

Review Article

Channeling Vision: Ca_v1.4—A Critical Link in Retinal Signal Transmission

D. M. Waldner ¹, N. T. Bech-Hansen,² and W. K. Stell³

¹Department of Neuroscience, Cumming School of Medicine, University of Calgary, Calgary, AB, Canada

²Department of Medical Genetics and Department of Surgery, Alberta Children's Hospital Research Institute, and Hotchkiss Brain Institute, Cumming School of Medicine, University of Calgary, Calgary, AB, Canada

³Department of Cell Biology and Anatomy and Department of Surgery, Hotchkiss Brain Institute, and Alberta Children's Hospital Research Institute, Cumming School of Medicine, University of Calgary, Calgary, AB, Canada

Correspondence should be addressed to D. M. Waldner; derek.waldner@ucalgary.ca

Received 2 January 2018; Accepted 15 February 2018; Published 9 May 2018

Academic Editor: Christina Zeitz

Copyright © 2018 D. M. Waldner et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Voltage-gated calcium channels (VGCC) are key to many biological functions. Entry of Ca²⁺ into cells is essential for initiating or modulating important processes such as secretion, cell motility, and gene transcription. In the retina and other neural tissues, one of the major roles of Ca²⁺-entry is to stimulate or regulate exocytosis of synaptic vesicles, without which synaptic transmission is impaired. This review will address the special properties of one L-type VGCC, Ca_v1.4, with particular emphasis on its role in transmission of visual signals from rod and cone photoreceptors (hereafter called “photoreceptors,” to the exclusion of intrinsically photoreceptive retinal ganglion cells) to the second-order retinal neurons, and the pathological effects of mutations in the *CACNA1F* gene which codes for the pore-forming α_{1F} subunit of Ca_v1.4.

1. History

The original discovery of Ca_v1.4 is intimately associated with the phenotypic and genotypic characterization of a family of inherited retinal disorders, congenital stationary night blindness (CSNB), a subset of which (CSNB2A) is caused by mutations in *CACNA1F*. CSNB first appeared in historical records in the 17th century associated with a Frenchman named Jean Nougaret for whom an extensive genealogical pedigree was available, implicating the disease as an autosomal dominant inherited condition [1]. Additional cases of hereditary night blindness with recessive and X-linked inheritance patterns were reported subsequently [2–5]. Widespread clinical use and understanding of electroretinography (ERG) in the mid-20th century allowed clinicians to begin separating CSNB into distinct clinical entities. Schubert and Bornschein first reported a subtype of CSNB in which the scotopic, negative-going a-wave was normal, while the positive-going b-wave was significantly attenuated [6]. CSNB with this electronegative ERG response (i.e., with a b-wave significantly smaller in amplitude than the a-wave) would be known thereafter

as “Schubert-Bornschein-type CSNB,” to distinguish it from “Riggs-type CSNB,” in which both a- and b-waves exhibit decreased amplitude [7]. Phenotypic heterogeneity of individuals with Schubert-Bornschein-type CSNB led Miyake to separate the disorder into “complete” (with *total* loss of rod function) and “incomplete” (with *partial* rod function) subtypes in 1986 [8]. This insight would subsequently be validated genetically, following comprehensive linkage analysis, with distinct gene loci identified for the causative mutations in “complete” and “incomplete” X-linked CSNB [9]. Large-scale DNA sequencing of Xp11.23, the genomic region implicated in X-linked incomplete CSNB (CSNB2A), led to the simultaneous identification by two research groups of mutations in the recently discovered gene “*CACNA1F*” in individuals with the condition [10, 11]. *CACNA1F* had been annotated in a publication only one-year prior, where segments of sequence were recognized as having high homology to known VGCC pore-forming α_1 subunits [12]. *CACNA1F* is now known to code for α_{1F} , the primary subunit of the voltage-gated L-type calcium channel Ca_v1.4.

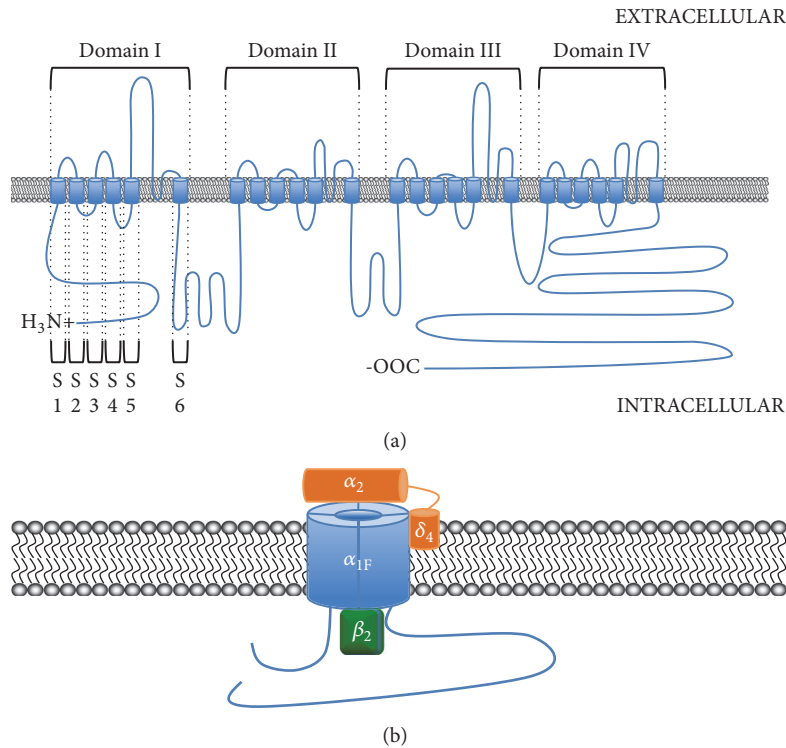


FIGURE 1: (a) Protein topology of $\text{Ca}_v1.4 \alpha_{1F}$ subunit. (b) Schematic of $\text{Ca}_v1.4$, including accessory subunits β_2 and $\alpha_2\delta_4$.

2. Molecular Properties, Distribution, and Function

2.1. CACNA1F , α_{1F} , and $\text{Ca}_v1.4$. *CACNA1F* is a 48 exon gene encoding α_{1F} , a 1977 amino-acid protein with the distinct structural characteristics typical of VGCC α_1 subunits [11, 13, 14]. These include intracellular N- and C-termini flanking four homologous domains, each of which consists of six transmembrane alpha helix segments (Figure 1(a); for a review of voltage-gated calcium channels, see Zamponi et al., 2015 [15]). α_{1F} is one of ten human α_1 VGCC subunits, which are the primary determinants of Ca_v channel characteristics [16]. In addition to the pore-forming α_1 subunits, VGCCs are also composed of several other accessory subunits that may differ in subtype, altering channel-trafficking, kinetics, and other properties. These include β , $\alpha_2\delta$, and γ subunits, which, together with α_1 , comprise VGCCs in a 1:1:1:1 ratio [16] (Figure 1(b)). By conventional nomenclature VGCCs are named for their α_1 subunit; all VGCCs that have α_{1F} as their pore-forming subunit are classified as $\text{Ca}_v1.4$ channels, though the accessory subunits may differ [16]. *In vivo*, $\text{Ca}_v1.4$ channels have been shown to primarily include the subunit subtypes β_2 and $\alpha_2\delta_4$ [17–21]. Gamma (γ) subunits in $\text{Ca}_v1.4$, if present, have not yet been identified.

2.2. $\text{Ca}_v1.4$ Distribution and Expression. $\text{Ca}_v1.4$ is primarily expressed in the retina, though unique splice variants of *CACNA1F* (with significantly different functions) are also expressed in B- and T-lymphocytes and possibly other immune cells [10, 22]. Broader tissue expression of $\text{Ca}_v1.4$

has been suggested from analysis of expressed sequence tags (EST), but immunohistochemical experiments have largely failed to confirm these suggestions [23]. The apparent presence of the channel in these unconfirmed locations might be due to the presence of circulating immune cells in these tissues when they were sampled for EST analysis [24]. Furthermore, even in the retina some localizations of $\text{Ca}_v1.4$ remain controversial. Early reports identified α_{1F} -like immunoreactivity and *Cacnalf* mRNA in the inner, as well as the outer, plexiform layer [13, 25, 26], whereas more recent immunohistochemical evidence suggests exclusive expression in photoreceptor terminals of the OPL [27–30]. Recent electrophysiological recordings of bipolar cell axon terminals of the IPL in *Cacnalf*-deficient mouse lines suggest that $\text{Ca}_v1.4$ contributes slightly to calcium influx in those cells, though this may be due to alterations in retinal circuitry or bipolar cell homeostasis resulting from dysfunctional photoreceptor synaptic transmission [31]. Thus, the localization of $\text{Ca}_v1.4$ beyond the photoreceptors of the retina remains to be resolved definitively, due in part to difficulties in creating subtype-specific antibodies or possibly very low-level protein expression in bipolar cell terminals.

2.3. $\text{Ca}_v1.4$ Function: The Ribbon Synapse and Phototransduction. The retina-specific expression of $\text{Ca}_v1.4$ and the visual dysfunction resulting from its disruption (CSNB2A) allude to the crucial role of this particular channel in visual signal transduction. At photoreceptor terminals, $\text{Ca}_v1.4$ channels are an integral component of the characteristic “ribbon synapses,” which are responsible for neurotransmission at

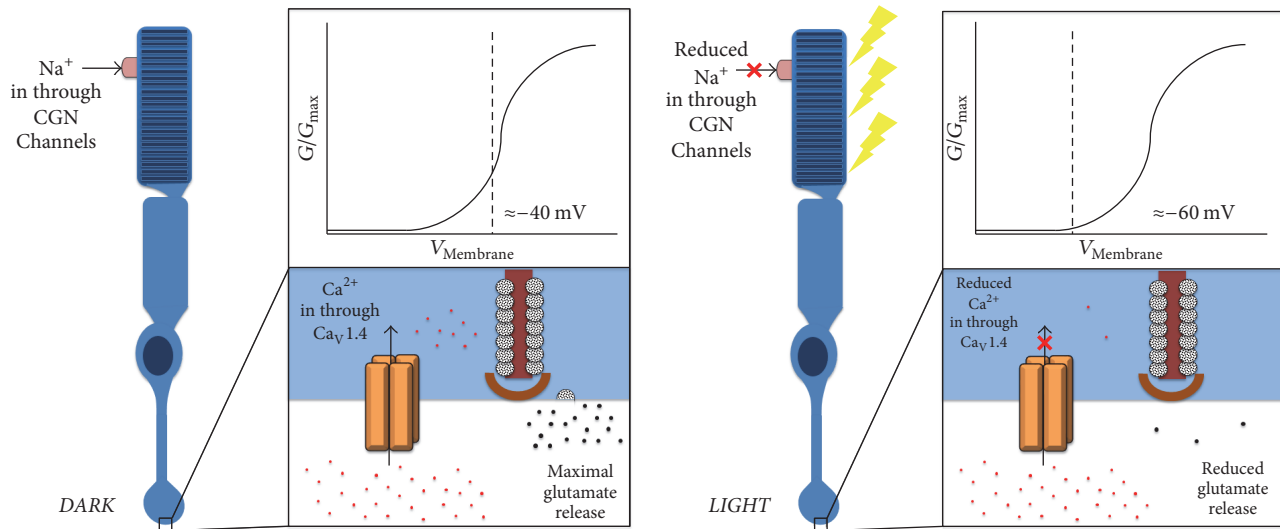


FIGURE 2: Effect of phototransduction on photoreceptor membrane potential, calcium influx through $Ca_V1.4$ and glutamate release at the ribbon synapse. In complete darkness, sodium ion influx through open CGN channels in photoreceptor outer segments maintains the membrane potential at ~ -40 mV, where channel conductance (G/G_{max}) and glutamate release are at their physiological maxima. Phototransduction in response to increased illumination causes graded decreases in sodium ion influx through CGN channels, a hyperpolarization of the photoreceptor membrane, a decrease in calcium ion influx into the presynaptic terminal and resultant decreased glutamate release.

