



Complete Genome Sequences of Mannanase-Producing *Bacillus* and *Niallia* Strains Isolated from the Intestine of the Black Tiger Shrimp (*Penaeus monodon*)

Witida Sathitkowitzai,^a  Thidathip Wongsurawat,^b Piroon Jenjaroenpun,^b Pacharaporn Angthong,^a Jiratchaya Nuanpirom,^c Ponsit Sathapondecha,^c  Wanilada Rungrasamee^a

^aMicroarray Research Team, National Center for Genetic Engineering and Biotechnology, Khlong Nueng, Khlong Luang, Thailand

^bDivision of Bioinformatics and Data Management for Research, Research Group and Research Network Division, Research Department, Faculty of Medicine, Siriraj Hospital, Mahidol University, Bangkok, Thailand

^cCenter for Genomics and Bioinformatics Research, Division of Biological Science, Faculty of Science, Prince of Songkla University, Hat Yai, Songkhla, Thailand

ABSTRACT Here, we report the complete genome sequences of mannanase-producing bacteria, namely, *Niallia* sp. strain Man26 and *Bacillus subtilis* strain Man122, isolated from the intestine of *Penaeus monodon*, the black tiger shrimp. Mannanases are used in various industries, such as food, animal feed, and biorefinery, to hydrolyze mannan to oligomers and mannose.

Mannanase is a hydrolytic enzyme for the degradation of mannan and hetero-mannan, a hemicellulose component of the plant cell wall, to generate manno-oligosaccharide (MOS), which is an important prebiotic for animals (1). Many bacteria are known to be able to produce mannanase, including Gram-positive bacteria such as *Bacillus* (2–7).

In this study, mannanase-producing *Bacillus* and *Niallia* strains were isolated from the intestines of black tiger shrimp (*Penaeus monodon*) reared at a BIOTEC shrimp facility (Thailand). Shrimp intestine was minced into small pieces and added to 200 μ L of 1 \times phosphate buffer (pH 7.4). The culture broth was serially diluted and plated on selective M9 medium containing 1% MOS from copra meal. Bacterial colonies were screened for their ability to produce mannanase using the Congo red staining method on locust bean gum agar plates (8, 9).

Bacteria were cultured in Luria-Bertani broth at 30°C for 48 h with shaking at 250 rpm. For each isolate, 3 mL was collected for DNA extraction using the ZymoBIOMICS DNA mini-prep kit (Zymo Research, USA) according to the manufacturer's protocol. The bacterial genome sequencing was performed using both Illumina and Nanopore platforms. For short reads, paired-end 2 \times 150-bp sequencing libraries were constructed using the NEBNext Ultra II DNA library preparation kit and sequenced with an Illumina NovaSeq sequencer. The sequencing adapters were trimmed using Fastp v0.19.5, and the quality of cleaned reads was determined using FastQC v0.11.9 (10). For long reads, transposase-based DNA library preparation was applied using the rapid barcoding kit (RBK004; Oxford Nanopore Technologies [ONT]). The DNA library was loaded on a MinION flow cell v106 (R10.3) and sequenced with a MinION Mk1C sequencer for 48 h. The raw signals were obtained, base called, and demultiplexed using Guppy v5.0.16 with the super accurate model (`-c dna_r10.3_450bps_sup.cfg -r -trim_barcodes -barcode_kits SQK-RBK004`), followed by adapter trimming using Porechop v0.2.4 software (11). The quality of ONT raw reads was determined with NanoPlot v1.28.1 (12). The raw read filtering was based on a mean quality score of 8 using NanoFilt v2.5.0, and only reads with lengths of 1,000 bases were stored for the *de novo* assembly. The genomes were constructed by hybrid assembly together with correction, circularization, and rotation using Unicycler v0.4.4 (13). Here, we report the

Editor Julie C. Dunning Hotopp, University of Maryland School of Medicine

Copyright © 2022 Sathitkowitzai et al. This is an open-access article distributed under the terms of the [Creative Commons Attribution 4.0 International license](https://creativecommons.org/licenses/by/4.0/).

Address correspondence to Wanilada Rungrasamee, wailada.run@biotec.or.th.

The authors declare no conflict of interest.

Received 11 February 2022

Accepted 8 May 2022

Published 26 May 2022

TABLE 1 Relevant statistics for sequencing and assembly of two mannanase-producing bacteria isolated from intestine of the black tiger shrimp (*P. monodon*) in Thailand

Parameter	Data for strain:	
	Man26	Man122
GenBank accession no.	CP095743, CP095744, CP095745, CP095746, CP095747, CP095748	CP091872
No. of short reads	6,164,481	5,731,944
No. of long reads	182,103	62,327
No. of assembled contigs	6	1
N_{50} (bp)	3,887,076	4,105,902
Genome size (bp)	5,714,135	4,105,902
GC content (%)	38.1	43.8
No. of coding sequences	5,705	4,070
Coding proportion (%)	83.6	87.8
No. of rRNAs	29	30
No. of tRNAs	130	87

complete genome sequences of *Niallia* sp. strain Man26 (GC content, 38.1%) and *Bacillus* sp. strain Man122 (GC content, 43.8%). The genomes were annotated using the NCBI Prokaryotic Genome Annotation Pipeline (PGAP) v4.11 (14). The assembly quality assessment by QUAST v5.0.2 (15) and associated statistics are reported in Table 1. Default parameters were used except where otherwise noted.

