

RESEARCH ARTICLE

Vascular endothelial growth factor-A promoter polymorphisms, circulating VEGF-A and survival in acute coronary syndromes

Barry R. Palmer^{1,2*}, Melinda A. Paterson¹, Chris. M. Frampton¹, Anna P. Pilbrow¹, Lorraine Skelton¹, Chris J. Pemberton¹, Robert N. Doughty³, Chris J. Ellis³, Richard W. Troughton¹, A. Mark Richards^{1,4}, Vicky A. Cameron¹

1 Department of Medicine, Christchurch Heart Institute, University of Otago Christchurch, Christchurch, New Zealand, **2** School of Health Sciences, College of Health, Massey University, Wellington, New Zealand, **3** Faculty of Medicine and Health Sciences, Department of Medicine, University of Auckland, Auckland, New Zealand, **4** Cardiovascular Research Institute, National University of Singapore, Singapore, Singapore

* b.palmer@massey.ac.nz



OPEN ACCESS

Citation: Palmer BR, Paterson MA, Frampton CM, Pilbrow AP, Skelton L, Pemberton CJ, et al. (2021) Vascular endothelial growth factor-A promoter polymorphisms, circulating VEGF-A and survival in acute coronary syndromes. PLoS ONE 16(7): e0254206. <https://doi.org/10.1371/journal.pone.0254206>

Editor: Andreas Zirlik, Medizinische Universitat Graz, AUSTRIA

Received: July 15, 2020

Accepted: June 22, 2021

Published: July 14, 2021

Copyright: © 2021 Palmer et al. This is an open access article distributed under the terms of the [Creative Commons Attribution License](https://creativecommons.org/licenses/by/4.0/), which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: Data to reproduce the findings in this report are available on reasonable request from the Christchurch Heart Institute database. The data is owned by the Christchurch Heart Institute and requests for data access should be sent to Barbara Neame, CHI Database Manager, Barbara.Neame@cdhb.health.nz.

Funding: This study was supported by grants from the Heart Foundation of New Zealand (HFNZ1417) to BRP, and the Health Research Council of New

Abstract

Background

Development of a competent collateral circulation in established coronary artery disease is cardio-protective. The vascular endothelial growth factor (VEGF) system plays a key role in this process. We investigated the prognostic performance of circulating VEGF-A and three genetic variants in the *VEGFA* gene in a clinical coronary cohort.

Methods and results

The Coronary Disease Cohort Study (CDCS) recruited 2,140 patients, with a diagnosis of acute coronary syndrome (ACS), after admission to Christchurch or Auckland City Hospitals between July 2002 and January 2009. We present data for 1927 patients from the cohort genotyped for three SNPs in the VEGF-A gene, rs699947 (C-2578A), rs2010963 (C405G) and rs3025039 (C936T). Plasma VEGF-A concentrations were assayed in a subgroup (n = 550) of CDCS patients (geometric mean 36.6 [34.7–38.5] pg/ml). VEGF-A levels correlated with patient heart rate at baseline (p = 0.034). None of rs699947, rs3025039, nor rs2010963 genotypes were significantly associated with VEGF-A levels, but rs3025039 genotype was positively associated with collateral vessels perfusion according to the Rentrop classification (p = 0.01) and baseline natriuretic peptide levels (p < 0.05). Survival in the CDCS cohort was independently associated with baseline VEGF-A levels and (in males) with rs699947 genotype.

Conclusions

This study is strongly suggestive that VEGF-A levels have value as a prognostic biomarker in coronary heart disease patients and SNPs in *VEGFA* deserve further investigation as prognostic markers and indicators of angiogenic potential influencing the formation of collateral circulation.

Zealand (HRC 02/152) to AMR. RND and AMR hold the NZ Heart Foundation Chair of Heart Health and Chair of Cardiovascular Studies, respectively. The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: The authors have declared that no competing interests exist.

Introduction

An important process ameliorating adverse outcomes after acute coronary occlusion is the formation of a functional collateral circulation around blocked arteries. Well-developed coronary collateral arteries are associated with improved survival in patients with coronary artery disease [1]. VEGF-A is a key factor in forming new blood vessels (angiogenesis) and collateral circulation (arteriogenesis), mediated by VEGF-A binding to the receptors VEGFR-1 (Flt-1) and VEGFR-2 (KDR) [2]. Polymorphisms in VEGF genes influence the expression of VEGF-A [3–5], and therefore possibly ability to form collateral circulation. A diverse set of single nucleotide polymorphisms (SNPs) at a variety of loci are associated with circulating VEGF-A concentrations [6]. *VEGFA* SNPs have been associated with CHD susceptibility [7,8]. VEGF-A is the prototype member of the VEGF family and was first cloned in 1989 [9]. Four other members of the human VEGF family have been identified: VEGF-B, VEGF-C (also called VEGF-2), VEGF-D, and placental growth factor (PlGF). VEGF-A is a homodimeric 34–42 kDa, heparin-binding glycoprotein with potent angiogenic, mitogenic and vascular permeability-enhancing activities specific for endothelial cells [2]. The well characterized heparin-binding isoform, VEGF165, is the principal effector of VEGF action [10].

VEGF-triggered downstream signaling in the vascular endothelium is mediated by three receptors, VEGFR-1, VEGFR-2, and VEGFR-3 (also known as Flt-4), belonging to the class III subfamily of receptor tyrosine kinases. VEGF-A binds to both VEGFR-1 and VEGFR-2. All three receptors contain seven immunoglobulin-like repeats in their extracellular domain and kinase insert domains in their intracellular region. They regulate VEGF family-mediated vasculogenesis, angiogenesis, and lymphangiogenesis and also mediate neurotrophic activity and regulate hematopoietic development. VEGFR-2 is thought to be the primary inducer of VEGF-mediated blood vessel growth, while VEGFR-3 plays a significant role in VEGF-C and VEGF-D-mediated lymphangiogenesis [11,12].

Anti-VEGF treatment as anti-angiogenic therapy in cancer is associated with an increased risk of adverse effects including MI, hypertension, proteinuria, arterial thromboembolism, cardiac ischemia and dysfunction affecting 1–15% of treated patients [13,14]. Conversely, VEGF-A might play a culprit role in atherogenesis and plaque instability via proinflammatory and angiogenic mechanisms [15,16]. However the contribution of VEGF to atherogenesis has been challenged. A *VEGFA* polymorphism associated with higher *VEGFA* expression was found to be associated with a lower risk of coronary artery disease in an epidemiological study [17]. Plasma levels of VEGF-A are increased in stable post-myocardial infarction (post-MI) patients compared to controls, correlating with inflammatory cytokines, but not atherosclerotic burden [18]. The sustained elevation in *VEGFA* expression during atherogenesis may be secondary, rather than causative, to inflammation and hypoxia in evolving lesions [2]. This study investigated the utility of three VEGF gene variants, previously shown to affect VEGF expression, and baseline plasma VEGF-A levels as prognostic markers in a well-characterized cohort of acute coronary syndromes patients (ACS) over prolonged follow up [19–22].

Materials and methods

Study population—Coronary disease cohort study

The Coronary Disease Cohort Study (CDCS) recruited 2140 patients, with a diagnosis of ACS, after admission to Christchurch or Auckland City Hospitals, New Zealand, between July 2002 and January 2009. Inclusion criteria included ischemic discomfort plus one or more of ECG change (ST-segment depression or elevation of ≥ 0.5 mm, T-wave inversion of ≥ 3 mm in ≥ 3 leads, or left bundle branch block), elevated levels of cardiac markers, a history of coronary

disease, age of ≥ 65 years, and a history of diabetes or vascular disease [23]. Patients with serious co-morbidity (e.g. end-stage renal failure, cancer), that limited their life expectancy to < 3 years, were excluded. Recruitment included a wide spectrum of age, both genders and significant sub-groups with established risk factors for CHD including hypertension and type II diabetes. Plasma for VEGF-A assay was collected at a baseline clinic, a median of 18 days after the index admission for ACS. We collected demographic and clinical data at baseline, including blood pressure, height, weight, ECG, echocardiography, family and personal medical history and medication regimes. Plasma samples were also assayed for natriuretic peptides and other analytes. Patients were followed for a median of 5.04 (0.11–9.49) years. Patients attended follow-up clinics 3–5 months and 12–14 months post-onset of ACS and participants completed questionnaires at 2 and 3 years post-discharge. Ethnicity was self-declared and categorized as Maori/Pacifika (Pacific Islander), European, Other or Unknown. Standardized transthoracic echocardiography was performed at baseline and at each follow-up clinic at Christchurch Hospital and University of Auckland clinics as described previously [19,24]. The study was approved by the New Zealand (NZ) Multi-Region Ethics Committee, retrospectively registered at the Australian New Zealand Clinical Trials Registry (ACTRN12605000431628 on 16 September 2005), and all participating patients provided written, informed consent.