photoreceptor and bipolar cell synapses. Synaptic ribbons are electron-dense, specialized protein complexes that recruit large numbers of glutamate-filled vesicles near the synaptic terminal, allowing for sustained vesicular release in response to sustained calcium influx [32, 33]. Unlike conventional neurons, rod and cone photoreceptors exhibit tonic glutamate release with maximal exocytosis in complete darkness and a graded decrease with increasing illumination. The phototransduction cascade is such that absorption of a photon within a photoreceptor outer segment causes a relative hyperpolarization (decrease in sustained depolarization) of the photoreceptor cell membrane, thereby decreasing the activation of the voltage-sensitive $Ca_V1.4$ channels and lessening calcium influx within the presynaptic terminal (Figure 2; for a review of phototransduction, see Yau, 1994 [34]). The proper assembly and function of the ribbon synapse involves a multitude of proteins and protein-protein interactions, which, if disrupted, can cause a loss of visual function (for a review of protein constituents and interactions of the photoreceptor ribbon synapse, see Mercer and Thoreson, 2011 [35]). The unique coupling of phototransduction and the ribbon synapse is responsible for the extraordinary sensitivity of the visual system and requires the unique properties of $Ca_V1.4$ for optimal function.

2.4. Electrophysiology of Photoreceptors and Heterologous $Ca_V1.4$ Expression Systems. Before $Ca_V1.4$ had been discovered and established as the functional calcium channel of photoreceptor terminals, electrophysiological recordings in nonmammalian photoreceptor inner segments (used for their comparatively large size) had revealed calcium currents with distinctive dynamics (reviewed in Doering et al.,

2007 [24]). I_{Ca} was shown to activate rapidly above roughly -45 to -40 mV, show sensitivity to typical L-type channel agonists and antagonists, and exhibit comparatively little voltage-dependent inactivation and no calcium-dependent inactivation (CDI) under typical recording conditions (although slight CDI was observable with reduced EGTA calcium buffering) [41, 42]. Many neuromodulators were shown to influence this photoreceptor calcium current, via mechanisms that remain largely unresolved today (for a review of these modulatory compounds and their effects, see Krizaj and Copenhagen 2002 [43]).

Following the identification of the *CACNA1F* gene, electrophysiological recordings in heterologous systems expressing $Ca_V1.4$ verified it as the channel mainly responsible for the unique calcium current observed in these earlier studies. The biophysical properties of $Ca_V1.4$ in these expression systems have been extensively reviewed [24] and so will be mentioned only briefly here. α_{1F} transiently expressed in human embryonic kidney cells with accessory subunits $\beta_{3/2a}$ and $\alpha_2\delta_1$ form a $Ca_V1.4$ channel that (1) is activated near -40 mV, the resting membrane potential of photoreceptors [23, 44, 45]; (2) exhibits the smallest unitary conductance of all known VGCCs, and an exceptionally low open probability [46]; (3) is inactivated as a function of voltage (voltage-dependent inactivation, recorded with Ba^{2+} as charge-carrier) more slowly than $Ca_V1.2$ and $Ca_V1.3$ [23, 44, 45]; and (4) exhibits almost no calcium-dependent inactivation [23, 44, 45, 47]. These distinctive properties are all essential to maintain the constant calcium influx without inactivation that is necessary for sustained glutamate release from rods and cones.

3. The Complexity of the Photoreceptor Calcium Current and Ca_v1.4 In Vivo

While heterologous systems are valuable for determining basic kinetic properties of Ca_v1.4, none of them has yet captured fully the complexities of the photoreceptor calcium current. *In vivo*, retinal photoreceptors express simultaneously several splice isoforms of Ca_v1.4 with different kinetics [48, 49] and associate with unique modulatory subunits (discussed below: *Ca_v1.4 Subunit Composition*) [17]. Additional factors, including physiological temperature (37°C) [50], calmodulin (CaM) coexpression [51–53], and calcium-binding protein 4 (CaBP4) coexpression [54, 55], also significantly modulate biophysical parameters of the channel. Thus, descriptions of Ca_v1.4 in all heterologous expression systems to date likely differ significantly from those of the channel as it exists *in vivo*.

3.1. CACNA1F Splice Isoforms. Over 20 splice isoforms of CACNA1F have been identified, several of which have significantly disparate dynamics [48, 49]. Most remarkably, several splice variants exist in which the C-terminus is truncated or otherwise disrupted. Tan et al. identified a splice variant incorporating an alternate 43rd exon, containing a stop codon that truncates ~1/2 of the C-terminus and represents 13.6% of the total CACNA1F transcript identified in their screening experiment of a human retina cDNA library [49]. Haeseleer et al. additionally identified two transcripts in which the C-terminal 47th exon is absent [48]. Investigations of the biophysical properties of the Ca_v1.4 isoforms encoded by these transcripts revealed significant differences from the wild-type, including a hyperpolarizing shift in activation voltage (compared to the full-length channel), and robust calcium-dependent inactivation. These data support models of ICDI/CTM regulation of Ca_v1.4 (discussed below: *C-Terminal Regulation and Calmodulin*) and suggest that coexpression of alternative splice variants results in a complex calcium current in photoreceptor terminals *in vivo*.

In extraretinal locations, unique splice variants of CACNA1F have also been identified in T-lymphocytes, which substantially alter the topology and function of the Ca_v1.4 channel. One of these in particular lacks a significant portion of the C-terminus and domain IV (including this domain's voltage-sensing segment) making this isoform unlikely to function as a voltage-gated channel. These channels have therefore been proposed to function via alternative gating mechanisms, likely via direct intracellular signaling from T-cell receptors [22]. Investigations in *Cacna1f* knockout mice have shown that Ca_v1.4 is critical for the survival, proliferation, and signaling of CD4⁺ and CD8⁺ T-cells, suggesting an essential role for these unique splice isoforms in normal functioning of the immune system [56].

3.2. Ca_v1.4 Subunit Composition. As previously mentioned, α_{1F} associates *in vivo* with β₂ and α₂δ₄ subunits, which regulate and modulate the calcium conductance of Ca_v1.4 [17–21]. The predominant β₂ splice variant in retinal Ca_v1.4 complexes, β_{2X13}, is unique in that it includes the N-terminal palmitoylation sites found in β_{2a}, but an alternate exon 7

(7B instead of 7A, found in β_{2a}) results in a shorter HOOK domain, associated with modulating VGCC inactivation kinetics [57]. This variant has been shown to increase voltage-dependent inactivation (VDI) of the Ca_v1.4 channel, in contrast with channels including the β_{2a} subunit, although the physiological significance of this effect is unclear [17]. Ca_v1.4 channels that include the α₂δ₄ subunit have been shown to exhibit more positive voltage-dependent activation and a lack of calcium-dependent facilitation of currents, compared to channels including the α₂δ₁ subunit [17]. Subunit complex α₂δ₄ has recently been shown to regulate the trafficking of α_{1F} subunits to the presynaptic membrane, thus regulating channel and therefore calcium current density in the synaptic terminal. Further, α₂δ₄ was shown to be critical for the formation of synapses between rod (but not cone) photoreceptors and the signal relaying ON-bipolar cells of the inner retina (see *Physiological and Morphological Observations in CSNB2A Model Retinas*) [58]. These data suggest, as expected, that the auxiliary subunit composition of Ca_v1.4 is important in shaping its unique biophysical properties.

3.3. C-Terminal Regulation and Calmodulin. The absence of calcium-dependent inactivation (CDI) observed in early characterizations of Ca_v1.4 suggested that the channel differed from other L-type VGCCs in C-terminal regulation [23, 44, 45]. CDI in Ca_v1.2 channels has been shown to be a function of the proximal C-terminal EF-hand, Pre-IQ and IQ motifs, which are necessary for interaction with the calcium-binding protein, calmodulin (CaM). In this model, the Pre-IQ and/or IQ domains are constitutively associated with apoCaM, which undergoes a conformational change upon binding calcium ions when they enter the channel pore. This conformational rearrangement of CaM is relayed to the channel core via the EF-hand motif and then inactivates the VGCC, possibly through interaction with the I-II intracellular linker [59–63]. The lack of CDI in Ca_v1.4 channel was initially confounding, as Ca_v1.4 exhibits high sequence homology with other Ca_v1 channels in these proximal C-terminal motifs [64–66].

In 2006, two groups independently showed that the relative dearth of CDI in Ca_v1.4, in heterologous expression systems, is a function of the distal C-terminus which functions as an autoinhibitory domain. The distal C-terminus was shown via fluorescence resonance energy transfer (FRET) experiments to interact directly with regions of the proximal C-terminus that are necessary for their CDI-mediating interaction with CaM. Singh et al. showed that this interaction also caused a positive shift in the voltage-dependence of activation and thus named the distal C-terminal region the “C-terminal modulator” (CTM), in contrast to the “inhibitor of calcium-dependent inactivation” (ICDI) term introduced by the concurrent discoverers, Wahl-Schott et al. [47, 65].

Whether the ICDI/CTM prevented CDI by competing with CaM binding directly, or by allosterically inhibiting transduction of CaM-mediated conformational changes in the inactivation machinery was not initially clear. Subsequent research has shown that the ICDI/CTM does indeed compete with CaM for occupancy within the proximal C-terminal

domains, but incompletely, such that physiological changes in CaM levels within the cell are functionally relevant [52]. The preassociation of apoCaM with the proximal C-terminal domains of Ca_v1.3 and chimeric Ca_v1.3/1.4 channels modulates channel properties, causing an increased channel open probability and slowed voltage-dependent inactivation [51, 53]; therefore, variability of expression of CaM in competition with the ICDI/CTM domain likely shapes the population dynamics of the Ca_v1.4 calcium current within a cell, both by apoCaM modulation and CaM-mediated CDI in bound channels [52].

Additional complexity is added by the recently discovered protein kinase A phosphorylation of the Ca_v1.4 ICDI/CTM domain. Phosphorylation of S1883 prevents ICDI/CTM interaction with the IQ domain, allowing CaM to bind and thereby increase channel open probability and confer CDI [67]. As PKA activity is dopamine-dependent, and retinal dopamine release is increased by illumination, Ca_v1.4 phosphorylation may be an important physiological mechanism for the regulation of visual sensitivity [68–71]. As this scheme has only recently been identified in a heterologous expression system, additional research is expected to provide further insight into functional consequences *in vivo*.

