *Immunity* (2013) **38**:13–25. doi:10.1016/j.immuni.2013.01.004
45. Rochman Y, Spolski R, Leonard WJ. New insights into the regulation of T cells by gamma(delta) family cytokines. *Nat Rev Immunol* (2009) **9**:480–90. doi:10.1038/nri2580

46. Steel JC, Waldmann TA, Morris JC. Interleukin-15 biology and its therapeutic implications in cancer. *Trends Pharmacol Sci* (2012) **33**:35–41. doi:10.1016/j.tips.2011.09.004
47. Lu J, Giuntoli RL, Omiya R, Kobayashi H, Kennedy R, Celis E. Interleukin 15 promotes antigen-independent in vitro expansion and long-term survival of antitumor cytotoxic T lymphocytes. *Clin Cancer Res* (2002) **8**:3877–84.
48. Montes M, Rufer N, Appay V, Reynard S, Pittet MJ, Speiser DE, et al. Optimum in vitro expansion of human antigen-specific CD8 T cells for adoptive transfer therapy. *Clin Exp Immunol* (2005) **142**:292–302. doi:10.1111/j.1365-2249.2005.02914.x
49. Li Y, Liu S, Hernandez J, Vence L, Hwu P, Radvanyi L. MART-1-specific melanoma tumor-infiltrating lymphocytes maintaining CD28 expression have improved survival and expansion capability following antigenic restimulation in vitro. *J Immunol* (2010) **184**:452–65. doi:10.4049/jimmunol.0901101
50. Dowlatshahi M, Huang V, Gehad AE, Jiang Y, Calarese A, Teague JE, et al. Tumor-specific T cells in human Merkel cell carcinomas: a possible role for Tregs and T-cell exhaustion in reducing T-cell responses. *J Invest Dermatol* (2013) **133**:1879–89. doi:10.1038/jid.2013.75
51. French JD, Roark CL, Born WK, O'Brien RL. $\{\gamma\delta\}$ T cell homeostasis is established in competition with $\{\alpha\beta\}$ T cells and NK cells. *Proc Natl Acad Sci U S A* (2005) **102**:14741–6. doi:10.1073/pnas.0507520102
52. Baccala R, Witherden D, Gonzalez-Quintal R, Dummer W, Surh CD, Havran WL, et al. Gamma delta T cell homeostasis is controlled by IL-7 and IL-15 together with subset-specific factors. *J Immunol* (2005) **174**:4606–12. doi:10.4049/jimmunol.174.8.4606
53. Izumi T, Kondo M, Takahashi T, Fujieda N, Kondo A, Tamura N, et al. Ex vivo characterization of gammadelta T-cell repertoire in patients after adoptive transfer of Vgamma9Vdelta2 T cells expressing the interleukin-2 receptor beta-chain and the common gamma-chain. *Cytotherapy* (2013) **15**:481–91. doi:10.1016/j.jcyt.2012.12.004
54. Sallusto F, Lenig D, Förster R, Lipp M, Lanzavecchia A. Two subsets of memory T lymphocytes with distinct homing potentials and effector functions. *Nature* (1999) **401**:708–12. doi:10.1038/44385
55. van Baarle D, Tsegaye A, Miedema F, Akbar A. Significance of senescence for virus-specific memory T cell responses: rapid ageing during chronic stimulation of the immune system. *Immunol Lett* (2005) **97**:19–29. doi:10.1016/j.imlet.2004.10.003
56. Voehringer D, Koschella M, Pircher H. Lack of proliferative capacity of human effector and memory T cells expressing killer cell lectin like receptor G1 (KLRG1). *Blood* (2002) **100**:3698–702. doi:10.1182/blood-2002-02-0657
57. Eberl M, Engel R, Aberle S, Fisch P, Jomaa H, Pircher H. Human Vgamma9/Vdelta2 effector memory T cells express the killer cell lectin-like receptor G1 (KLRG1). *J Leukoc Biol* (2005) **77**:67–70. doi:10.1189/jlb.0204096