Clinical events

Clinical events were determined from recruitment questionnaires, planned follow-up clinic visits, consultation of patient notes, the NZ Ministry of Health and hospital Patient Management System databases, linked using the National Health Index number for each patient. Survival times were calculated from the date of index admission. The investigation conforms to the principles outlined in the Declaration of Helsinki and Title 45, U.S. Code of Federal Regulations, Part 46.

Analyte measurements

Plasma samples were collected and stored in sealed tubes at -80°C as previously described [24]. VEGF-A was analyzed using a chemiluminescent quantitative sandwich enzyme immunoassay (R&D Systems, Inc. Minneapolis). Assay of baseline plasma samples for sFlt-1 (sVEGFR1) in this cohort has been described elsewhere [24]. Circulating levels of natriuretic peptides were assayed as previously described [25]. Interassay variation ranged from 3.3% (NT-proBNP) to 11.8% (BNP). Levels of VEGF-A at baseline were assayed in baseline plasma samples from 550 CDCS participants with the earliest recruitment dates in the cohort in order to maximize numbers of events on follow up for inclusion in survival analyses (recruited between July 2002 and August 2007).

Evaluation of angiographic data

In the Christchurch subgroup of the cohort, collateral vessels were graded according to the Rentrop classification: 0: no filling of any collateral vessels, 1: filling of side branches of the artery to be perfused by collateral vessels without visualization of the epicardial segment; 2: partial filling of the epicardial artery by collateral vessels; and 3: complete filling of the epicardial artery by collateral vessels [26]. Those performing the grading were blinded to other clinical data. Coronary artery anatomy, severity of coronary stenoses and the myocardium at risk were assessed according to the Brandt score [27] in the same subgroup.

DNA extraction and SNP genotyping

DNA samples were obtained for 2067 CDCS cohort patients. Extraction of genomic DNA for genotyping was performed as described previously [22,24]. DNA samples were genotyped for the rs699947 (C-2578A, assay ID C_8311602_10), rs2010963 (C405G, assay ID C_8311614_10) and rs3025039 (C936T, assay ID C_16198794_10) polymorphisms in *VEGFA* using 5 μ L reaction volumes in 384-well plates with allele-specific TaqMan genotyping probes (ThermoFisher Scientific). Genotyping reactions including 1x Roche LightCycler 480 Probes Master mix and 100ng of genomic DNA were performed in a Roche LC480 (Roche Diagnostics Ltd., Rotkreuz, Switzerland) as described elsewhere [24]. Linkage disequilibrium data as determined for the 3 SNPs is shown in [S1 Table](#). As quality control a random selection of 10% of samples were re-genotyped with 100% concordance with the original genotypes.

Statistical analysis

Univariate analyses were performed to test for associations between SNP genotype and demographics, analyte levels and echocardiographic measurements using χ^2 and ANOVA tests with Bonferroni correction. Skewed data were log-transformed before analysis and geometric means with 95% confidence intervals reported and adjusted for age, and the time between index admission and baseline sampling. The survival of stratified groups was compared using Kaplan-Meier analysis and the log-rank test. Independent associations between genotype and survival were tested using Cox proportional hazards multivariate analysis including the following established predictors: age [28,29], gender [30], previous MI [29], antecedent hypertension [31], β -blocker treatment [32], physical activity [33], and NT-proBNP levels [25]. Multivariate linear regression models were based on covariates showing univariate association with VEGF-A.

Assuming an overall mortality of 32% in CDCS, the study had 90% power to detect a hazard ratio (HR) >1.46 as statistically significant (two tailed $\alpha < 0.05$) for analysis of rs699947 genotypes (MAF = 0.491) and for rs3025039 (MAF = 0.140) >90% power to detect an odds ratio of 1.60. For patients assayed for VEGF-A there was 90% power to detect a difference of 10 pg/ml as statistically significant (two tailed $\alpha < 0.05$, assuming SD = 35). Ethnicity was self-declared and categorized as Maori/Pacific Islander, European, Other or Unknown, or in some analyses European versus Non-European. An additive genetic model was used unless stated otherwise. All analyses were performed using SPSS version 25 (IBM, Armonk, USA). There was no imputation of missing data with approximately 1% of cases missing measures utilised in the multivariable Cox regression analyses of the VEGF SNP genotypes. The baseline characteristics of the $n = 550$ cohort with VEGF-A plasma samples are compared with the remainder of the study group. Unadjusted p-values without correction for multiple testing are reported. Statistical significance was set at the 5% level ($p < 0.05$).

Results

VEGF SNP genotypes and CDCS cohort data

Baseline characteristics of the CDCS cohort are summarized in [Table 1](#). Proportions of self-declared Ethnicity were found to be Maori/Pacific (6.3%), European (87.2%), Other (3.6%) or Unknown (2.9%) (see [Table 1](#)). Genotypes were obtained for 1927 patients from the CDCS cohort for all three SNPs, rs3025039 (C936T), rs2010963 (C405G) and rs699947 (C-2578A) in *VEGFA*. Genotype frequencies were rs3025039: CC, 73.9%; CT, 24.2% and TT, 1.9% (minor allele frequency [MAF] = 0.140); rs2010963 GG, 46.7%; GC 43.6% and CC 9.7% (MAF = 0.315); rs699947 CC 26.2%; CA 49.4% and AA 24.4% (MAF = 0.491) and all genotypes

Table 1. Baseline characteristics of the CDCS cohort.

Baseline characteristics	n	Mean \pm SE or n (%)
Male Gender	1927	1378 (71.5%)
Index event diagnosis: Unstable Angina	1927	516 (26.8%)
ST-elevation MI	1927	427 (22.2%)
Non-ST-elevation MI	1927	984 (51.1%)
Age at baseline (years)	1927	66.7 \pm 0.28
Ethnicity (European, Maori & Pasifika, Other, Unknown)	1927	87.2%,6.3%,3.6%,2.9%
Previous MI	1909	569 (29.5%)
Previous Heart Failure	1914	183 (9.5%)
Antecedent Hypertension	1907	999 (51.8%)
Type II diabetes	1918	312 (16.2%)
Renal disease	1908	190 (9.9%)
BMI (kg/m ²)	1907	27.4 \pm 0.11
Tobacco Use (never smoked)	1923	703 (36.5%)
Alcohol Use (non-drinker)	1921	494 (25.6%)
LVEF	1857	57.4% \pm 0.28
Discharge Medications		
ACE inhibitor	1927	1098 (57.0%)
β -blocker	1927	1680 (87.2%)
Diuretic	1927	528 (27.4%)
Statin	1927	1706 (88.5%)
Dual antiplatelet therapy	1927	1006 (52.3%)

<https://doi.org/10.1371/journal.pone.0254206.t001>

conformed to the Hardy-Weinberg equilibrium ($p \geq 0.769$). MAF did not differ between ethnic groups ($p \geq 0.280$). Baseline patient characteristics for each genotype group are shown in Table 2 (rs699947) and S3 & S4 Tables (rs3025039 and rs2010963). Patients with genotype rs699947 AA were physically active less frequently than other genotype groups (Table 2). Scoring of angiograms from patients in the cohort for collateral circulation using the Rentrop grading system revealed that rs3025039 was significantly associated with Rentrop score ($n = 587$ with data for both), with patients with TT genotype having higher scores than the CC or CT genotype groups ($p = 0.032$) (S2 Table). No significant associations with baseline characteristics and other SNP genotypes were observed.