3.4. Calcium-Binding Protein 4 (CaBP4). The function of a second protein modulator, calcium-binding protein 4 (CaBP4), also appears to be essential for Ca_v1.4 function *in vivo*. CaBP4 is a one member of a subfamily of calmodulin-like proteins implicated in the regulation of voltage-gated calcium channels and inositol triphosphate receptors [72–74]. Mutations in CaBP4 cause autosomal recessive CSNB2 in humans, and mouse CaBP4-KO models exhibit a retinal phenotype similar to that of Ca_v1.4-KO mice [55, 75]. Research has shown that CaBP4 also binds to the C-terminus of the Ca_v1.4 channel, and in ICDI/CTM-truncation mutants it prevents CaM-mediated CDI [54, 55]. When ICDI/CTM is present, CaBP4 hyperpolarizes the channel-activation voltage, countering the positive shift attributed to the ICDI/CTM itself, and thus allowing Ca_v1.4 to function optimally at the physiological photoreceptor membrane potential [54]. Despite this apparent neutralization, both CaBP4 and the ICDI/CTM domain are necessary for proper channel function, as deleterious mutations in either one of them independently causes CSNB2 (see below: *CSNB2-Causing Mutations*). This suggests that the distal C-terminus may have an additional, essential role in channel function, beyond inhibition of CDI and channel modulation.

Structural studies have shown that calcium-bound CaBP4 tightly binds the IQ-motif, thereby likely competitively inhibiting both CaM and ICDI/CTM binding, whereas calcium-free/Mg²⁺-bound CaBP4 does not. Therefore, Park et al. suggest that, with high intracellular calcium levels in darkness, CaBP4 simultaneously prevents CaM-mediated CDI and ICDI/CTM-mediated positive shifts in activation voltage, thus promoting the open state. In light, the decrease in cytosolic calcium destabilizes the CaBP4-IQ interaction, allowing for ICDI/CTM and/or CaM binding and rapid channel closure [76]. Alternatively, Yang et al. argue that the higher intracellular concentrations and binding affinity

of CaM suggest an allosteric mechanism, and they provide compelling evidence that CaBP4 interactions with alternative, less competitive sites are sufficient for CaBP4 regulatory functions with channel-bound CaM [77]. Further research is required to determine how the complex interplays among CaM, CaBP4, the ICDI/CTM domain, and PKA phosphorylation of Ca_v1.4 shapes the calcium current in photoreceptors for optimal visual function. For a summary of the effects of these interactions on CDI under various physiological circumstances, see Figure 3.

4. Ca_v1.4 Dysfunction in Disease

4.1. Congenital Stationary Night Blindness 2 (CSNB2) Characteristics. Congenital stationary night blindness 2 (CSNB2, also known as incomplete CSNB) is an X-linked (CSNB2A) or autosomal (CSNB2B) recessive, minimally progressive disorder with substantial clinical heterogeneity. Symptoms may include nystagmus, strabismus, night blindness, photophobia, colour vision defects, and decreased visual acuity, the presence and severity of which vary greatly from patient to patient [78–80]. In fact, recent articles have emphasized that, given the low prevalence and impact of night vision problems in individuals with the condition (only affecting 54% in a recent analysis of 101 Dutch patients, [78, 81]), “night blindness” is a misnomer, suggesting “congenital rod-cone synaptic dysfunction” or the historical term “Åland Eye Disease” as viable alternatives [39, 82]. Electroretinography (ERG) and mutation analyses are the definitive tools for diagnosing CSNB2. Patients exhibit a characteristic reduction in scotopic b-wave amplitude, with preservation of the a-wave, indicating normal rod phototransduction with disrupted synaptic transmission [8, 38]. Photopic ERG (30 Hz-flicker) responses are also characteristically abnormal, making it possible to distinguish between CSNB2 and similar retinal afflictions including CSNB1, rod dystrophies, and cone dystrophies [83]. Observations in CSNB2 patients via optical coherence tomography (OCT) have revealed several anatomical abnormalities, including thinning of inner and outer retinal layers, foveal hypoplasia, reduced length of foveal cone outer segments, and suggested abnormal synaptogenesis [39, 84, 85]. Estimating the prevalence of CSNB2 is complicated by the aforementioned symptomatic variability, together with the historically rare use of electroretinography in common clinical practice. Nonetheless, a conservative estimate of 1 in 22,000 live-born males was calculated on the basis of the ratio of identified CSNB2-affected to all live-born males in Denmark between 1980 and 2009 [39]. Recent ERG analyses of heterozygous female carriers of *CACNA1F* mutations suggest that while the affected individuals lack subjective awareness, their objective visual function is mildly compromised [86, 87]. These data align with observations of heterozygous *Cacna1f*^{+/-} mice, which exhibit reduced visual function and mosaic abnormalities of retinal structure [86, 88]. It should also be noted that, despite the restricted expression of Ca_v1.4, mutations in *CACNA1F* have been implicated in schizophrenia by haplotype-based haplotype relative (HHRR) analysis, but further work is needed to substantiate this correlation and determine whether the mutations cause or increase risk of the disorder [89].

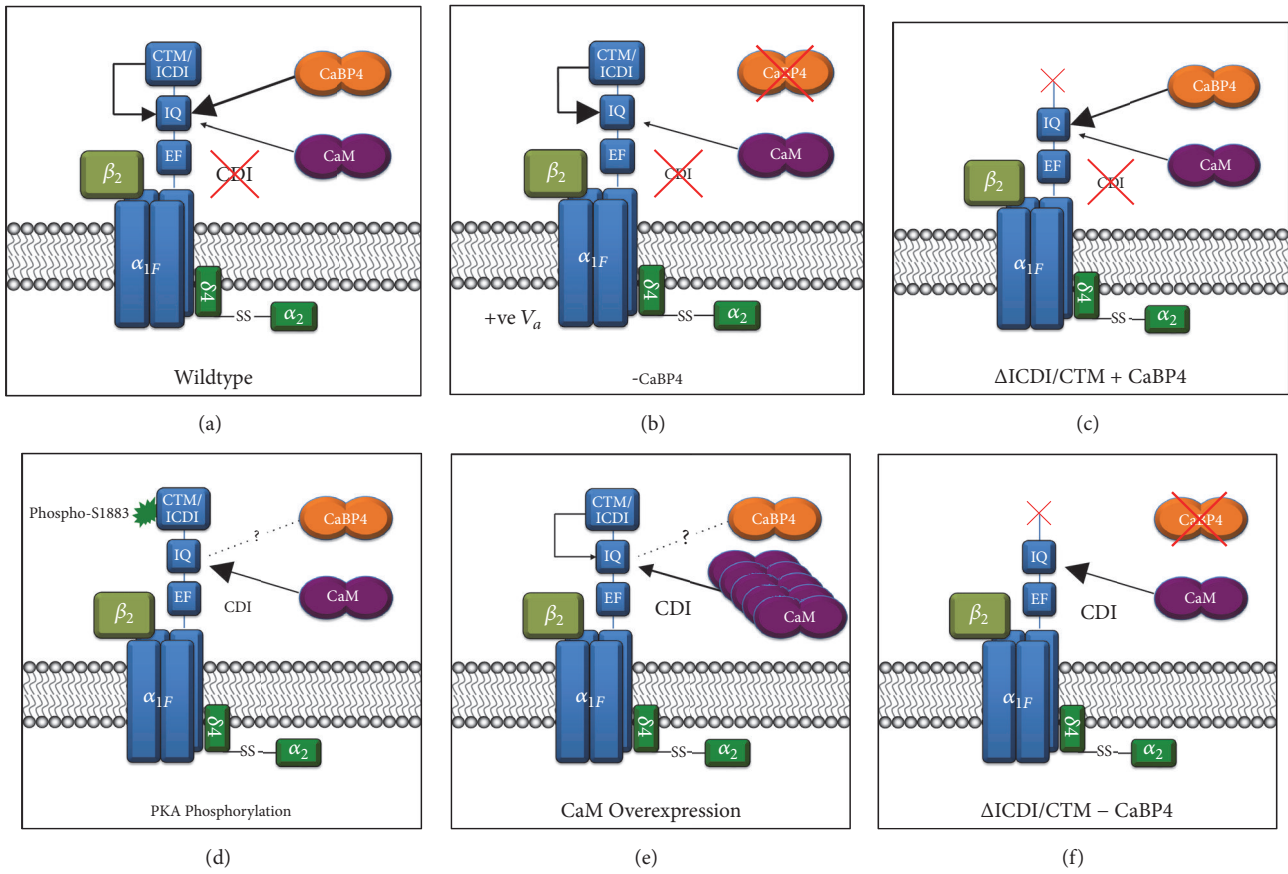


FIGURE 3: Mechanisms of C-terminal regulation. (a) In the wild-type $Ca_v1.4$ channel, with CaM and CaBP4 coexpressed, CaBP4 and the ICDI/CTM competitively interact with the proximal C-terminus, preventing CaM-mediated calcium-dependent inactivation (CDI). (b) When CaBP4 is not present, the ICDI/CTM still competitively inhibits CaM-mediated CDI, but the interaction between the ICDI/CTM and proximal C-terminus causes a significant positive shift in the activation voltage. (c) In the absence of the ICDI/CTM, CaBP4 prevents CDI, and the half-maximal voltage is unaffected, suggesting an additional crucial role for the ICDI/CTM *in vivo*. (d) The ICDI/CTM can be phosphorylated at serine 1883 by protein kinase A (PKA), allowing CaM to facilitate CDI and modulate channel properties. (e) Overexpression of CaM results in CaM's outcompeting ICDI/CTM for binding to the proximal C-terminus, thus facilitating CDI. The effect of CaBP4 in this circumstance has yet to be investigated. (f) In the absence of both CaBP4 and the ICDI/CTM, CaM is not competitively inhibited from interaction with the proximal C-terminus; thus, it confers CDI.

4.2. CSNB2-Causing Mutations. Mutations associated with CSNB2 have been identified in two genes, *CACNA1F* and *CABP4*, which cause CSNB2A and CSNB2B, respectively [10, 11, 75]. Mutations in *CACNA1F* are more prevalent by far, being associated with >90% of CSNB2 cases [38]. Mutations in an additional gene, *CACNA2D4* (which codes for the $\alpha_2\delta_4$ subunit of $Ca_v1.4$), have also been implicated in very rare cases of a CSNB2-like disorder, which may be reclassified as a CSNB2-subtype in time [21, 38, 90]. Given that CaBP4 and $\alpha_2\delta_4$ are important modulators of $\alpha_{1F}/Ca_v1.4$ function, all CSNB2-causing mutations may cause disease by deleteriously affecting the photoreceptor presynaptic calcium current [17, 54]. In addition to CSNB2, *CACNA1F* mutations are also known to cause X-linked Cone-Rod Dystrophy 3 (CORDX3) [91, 92], Åland Eye Disease [39, 93, 94], and even a progressive retinal/optic disc atrophy [95]. Similarities between these allelic variants have led some to suggest that a degree of unification may be necessary [39].

