An update on the roles of immune system-derived microRNAs in cardiovascular diseases

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emerging field of cardioimmunology has demonstrated how cells of the immune system play vital roles in the path- ogenesis of CVD. MicroRNAs (miRNAs) are critical regulators of cellular identity and function. Cell-intrinsic, as well as cell-extrinsic, roles of immune and inflammatory cell-derived miRNAs have been, and continue to be, exten- sively studied. Several 'immuno-miRNAs' appear to be specifically expressed or demonstrate greatly enriched ex- pression within leucocytes. Identification of miRNAs as critical regulators of immune system signalling pathways has posed the question of whether and how targeting these molecules therapeutically, may afford opportunities for dis- ease treatment and/or management. As the field of cardioimmunology rapidly continues to advance, this review dis- cusses findings from recent human and murine studies which contribute to our understanding of how leucocytes of innate and adaptive immunity are regulated—and may also regulate other cell types, via the actions of the miRNAs they express, in the context of CVD. Finally, we focus on available information regarding miRNA regulation of regu- latory T cells and argue that targeted manipulation of miRNA regulated pathways in these cells may hold therapeu- tic promise for the treatment of CVD and associated risk factors.

Keywords

Heart • Cardiovascular disease • Cardioimmunology • Immunology • MicroRNA • TREGS

1. Introduction

Cardiovascular disease (CVD) is the leading cause of pre-mature death with ${\sim}17.8$ million deaths worldwide in 2017.¹ With approximately half a billion people afflicted by CVD globally, it also constitutes a major so-cioeconomic burden, accruing substantial direct costs to healthcare systems as well as indirect costs of reduced productivity and days of work lost to illness.¹

The heart and vasculature are primarily composed of nonhaematopoietically derived cell types including cardiomyocytes, fibroblasts, endothelial cells (ECs), smooth muscle cells, pericytes, and stroma. However, the presence of leucocytes within cardiovascular tissue has long been acknowledged and all the major immune cell subtypes, including lymphocyte and myeloid-derived populations, are found within cardiovascular tissues.^{2,3} Some of these populations are resident cells, while infiltrating immune cells contribute to changes in the structural and functional properties of the heart and vasculature and may amplify CVD progression. Over the last two decades, the broad roles played by cells of the immune response in cardiovascular biology have begun to be appreciated. These range from roles in heart development, to maintenance of vascular homoeostasis and steady state physiology through to critical roles in directing and protecting from disease-related pathology. Interested readers are directed to specialized recent reviews on the field, provided by Swirski and Nahrendorf² and Strassheim *et al.*⁴

Understanding the critical regulators of immune cell activity and function is key to appreciate how immune responses which underlie or exacerbate certain diseases might be corrected to cure disease. MicroRNAs (miRNAs) are among the most important regulators of immune cellular function and identity. This review aims to provide an up-to-date discussion of available literature, which investigates immune cell-derived

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miRNAs and their roles in regulating immune responses in the setting of CVD and its associated risk factors. We additionally highlight putative roles played by immune system-derived miRNAs delivered to non-immune cardiovascular cell types via extracellular vesicles (EVs). Finally, we discuss miRNA regulation of regulatory T cells (T_{REGS}) and suggest that the manipulation of miRNA pathways in these cells may prove beneficial in the context of T_{REGS} immunotherapy and CVD. A reference for abbreviations used throughout this manuscript can be found in Table 1.

2. Immune system enriched miRNAs: roles in immune cell development and function

MicroRNAs play essential roles in governing immune system development, homoeostasis, and activation.⁵ The canonical pathways of miRNA biogenesis are well recognized but more recently, non-canonical biogenesis pathways and novel miRNA regulatory mechanisms have also been described (*Figure 1*). Dysregulated expression of miRNAs during disease settings including CVD, autoimmunity, and tumourigenesis has been extensively reported,^{12,13} and alterations to the expressions of many of these miRNAs may serve useful roles as biomarkers for diagnostic purposes and/or for monitoring disease progression.¹³ Many individual miRNAs highly expressed by immune system cells—the so-called 'immuno-miRNAs', play unique roles in directing immune system responses and additional novel players are often described. However, some of these serve particularly critical roles and warrant a brief introduction.