VEGFA SNP genotypes and analyte measurements

Baseline plasma samples from 550 CDCS patients were assayed for VEGF-A (geometric mean 36.8 ± 1.46 pg/ml). Baseline characteristics of this group are summarized and compared to the remainder of the cohort in S2 Table, some patient characteristics differed ($p < 0.05$) between these groups including age and ethnicity. Patients with above-median VEGF-A levels were older (mean age: above median 70.5 ± 0.94 , below median 67.5 ± 1.04 years, $p = 0.038$), had greater waist measurements (mean: above median 95.9 ± 1.08 , below median 92.0 ± 1.00 cm, $p = 0.008$) and trended towards greater BMI (mean: above median 27.8 ± 0.40 , below median 26.8 ± 0.38 , $p = 0.059$). VEGF-A levels were weakly correlated with patient heart rate at baseline ($n = 512$, $r = 0.094$, $p = 0.034$). None of the genotypes for rs699947, rs3025039 or rs2010963 were significantly associated with VEGF-A levels. Baseline BNP ($n = 1894$; CC 17.4 ± 1.02 , CT 16.7 ± 1.04 , TT 13.2 ± 1.13 pmol/L, $p = 0.012$) and NT-proBNP levels ($n = 1908$; CC 77.3 ± 1.03 , CT 75.4 ± 1.05 ; TT 60.7 ± 1.18 pmol/L, $p = 0.038$) adjusted for age and time to plasma sampling, were significantly associated with rs3025039 genotype.

Table 2. Baseline characteristics, drug treatment and neurohormonal data for CDCS patients stratified by VEGF SNP genotype group.

VEGFA C-2578A rs699947 Genotype							
	n	CC	n	CA	n	AA	p
Age (years) [§]	50530	66.0±0.55	949	66.8±0.40	469	67.2±0.57	0.257
Male Gender	505	349 (69.1%)	951	684 (71.9%)	471	342 (72.6%)	0.411
BMI (kg/m ²) [§]	495	27.6±0.24	938	27.3±0.15	463	27.9±0.24	0.138
Physical Activity [‡]	466	1,17.8%; 2, 14.2%; 3, 11.8%; 4,56.2%; 3; 3	886	1,22.0%; 2,10.3%; 3,13.5%; 4,54.2%	436	1,22.7%; 2,13.8%; 3,16.7%; 4, 46.8%	0.010
LVEF	492	57.7±0.57	914	57.4±0.39	451	57.1±0.60	0.743
History							
Previous Myocardial Infarction [§]	502	143 (28.5%)	943	271 (28.7%)	464	155 (33.4%)	0.149
Hypertension [§]	502	263 (52.4%)	942	485 (51.5%)	463	251 (54.2%)	0.630
Diabetes [§]	504	82 (16.3%)	948	147 (15.5%)	466	83 (17.8%)	0.544
Renal Disease [§]	501	49 (9.8%)	943	92 (9.8%)	464	49 (10.6%)	0.883
Alcohol (Non-Drinkers) [§]	505	123 (24.4%)	951	257 (27.0%)	471	114 (24.2%)	0.002
Analytes							
Plasma Creatinine ^{§§} (mmol/l)	490	94.1 (91.7–96.5)	919	94.1 (92.5–95.7)	457	94.5 (92.4–96.8)	0.322
BNP (pmol/L) ^{§§}	501	16.2 (15.0–17.6)	941	16.8 (15.9–17.9)	468	18.4 (17.0–20.0)	0.080
NT-proBNP (pmol/L) ^{§§}	501	73.8 (67.2–81.0)	941	74.7 (69.6–80.0)	468	83.9 (76.0–92.6)	0.108
sFlt-1 (pg/mL)	121	109 (101–118)	236	105 (99.1–111)	136	106 (98.1–115)	0.710
VEGF-A (pg/mL)	131	37.8(33.6–42.6)	264	36.7 (34.1–39.5)	155	35.4 (32.2–38.8)	0.658
Discharge Medications							
ACE inhibitor [§]	505	301 (59.6%)	949	541 (57.0%)	469	254 (54.2%)	0.230
β-blocker [§]	505	443 (87.7%)	949	835 (88.0%)	469	399 (85.1%)	0.279
Diuretic [§]	505	146 (28.9%)	949	254 (26.8%)	469	127 (27.1%)	0.672
Statin [§]	505	452 (89.5%)	949	839 (88.4%)	469	412 (87.8%)	0.704
Amiodarone	505	21 (4.2%)	952	51 (5.4%)	471	24 (5.1%)	0.530
Clopidogrel	505	278 (55.0%)	952	511 (53.7%)	472	231 (49.0%)	0.190
Angiographic measures							
Rentrop score	140	0.51±0.07	315	0.47±0.04	131	0.53±0.07	0.310
Brandt score	264	3.31±0.19	526	3.37±0.14	231	3.32±0.21	0.821
Vessel Disease	264	2.09±0.06	529	1.99±0.04	233	2.09±0.06	0.305
Median Follow-Up (years) [§]	505	5.13 (0.14–9.48)	949	5.23 (0.11–9.49)	469	5.19 (0.14–9.49)	

[§]Means (SEM) or occurrence (percentage);

^{§§}Geometric mean (95% confidence interval);

[§]Median (range).

[‡]Score of 1 = sedentary, 2 = <30 minutes activity on >2 days/week, 3 = ≥30 minutes on 2 days/week, 4 = ≥30 minutes on ≥3 days/week.

<https://doi.org/10.1371/journal.pone.0254206.t002>

Clinical outcome in the CDCS cohort

Patients with above-median VEGF-A levels were more likely to die during the follow-up period (all-cause mortality: above median 34.7%, below median 24.4%, $p < 0.001$, events = 161) (Fig 1). High VEGF-A levels were associated with higher mortality due to cardiovascular causes (CVD mortality: above median 36.6%, below median 16.7%, $p < 0.001$, events = 70). The subgroup of the cohort with both high VEGF-A and NT-proBNP levels had the greatest mortality, but only just significantly more than those with above median NT-proBNP, but low VEGF-A ($p = 0.04$) (S1 Fig). VEGF-A level at baseline was also an independent predictor of mortality in Cox proportional hazards models, both with a minimal set of covariates (Table 3),