While all known CSNB2-causing mutations affect presynaptic proteins associated with the photoreceptor calcium current, CSNB1 (also known as complete congenital stationary night blindness) is caused by deleterious mutations in essential postsynaptic proteins of ON-bipolar cells. Accordingly, these patients exhibit visual defects and ERG responses consistent with a loss of ON-bipolar cell function, but an intact OFF-bipolar cell pathway [38]. Though outside the scope of this review, studies of CSNB1 animal models have been valuable in determining the mechanisms underlying the ON-bipolar pathway (for a comprehensive review of CSNB1, and other CSNB subtypes, see Zeitz et al. [38]).

Over 140 mutations in *CACNA1F* have been described (see Figure 4; for a detailed review of all known CSNB-causing mutations identified before 2015, see Zeitz et al. [38]). These have been insightfully divided into three categories: (1) loss-of-function mutations; (2) gain-of-function mutations; and (3) ICDI/C-terminal modulator- (CTM-)

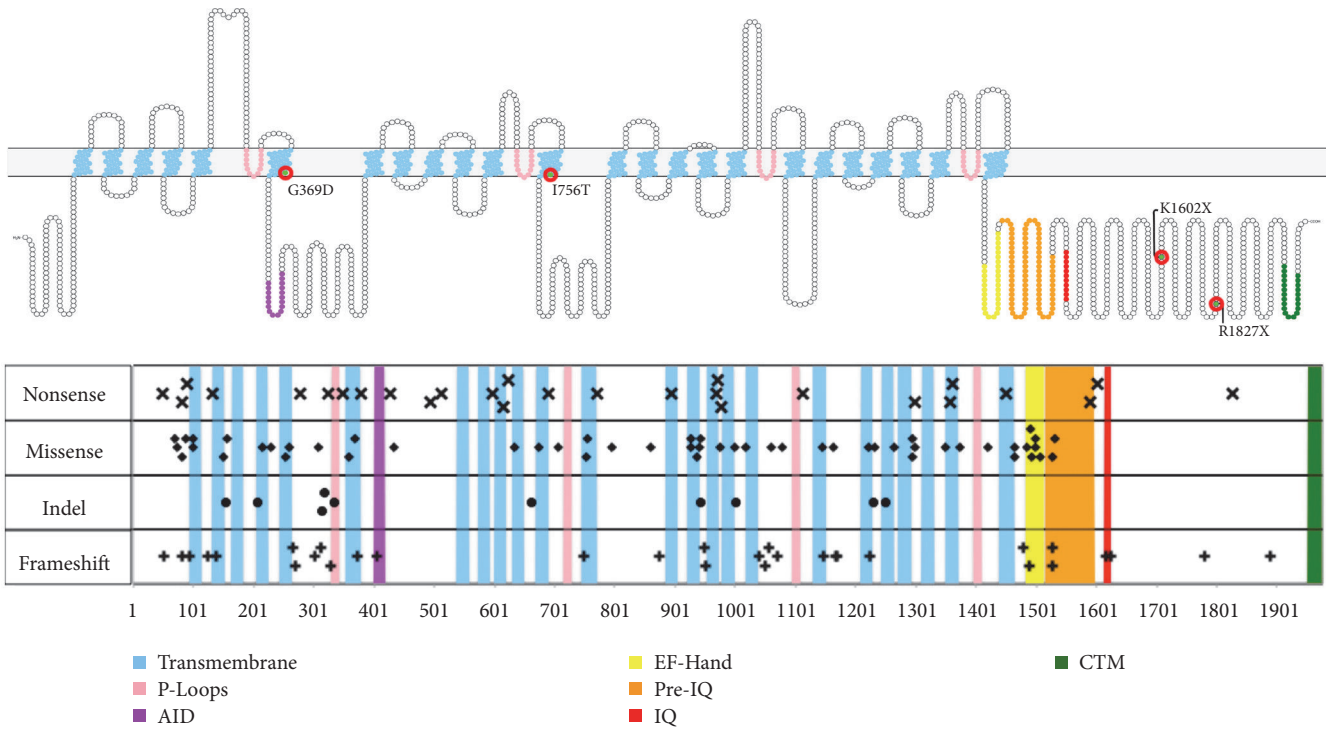


FIGURE 4: CSNB2A-causing mutations in *CACNA1F* and their locations in the α_{1F} subunit (Isoform 1, NCBI Reference Sequences: NM_005183.3 (mRNA), NP_005174.2 (Protein), UNIPROT Identifier: O60840-1) (nomenclature of α_{1F} mutations differs between laboratories, as groups have used two distinct protein isoforms of different lengths (due to an 11 amino-acid difference in exon 9 beginning at residue 426). Consequently, mutations occurring after amino-acid 426 may be differentially annotated between publications (ex. R1827X [36] versus R1816X [37]). Readers should be aware of this discrepancy between publications, and authors should clearly indicate the particular isoform used when annotating mutations). Compiled from Zeitz et al. [38] and Hove et al. [39]. AID: alpha-interaction domain; CTM: C-terminal modulator/inhibitor of calcium-dependent inactivation. Mutations specifically addressed in the text are highlighted. α_{1F} topographical visualization generated with Protter [40].

impairing mutations [36, 38, 39]. Diversity in the last two types of mutation certainly contributes to some of the clinical variability observed, but interestingly, substantial symptomatic variability is observed in patients with similar or even identical mutations [80, 96]. This may be attributable to the effects of other modulating genetic or environmental factors [79, 80, 96].

4.3. Effects of Mutations on $Ca_v1.4$ Function. The effects of various CSNB2A-causing *CACNA1F* mutations on $Ca_v1.4$ channel function have been investigated in several heterologous expression systems, in part to elucidate the pathogenic mechanisms contributing to CSNB2A variability. All nonsense mutations within 50–55 amino acids of a downstream splice site are expected to undergo nonsense-mediated decay, leading to a loss of $Ca_v1.4$ channel expression, although several missense mutations have also been shown to abolish calcium influx with wild-type-like expression levels [36, 97–99]. Biochemical mechanisms underlying the loss of calcium influx with several of these point mutations are reviewed in Stockner and Koschak, 2013 [36]. Of the mutations characterized to date that cause significant changes in channel kinetics without abolishing measurable current, all cause a negative shift in activation voltage, which may cause a

detrimental reduction in the dynamic range of photoreceptor responses or degeneration as a consequence of the sustained high levels of intracellular calcium. The most drastic of these, I745T, is associated with a particularly severe CSNB2A phenotype and significant retinal disruption [27, 28, 87, 100, 101]. The G369D mutation was reported by Hoda et al. to cause a significant (–13.7 mV) shift in activation voltage, in contrast to a previous report in which no such shift was found [23, 98]. The authors attributed this to electrostatic variability in vicinity of the mutation, due to slight differences in primary structure in the α_{1F} constructs that they used [23, 98]. This suggests that subtle electrostatic differences can drastically alter channel dynamics and may contribute to clinical variability in CSNB2A patients. The putative effect of R1827X, K1602X, and other C-terminal truncation mutations is the abolition of C-terminal regulation, discussed above (*C-Terminal Regulation and Calmodulin*).

5. Insights from CSNB2A Animal Models

5.1. Overview of CSNB2 Models. Seven animal models with mutations in *Cacnalf* have been identified or created, revealing many insights into the physiological role of $Ca_v1.4$. The first definitive model with a mutation in *Cacnalf* was

published in 2005, with an insertion in exon 7 leading to a premature truncation at amino-acid G305. This *Cacnalf*^{G305X} mouse, in contrast to the human condition, exhibits a complete loss of both scotopic and photopic ERG b-waves [102]. To date, no behavioral or electrophysiological tests have revealed any visual function in the *Cacnalf*^{G305X} or in another loss-of-function mouse model, *Cacnalf*^{Δ14-17} (created by excision of exons 14–17, also leading to premature truncation) [103]. Visual function in these mice has been collectively evaluated by optokinetic response analysis [104], analysis of cortical visual evoked potentials [102], multiunit activity recording from the superior colliculus [102], electroretinography [100, 102], a visually guided water-maze behavioral task [86], and multielectrode array analyses of full-field flash ganglion cell responses in isolated retinas [105]. Similar results have been observed in two “wait until dark” (*wud*) zebrafish, which have premature truncations in *cacnalf*, one of two zebrafish paralogs of human *CACNAIF*. These zebrafish have severely attenuated ERG a- and b-waves, exhibit no visual startle response, and do not follow visual stimuli, indicating that they are completely blind [106].

Why these models appear to lack detectable visual function while loss-of-function *CACNAIF* mutations in CSNB2-affected humans (~50% of known mutations) cause only a variable reduction in visual acuity (among other symptoms) is unclear. Expression of a second, compensatory voltage-gated calcium channel (most likely Ca_v1.3) is the most plausible explanation, though this has yet to be shown definitively in the human retina. In *Cacnald*-KO (α_{1D} /Ca_v1.3) mice, there is no observed change in performance in visually guided behavioral tasks, but a mild decrease in photoreceptor ribbon synapse density and disrupted ribbon morphology has been reported by two groups. ERG analyses of retinas in these mice, however, have provided conflicting reports. A recent study of *Cacnald*-KO mice reported decreased ERG a-wave, b-wave, and oscillatory potential (associated with amacrine cell function) amplitudes, in contrast to a previous study in which no such reductions were found [30, 107]. ERG analyses of *Cacnalf*/*Cacnald* double-KO mice showed no significant differences between these and *Cacnalf*-KO mice [108]. These discrepancies warrant further investigation into the role of Ca_v1.3 in the photoreceptor synaptic terminal.

In contrast to mouse and zebrafish models, premature truncation of *Cacnalf* in a spontaneous rat model (*Cacnalf*^{R981X}) causes impaired but not absent visual function, as well as a reported effect on several nonvisual behaviors [109, 110]. Interestingly, this model also exhibits decreased skeletal muscle endurance and contractility, leading the authors to suggest a role for Ca_v1.4 outside of the retina and immunocytes in rat [111].

Two additional mouse models of CSNB2 with unique mutations also have been reported and shown to exhibit unique phenotypic characteristics. The *nob2* (*no b-wave 2*) mouse was originally discovered as part of an ERG-based screen at the Jackson Laboratory for spontaneous retinal mutants and predicted to cause a complete loss of Ca_v1.4 expression due to the insertion of a transposable

element in *Cacnalf* exon 2 [112, 113]. Differences in phenotype between this mouse and the previously described *Cacnalf*^{G305X} knockout model, including an observable ERG b-wave in *nob2* [112, 114], led to the identification of an alternate *Cacnalf* splice variant, which results in reduced (~10%) expression of wild-type-like Ca_v1.4 in the *nob2* retina [115]. The photopic and scotopic contrast sensitivities of the optokinetic response in *nob2* mice are only slightly below those of wild-type, suggesting that low-level expression of α_{1F} is sufficient for near-optimal visual function, despite the persistence of other morphological anomalies in these retinas [104, 115, 116].

