



# Optogenetic interrogation of cell signalling: human neuropsin (hOPN5) represents a potent tool for controlling the Gq pathway with light

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G protein coupled receptors (GPCR) are ubiquitously expressed cell-surface receptors involved in the regulation of most physiological processes. This universality makes GPCR extremely attractive as both research tools and pharmacological targets. Upon stimulation of a particular GPCR, distinct intracellular signalling pathways are activated via heterotrimeric G proteins, themselves divided into four major families, Gi/o, Gq/11, Gs and G12/13.

GPCR signalling is classically modulated by pharmacological agents, lacking spatial and temporal resolution. This prevents the study of cell-type- and compartment-specific signalling on physiologically relevant timescales. Optogenetics can overcome these limitations by combining cell-type-specific genetic targeting with spatiotemporally precise optical stimulation. The first optogenetic studies were performed in 1988 by Khorana and colleagues that heterologously expressed bovine rhodopsin in *Xenopus* oocytes to elicit light-induced inward currents [4]. In 2003, Zemelman et al. optically triggered action potentials in vertebrate neurons co-expressing *Drosophila* arrestin-2, rhodopsin and the  $\alpha$ -subunit of the cognate G protein [10]. However, applications of optogenetically controlled GPCR (opto-GPCR) remained scarce [3, 5], probably due to the complexity of intracellular signalling, difficulty of exogenous expression and the concurrent rise of microbial one-component optogenetics.

Ideally, opto-GPCR should be non-promiscuous and specific for only one G protein signalling pathway. Further, they should be easily switched between G protein activated

and deactivated states and effectively drive downstream signalling at low, non-phototoxic light levels. Recently, opto-GPCR optogenetics was fostered by the identification of novel rhodopsins across different vertebrate and invertebrate species as well as the increasing availability of high-resolution GPCR structures in the active and inactive states.

One interesting candidate for optogenetic applications is the bistable mammalian neuropsin (OPN5) [7, 9] found in neuronal tissue and shown to be involved in photoentrainment and thermogenesis. In a recent ground-breaking study published in *Nature Communications*, Wagdi et al. elegantly showed that human OPN5 (hOPN5) signals specifically through Gq [8] and lacks promiscuity as found in most other Gq-coupled opto-GPCR, including melanopsin [1, 6]. In a separate study, Dai et al. controlled Gq signalling with the chicken orthologue of OPN5 [2], confirming OPN5's utility as a potent optogenetic tool.

To dissect hOPN5 signalling, Wagdi et al. first expressed hOPN5 in HEK cells and determined Gq signalling by IP1 levels, a degradation product of IP3, as well as by semi-quantitative Ca<sup>2+</sup> imaging (Fig. 1). The absence of Gi/o signalling found in most other Gq-coupled opto-GPCR was convincingly shown in HEK293-GIRK cells expressing a G protein coupled inwardly-rectifying potassium channel (GIRK) modulated by both, Gi/o and Gq. The authors then assessed optogenetic control of beating rate and contractility in diverse muscle cells. Light stimulation of embryonic stem cell-derived cardiomyocytes and of intact mouse hearts expressing hOPN5 equally led to positive chronotropic effects, which were completely abolished by application of a specific Gq blocker. hOPN5 activation also caused positive inotropic effects in murine adult cardiomyocytes and directly induced Gq-mediated contractions in small intestine, bladder and uterine smooth muscle cells. Finally, hOPN5 stimulation was employed in a biotechnological application by establishing an all-optical high-throughput screening (HTS) assay on Gq-coupled

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