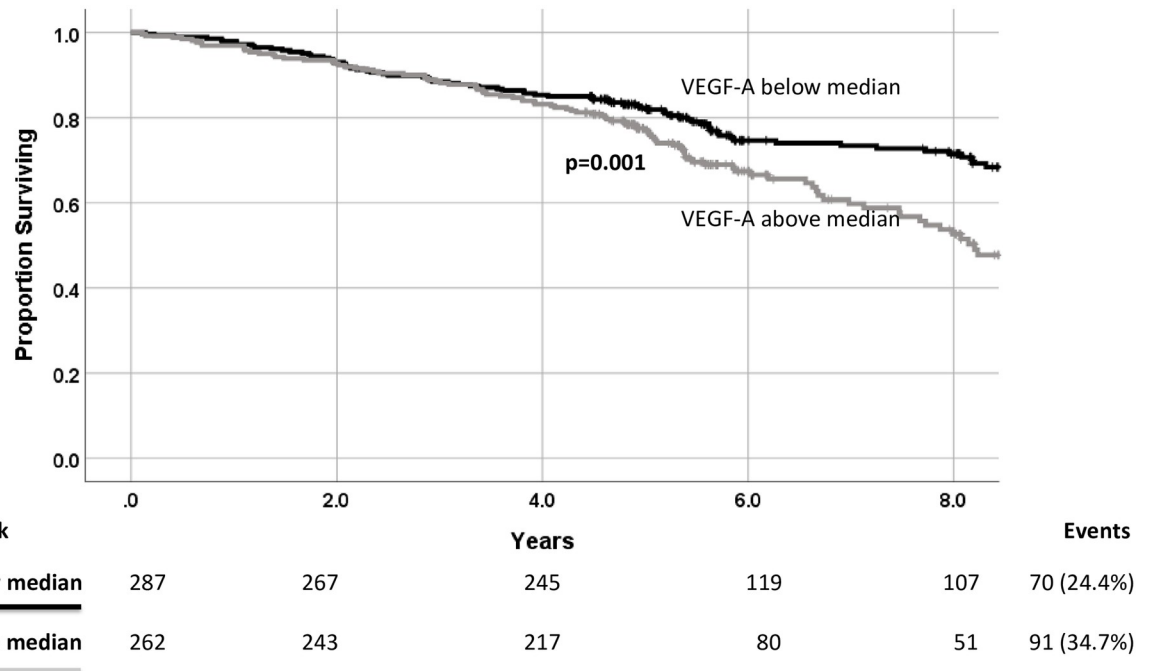


Fig 1. Kaplan-Meier survival plot for survival versus all-cause death in the CDCS cohort stratified by above and below median baseline VEGF-A levels.

<https://doi.org/10.1371/journal.pone.0254206.g001>

including NT-proBNP and sFlt-1, and the same covariates as used to assess rs699947 as a predictor. ROC analysis showed that baseline plasma VEGF-A was a poorer predictor of survival at 5 years when compared to NT-proBNP and sFlt-1 (Fig 2).

Survival in the male subgroup of the CDCS cohort was significantly associated with rs699947 genotype in a multivariate Cox proportional hazards model of survival including rs699947 genotype and diabetic status (Table 4). This analysis revealed that the association of rs699947 genotype with mortality in males was dependent on an interaction with diabetic status. Mortality was associated with genotype ($p = 0.008$) in male patients without type II

Table 3. Cox’s proportional hazards regression model including baseline VEGF-A and sFlt-1 levels for mortality in the CDCS cohort (n = 458, 139 deaths).

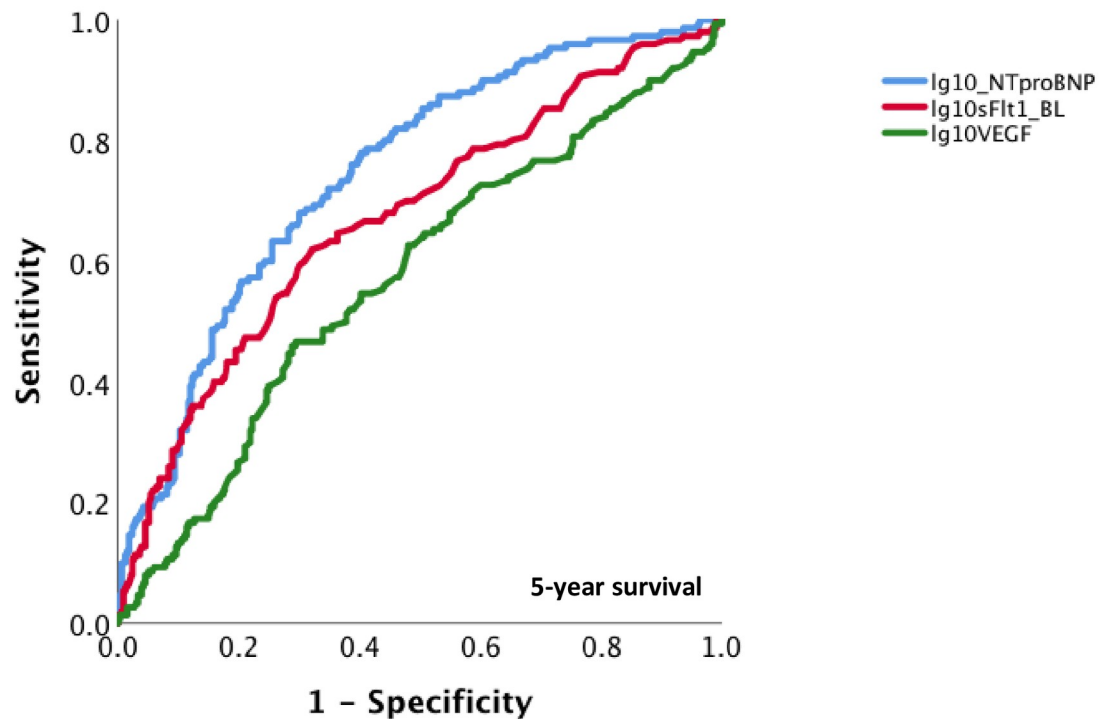
	Coefficient	SE	Wald	Significance	Hazard Ratio	95% CI for HR	
						Lower	Upper
Age at index admission	0.04	0.01	10.7	0.001	1.04	1.02	1.07
Log ₁₀ NT-proBNP at baseline [§]	0.73	0.27	7.45	0.006	2.08	1.23	3.51
Log ₁₀ VEGF-A at baseline [§]	0.77	0.30	6.62	0.010	2.16	1.20	3.89
Log ₁₀ sFlt-1 at baseline [§]	1.13	0.47	5.85	0.016	3.10	1.24	7.74
Physical Activity (scale 1–4) ^{§§}	-0.35	0.07	24.8	<0.001	0.70	0.61	0.81
Previous Myocardial Infarction	0.54	0.18	8.66	0.003	1.71	1.20	2.44
Atrial Fibrillation at baseline	0.22	0.11	4.03	0.045	1.24	1.01	1.54
Cockcroft-Gault—eGFR	-0.01	0.004	11.6	0.001	0.99	0.98	0.99

[§]Hazard Ratio represents the change in risk for every 10-fold increase in analyte level.

^{§§}Score of 1 = sedentary, 2 = <30 minutes activity on >2 days/week, 3 = ≥30 minutes on 2 days/week, 4 = ≥30 minutes on ≥3 days/week.

Abbreviations: CI = confidence interval, eGFR = estimated glomerular filtration rate, HR = hazard ratio, MI = myocardial infarction, NT-proBNP = amino-terminal pro-b-type natriuretic peptide, sFlt-1 = soluble fms-like tyrosine kinase-1.

<https://doi.org/10.1371/journal.pone.0254206.t003>



Analyte	AUC	95% CI for AUC	p-value
VEGF-A at baseline	0.572 ± 0.029	0.516-0.628	P=0.011
sFlt-1 at baseline	0.672 ± 0.027	0.619-0.725	p<0.001
NT-proBNP at baseline	0.745 ± 0.024	0.699-0.792	p<0.001

Fig 2. Receiver-operator curve analysis of VEGF-A, sFlt-1 and NT-proBNP as predictors of survival at 5 years in the CDCS cohort.

<https://doi.org/10.1371/journal.pone.0254206.g002>

diabetes, but not in those with diabetes (in whom mortality was higher overall than in non-diabetic participants) ($p = 0.722$) (Fig 3). Survival in the female patients from the CDCS cohort was not associated with rs699947 genotype ($p = 0.731$), even when stratified by diabetic status ($p > 0.176$). No other significant associations between SNPs and clinical outcomes were observed.

Discussion

The characterization of genetic polymorphisms of the *VEGFA* gene that influence the gene's expression [3,4] and the central role of VEGF-A in blood vessel formation have made the gene an object of interest in studies of susceptibility to, and progression of, cardiovascular disease [34,35]. We found one of three SNPs assayed, rs699947, was an independent predictor of mortality in male non-diabetic participants in the CDCS cohort. Allele frequencies for all three SNPs genotyped in this study agreed closely with those reported for the CEU and GBR populations [36]. Reports of gender-specific expression of human VEGF-A are available [37], but not to our knowledge rs699947 x gender-specific associations with heart pathophysiology.