The *Cacnalf*^{I756T} mouse represents the only *Cacnalf* missense mutant model described to date. This mutant was engineered to emulate a CSNB2-causing human mutation, discovered in a New Zealand family with particularly severe clinical characteristics, including nonocular neurological abnormalities and visual defects in heterozygous female carriers (rarely reported with *CACNAIF* mutations)¹⁰¹. Electrophysiological analysis of the resultant Ca_v1.4 channel in a heterologous system revealed a drastic change in the activation voltage, leading to a reduced dynamic range (described above: *Effects of Mutations on Ca_v1.4 Function*). ERG recordings from *Cacnalf*^{I756T} mice and CSNB2 patients carrying the analogous mutation are similarly affected (unlike other *Cacnalf* mutant models), with a residual but significantly attenuated b-wave, reduced a-wave, and other characteristics generally absent from other CSNB2A models [28, 101]. Interestingly, Knoflach et al. have reported an upregulation of *Cacnald* mRNA (α_{1D} /Ca_v1.3) in the *Cacnalf*^{I756T} mouse retina, providing some animal evidence for the mechanism that might contribute to residual vision in human CSNB2A patients [28].

5.2. Physiological and Morphological Observations in CSNB2A Model Retinas. Animal models have also allowed for extensive descriptions of physiological changes in response to mutations in *Cacnalf*. In retinas of all *Cacnalf*-KO models, synaptic ribbons of the photoreceptor synaptic terminals (discussed above: *Ca_v1.4 Function: The Ribbon Synapse and Phototransduction*) never anchor to the presynaptic membrane or elongate into their characteristic morphology, according to electron microscopical and immunohistochemical observations [27, 106, 117, 118]. Comparisons to *CaBP4*-KO and *Cacnalf*^{I756T} retinas suggest that the synthesis of α_{1F} protein and the assembly of complete Ca_v1.4 channels in the membrane (regardless of kinetics) are necessary for the initial formation of photoreceptor presynaptic terminals, until P13 after which biophysical properties become important for proper maturation as these models exhibit typical retinal development until eye opening. Ca_v1.4 may play a scaffolding role for synaptic elements, including PSD-95 and PMCA, which help develop the presynaptic ribbon [27].

Recent insights from mouse models support this scaffolding role for Ca_v1.4, particularly in rod synaptogenesis. Ca_v1.4 appears to coordinate the membrane localization of the extracellular protein ELFN1, which associates specifically with the $\alpha_2\delta_4$ subunit of the channel. ELFN1 is a

trans-synaptic protein, which facilitates synaptic alignment with the metabotropic glutamate receptor mGluR6 in rod ON-bipolar cells. In the absence of $Ca_v1.4$, the $\alpha_2\delta_4$ subunit, or ELFN1 itself, rod synapses do not form. Remarkably, neither $\alpha_2\delta_4$ nor ELFN1 is necessary for cone synaptogenesis; this suggests that cones may use an alternative, as yet undiscovered, molecular mechanism for synaptogenesis [58, 119].

The disruption of the presynaptic terminal and the absence or irregularity of calcium signaling causes complete failure (in loss-of-function [KO]) or severe aberration (in I756T) of synaptogenesis between the photoreceptors and second-order neurons (bipolar and horizontal cells) [27, 102, 117, 118]. In response, normally rod-connected dendrites of both bipolar and horizontal cells form abnormal extensions into the outer nuclear layer (“sprouting”), and photoreceptor terminals have been reported to retract from the outer plexiform layer into the outer nuclear layer [100, 116, 118]. Despite their abnormal course and destination, sprouted dendrites have been reported to form ectopic synapses in the ONL as revealed by their close apposition to photoreceptor processes containing any of several presynaptic elements, including Bassoon [100, 102, 118], RIBEYE [27, 88, 118], Cplx4 [86, 118], vGlut1 [86, 118], Cplx3 [86], Piccolo [88, 100], VAMP2 [118], and synaptophysin [118]. Despite the presence of vesicle-release machinery at these ectopic synapse-like associations, no evidence of synaptic transmission has been reported to date [118]. Abnormal bipolar cell synapses in the ONL of a human CSNB2A patient have also been suggested from abnormalities observed using optical coherence tomography [84]. Comparisons to animal models of other retinal diseases suggest that bipolar and horizontal cell dendrites may sprout to seek contact with active presynaptic cells. In *CNGA3*-KO mice, in which cone phototransduction is disrupted, cone bipolar cells form ectopic synapses with nearby rods. In *rhodopsin*-KO mice, in which rod phototransduction is disrupted, rod bipolar cells form ectopic synapses with cones. In *CNGA3*^{-/-}/*Rho*^{-/-} double-KO mice, no ectopic synapses are observed by P46 [120].

CSNB2A is commonly referred to as a nonprogressive (“stationary”) condition. However, both *Cacnalf*-KO and *Cacnalf*^{I756T} mice exhibit progressive photoreceptor dystrophy and degeneration. Histological images showing thinning of the ONL and TUNEL-labeling of presumably apoptotic cells, as well as photoreceptor-specific immunolabeling suggest that photoreceptors are gradually lost in all CSNB2A mouse models, with *Cacnalf*^{I756T} retinas exhibiting the most rapid degeneration [86, 100, 117, 118]. These data are supported by longitudinal ERG analyses, which have shown significant declines in a-wave amplitudes between one and eight months in both mutants, but much more dramatic reduction in *Cacnalf*^{I756T} retinas [100]. Immunohistochemical visualization of adult cones reveals general dystrophy, absent (KO) or abnormal (I756T) terminals, and axonal abnormalities including varicosities and sprouting [86, 88, 100, 117, 118]. We have recently shown that sprouted PKC α + rod bipolar cells form ectopic contacts with these dystrophic cones in the *Cacnalf*-KO retina, which increasingly consolidate as these mice age [88]. These contacts, along with the cone

axonal sprouting (which have not been described in any other synaptopathy model), are particularly intriguing, as they suggest that many cones retain the potential for synaptic plasticity beyond the normal period of photoreceptor synaptogenesis [88, 118]. For a summary of known major morphophysiological abnormalities in the mouse *Cacnalf*-KO retina, see Figure 5.

The phenotype of *Cacnalf*-KO mice makes this model uniquely suited to answer an outstanding question in retinal neuroscience: can photoreceptors form functional synapses with inner retinal neurons, after the normal period of development has ceased? This question is especially pertinent now, following the recent evidence that transplanted photoreceptor precursors do not integrate into the circuitry of degenerating retinas, as was originally believed [121]. Photoreceptors in *Cacnalf*-KO retinas are apparently “invisible” to second-order neurons, yet the inner retina exhibits few of the degenerative changes observed in models of primarily photoreceptor degeneration [122]. Therefore, *Cacnalf*-KO animal models may be useful tools for identifying the factors that are critical for normal photoreceptor synaptogenesis, and further investigation of these models may offer, not only insight into potential clinical interventions for patients with CSNB2A, but also hope for the millions living with photoreceptor-destroying degenerative retinal diseases.

6. Conclusion

$Ca_v1.4$ is a highly specialized voltage-gated calcium channel that has evolved unique characteristics for its distinct role within the nervous system. The remarkable sensitivity of the visual system is dependent on $Ca_v1.4$, as evidenced by the vision losses in patients with CSNB2A and other clinical conditions caused by mutations in *CACNA1F* (e.g., Åland Eye Disease); accumulating evidence suggests that this channel may also serve yet unknown or poorly understood functional roles in other systems. Research into the biophysical effects of mutations, in *CACNA1F* and mechanisms of $Ca_v1.4$ regulation and gating, has been enlightening for our understanding of voltage-gated calcium channels, CSNB2A, and retinal circuitry as a whole. In particular, the recent use of animal models to investigate physiological changes in response to known CSNB2A-causing mutations has provided a wealth of knowledge into the functional roles of $Ca_v1.4$ (and other VGCCs) in the nervous system. Continuing to unearth the complexities of neural circuitry, by exploiting these models as unique and powerful research tools, brings us closer to devising life-changing clinical therapies for CSNB2A and other retinal degenerative diseases.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Authors' Contributions

N. T. Bech-Hansen and W. K. Stell contributed equally to this review.

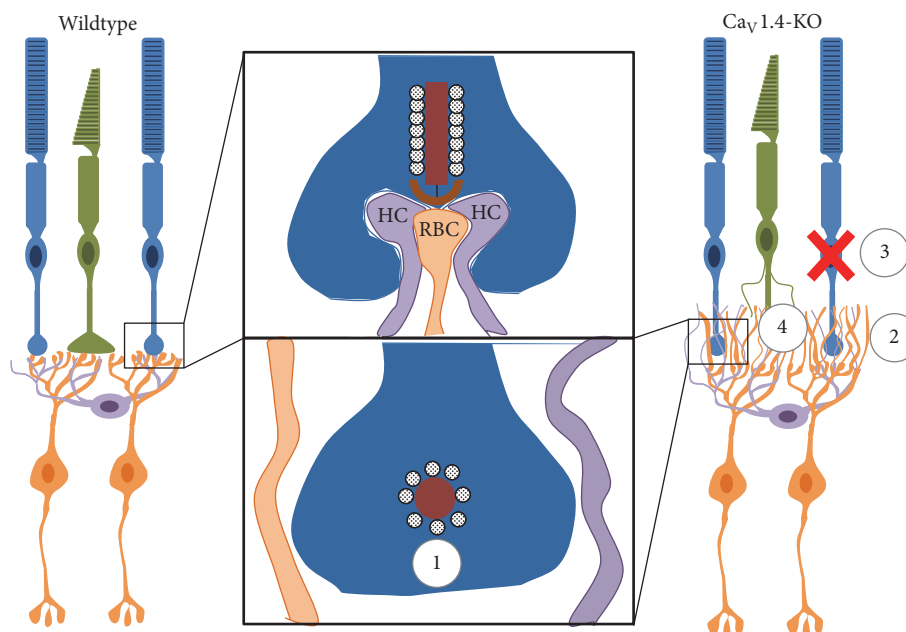


FIGURE 5: Known morphophysiological abnormalities in *Cacnalf*-KO mouse retinas (lacking $Ca_v1.4$). (1) The synaptic ribbon fails to mature or anchor to the presynaptic membrane; (2) normally rod-contacting bipolar and horizontal cell dendrites sprout past the outer plexiform layer, into the outer nuclear layer, potentially seeking presynaptic partners; (3) photoreceptors (both rods and cones) gradually die, leading to loss of nuclei in the outer nuclear layer; and (4) cone photoreceptors exhibit aberrant axonal morphology, including axonal sprouting.

Acknowledgments

The authors acknowledge support from the Canadian Institutes for Health Research (Operating Grant FRN 111192 to N. T. Bech-Hansen), the Foundation Fighting Blindness-Canada (FFB-EyeGeye Research Training Fund, to W. K. Stell), NSERC (Discovery Grant RGPIN/131-2013, to W. K. Stell), the University of Calgary (Lions Sight Centre Fund, to N. T. Bech-Hansen and W. K. Stell), and donations from Alberta Ride for Sight. The authors would also like to thank Dr. Robert J. French (University of Calgary) for helpful input, advice, and feedback.