58. Vermijlen D, Ellis P, Langford C, Klein A, Engel R, Willmann K, et al. Distinct cytokine-driven responses of activated blood gammadelta T cells: insights into unconventional T cell pleiotropy. *J Immunol* (2007) **178**:4304–14. doi:10.4049/jimmunol.178.7.4304
59. Dustin ML, Tseng SY, Varma R, Campi G. T cell-dendritic cell immunological synapses. *Curr Opin Immunol* (2006) **18**:512–6. doi:10.1016/j.coi.2006.05.017
60. Salomon B, Bluestone JA. Complexities of CD28/B7: CTLA-4 costimulatory pathways in autoimmunity and transplantation. *Annu Rev Immunol* (2001) **19**:225–52. doi:10.1146/annurev.immunol.19.1.225
61. Zou W. Immunosuppressive networks in the tumour environment and their therapeutic relevance. *Nat Rev Cancer* (2005) **5**:263–74. doi:10.1038/nrc1586
62. Abe Y, Muto M, Nieda M, Nakagawa Y, Nicol A, Kaneko T, et al. Clinical and immunological evaluation of zoledronate-activated Vgamma9gammadelta T-cell-based immunotherapy for patients with multiple myeloma. *Exp Hematol* (2009) **37**:956–68. doi:10.1016/j.exphem.2009.04.008
63. Nakajima J, Murakawa T, Fukami T, Goto S, Kaneko T, Yoshida Y, et al. A phase I study of adoptive immunotherapy for recurrent non-small-cell lung cancer patients with autologous gammadelta T cells. *Eur J Cardiothorac Surg* (2010) **37**:1191–7. doi:10.1016/j.ejcts.2009.11.051
64. Bennouna J, Levy V, Sicard H, Senellart H, Audrain M, Huret S, et al. Phase I study of bromohydrin pyrophosphate (BrHPP, IPH 1101), a Vgamma9Vdelta2 T lymphocyte agonist in patients with solid tumors. *Cancer Immunol Immunother* (2010) **59**:1521–30. doi:10.1007/s00262-010-0879-0
65. Kobayashi H, Tanaka Y, Shimmura H, Minato N, Tanabe K. Complete remission of lung metastasis following adoptive immunotherapy using activated autologous gammadelta T-cells in a patient with renal cell carcinoma. *Anticancer Res* (2010) **30**:575–9.
66. Noguchi A, Kaneko T, Kamigaki T, Fujimoto K, Ozawa M, Saito M, et al. Zoledronate-activated Vgamma9gammadelta T cell-based immunotherapy is feasible and restores the impairment of gammadelta T cells in patients with solid tumors. *Cytotherapy* (2011) **13**:92–7. doi:10.3109/14653249.2010.515581
67. Sakamoto M, Nakajima J, Murakawa T, Fukami T, Yoshida Y, Murayama T, et al. Adoptive immunotherapy for advanced non-small cell lung cancer using zoledronate-expanded gammadelta T cells: a phase I clinical study. *J Immunother* (2011) **34**:202–11. doi:10.1097/CJI.0b013e318207ecfb
68. Couzin-Frankel J. Breakthrough of the year 2013. Cancer immunotherapy. *Science* (2013) **342**:1432–3. doi:10.1126/science.342.6165.1432

Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

Received: 04 June 2014; paper pending published: 30 June 2014; accepted: 06 July 2014; published online: 23 July 2014.

Citation: Khan MWA, Curbishley SM, Chen H-C, Thomas AD, Pircher H, Mavilio D, Steven NM, Eberl M and Moser B (2014) Expanded human blood-derived $\gamma\delta$ T cells display potent antigen-presentation functions. *Front. Immunol.* **5**:344. doi:10.3389/fimmu.2014.00344

This article was submitted to *Immunotherapies and Vaccines*, a section of the journal *Frontiers in Immunology*.

Copyright © 2014 Khan, Curbishley, Chen, Thomas, Pircher, Mavilio, Steven, Eberl and Moser. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.