Table 4. Cox's proportional hazards regression model for mortality in male patients from the CDCS cohort (n = 1215, 249 deaths).

	Coefficient	SE	Wald	Significance	Hazard Ratio	95% CI for HR	
						Lower	Upper
Age at index admission	0.06	0.01	46.4	<0.001	1.06	1.05	1.08
Log ₁₀ NT-proBNP at baseline [§]	1.35	0.22	39.1	<0.001	3.86	2.3	5.90
ACE inhibitor at discharge	0.15	0.15	0.01	0.920	0.99	0.74	1.32
Amiodarone treatment at discharge	0.69	0.30	5.33	0.021	2.00	1.11	3.61
β-blocker treatment at discharge	0.30	0.21	2.02	0.155	1.35	0.89	2.03
Statin treatment at discharge	0.13	0.19	0.46	0.499	1.13	0.79	1.63
Physical Activity (scale 1–4) ^{§§}	-0.17	0.06	8.86	0.003	0.84	0.75	0.94
Previous Myocardial Infarction	0.46	0.15	10.0	0.002	1.58	1.19	2.11
Previous Heart Failure	0.25	0.18	1.88	0.170	1.29	0.90	1.85
Antecedent Hypertension	0.14	0.14	1.07	0.302	1.15	0.88	1.51
Previous Cancer	0.07	0.17	0.16	0.694	1.07	0.77	1.49
Previous renal disease	0.55	0.17	10.4	0.001	1.73	1.24	2.42
Previous pulmonary disease	0.39	0.16	5.65	0.017	1.47	1.07	2.02
Atrial fibrillation	0.45	0.16	7.55	0.006	1.57	1.14	2.16
LVEF	0.00	0.01	0.00	0.986	1.00	0.99	1.01
NYHA score			9.58	0.023			
I v II	0.37	0.18	4.17	0.041	0.69	0.49	0.99
I v III	0.15	0.19	0.63	0.428	1.16	0.80	1.68
I v IV	0.24	0.21	1.30	0.25	1.27	0.84	1.90
BMI	0.004	0.02	0.04	0.845	1.00	0.97	1.04
Time to sampling	0.01	0.01	3.19	0.074	1.01	1.00	1.02
Ethnicity European v non-European	-0.26	0.26	1.02	0.313	0.77	0.46	1.28
Type II Diabetes	0.44	0.18	5.91	0.015	1.56	1.09	2.22
rs699947 genotype CC/CA v AA	0.80	0.34	5.60	0.018	2.23	1.15	4.35
Type II Diabetes x rs699947 genotype	0.92	0.37	6.22	0.013	2.52	1.22	5.20

[§]Hazard Ratio represents the change in risk for every 10-fold increase in BNP level.

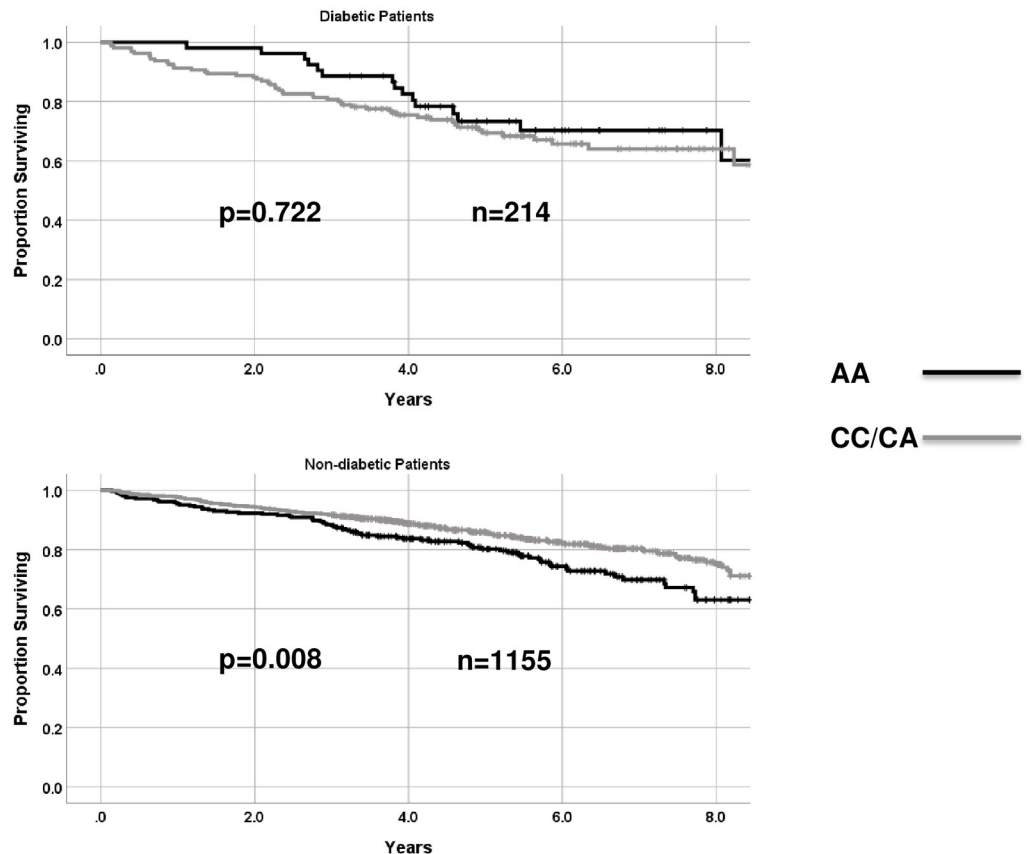
^{§§}Score of 1 = sedentary, 2 = <30 minutes activity on >2 days/week, 3 = ≥30 minutes on 2 days/week, 4 = ≥30 minutes on ≥3 days/week.

Abbreviations: BMI = body mass index, CI = confidence interval, HR = hazard ratio, LVEF = left ventricular ejection fraction, NT-proBNP = amino-terminal pro-b-type natriuretic peptide, NYHA = New York Heart Association.

<https://doi.org/10.1371/journal.pone.0254206.t004>

VEGF-A release and angiogenesis has been shown to be stimulated by estradiol in human mesenchymal stem cells, that have the potential to differentiate into myocytes [38]. There is at least one other report of a SNP associated with VEGF-A expression interacting with metabolic syndrome status [39]. There are some reports that VEGF-A expression may be altered in diabetes [40], and diabetes and SNP genotype appear to interact to influence angiogenesis [41,42], which would fit with our observations. It would seem that the influence of rs699947 in normal VEGF angiogenic function is deranged in diabetes, and potentially more so in males than females, leading to greater plaque formation and instability. This hypothesis requires testing in more detailed studies.

The mean levels of VEGF-A we measured in our cohort were very similar or slightly elevated compared to those reported in healthy controls [43,44]. Above median levels of VEGF-A were found to be associated with mortality. This is in agreement with others [45]. While it has been hypothesized that the VEGF pathway is protective through its promotion of angiogenesis, and therefore countering the ischemic effects of coronary atherosclerosis, evidence exists that excess VEGF expression contributes to atherosclerotic plaque formation [2,16]. This suggests



Patients at Risk		Events				
No diabetes	AA 286	264	198	97	37	72 (25.1%)
	CC/CA 869	820	590	322	90	146 (16.8%)
Diabetes	AA 53	52	40	19	7	14 (26.4%)
	CC/CA 161	142	99	46	14	50 (31.1%)

Fig 3. Kaplan-Meier survival versus all-cause death in male patients from the CDCS cohort for a) non-diabetic patients and b) type II diabetic patients stratified by rs699947 genotype CC/CA v AA.