References

- [1] F. D. Carroll and C. Haig, "Congenital Stationary Night Blindness without Ophthalmoscopic or Other Abnormalities," *Transactions of the American Ophthalmological Society*, vol. 50, pp. 193–209, 1952.
- [2] F. Moro, S. Li Volti, S. Tomarchio, and F. Mollica, "X-linked recessive myopia associated with nyctalopia in a Sicilian family," *Ophthalmic Paediatrics and Genetics*, vol. 1, no. 3, pp. 173–176, 1982.
- [3] J. François, *Heredity in ophthalmology*, 400 St. Louis, Mosby, 1961.
- [4] V. J. Gassler, "Ueber eine bis jetzt nicht bekannte recessive Verknüpfung von hochgradiger Myopie mit angeborener Hemeralopie," *Orell Füssli*, 1925.
- [5] F. C. Donders, "Torpeur de la retine congenitale e hereditaire," *Ann Ocul (Paris)*, vol. 34, pp. 270–273, 1855.
- [6] G. Schubert, H. Bornschein, and Ophthalmologica, "Ophthalmologica," *Journal International d'Ophthalmologie. International Journal of Ophthalmology*, vol. 123, pp. 396–413.
- [7] L. A. Riggs, "Electroretinography in cases of night blindness," *American Journal of Ophthalmology*, vol. 38, no. 1, pp. 70–78, 1954.
- [8] Y. Miyake, K. Yagasaki, M. Horiguchi, Y. Kawase, and T. Kanda, "Congenital stationary night blindness with negative electroretinogram. A new classification," *Archives of ophthalmology (Chicago, Ill. : 1960)*, vol. 104, no. 7, pp. 1013–1020, 1986.
- [9] K. M. Boycott, W. G. Pearce, M. A. Musarella et al., "Evidence for genetic heterogeneity in X-linked congenital stationary night blindness," *American Journal of Human Genetics*, vol. 62, no. 4, pp. 865–875, 1998.
- [10] T. M. Strom, G. Nyakatura, E. Apfelstedt-Sylla et al., "An L-type calcium-channel gene mutated in incomplete X-linked congenital stationary night blindness," *Nature Genetics*, vol. 19, no. 3, pp. 260–263, 1998.
- [11] N. T. Bech-Hansen, M. J. Naylor, T. A. Maybaum et al., "Loss-of-function mutations in a calcium-channel $\alpha 1$ -subunit gene in Xp11.23 cause incomplete X-linked congenital stationary night blindness," *Nature Genetics*, vol. 19, no. 3, pp. 264–267, 1998.
- [12] S. E. Fisher, A. Ciccociola, K. Tanaka et al., "Sequence-based exon prediction around the synaptophysin locus reveals a gene-rich area containing novel genes in human proximal Xp," *Genomic*, vol. 45, no. 2, pp. 340–347, 1997.
- [13] M. J. Naylor, D. E. Rancourt, and N. T. Bech-Hansen, "Isolation and characterization of a calcium channel gene, *Cacnalf*, the murine orthologue of the gene for incomplete X-linked congenital stationary night blindness," *Genomics*, vol. 66, no. 3, pp. 324–327, 2000.
- [14] W. R. Taylor and C. Morgans, "Localization and properties of voltage-gated calcium channels in cone photoreceptors of *Tupaia belangeri*," *Visual neuroscience*, vol. 15, no. 3, pp. 541–552, 1998.

- [15] G. W. Zamponi, J. Striessnig, A. Koschak, and A. C. Dolphin, "The physiology, pathology, and pharmacology of voltage-gated calcium channels and their future therapeutic potential," *Pharmacological Reviews*, vol. 67, no. 4, pp. 821–870, 2015.
- [16] W. A. Catterall, "Voltage-gated calcium channels," *Cold Spring Harbor Perspectives in Biology*, vol. 3, no. 8, Article ID a003947, 2011.
- [17] A. Lee, S. Wang, B. Williams et al., "Characterization of Cav1.4 complexes ($\alpha_{1.4}$, β_2 , and $\alpha_2\delta_4$) in HEK293T cells and in the retina," *Journal of Biological Chemistry*, vol. 290, no. 3, pp. 1505–1521, 2015.
- [18] K. A. Wycisk, B. Budde, S. Feil et al., "Structural and Functional Abnormalities of Retinal Ribbon Synapses due to *Cacna2d4* Mutation," *Investigative Ophthalmology & Visual Science*, vol. 47, no. 8, pp. 3523–3530, 2006.
- [19] S. L. Ball, P. A. Powers, H-S. Shin et al., "Role of the β_2 subunit of voltage-dependent calcium channels in the retinal outer plexiform layer," *Investigative Ophthalmology & Visual Science*, vol. 43, no. 5, pp. 1595–1603, 2002.
- [20] L. P. De Sevilla Müller, J. Liu, A. Solomon, A. Rodriguez, and N. C. Brecha, "Expression of voltage-gated calcium channel $\alpha_2\delta_4$ subunits in the mouse and rat retina," *The Journal of Comparative Neurology*, vol. 521, no. 11, pp. 2486–2501, 2013.
- [21] R. Ba-Abbad, G. Arno, K. Carss et al., "Mutations in *CACNA2D4* Cause Distinctive Retinal Dysfunction in Humans," *Ophthalmology*, vol. 123, no. 3 article no. e662, pp. 668–671, 2016.
- [22] M. F. Kotturi and W. A. Jefferies, "Molecular characterization of L-type calcium channel splice variants expressed in human T lymphocytes," *Molecular Immunology*, vol. 42, pp. 1461–1474, 2005.
- [23] J. E. McRory, J. Hamid, C. J. Doering et al., "The CACNA1F Gene Encodes an L-Type Calcium Channel with Unique Biophysical Properties and Tissue Distribution," *The Journal of Neuroscience : the Official Journal of the Society for Neuroscience*, vol. 24, no. 7, pp. 1707–1718, 2004.
- [24] C. J. Doering, J. B. Peloquin, and J. E. McRory, "The $Ca_{(v)}1.4$ calcium channel: more than meets the eye," *Channels (Austin, Tex.)*, vol. 1, pp. 3–10, 2007.
- [25] A. Berntson, W. R. Taylor, and C. W. Morgans, "Molecular identity, synaptic localization, and physiology of calcium channels in retinal bipolar cells," *Journal of Neuroscience Research*, vol. 71, no. 1, pp. 146–151, 2003.
- [26] C. W. Morgans, "Localization of the α_{1F} Calcium Channel Subunit in the Rat Retina," *Investigative Ophthalmology & Visual Science*, vol. 42, pp. 2414–2418, 2001.
- [27] X. Liu, V. Kerov, F. Haeseleer et al., "Dysregulation of $Ca_v1.4$ channels disrupts the maturation of photoreceptor synaptic ribbons in congenital stationary night blindness type 2," *Channels (Austin, Tex.)*, vol. 7, no. 6, pp. 514–523, 2013.
- [28] D. Knoflach, V. Kerov, S. B. Sartori et al., "Cav1.4 IT mouse as model for vision impairment in human congenital stationary night blindness type 2," *Channels (Austin, Tex.)*, vol. 7, pp. 503–513, 2013.
- [29] D. Specht, S. Tom Dieck, J. Ammermüller, H. Regus-Leidig, E. D. Gundelfinger, and J. H. Brandstätter, "Structural and functional remodeling in the retina of a mouse with a photoreceptor synaptopathy: Plasticity in the rod and degeneration in the cone system," *European Journal of Neuroscience*, vol. 26, no. 9, pp. 2506–2515, 2007.
- [30] L. Shi, J. Y. Chang, F. Yu, M. L. Ko, and G. Y. Ko, "The contribution of L-type Cav1.3 channels to Retinal Light Responses," *Frontiers in Molecular Neuroscience*, vol. 10, p. 394, 2017.
- [31] Q. Lu, *Characterization of High-Voltage Activated Calcium Currents in Retinal Bipolar Cells PhD thesis [Ph.D. thesis]*, Wayne State University, 2013.
- [32] F. Schmitz, "The Making of Synaptic Ribbons: How They Are Built and What They Do," *The Neuroscientist: a Review Journal Bringing Neurobiology, Neurology and Psychiatry*, vol. 15, no. 6, pp. 611–624, 2009.
- [33] R. Heidelberger, W. B. Thoreson, and P. Witkovsky, "Synaptic transmission at retinal ribbon synapses," *Progress in Retinal and Eye Research*, vol. 24, no. 6, pp. 682–720, 2005.
- [34] K. W. Yau, "Phototransduction mechanism in retinal rods and cones. The Friedenwald Lecture," *Investigative Ophthalmology & Visual Science*, vol. 35, pp. 9–32, 1994.
- [35] A. J. Mercer and W. B. Thoreson, "The dynamic architecture of photoreceptor ribbon synapses: Cytoskeletal, extracellular matrix, and intramembrane proteins," *Visual Neuroscience*, vol. 28, no. 6, pp. 453–471, 2011.
- [36] T. Stockner and A. Koschak, *Biochimica et biophysica acta* 1828, pp. 1598–1607, 2013.
- [37] K. Wutz, C. Sauer, E. Zrenner et al., "Thirty distinct CACNA1F mutations in 33 families with incomplete type of XLCSNB and *Cacn1f* expression profiling in mouse retina," *European Journal of Human Genetics*, vol. 10, pp. 449–456, 2002.
- [38] C. Zeitz, A. G. Robson, and I. Audo, "Congenital stationary night blindness: An analysis and update of genotype-phenotype correlations and pathogenic mechanisms," *Progress in Retinal and Eye Research*, vol. 45, pp. 58–110, 2015.
- [39] M. N. Hove, K. Z. Kilic-Biyik, A. Trotter et al., "Clinical Characteristics, Mutation Spectrum, and Prevalence of Åland Eye Disease/Incomplete Congenital Stationary Night Blindness in Denmark," *Investigative Ophthalmology & Visual Science*, vol. 57, pp. 6861–6869, 2016.
- [40] U. Omasits, C. H. Ahrens, S. Müller, and B. Wollscheid, "Protter: interactive protein feature visualization and integration with experimental proteomic data," *Bioinformatics (Oxford, England)*, vol. 30, no. 6, pp. 884–886, 2014.
- [41] M. F. Wilkinson and S. Barnes, "The dihydropyridine-sensitive calcium channel subtype in cone photoreceptors," *The Journal of General Physiology*, vol. 107, no. 5, pp. 621–630, 1996.
- [42] D. P. Corey, J. M. Dubinsky, and E. A. Schwartz, "The calcium current in inner segments of rods from the salamander (*Ambystoma tigrinum*) retina," *The Journal of Physiology*, vol. 354, pp. 557–575, 1984.
- [43] D. Krizaj and D. R. Copenhagen, "Calcium regulation in photoreceptors," *Frontiers in Bioscience: A Journal and Virtual Library*, vol. 7, pp. d2023–d2044, 2002.
- [44] L. Baumann, A. Gerstner, X. Zong, M. Biel, and C. Wahl-Schott, "Functional characterization of the L-type Ca^{2+} channel Cav1.4 α from mouse retina," *Investigative Ophthalmology & Visual Science*, vol. 45, no. 2, pp. 708–713, 2004.
- [45] A. Koschak, D. Reimer, D. Walter et al., "Cav1.4 α subunits can form slowly inactivating dihydropyridine-sensitive L-type Ca^{2+} channels lacking Ca^{2+} -dependent inactivation," *The Journal of Neuroscience : the Official Journal of the Society for Neuroscience*, vol. 23, pp. 6041–6049, 2003.
- [46] C. J. Doering, J. Hamid, B. Simms, J. E. McRory, and G. W. Zamponi, "Ca_v1.4 encodes a calcium channel with low open probability and unitary conductance," *Biophysical Journal*, vol. 89, pp. 3042–3048, 2005.
- [47] A. Singh, D. Hamedinger, J. C. Hoda et al., "C-terminal modulator controls Ca^{2+} -dependent gating of $Ca(v)1.4$ L-type