2.1 miRNA-17-92 cluster

The cluster encodes six unique miRNAs: miR-17, miR-18a, miR-19a, miR-20a, miR-19b-1, and miR-92a-1 and plays multiple, non-redundant roles in B cell development, supportive of its actions as an oncogene promoting B cell lymphomas.¹⁴ Other critical roles include regulation of T cell activation, maturation and development of T-helper subsets (Th) and T_{REGS}, and additional important functions in monocyte differentiation to macrophages and macrophage activation pathways.¹⁵

2.2 miRNA-21

Expressed in multiple cell types in addition to leukocytes, up-regulation of miR-21 (referring to the predominantly expressed mature strand miR-21-5p) is associated with exposure to inflammatory stimuli and immune cell activation.¹⁶ Additionally, miR-21 functions as an oncogene and its overexpression results in B cell lymphoma in mice,¹⁷ supporting the association of miR-21 with the development of many cancers.¹⁸ Aging is associated with increased expression of miR-21 in T cells, resulting in failure to generate memory T cells, contributing to aging-related immune system dysfunction (immunosenescence).¹⁹

2.3 miRNA-142

miRNA-142 is a highly evolutionarily conserved miRNA, widely considered to be haematopoietic-lineage specific or enriched.^{20,21} Expression of mature isoforms of this miRNA from both arms of the miR-142 precursor (miR-142-3p and miR-142-5p) has been extensively validated. Multiple roles in the development of haematopoietic lineages and immune system function are attributed to these molecules.^{22–28} The regulation of lineage-enriched expression of miR-142 isoforms may be related to the association of *MIR142* promotor activity with super-

2.4 miRNA-146a

miRNA-146a is a powerful negative regulator of both innate and adaptive immune responses.^{32–34} Expression of miR-146a is driven by the NF-kB signalling pathway³⁵ and is correlated with immune cell activation via T cell receptor signalling or Toll-like receptor activation. Critically, miR-146a-5p functions as a negative feedback regulator, down-regulating NF-kB signalling transducers IRAK1 and TRAF6 to limit cellular activation and its own expression levels.^{33,34}

2.5 miRNA-155

miRNA-155 is a master regulator of immune system responses. Mice deficient for miR-155 have severely defective lymphoid and myeloid cell development and homoeostasis.^{36,37} Activation of immune cells is additionally promoted by miR-155.³⁸ Often, this has been linked to repression of pathways involving the canonical miR-155-5p target, suppressor of cytokine signalling 1 (SOCS1), a potent inhibitor of Janus kinase (JAK)-signal transducer and activator of transcription (STAT) signalling pathways.^{39–41}

2.6 miRNA-181

Demonstration that miR-181 is highly and specifically expressed in haematopoietic progenitor cells gave one of the first indications that miRNAs might play critical roles in immune cell lineage differentiation.²⁰ Subsequently, miR-181 family members (miR-181a-1, miR-181a-2, miR-181b-1, miR-181b-2, miR-181c, and miR-181d) have been shown to play critical roles in lymphoid and myeloid lineage differentiation and act as a metabolic rheostat for the immune system.^{42–44} Down-regulation of miR-181 family member expression may underlie reduced or dysregulated immune system function associated with aging and immunosenescence.^{45,46}

2.7 miRNA-223

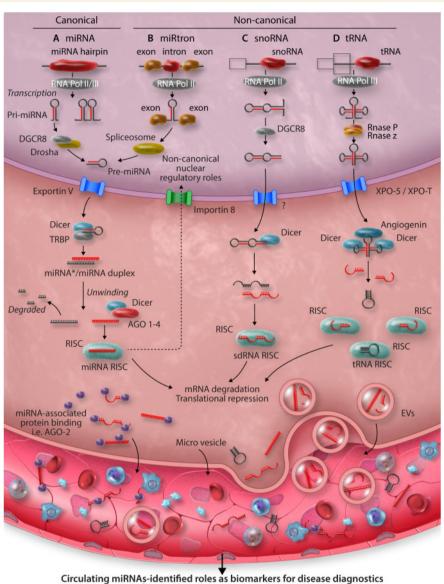
Particularly critical in the regulation of innate immunity,^{47,48} expression of miR-223 within the immune system is restricted largely to myeloidderived cell types, in which miR-223-3p acts as a critical mediator of both myeloid cell differentiation and activation.⁴⁷