<https://doi.org/10.1371/journal.pone.0254206.g003>

high levels of VEGF-A may be associated with elevated risk, perhaps by the mechanism of exacerbating plaque instability. Alternatively, high VEGF-A levels may represent a restorative, but inadequate, response to tissue ischemia from myocardial tissue with inadequate perfusion. There are reports of correlation of levels of VEGF-A and inflammatory markers [18], suggesting similar phenotypic profiles for effectors of the VEGF and inflammatory systems in patients with post-acute CHD. Mortality due to non-cardiovascular causes did not appear to be elevated in the subgroup of the cohort with high VEGF-A levels, as might be expected due to the role of VEGF-A in tumour angiogenesis [46]. The findings that high levels of VEGF-A are associated with increased mortality in this cohort are consistent with rs699947 having been previously implicated in the pathogenesis of CHD [47] and being a predictor independent of NT-proBNP and sFlt-1 suggests a complex regulatory network, although natriuretic peptides and VEGFA may respond to similar signalling is suggested by S1 Fig.

We also found that rs3025039 was significantly associated with Rentrop score, a measure of collateral vessel development. While the rs3025039 TT genotype group had higher levels of VEGF-A, but not significantly so, as others have found [48,49]. Other reports have also suggested a link between VEGF gene variants and angiographic measurements in CVD [50,51]. Those studies focused on stent restenosis and carotid artery stenosis, whereas our measurements examined the extent of coronary vessel disease and the degree to which collateral vessels were perfused. The polymorphisms that had significant clinical association (rs3025039 and rs699947) may be altering expression of VEGF-A at the cardiac tissue level, but significant SNP-associated differences were not detectable in circulating levels in this cohort due to factors associated with differential response to the coronary event and varying medication regimes.

Limitations of the study include: 1) Missing data for some parameters limited the power of this study to explore their association with genotype and VEGF levels; 2) The study cohort was dominated by ethnic Europeans and the results cannot be extrapolated to other populations with any certainty; 3) Blood samples were collected at varying times after the index event in order to avoid major influences from the acute event on plasma analytes, while this variable may have affected levels of these analytes, adjustment for time to sampling was included in statistical analysis in an effort to mitigate this; 4) The assay for VEGF-A was only conducted on a minority of the total CDCS cohort, which differed in age and ethnic profile from the remainder of the cohort, these covariates were included in multivariate analyses to correct for this; 5) The CDCS cohort was recruited over 10 years ago and therefore limited availability of recent treatment regimes, such as dual anti-platelet therapy (received by 54% of the CDCS cohort) may have affected clinical outcome endpoints.

Conclusion

In summary, we report associations between plasma levels of VEGF-A, some of its genetic variants, patient characteristics and outcome measures. This, along with our recent report of similar findings with levels of sFlt-1, the soluble version of the VEGFR1 receptor [24], indicates that both plasma analytes and genetic markers from the VEGF system may have prognostic value in patients with cardiovascular disease. These observations suggest further investigation in larger, more diverse cohorts would be of value.

Supporting information

S1 Fig. Kaplan-Meier survival plot for survival versus all-cause death in the CDCS cohort stratified by groups formed by the combination of median splits for baseline VEGF-A and NT-proBNP levels.

(TIFF)

S1 Table. Linkage disequilibrium data (R^2 data) for SNPs genotyped in this study.

(PDF)

S2 Table. Baseline characteristics of the CDCS cohort stratified by whether patient samples were assayed for VEGF-A or not.

(PDF)

S3 Table. CDCS cohort patient characteristics stratified by rs3025039 genotype.

(PDF)

S4 Table. CDCS cohort patient characteristics stratified by rs2010963 genotype.

(PDF)

Acknowledgments

The authors thank participants in the study, Christchurch Heart Institute Translational Biodiscovery unit staff for the hormone and biochemical assays, and study coordinators of the Christchurch Heart Institute and Auckland Cardiovascular Research Group for assistance with the recruitment and follow-up of the patients.

Author Contributions

Conceptualization: Barry R. Palmer, Robert N. Doughty, A. Mark Richards, Vicky A. Cameron.

Data curation: Barry R. Palmer, Chris. M. Frampton, Anna P. Pilbrow, Lorraine Skelton, Vicky A. Cameron.

Formal analysis: Barry R. Palmer, Chris. M. Frampton, Anna P. Pilbrow.

Funding acquisition: Barry R. Palmer, Robert N. Doughty, A. Mark Richards, Vicky A. Cameron.

Investigation: Barry R. Palmer, Melinda A. Paterson, Chris J. Pemberton, Robert N. Doughty, Chris J. Ellis, Richard W. Troughton, A. Mark Richards.

Methodology: Melinda A. Paterson, Anna P. Pilbrow, Lorraine Skelton, Chris J. Pemberton, Robert N. Doughty, Chris J. Ellis, Richard W. Troughton, A. Mark Richards, Vicky A. Cameron.

Project administration: Barry R. Palmer, Lorraine Skelton, Robert N. Doughty, A. Mark Richards.

Resources: Chris J. Pemberton.

Supervision: Barry R. Palmer, Anna P. Pilbrow, Lorraine Skelton, Chris J. Pemberton, Robert N. Doughty, Chris J. Ellis, Richard W. Troughton, A. Mark Richards, Vicky A. Cameron.

Writing – original draft: Barry R. Palmer, Anna P. Pilbrow, A. Mark Richards, Vicky A. Cameron.

Writing – review & editing: Barry R. Palmer, Melinda A. Paterson, Chris. M. Frampton, Anna P. Pilbrow, Lorraine Skelton, Chris J. Pemberton, Robert N. Doughty, Chris J. Ellis, Richard W. Troughton, A. Mark Richards, Vicky A. Cameron.

References

1. Meier P, Gloekler S, Zbinden R, Beckh S, de Marchi SF, Zbinden S, et al. Beneficial effect of recruitable collaterals: a 10-year follow-up study in patients with stable coronary artery disease undergoing quantitative collateral measurements. *Circulation*. 2007; 116(9):975–83. <https://doi.org/10.1161/CIRCULATIONAHA.107.703959> PMID: 17679611.
2. Yla-Herttuala S, Rissanen TT, Vajanto I, Hartikainen J. Vascular endothelial growth factors: biology and current status of clinical applications in cardiovascular medicine. *Journal of the American College of Cardiology*. 2007; 49(10):1015–26. <https://doi.org/10.1016/j.jacc.2006.09.053> PMID: 17349880.
3. Watson CJ, Webb NJ, Bottomley MJ, Brenchley PE. Identification of polymorphisms within the vascular endothelial growth factor (VEGF) gene: correlation with variation in VEGF protein production. *Cytokine*. 2000; 12(8):1232–5. <https://doi.org/10.1006/cyto.2000.0692> PMID: 10930302.
4. Renner W, Kotschan S, Hoffmann C, Obermayer-Pietsch B, Pilger E. A common 936 C/T mutation in the gene for vascular endothelial growth factor is associated with vascular endothelial growth factor plasma levels. *Journal of vascular research*. 2000; 37(6):443–8. <https://doi.org/10.1159/000054076> PMID: 11146397.