Data availability. All genome sequences, including chromosomes and plasmids, were deposited in the NCBI database under BioProject accession number PRJNA799131, including raw reads from Illumina and ONT sequencing under SRA accession numbers SRR17687495 and SRR17687496 for Man26 and SRR17701557 and SRR17701558 for Man122, respectively. In addition, the assembled contigs were deposited in GenBank under accession numbers CP095743, CP095744, CP095745, CP095746, CP095747, and CP095748 for Man26 and CP091872 for Man122.

ACKNOWLEDGMENTS

This work received funding support from a midcareer fellowship grant at the National Center for Genetic Engineering and Biotechnology (grant P1652214) and NSRF via the Program Management Unit for Human Resources and Institutional Development, Research, and Innovation (grant B05F640108).

REFERENCES

- Zhang R, Li X-Y, Cen X-L, Gao Q-H, Zhang M, Li K-Y, Wu Q, Mu Y-L, Tang X-H, Zhou J-P, Huang Z-X. 2021. Enzymatic preparation of manno-oligosaccharides from locust bean gum and palm kernel cake, and investigations into its prebiotic activity. *Electron J Biotechnol* 49:64–71. <https://doi.org/10.1016/j.ejbt.2020.11.001>.
- Liu H-X, Gong J-S, Li H, Lu Z-M, Li H, Qian J-Y, Xu Z, Shi J-S. 2015. Biochemical characterization and cloning of an endo-1,4- β -mannanase from *Bacillus subtilis* YH12 with unusually broad substrate profile. *Process Biochem* 50:712–721. <https://doi.org/10.1016/j.procbio.2015.02.011>.
- Seesom W, Thongket P, Yamamoto T, Takenaka S, Sakamoto T, Sukhumsirichart W. 2017. Purification, characterization, and overexpression of an endo-1,4- β -mannanase from thermotolerant *Bacillus* sp. SWU60. *World J Microbiol Biotechnol* 33:53. <https://doi.org/10.1007/s11274-017-2224-7>.
- Zhang W, Liu Z, Zhou S, Mou H, Zhang R. 2019. Cloning and expression of a β -mannanase gene from *Bacillus* sp. MK-2 and its directed evolution by random mutagenesis. *Enzyme Microb Technol* 124:70–78. <https://doi.org/10.1016/j.enzmictec.2019.02.003>.
- Singh S, Singh G, Khatri M, Kaur A, Arya S. 2019. Thermo and alkali stable β -mannanase: characterization and application for removal of food (mannans based) stain. *Int J Biol Macromol* 134:536–546. <https://doi.org/10.1016/j.ijbiomac.2019.05.067>.
- Titapoka S, Keawsompong S, Haltrich D, Nitisinprasert S. 2008. Selection and characterization of mannanase-producing bacteria useful for the formation of prebiotic manno-oligosaccharides from copra meal. *World J Microbiol Biotechnol* 24:1425–1433. <https://doi.org/10.1007/s11274-007-9627-9>.
- Tuntrakool P, Keawsompong S. 2018. Kinetic properties analysis of beta-mannanase from *Klebsiella oxytoca* KUB-CW2-3 expressed in *Escherichia coli*. *Protein Expr Purif* 146:23–26. <https://doi.org/10.1016/j.pep.2018.01.009>.
- Gessesse A, Gashe BA. 1997. Production of alkaline xylanase by an alkaliphilic *Bacillus* sp. isolated from an alkalinesoda lake. *J Appl Microbiol* 83:402–406. <https://doi.org/10.1046/j.1365-2672.1997.00242.x>.
- Ghazala I, Haddar A, Romdhane M, Ellouz-Chaanouni S. 2016. Screening and molecular identification of new microbial strains for production of enzymes of biotechnological interest. *Braz Arch Biol Technol* <https://doi.org/10.1590/1678-4324-2016150152>.
- Andrews S. 2010. FastQC: a quality control tool for high throughput sequence data. <http://www.bioinformatics.babraham.ac.uk/projects/fastqc>.
- Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Completing bacterial genome assemblies with multiplex MinION sequencing. *Microb Genom* 3:e000132. <https://doi.org/10.1099/mgen.0.000132>.
- De Coster W, D'Hert S, Schultz DT, Cruts M, Van Broeckhoven C. 2018. NanoPack: visualizing and processing long-read sequencing data. *Bioinformatics* 34:2666–2669. <https://doi.org/10.1093/bioinformatics/bty149>.

13. Wick RR, Judd LM, Gorrie CL, Holt KE. 2017. Unicycler: resolving bacterial genome assemblies from short and long sequencing reads. *PLoS Comput Biol* 13:e1005595. <https://doi.org/10.1371/journal.pcbi.1005595>.
14. Tatusova T, DiCuccio M, Badretdin A, Chetvernin V, Nawrocki EP, Zaslavsky L, Lomsadze A, Pruitt KD, Borodovsky M, Ostell J. 2016. NCBI Prokaryotic Genome Annotation Pipeline. *Nucleic Acids Res* 44: 6614–6624. <https://doi.org/10.1093/nar/gkw569>.
15. Gurevich A, Saveliev V, Vyahhi N, Tesler G. 2013. QUAST: quality assessment tool for genome assemblies. *Bioinformatics* 29:1072–1075. <https://doi.org/10.1093/bioinformatics/btt086>.