3 miRNA-mediated regulation of immunity and CVD risk factors

3.1 Hypertension

While immune cell contribution to hypertension was discovered many years ago, it was not until the mid-2000s that a ground-breaking study by Guzik *et al.* showed that $Rag1^{-/-}$ mice lacking T and B cells were protected against angiotensin II (Ang-II) and deoxycorticosterone acetate salt-induced hypertension. Moreover, the adoptive transfer of T but not B cells was able to restore the hypertensive abnormalities seen in the control groups, suggesting that T cells are critical mediators of hypertension.⁴⁹

In a murine model of Ang-II-induced hypertension, levels of the miR-199/214 family member miR-214-3p were up-regulated eight-fold in perivascular tissue of Ang-II-infused mice compared to wild type (WT)



and / or monitoring progression / severity

Figure 1 Overview of mammalian miRNA biogenesis pathways and functions. (A) Canonically, microRNAs are generated following RNA polymerase II/III dependant transcription as primary transcripts (pri-miRNAs). Cleavage of the pri-miRNA to a stem-loop, pre-miRNA follows within the nucleus, orchestrated by the microprocessor complex comprising the RNAse-III like enzyme, Drosha, and a dsDNA binding co-factor—DiGeorge syndrome chromosomal region 8 (DGCR8). Next, pre-miRNAs are translocated to the cytoplasm via the RAS-related nuclear protein-guanosine-5'-triphosphate-ase (Ran-GTP) dependant, Exportin V nuclear transporter. A final cleavage of pre-miRNA to a duplex containing the mature microRNA sequences is undertaken by Dicer. Dicer also integrates the duplex within an Argonaut protein family member (AGO1-4 in mammals) wherein, one strand of the duplex is typically degraded, while the other is selected to remain AGO-associated, forming the microRNA-induced silencing complex (miRISC). It is the miRISC, which facilitates localization and binding to mRNA target sequences. Finally, following formation of the miRISC: mRNA complex, enzymatic degradation of the mRNA transcript or its direct translational inhibition occurs, epigenetically altering gene expression by modifying the dynamics of protein synthesis. While all mature microRNAs are thought to be generated within the cytoplasm, nuclear import of AGO2 loaded with microRNA can occur in an Importin 8-dependant manner and described nuclear functions of microRNAs includes post-transcriptional gene-silencing, transcriptional gene-silencing, and transcriptional gene activation, via binding to recently transcribed RNAs, or through direct chromatin interactions with gene promotor or enhancer regions.^{6–8} (B) MiRtrons are derived from sequences spliced within the introns of coding genes. In the nucleus, processing of miRtrons bypasses the requirement for conventional Drosha/DGCR8 cleavage. The pre-miRNA is derived from spliceosome activity.^{9,10} (C) Small nucleolar RNAs undergo splicing and debranching in the nucleus before being exported to the cytoplasm via an unknown nuclear transporter. Here, they undergo final cleavage by Dicer before being incorporated into the RISC. (D) RNA polymerase III transcribed tRNA undergo processing by RNases, which cleave the 5' and 3' ends allowing its transport into the cytoplasm via Exportin-t. tRNAs undergo further cleavage by Dicer and Angiogenin before becoming incorporated into the tRNA RISC.¹¹ Mature miRNAs can be released into the circulation via the excretion of EVs, exosomes, budding of microvesicles, or via proteins. In the circulation they can be readily detected in biological fluids, such as blood, urine, and saliva, thus acting as biomarkers of disease, creating unique miRNA signatures in response to disease.