5. Pare-Brunet L, Glubb D, Evans P, Berenguer-Llergo A, Etheridge AS, Skol AD, et al. Discovery and functional assessment of gene variants in the vascular endothelial growth factor pathway. *Human mutation*. 2014; 35(2):227–35. Epub 2013/11/05. <https://doi.org/10.1002/humu.22475> PMID: 24186849.
6. Choi SH, Ruggiero D, Sorice R, Song C, Nutile T, Vernon Smith A, et al. Six Novel Loci Associated with Circulating VEGF Levels Identified by a Meta-analysis of Genome-Wide Association Studies. *PLoS Genet*. 2016; 12(2):e1005874. Epub 2016/02/26. <https://doi.org/10.1371/journal.pgen.1005874> PMID: 26910538.
7. Han X, Liu L, Niu J, Yang J, Zhang Z. Association between VEGF polymorphisms (936c/t, -460t/c and -634g/c) with haplotypes and coronary heart disease susceptibility. *Int J Clin Exp Pathol*. 2015; 8(1):922–7. Epub 2015/03/11. PMID: 25755796.
8. Kalayi Nia S, Ziaee S, Boroumand MA, Sotudeh Anvari M, Pourgholi L, Jalali A. The impact of vascular endothelial growth factor +405 C/G polymorphism on long-term outcome and severity of coronary artery disease. *J Clin Lab Anal*. 2017; 31(4). Epub 2016/10/06. <https://doi.org/10.1002/jcla.22066> PMID: 27704620.
9. Leung DW, Cachianes G, Kuang WJ, Goeddel DV, Ferrara N. Vascular endothelial growth factor is a secreted angiogenic mitogen. *Science (New York, NY)*. 1989; 246(4935):1306–9. Epub 1989/12/08. <https://doi.org/10.1126/science.2479986> PMID: 2479986.
10. Staton CA, Kumar I, Reed MW, Brown NJ. Neuropilins in physiological and pathological angiogenesis. *The Journal of pathology*. 2007; 212(3):237–48. <https://doi.org/10.1002/path.2182> PMID: 17503412.
11. Koch S, Claesson-Welsh L. Signal transduction by vascular endothelial growth factor receptors. *Cold Spring Harb Perspect Med*. 2012; 2(7):a006502. Epub 2012/07/05. <https://doi.org/10.1101/cshperspect.a006502> PMID: 22762016.
12. Ku DD, Zaleski JK, Liu S, Brock TA. Vascular endothelial growth factor induces EDRF-dependent relaxation in coronary arteries. *Am J Physiol*. 1993; 265(2 Pt 2):H586–92. Epub 1993/08/01. <https://doi.org/10.1152/ajpheart.1993.265.2.H586> PMID: 8368362.
13. Faruque LI, Lin M, Battistella M, Wiebe N, Reiman T, Hemmelgarn B, et al. Systematic review of the risk of adverse outcomes associated with vascular endothelial growth factor inhibitors for the treatment of cancer. *PloS one*. 2014; 9(7):e101145. Epub 2014/07/06. <https://doi.org/10.1371/journal.pone.0101145> PMID: 24988441.
14. Abdel-Qadir H, Ethier JL, Lee DS, Thavendiranathan P, Amir E. Cardiovascular toxicity of angiogenesis inhibitors in treatment of malignancy: A systematic review and meta-analysis. *Cancer Treat Rev*. 2017; 53:120–7. Epub 2017/01/21. <https://doi.org/10.1016/j.ctrv.2016.12.002> PMID: 28104567.
15. Matsumoto T, Mugishima H. Signal transduction via vascular endothelial growth factor (VEGF) receptors and their roles in atherogenesis. *J Atheroscler Thromb*. 2006; 13(3):130–5. Epub 2006/07/13. <https://doi.org/10.5551/jat.13.130> PMID: 16835467.
16. Inoue M, Itoh H, Ueda M, Naruko T, Kojima A, Komatsu R, et al. Vascular endothelial growth factor (VEGF) expression in human coronary atherosclerotic lesions: possible pathophysiological significance of VEGF in progression of atherosclerosis. *Circulation*. 1998; 98(20):2108–16. Epub 1998/11/17. <https://doi.org/10.1161/01.cir.98.20.2108> PMID: 9815864.
17. Howell WM, Ali S, Rose-Zerilli MJ, Ye S. VEGF polymorphisms and severity of atherosclerosis. *Journal of medical genetics*. 2005; 42(6):485–90. <https://doi.org/10.1136/jmg.2004.025734> PMID: 15937083.
18. ErZen B, Silar M, Sabovic M. Stable phase post-MI patients have elevated VEGF levels correlated with inflammation markers, but not with atherosclerotic burden. *BMC Cardiovasc Disord*. 2014; 14:166. Epub 2014/11/25. <https://doi.org/10.1186/1471-2261-14-166> PMID: 25417001.
19. Palmer BR, Jarvis MD, Pilbrow AP, Ellis KL, Frampton CM, Skelton L, et al. Angiotensin-converting enzyme 2 A1075G polymorphism is associated with survival in an acute coronary syndromes cohort. *American Heart Journal* 2008; 156(4):752–8. <https://doi.org/10.1016/j.ahj.2008.06.013> PMID: 18926157
20. Ellis KL, Pilbrow AP, Frampton CM, Doughty RN, Whalley GA, Ellis CJ, et al. A Common Variant at Chromosome 9P21.3 Is Associated With Age of Onset of Coronary Disease but Not Subsequent Mortality. *Circ Cardiovasc Genet*. 2010; 3(3):286–93. <https://doi.org/10.1161/CIRCGENETICS.109.917443> PMID: 20400779
21. Palmer BR, Frampton CM, Skelton L, Yandle TG, Doughty RN, Whalley GA, et al. KCNE5 polymorphism rs697829 is associated with QT interval and survival in acute coronary syndromes patients. *J Cardiovasc Electrophysiol*. 2012; 23(3):319–24. Epub 2011/10/12. <https://doi.org/10.1111/j.1540-8167.2011.02192.x> PMID: 21985337.
22. Palmer BR, Slow S, Ellis KL, Pilbrow AP, Skelton L, Frampton CM, et al. Genetic polymorphism rs6922269 in the MTHFD1L gene is associated with survival and baseline active vitamin B12 levels in post-acute coronary syndromes patients. *PloS one*. 2014; 9(3):e89029. Epub 2014/03/13. <https://doi.org/10.1371/journal.pone.0089029> PMID: 24618918.