- Ca²⁺ channels,” *Nature Neuroscience*, vol. 9, no. 9, pp. 1108–1116, 2006.
- [48] F. Haeseleer, B. Williams, and A. Lee, “Characterization of C-terminal Splice Variants of Ca_v1.4 Ca²⁺ Channels in Human Retina,” *The Journal of Biological Chemistry*, vol. 291, pp. 15663–15673, 2016.
- [49] G. M. Tan, D. Yu, J. Wang, and T. W. Soong, “Alternative Splicing at C Terminus of Ca_v1.4 Calcium Channel Modulates Calcium-dependent Inactivation, Activation Potential, and Current Density,” *The Journal of Biological Chemistry*, vol. 287, pp. 832–847, 2012.
- [50] J. B. Peloquin, C. J. Doering, R. Rehak, and J. E. McRory, “Temperature dependence of Cav1.4 calcium channel gating,” *Neuroscience*, vol. 151, no. 4, pp. 1066–1083, 2008.
- [51] P. J. Adams, M. Ben-Johny, I. E. Dick, T. Inoue, and D. T. Yue, “Apocalmodulin itself promotes ion channel opening and Ca²⁺ regulation,” *Cell*, vol. 159, no. 3, pp. 608–622, 2014.
- [52] X. Liu, P. S. Yang, W. Yang, and D. T. Yue, “Enzyme-inhibitor-like tuning of Ca²⁺ channel connectivity with calmodulin,” *Nature*, vol. 463, pp. 968–972, 2010.
- [53] K. Griessmeier, H. Cuny, K. Rötzer et al., “Calmodulin is a functional regulator of Cav1.4 L-type Ca²⁺ channels,” *The Journal of Biological Chemistry*, vol. 284, no. 43, pp. 29809–29816, 2009.
- [54] L. Shaltiel, C. Pappas, S. Fenske et al., “Complex regulation of voltage-dependent activation and inactivation properties of retinal voltage-gated Cav1.4 L-type Ca²⁺ channels by Ca²⁺-binding protein 4 (CaBP4),” *The Journal of Biological Chemistry*, vol. 287, no. 43, pp. 36312–36321, 2012.
- [55] F. Haeseleer, Y. Imanishi, T. Maeda et al., “Essential role of Ca²⁺-binding protein 4, a Cav1.4 channel regulator, in photoreceptor synaptic function,” *Nature Neuroscience*, vol. 7, no. 10, pp. 1079–1087, 2004.
- [56] K. Omilusik, J. J. Priatel, X. Chen et al., “The Ca_v1.4 calcium channel is a critical regulator of T cell receptor signaling and naive T cell homeostasis,” *Immunity*, vol. 35, no. 3, pp. 349–360, 2011.
- [57] M. W. Richards, J. Leroy, W. S. Pratt, and A. C. Dolphin, “The HOOK-domain between the SH3 and the GK domains of Cavbeta subunits contains key determinants controlling calcium channel inactivation,” *Channels (Austin, Tex.)*, vol. 1, no. 2, pp. 92–101, 2007.
- [58] Y. Wang, K. E. Fehlhaber, I. Sarria et al., “The auxiliary calcium channel subunit $\alpha 2\delta 4$ is required for axonal elaboration, synaptic transmission, and wiring of rod photoreceptors,” *Neuron*, vol. 93, no. 6, Article ID e1356, pp. 1359–1374, 2017.
- [59] R. D. Zühlke, G. S. Pitt, K. Deisseroth, R. W. Tsien, and H. Reuter, “Calmodulin supports both inactivation and facilitation of L-type calcium channels,” *Nature*, vol. 399, pp. 159–162, 1999.
- [60] B. Z. Peterson, C. D. DeMaria, J. P. Adelman, and D. T. Yue, “Calmodulin is the Ca²⁺ sensor for Ca²⁺-dependent inactivation of L-Type calcium channels,” *Neuron*, vol. 22, pp. 549–558, 1999.
- [61] B. Z. Peterson, J. S. Lee, J. G. Mülle, Y. Wang, M. de Leon, and D. T. Yue, “Critical determinants of Ca(2+)-dependent inactivation within an EF-hand motif of L-type Ca(2+) channels,” *Biophysical Journal*, vol. 78, no. 4, pp. 1906–1920, 2000.
- [62] J. Kim, S. Ghosh, D. A. Nunziato, and G. S. Pitt, “Identification of the components controlling inactivation of voltage-gated Ca²⁺ channels,” *Neuron*, vol. 41, no. 5, pp. 745–754, 2004.
- [63] M. G. Erickson, H. Liang, M. X. Mori, and D. T. Yue, “FRET Two-Hybrid Mapping Reveals Function and Location of L-Type Ca²⁺ Channel CaM Preassociation,” *Neuron*, vol. 39, no. 1, pp. 97–107, 2003.
- [64] F. Van Petegem, F. C. Chatelain, and D. L. Minor Jr., “Insights into voltage-gated calcium channel regulation from the structure of the Ca_v1.2 IQ domain–Ca²⁺/calmodulin complex,” *Nature Structural & Molecular Biology*, vol. 12, pp. 1108–1115, 2005.
- [65] C. Wahl-Schott, L. Baumann, H. Cuny, C. Eckert, K. Griessmeier, and M. Biel, “Switching off calcium-dependent inactivation in L-type calcium channels by an autoinhibitory domain,” *Proceedings of the National Academy of Sciences of the United States of America*, vol. 103, no. 42, pp. 15657–15662, 2006.
- [66] J. Striessnig, H. J. Bolz, and A. Koschak, “Channelopathies in Cav1.1, Cav1.3, and Cav1.4 voltage-gated L-type Ca²⁺ channels,” *Pflügers Archiv - European Journal of Physiology*, vol. 460, no. 2, pp. 361–374, 2010.
- [67] L. Sang, I. E. Dick, and D. T. Yue, “Protein kinase A modulation of Ca_v1.4 calcium channels,” *Nature Communications*, vol. 7, no. 12239, 2016.
- [68] P. Witkovsky, “Dopamine and retinal function,” *Documenta Ophthalmologica. Advances in Ophthalmology*, vol. 108, no. 1, pp. 17–40, 2004 (Chinese).
- [69] P. Witkovsky, R. Gabriel, J. W. Haycock, and E. Meller, “Influence of light and neural circuitry on tyrosine hydroxylase phosphorylation in the rat retina,” *Journal of Chemical Neuroanatomy*, vol. 19, no. 2, pp. 105–116, 2000.
- [70] D. Parkinson and R. R. Rando, “Effect of light on dopamine turnover and metabolism in rabbit retina,” *Investigative Ophthalmology & Visual Science*, vol. 24, pp. 384–388, 1983.
- [71] P. M. Iuvone, C. L. Galli, C. K. Garrison-Gund, and N. H. Neff, “Light stimulates tyrosine hydroxylase activity and dopamine synthesis in retinal amacrine neurons,” *Science (New York, N.Y.)*, vol. 202, no. 4370, pp. 901–902, 1978.
- [72] F. Findeisen, C. H. Rumpf, and D. L. Minor Jr., “Apo States of Calmodulin and CaBP1 Control Ca_v1 Voltage-Gated Calcium Channel Function through Direct Competition for the IQ Domain,” *Journal of Molecular Biology*, vol. 425, no. 17, pp. 3217–3234, 2013.
- [73] S. Oz, V. Tsemakhovich, C. J. Christel, A. Lee, and N. Dascal, “CaBP1 regulates voltage-dependent inactivation and activation of Ca V1.2 (L-type) calcium channels,” *The Journal of Biological Chemistry*, vol. 286, no. 16, pp. 13945–13953, 2011.
- [74] N. Nejatbakhsh and Z. P. Feng, “Calcium binding protein-mediated regulation of voltage-gated calcium channels linked to human diseases,” *Acta Pharmacologica Sinica*, vol. 32, no. 6, pp. 741–748, 2011.
- [75] C. Zeitz, B. Kloeckener-Gruissem, and U. Forster, “Mutations in CABP4, the gene encoding the Ca²⁺-binding protein 4, cause autosomal recessive night blindness,” *American Journal of Human Genetics*, vol. 79, no. 4, pp. 657–667, 2006.
- [76] S. Park, C. Li, F. Haeseleer, K. Palczewski, and J. B. Ames, “Structural Insights into Activation of the Retinal L-type Ca²⁺ Channel (Cav1.4) by Ca²⁺-binding Protein 4 (CaBP4),” *The Journal of Biological Chemistry*, vol. 289, pp. 31262–31273, 2014.
- [77] P. S. Yang, M. B. Johny, and D. T. Yue, “Allostery in Ca²⁺ channel modulation by calcium-binding proteins,” *Nature Chemical Biology*, vol. 10, pp. 231–238, 2014.
- [78] M. M. C. Bijveld, R. J. Florijn, A. A. B. Bergen et al., “Genotype and phenotype of 101 dutch patients with congenital stationary night blindness,” *Ophthalmology*, vol. 120, no. 10, pp. 2072–2081, 2013.