mice. Moreover, under Ang-II infusion, miR-214^{-/-} mice showed blunted hypertension, vascular stiffening, and perivascular fibrosis when compared with WT controls. These effects mapped specifically to changes in the T cell transcriptome in the miR-214 $^{-/-}$ mice, which led to blunted T cell activation and infiltration into perivascular adipose tissue (AT). To assess the translational relevance of this, it was found that plasma miR-214-3p expression levels were increased in hypertensive patients.⁵⁰ In another murine study of Ang-II-induced hypertension, the knockout of miR-31 (miR-31^{-/-}), which maps specifically to T_{reg} and Th_{17} cells led to blunted hypertension and decreased cardiac hypertrophy and fibrosis. Furthermore, miR-31 ablation in mice resulted in a decrease of kidney infiltrating CD45⁺F4/80⁺ macrophages and an increase in kidney specific CD4⁺FoxP3⁺T_{REGS}. As a result, miR-31^{-/-} mice had reduced vascular and renal damage. These differences mapped specifically to miR-31-/mediated post-transcriptional regulation of its direct target protein phosphatase 6c.⁵¹

3.2 Obesity and epicardial AT

Obesity and diabetes are major risk factors for CVD. AT is an endocrine organ of paramount importance in the development of systemic inflammation, which affects cardiovascular cell metabolism, function, and disease progression. Furthermore, the immune system plays critical roles in the development and function of AT and obesity is increasingly considered to be an immunometabolic disease. Thickening of epicardial adipose tissue (EAT), which surrounds the heart and major vessels is associated with coronary artery disease (CAD). In patients with sudden-CAD associated death, levels of miR-34a-3p and miR-34a-5p in EAT were up-regulated.⁵² This is of particular interest as miR-34a has been associated with obesity-induced systemic inflammation via the suppression of antiinflammatory 'M2' macrophage polarization and increase in proinflammatory 'M1' macrophages and cytokines within visceral AT of murine models of diet-induced obesity.⁵³ Furthermore, whole-genome sequencing on EAT from CAD patients, shows that genes involved in antigen presentation, chemokine signalling, and systemic inflammation are up-regulated, whilst levels of miR-103-3p, which regulates Th₂ chemokines, like CCL13, were down-regulated.⁵⁴ Interestingly, the presence of EAT around the left atria (LAEAT) has been linked to the prevalence of atrial fibrillation. Using data from the miRhythm study and bioinformatic platforms, Tran et al. analysed miRNA levels associated with LAEAT from AF patients. Levels of miR-155-5p and miR-302a-3p were associated with LAEAT in AF. Both miRNAs are involved in the regulation of IL-8, which has been linked to leukocyte trafficking and activation.^{55,56} Furthermore, downstream predicted genes of both miRNAs were associated with cardiac hypertrophy and adipogenesis.⁴

4. miRNA-mediated regulation of immune cells during CVD and associated complications

4.1 Atherosclerosis

Atherosclerosis is considered to be an inflammatory, immune-mediated disease. Of the immune cells associated with atherosclerosis pathogenesis, macrophages play a central role in plaque formation and disease progression. As LDL retained beneath the endothelial wall becomes oxidized (oxLDL), it activates ECs and promotes the recruitment and infiltration of monocytes/macrophages. Uptake of oxLDL and internalization of apoptotic cell fragments by macrophages results in lipid accumulation and foam cell (FC) formation. The roles of miRNAs in the regulation of macrophages during atherosclerosis have been extensively investigated.⁵⁷ Recent work has continued to uncover novel miRNAmediated mechanisms of macrophage regulation and their atheroprotective and atherogenic effects (Figure 2). Utilizing publicly available miRNA array data, Zheng et al.⁵⁸ postulated that miR-133b-3p expression in macrophages serves a pro-atherogenic role by modulating expression of mastermind-like protein 1 (MAML1), tentatively supporting studies which highlight a role for NOTCH signalling in plaque leukocyte influx and atherosclerosis progression.⁵⁹⁻⁶¹ However, although the authors convincingly demonstrated regulation of MAML1 by miR-133b-3p, given the well-appreciated role of MAML1 as a transcriptional coactivator of NOTCH signalling, it is not immediately clear how derepression of MAML1 expression via down-regulation of miR-133b-3p might promote the associated down-regulation of various NOTCH pathway components-as implied by the authors. Therefore, further work which clarifies the exact relevance of the miR-133b-3p/MAML1 interaction in macrophages during atherosclerosis is required.