23. Ellis KL, Pilbrow AP, Frampton CM, Doughty RN, Whalley GA, Ellis CJ, et al. A Common Variant at Chromosome 9P21.3 Is Associated with Age of Onset of Coronary Disease but Not Subsequent Mortality. *Circ Cardiovasc Genet*. 2010; 3(3):286–93. <https://doi.org/10.1161/CIRCGENETICS.109.917443> PMID: 20400779.
24. Marks ECA, Wilkinson TM, Frampton CM, Skelton L, Pilbrow AP, Yandle TG, et al. Plasma levels of soluble VEGF receptor isoforms, circulating pterins and VEGF system SNPs as prognostic biomarkers in patients with acute coronary syndromes. *BMC Cardiovasc Disord*. 2018; 18(1):169. <https://doi.org/10.1186/s12872-018-0894-1> PMID: 30111293.
25. Richards AM, Nicholls MG, Espiner EA, Lainchbury JG, Troughton RW, Elliott J, et al. B-type natriuretic peptides and ejection fraction for prognosis after myocardial infarction. *Circulation*. 2003; 107(22):2786–92. <https://doi.org/10.1161/01.CIR.0000070953.76250.B9> PMID: 12771003.
26. Rentrop KP, Cohen M, Blanke H, Phillips RA. Changes in collateral channel filling immediately after controlled coronary artery occlusion by an angioplasty balloon in human subjects. *Journal of the American College of Cardiology*. 1985; 5(3):587–92. [https://doi.org/10.1016/s0735-1097\(85\)80380-6](https://doi.org/10.1016/s0735-1097(85)80380-6) PMID: 3156171.
27. Brandt PWT, Partridge JB, Wattie WJ. Coronary Arteriography—Method of Presentation of Arteriogram Report and a Scoring System. *Clin Radiol*. 1977; 28(4):361–5. [https://doi.org/10.1016/s0009-9260\(77\)80140-2](https://doi.org/10.1016/s0009-9260(77)80140-2) PMID: 872502
28. Michaels AD, Goldschlager N. Risk stratification after acute myocardial infarction in the reperfusion era. *Progress in cardiovascular diseases*. 2000; 42(4):273–309. <https://doi.org/10.1053/pcad.2000.0420273> PMID: 10661780.
29. Eagle KA, Lim MJ, Dabbous OH, Pieper KS, Goldberg RJ, Van de Werf F, et al. A validated prediction model for all forms of acute coronary syndrome: estimating the risk of 6-month postdischarge death in an international registry. *Jama*. 2004; 291(22):2727–33. <https://doi.org/10.1001/jama.291.22.2727> PMID: 15187054.
30. Blum A, Blum N. Coronary artery disease: Are men and women created equal? *Gen Med*. 2009; 6(3):410–8. <https://doi.org/10.1016/j.genm.2009.09.005> PMID: 19850237.
31. Richards AM, Nicholls MG, Troughton RW, Lainchbury JG, Elliott J, Frampton C, et al. Antecedent hypertension and heart failure after myocardial infarction. *Journal of the American College of Cardiology*. 2002; 39(7):1182–8. [https://doi.org/10.1016/s0735-1097\(02\)01737-0](https://doi.org/10.1016/s0735-1097(02)01737-0) PMID: 11923044
32. Gundersen T, Grottnum P, Pedersen T, Kjekshus JK. Effect of timolol on mortality and reinfarction after acute myocardial infarction: prognostic importance of heart rate at rest. *The American journal of cardiology*. 1986; 58(1):20–4. [https://doi.org/10.1016/0002-9149\(86\)90234-1](https://doi.org/10.1016/0002-9149(86)90234-1) PMID: 3524181.
33. Chow CK, Jolly S, Rao-Melacini P, Fox KA, Anand SS, Yusuf S. Association of diet, exercise, and smoking modification with risk of early cardiovascular events after acute coronary syndromes. *Circulation*. 121(6):750–8. <https://doi.org/10.1161/CIRCULATIONAHA.109.891523> PMID: 20124123.
34. Ma WQ, Wang Y, Han XQ, Zhu Y, Liu NF. Association of genetic polymorphisms in vascular endothelial growth factor with susceptibility to coronary artery disease: a meta-analysis. *BMC medical genetics*. 2018; 19(1):108. Epub 2018/07/06. <https://doi.org/10.1186/s12881-018-0628-3> PMID: 29973139.
35. Zhao X, Meng L, Jiang J, Wu X. Vascular endothelial growth factor gene polymorphisms and coronary heart disease: a systematic review and meta-analysis. *Growth Factors*. 2018; 36(3–4):153–63. Epub 2018/10/16. <https://doi.org/10.1080/08977194.2018.1477141> PMID: 30317903.
36. Machiela MJ, Chanock SJ. LDlink: a web-based application for exploring population-specific haplotype structure and linking correlated alleles of possible functional variants. *Bioinformatics*. 2015; 31(21):3555–7. Epub 2015/07/04. <https://doi.org/10.1093/bioinformatics/btv402> PMID: 26139635.
37. Mani C, Kochhar P, Ravikumar G, Dwarkanath P, Sheela CN, George S, et al. Placental expression of ENG, VEGF, and FLT: Gender-specific associations with maternal vitamin B12 status. *European journal of clinical nutrition*. 2019. Epub 2019/06/19. <https://doi.org/10.1038/s41430-019-0449-2> PMID: 31209272.
38. Mihai MC, Popa MA, Suica VI, Antohe F, Jackson EK, Simionescu M, et al. Mechanism of 17beta-estradiol stimulated integration of human mesenchymal stem cells in heart tissue. *Journal of molecular and cellular cardiology*. 2019; 133:115–24. Epub 2019/06/16. <https://doi.org/10.1016/j.yjmcc.2019.06.007> PMID: 31201797.
39. Ghazizadeh H, Avan A, Fazilati M, Azimi-Nezhad M, Tayefi M, Ghasemi F, et al. Association of rs6921438 A<G with serum vascular endothelial growth factor concentrations in patients with metabolic syndrome. *Gene*. 2018; 667:70–5. Epub 2018/05/08. <https://doi.org/10.1016/j.gene.2018.05.017> PMID: 29733969.
40. Bitar MS, Al-Mulla F. Upregulation of CREM/ICER suppresses wound endothelial CRE-HIF-1alpha-VEGF-dependent signaling and impairs angiogenesis in type 2 diabetes. *Dis Model Mech*. 2015; 8(1):65–80. Epub 2014/11/09. <https://doi.org/10.1242/dmm.017145> PMID: 25381014.

41. Lin TH, Wang CL, Su HM, Hsu PC, Juo SH, Voon WC, et al. Functional vascular endothelial growth factor gene polymorphisms and diabetes: effect on coronary collaterals in patients with significant coronary artery disease. *Clin Chim Acta*. 2010; 411(21–22):1688–93. Epub 2010/07/14. <https://doi.org/10.1016/j.cca.2010.07.002> PMID: 20621071.
42. Gala-Bladzinska A, Czech J, Braun M, Skrzypa M, Gargasz K, Mazur A, et al. Association of 18bp insertion/deletion polymorphism, at -2549 position of VEGF gene, with diabetic vascular complications in type 2 diabetes mellitus. *Adv Med Sci*. 2019; 64(1):137–43. Epub 2019/01/18. <https://doi.org/10.1016/j.advms.2018.08.011> PMID: 30654317.
43. Li H, Kantoff PW, Ma J, Stampfer MJ, George DJ. Prediagnostic plasma vascular endothelial growth factor levels and risk of prostate cancer. *Cancer Epidemiol Biomarkers Prev*. 2005; 14(6):1557–61. Epub 2005/06/09. <https://doi.org/10.1158/1055-9965.EPI-04-0456> PMID: 15941972.
44. Carilho R, de Carvalho M, Swash M, Pinto S, Pinto A, Costa J. Vascular endothelial growth factor and amyotrophic lateral sclerosis: the interplay with exercise and noninvasive ventilation. *Muscle Nerve*. 2014; 49(4):545–50. Epub 2013/07/23. <https://doi.org/10.1002/mus.23955> PMID: 23868282.
45. Eaton CB, Gramling R, Parker DR, Roberts MB, Lu B, Ridker PM. Prospective association of vascular endothelial growth factor-A (VEGF-A) with coronary heart disease mortality in southeastern New England. *Atherosclerosis*. 2008; 200(1):221–7. Epub 2008/02/12. <https://doi.org/10.1016/j.atherosclerosis.2007.12.027> PMID: 18261732.
46. Matsumoto K, Ema M. Roles of VEGF-A signalling in development, regeneration, and tumours. *J Biochem*. 2014; 156(1):1–10. Epub 2014/05/20. <https://doi.org/10.1093/jb/mvu031> PMID: 24839295.
47. Cui QT, Li Y, Duan CH, Zhang W, Guo XL. Further evidence for the contribution of the vascular endothelial growth factor gene in coronary artery disease susceptibility. *Gene*. 2013; 521(2):217–21. Epub 2013/04/03. <https://doi.org/10.1016/j.gene.2013.03.091> PMID: 23545315.
48. Dong PP. Association of vascular endothelial growth factor expression and polymorphisms with the risk of gestational diabetes mellitus. *J Clin Lab Anal*. 2019; 33(2):e22686. Epub 2018/10/24. <https://doi.org/10.1002/jcla.22686> PMID: 30350881.
49. Al-Habboubi HH, Sater MS, Almawi AW, Al-Khateeb GM, Almawi WY. Contribution of VEGF polymorphisms to variation in VEGF serum levels in a healthy population. *Eur Cytokine Netw*. 2011; 22(3):154–8. Epub 2011/10/11. <https://doi.org/10.1684/ecn.2011.0289> PMID: 21982816.
50. Osadnik T, Strzelczyk JK, Regula R, Bujak K, Fronczek M, Gonera M, et al. The Relationships between Polymorphisms in Genes Encoding the Growth Factors TGF-beta1, PDGFB, EGF, bFGF and VEGF-A and the Restenosis Process in Patients with Stable Coronary Artery Disease Treated with Bare Metal Stent. *PloS one*. 2016; 11(3):e0150500. Epub 2016/03/02. <https://doi.org/10.1371/journal.pone.0150500> PMID: 26930482.
51. Yadav BK, Yadav R, Chang H, Choi K, Kim JT, Park MS, et al. Genetic Polymorphisms rs699947, rs1570360, and rs3025039 on the VEGF Gene Are Correlated with Extracranial Internal Carotid Artery Stenosis and Ischemic Stroke. *Ann Clin Lab Sci*. 2017; 47(2):144–55. Epub 2017/04/27. PMID: 28442515.