- [79] C. Zeitz, "Molecular genetics and protein function involved in nocturnal vision," *Expert Review of Ophthalmology*, vol. 2, no. 3, pp. 467–485, 2007.
- [80] K. M. Boycott, W. G. Pearce, and N. T. Bech-Hansen, "Clinical variability among patients with incomplete X-linked congenital stationary night blindness and a founder mutation in *CACNA1F*," *Canadian Journal of Ophthalmology/Journal Canadien d'Ophthalmologie*, vol. 35, no. 4, pp. 204–213, 2000.
- [81] M. M. Bijveld, M. M. van Genderen, F. P. Hoeven et al., "Assessment of night vision problems in patients with congenital stationary night blindness," *PLoS One*, vol. 8, no. 5, Article ID e62927, 2013.
- [82] K. W. Littink, M. M. van Genderen, R. W. J. Collin et al., "A novel homozygous nonsense mutation in *CABP4* causes congenital cone-rod synaptic disorder," *Investigative Ophthalmology & Visual Science*, vol. 50, pp. 2344–2350, 2009.
- [83] I. Audo, A. G. Robson, G. E. Holder, and A. T. Moore, "The Negative ERG: Clinical Phenotypes and Disease Mechanisms of Inner Retinal Dysfunction," *Survey of Ophthalmology*, vol. 53, no. 1, pp. 16–40, 2008.
- [84] A. Vincent and E. Heon, "Outer retinal structural anomaly due to frameshift mutation in *CACNA1F* gene," *Eye (London, England)*, vol. 26, no. 9, pp. 1278–1280, 2012.
- [85] R. W. Chen, J. P. Greenberg, M. A. Lazow et al., "Autofluorescence imaging and spectral-domain optical coherence tomography in incomplete congenital stationary night blindness and comparison with retinitis pigmentosa," *American Journal of Ophthalmology*, vol. 153, no. 1, article no. e142, pp. 143–154, 2012.
- [86] S. Michalakis, L. Shaltiel, V. Sothilingam et al., "Mosaic synaptopathy and functional defects in *Cav1.4* heterozygous mice and human carriers of *CSNB2*," *Human Molecular Genetics*, vol. 23, no. 6, pp. 1538–1550, 2014.
- [87] A. Hemara-Wahanui, S. Berjukow, C. I. Hope et al., "A *CACNA1F* mutation identified in an X-linked retinal disorder shifts the voltage dependence of *Cav1.4* channel activation," *Proceedings of the National Academy of Sciences of the United States of America*, vol. 102, no. 21, pp. 7553–7558, 2005.
- [88] D. M. Waldner, N. C. Giraldo Sierra, S. Bonfield et al., "Cone dystrophy and ectopic synaptogenesis in a *Cacna1f* loss of function model of congenital stationary night blindness (*CSNB2A*)," *Channels (Austin, Tex.)*, vol. 12, no. 1, pp. 17–33, 2018.
- [89] J. Wei and G. P. Hemmings, "A further study of a possible locus for schizophrenia on the X chromosome," *Biochemical and Biophysical Research Communications*, vol. 344, no. 4, pp. 1241–1245, 2006.
- [90] K. A. Wycisk, C. Zeitz, S. Feil et al., "Mutation in the auxiliary calcium-channel subunit *CACNA2D4* causes autosomal recessive cone dystrophy," *American Journal of Human Genetics*, vol. 79, no. 5, pp. 973–977, 2006.
- [91] J. Hauke, A. Schild, A. Neugebauer et al., "A Novel Large In-Frame Deletion within the *CACNA1F* Gene Associates with a Cone-Rod Dystrophy 3-Like Phenotype," *PLoS One*, vol. 8, no. 10, Article ID e76414, 2013.
- [92] R. Jalkanen, M. Mänlyjärvi, R. Tobias et al., "X linked cone-rod dystrophy, *CORDX3*, is caused by a mutation in the *CACNA1F* gene," *Journal of Medical Genetics*, vol. 43, no. 8, pp. 699–704, 2006.
- [93] A. Vincent, T. Wright, M. A. Day, C. A. Westall, and E. Heon, "A novel p.Gly603Arg mutation in *CACNA1F* causes Åland island eye disease and incomplete congenital stationary night blindness phenotypes in a family," *Molecular Vision*, vol. 17, pp. 3262–3270, 2011.
- [94] R. Jalkanen, N. T. Bech-Hansen, R. Tobias et al., "A novel *CACNA1F* gene mutation causes Åland Island eye disease," *Investigative Ophthalmology & Visual Science*, vol. 48, no. 6, pp. 2498–2502, 2007.
- [95] M. Nakamura, S. Ito, C. H. Piao, H. Terasaki, and Y. Miyake, "Retinal and optic disc atrophy Associated with a *CACNA1F* mutation in a Japanese family," *Archives of Ophthalmology (Chicago, Ill. : 1960)*, vol. 121, no. 7, pp. 1028–1033, 2003.
- [96] N. Lodha, C. M. Loucks, C. Beaulieu, J. S. Parboosingh, and N. T. Bech-Hansen, "Congenital stationary night blindness: Mutation update and clinical variability," *Advances in Experimental Medicine and Biology*, vol. 723, pp. 371–379, 2012.
- [97] J. B. Peloquin, R. Rehak, C. J. Doering, and J. E. McRory, "Functional analysis of congenital stationary night blindness type-2 *CACNA1F* mutations F742C, G1007R, and R1049W," *Neuroscience*, vol. 150, no. 2, pp. 335–345, 2007.
- [98] J. C. Hoda, F. Zagherro, A. Koschak, and J. Striessnig, "Congenital stationary night blindness type 2 mutations S229P, G369D, L1068P, and W1440X alter channel gating or functional expression of $Ca_v1.4$ L-type Ca^{2+} channels," *The Journal of Neuroscienceneuroscience : the Official Journal of the Society for Neuroscience*, vol. 25, no. 1, pp. 252–259, 2005.
- [99] L. E. Maquat, "Nonsense-mediated mRNA decay: splicing, translation and mRNP dynamics," *Nature Reviews Molecular Cell Biology*, vol. 5, no. 2, pp. 89–99, 2004.
- [100] H. Regus-Leidig, J. Atorf, A. Feigenspan et al., "Photoreceptor degeneration in two mouse models for congenital stationary night blindness type 2," *PLoS One*, vol. 9, no. 1, Article ID e86769, 2014.
- [101] C. I. Hope, D. M. Sharp, A. Hemara-Wahanui et al., "Clinical manifestations of a unique X-linked retinal disorder in a large New Zealand family with a novel mutation in *CACNA1F*, the gene responsible for *CSNB2*," *Clinical & Experimental Ophthalmology*, vol. 33, no. 2, pp. 129–136, 2005.
- [102] F. Mansergh, N. C. Orton, J. P. Vessey et al., "Mutation of the calcium channel gene *Cacna1f* disrupts calcium signaling, synaptic transmission and cellular organization in mouse retina," *Human Molecular Genetics*, vol. 14, no. 20, pp. 3035–3046, 2005.
- [103] D. Specht, S. B. Wu, P. Turner et al., "Effects of presynaptic mutations on a postsynaptic *Cacna1s* calcium channel colocalized with mGluR6 at mouse photoreceptor ribbon synapses," *Investigative Ophthalmology & Visual Science*, vol. 50, no. 2, pp. 505–515, 2009.
- [104] N. Lodha, S. Bonfield, and N. C. Orton, "Congenital stationary night blindness in mice - a tale of two *Cacna1f* mutants," *Advances in Experimental Medicine and Biology*, vol. 664, pp. 549–558, 2010.
- [105] D. Knoflach, K. Schicker, M. Glösmann, and A. Koschak, "Gain-of-function nature of *Cav1.4* L-type calcium channels alters firing properties of mouse retinal ganglion cells," *Channels (Austin, Tex.)*, vol. 9, no. 5, pp. 298–306, 2015.
- [106] S. Jia, A. Muto, W. Orisme et al., "Zebrafish *cacna1fa* is required for cone photoreceptor function and synaptic ribbon formation," *Human Molecular Genetics*, vol. 23, no. 11, pp. 2981–2994, 2014.
- [107] P. Busquet, N. K. Nguyen, E. Schmid et al., "*CaV1.3* L-type Ca^{2+} channels modulate depression-like behaviour in mice independent of deaf phenotype," *The International Journal of Neuropsychopharmacology/Official Scientific Journal of the Collegium Internationale Neuropsychopharmacologicum (CINP)*, vol. 13, no. 4, pp. 499–513, 2010.

- [108] M. A. McCall, N. S. Peachey, and R. G. Gregg, *Eye, Retina, and Visual System of the Mouse*, R. W. Williams and L. M. Chalupa, Eds., MIT Press, 2007.
- [109] J. An, L. Wang, Q. Guo, L. Li, Xia F., and Z. Zhang, "Behavioral phenotypic properties of a natural occurring rat model of congenital stationary night blindness with *Cacnalf* mutation," *Journal of Neurogenetics*, vol. 26, no. 3-4, pp. 363–373, 2012.
- [110] Y. Gu, L. Wang, J. Zhou et al., "A naturally-occurring mutation in *Cacnalf* in a rat model of congenital stationary night blindness," *Molecular Vision*, vol. 14, pp. 20–28, 2008.
- [111] J. An, L. Zhang, B. Jiao et al., "*Cacnalf* gene decreased contractility of skeletal muscle in rat model with congenital stationary night blindness," *Gene*, vol. 562, no. 2, pp. 210–219, 2015.
- [112] B. Chang, J. R. Heckenlively, P. R. Bayley et al., "The *nob2* mouse, a null mutation in *Cacnalf*: Anatomical and functional abnormalities in the outer retina and their consequences on ganglion cell visual responses," *Visual Neuroscience*, vol. 23, no. 1, pp. 11–24, 2006.
- [113] B. Chang, N. L. Hawes, R. E. Hurd, M. T. Davisson, S. Nusinowitz, and J. R. Heckenlively, "Retinal degeneration mutants in the mouse," *Vision Research*, vol. 42, no. 4, pp. 517–525, 2002.
- [114] M. Yu and N. S. Peachey, "Attenuation of oscillatory potentials in *nob2* mice," *Documenta Ophthalmologica. Advances in Ophthalmology*, vol. 115, no. 3, pp. 173–186, 2007.
- [115] C. J. Doering, R. Rehak, S. Bonfield et al., "Modified Cav1.4 expression in the *Cacnalf^{nob2}* mouse due to alternative splicing of an ETn inserted in exon 2," *PloS One*, vol. 3, no. 7, Article ID e2538, 2008.
- [116] P. R. Bayley and C. W. Morgans, "Rod bipolar cells and horizontal cells form displaced synaptic contacts with rods in the outer nuclear layer of the *nob2* retina," *The Journal of Comparative Neurology*, vol. 500, no. 2, pp. 286–298, 2007.
- [117] M. A. Raven, N. C. Orton, H. Nassar et al., "Early afferent signaling in the outer plexiform layer regulates development of horizontal cell morphology," *The Journal of Comparative Neurology*, vol. 506, no. 5, pp. 745–758, 2008.
- [118] N. Zabouri and S. Haverkamp, "Calcium channel-dependent molecular maturation of photoreceptor synapses," *PloS One*, vol. 8, no. 5, Article ID e63853, 2013.
- [119] Y. Cao, I. Sarria, and K. E. Fehlbauer, "Mechanism for Selective Synaptic Wiring of Rod Photoreceptors into the Retinal Circuitry and Its Role in Vision," *Neuron*, vol. 87, no. 6, pp. 1248–1260, 2015.
- [120] S. Haverkamp, S. Michalakis, E. Claes et al., "Synaptic plasticity in CNGA3^{-/-} mice: Cone bipolar cells react on the missing cone input and form ectopic synapses with rods," *The Journal of Neuroscience: the Official Journal of the Society for Neuroscience*, vol. 26, no. 19, pp. 5248–5255, 2006.
- [121] M. S. Singh, J. Balmer, A. R. Barnard et al., "Transplanted photoreceptor precursors transfer proteins to host photoreceptors by a mechanism of cytoplasmic fusion," *Nature Communications*, vol. 7, Article ID 13537, 2016.
- [122] B. W. Jones, R. L. Pfeiffer, W. D. Ferrell, C. B. Watt, M. Marmor, and R. E. Marc, "Retinal remodeling in human retinitis pigmentosa," *Experimental Eye Research*, vol. 150, pp. 149–165, 2016.