miRNA-17-5p-a potentially useful, circulating biomarker for monitoring atherosclerosis severity,⁶² has also been implicated in the regulation of inflammation and macrophage lipid accumulation during atherosclerosis, through the direct regulation of ATP-binding cassette transporter A1 (ABCA1), a critical component of cellular reverse cholesterol transport/lipid efflux.⁶³ Down-regulation of ABCA1 leads to accumulation of intracellular lipids, promoting FC generation during atherosclerosis. miRNA-17-5p joins several additional miRNAs demonstrated to regulate ABCA1 in macrophages, thereby limiting reverse cholesterol transport,^{57,64} including the miR-33 family.^{65,66} miRNA-33-5p also regulates macrophage expression of peroxisome proliferatoractivated receptor gamma co-activator 1-alpha, a master regulator of mitochondrial biogenesis and energy metabolism-therefore limiting cholesterol efflux via ABCA1 by restricting production of ATP.⁶⁷ Inhibition of cholesterol efflux can promote formation of lipid-rich cell membrane rafts, promoting pro-inflammatory extracellular signalling pathways and inflammation.⁶⁸ Alongside enhanced plaque instability, miR-205-5p was recently shown to direct lipid raft formation in macrophages, promoting pro-inflammatory macrophage phenotypes,⁶⁹ expanding the understanding of miRNA-mediated control of macrophage lipid transport and its importance during atherosclerosis.

Defective macrophage clearance of other apoptotic macrophages and FCs (efferocytosis) may also underlie inflammation and atherosclerosis progression. Down-regulation of miR-378a-3p in aortic macrophages of hyperlipidemic Apoe^{-/-} mice was shown to result in de-repression of signal regulatory protein α (SIRP α),⁷⁰ the transmembrane signalling receptor for CD47 (integrin-associated protein). The CD47: SIRPa axis act as a don't eat me signal and up-regulation of this pathway prevents clearance of apoptotic cells within plaque lesions, accelerating atherosclerosis progression. Dysregulation of the CD47: SIRPa axis could also underlie atherosclerosis progression in humans, as was implied by the observation that the long non-coding RNA (IncRNA) myocardial infarction associated transcript (MIAT) was significantly higher in the circulating sera of atherosclerosis patients and those with symptoms of vulnerable atherosclerotic plaque, compared to controls, indicating that MIAT may also serve as a useful biomarker of plaque stability and atherosclerosis severity.⁷¹ In this study, the authors suggest that MIAT acts as a competing endogenous RNA (ceRNA) for miR-149-5p in macrophages and contains complementary miR-149-5p binding sequences which allow MIAT to 'sponge' miR-149-5p away from CD47, de-repressing CD47 expression. This study appears to convincingly demonstrate a functional role for this

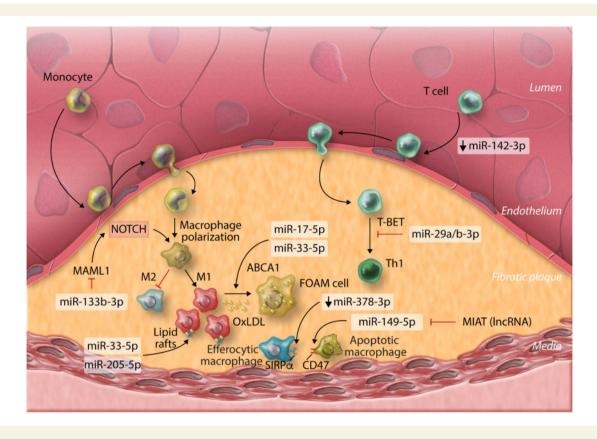


Figure 2 MicroRNAs involved with plaque-associated immune cells in atherosclerosis and their functions. OxLDL, oxidized low-density lipoprotein; lncRNA, long non-coding ribonucleic acid; MIAT, myocardial infarction associated transcript.

interaction, using ceRNA overexpression systems and *in vitro* miRNA manipulation to probe the nature of this dynamic in macrophage cell lines. However, the degree to which ceRNAs truly act to regulate endogenous miRNA activity through modulation of bioavailability for molecular target interaction is controversial (reviewed by Thomson and Dinger⁷²). In their article, the authors do not investigate the limitations that physiological expression levels of both molecules may impose upon the outcome of the interaction during the real-world context of disease. This general criticism applies to many attempts to characterize cell-endogenous physiological regulators of miRNA activity. However, the concept presented in this work highlights the importance of considering the complex components of networks that regulate the functional activity of specific miRNAs in a given cell type.

Additional leukocytes also serve critical roles in the pathophysiology of atherosclerosis. T cells and particularly the CD4⁺ lineage are important players in atherosclerosis pathogenesis and the influx of CD4⁺ T cells to the vascular walls may be regulated by alterations to cytoskeleton dynamics as a result of aberrant expression of miR-142-3p.⁷³ CD4⁺ Th₁ responses, directed by the Th₁ master T-box transcriptional regulators T-BET and (to some extent) Eomesodermin (EOMES) are the most prevalent Th subset in atherosclerotic plaques and promote atherosclerosis progression and severity.^{74–76} miRNA-29 seed-family members miR-29a-3p and miR-29b-3p act to restrict T-BET and EOMES expression in T cells⁷⁷ and may therefore putatively alleviate atherosclerosis and enhance plaque stability by directly inhibiting Th₁ development. However, evidence that global antagonism of miR-29 members using locked nucleic acids (LNAs) decreases atherosclerotic lesion size and promotes plaque remodelling and stability in mice,⁷⁸ suggests that in certain cell types, miR-29 family members may actually contribute to disease progression. However, in this study, the impact of LNA-miR-29 on the development of Th1 responses was not specifically addressed. More studies are required to fully define the contribution of T-cell-derived miR-29 family members to the promotion of plaque-associated Th₁ development, both during human and murine disease, in order to guide potential therapeutic strategies.

4.2 Myocardial infarction

Monocytes and macrophages are critical mediators of inflammation and promote cardiomyocyte apoptosis during the early inflammatory response post-MI, but also contribute to later stages of left ventricular remodelling and dilation, eventually leading to HF.⁷⁹ miRNA-21-5p may be a critical endogenous mediator, which limits the polarization of proinflammatory macrophages in the heart, post-MI. miRNA-21-5p expression was up-regulated rapidly in infarct areas of murine cardiac tissue, however, MI-induction in mice deficient for miR-21 (miR-21^{-/-}) resulted in worse survival rates and greater infarct size and scarring.⁸⁰ Macrophage expression of miR-21-5p directly restricted inflammatory activation and cytokine production by targeting Kelch repeat and BTB (POZ) domain containing 7 (KBTBD7), a transcriptional activator involved in mitogen-activated protein kinase signalling. In the absence of miR-21-5p, KBTBD7 overexpression enhanced macrophage activation by promoting p38 and NF-kB activation.⁸⁰ Promotion of miR-21 regulatory mechanisms through intravenous delivery of nanoparticles (NPs)

containing miR-21-5p mimics, targeted to cardiac macrophages shortly after MI-induction may therefore be a therapeutic strategy to limit pro-inflammatory/promote reparative, wound healing macrophage phenotypes, leading to attenuation of pathological remodelling.⁸¹ Therapeutically targeting miR-375-3p in macrophages may be an additional means of modulating pro-inflammatory macrophage development post-MI, via activation of 3-phosphoinositide-dependent protein kinase 1 and reduction of pro-inflammatory cytokine expression.⁸² Additionally, expression of a superoxide producing enzyme, NADPH oxidase 2 (NOX2) is enhanced in cardiomyocytes and macrophages post-MI⁸³ and promotes macrophage inflammatory polarization and cytokine expression⁸⁴ and is associated with adverse cardiac remodelling post-MI.⁸⁵ In both human and murine macrophages, miR-106b-5p, miR-148b-3p, and miR-204-5p were shown to suppress NOX2 and targeted up-regulation of these miRNAs through pH-sensitive polyketal NP-specific delivery to macrophages significantly improved infarct size and cardiac function post-MI in mice, highlighting another potential therapeutic axis for management of MI⁸⁶ (Figure 3A).

4.3 Myocarditis

Myocarditis is an important cause of sudden cardiac death and heart failure, particularly in young, otherwise healthy patients. A breakdown in tolerogenic immune mechanisms is apparent during myocarditis and may particularly underpin autoimmune-mediated myocarditis. For example, miR-98-5p expression is enhanced in B cells during experimental autoimmune myocarditis (EAM) of mice and acts to suppress regulatory IL-10 expression.⁸⁷ Likewise, polarization of tolerogenic dendritic cells, which promote induction of T_{REGS} may be inhibited in EAM as a result of miR-223-3p down-regulation, de-repressing NLRP3 inflammasome expression.⁸⁸ However, the study of DCs in this work relies entirely on the use of murine bone marrow-derived dendritic cells generated through in vitro culture, potentially limiting conclusions regarding translational relevance in humans. None-the-less, T_{REGS} are generally thought to be cardioprotective during myocarditis while effector CD4⁺ T cells are critical players in myocarditis pathogenesis.⁸⁹ Th₁₇ effectors play a particularly pathogenic role in later or chronic stages of autoimmune-related myocarditis progression and in the progression to dilated cardiomyopathy (DCM), while the balance of Th_{17} to T_{REG} cells also serves to modulate myocarditis severity, with reduced T_{REGS} leading to more profound Th₁₇ responses and more severe disease.⁸⁹ As reported for inflammatory conditions of other solid organs,³⁹ miR-155-5p plays a critical role in regulating Th_{17}/T_{REG} balance during murine EAM, in which up-regulation of miR-155-5p in cardiac CD4⁺ T cells following induction of EAM promotes Th_{17} responses, while inhibiting generation and responses of ${\sf T}_{\sf REGS.}^{}^{90}$ Additional evidence from Coxsackievirus-B3 murine models of viral myocarditis also supports a pathogenic role for up-regulated miR-155-5p in dysfunctional activation of T cells during acute myocarditis, as well as in promoting monocyte/ macrophage cardiac infiltration and inflammatory 'M1' cardiac macrophage polarization.^{91,92} Moreover, up-regulated expression of miR-155-5p, as well as the immune-related miRNAs miR-146b-5p and miR-21-5p were observed in human ventricular septal biopsies of acute viral myocarditis patients.⁹¹ Targeting miR-155-5p expression may therefore hold therapeutic promise for the treatment of myocarditis in certain settings. However, a pathogenic role for miR-155-5p during myocarditis may not be absolute. Natural Killer (NK) cells are innate mediators of viral clearance and are protective in viral myocarditis, with roles in restricting autoimmune inflammation during EAM also reported.⁹³ NK cell activation is positively correlated with expression of miR-155-5p,

and promotes NK cell expression of IFN γ , partially through repression of the inositol phosphatase SHIP1.⁹⁴ During murine viral myocarditis, increased NK expression of miR-155-5p and enhanced NK cell activation is apparent—promoting viral clearance and reducing cardiac immune cell infiltration and inflammatory cytokine production⁹⁵ (*Figure 3B*).

4.4 Cardiac hypertrophy

Leukocyte infiltration into cardiac tissue exacerbates cardiovascular dysfunction in in vivo models of pressure overload-induced hypertrophy. miRNA-155-5p was increased in cardiac biopsies of hypertrophic aortic stenosis patients and correlated negatively with cardiac function yet positively with wall thickness relative to non-hypertrophic controls undergoing coronary bypass artery grafting.⁴¹ Furthermore, in two different murine models of pressure overload, genetic ablation, or pharmacological inhibition of miR-155-5p led to a decrease in cardiac infiltrating leukocytes, particularly CD68⁺CD11b⁺F4/80⁺ monocytes/macrophages. This was accompanied by preserved cardiac function and the absence of hypertrophy, both at the physiological and gene level. These affects were shown to be macrophage specific, as adenoviral-mediated inhibition of miR-155-5p in cardiomyocytes of WT mice did not prevent hypertrophy, whilst in bone marrow adoptive transfer experiments, the presence of miR-155^{-/-} macrophages in WT mice was able to protect against hypertrophy. SOCS1 was the main target of miR-155-5p in macrophages postulated to drive hypertrophy; in the absence of miR-155-5p, SOCS1 overexpression inhibited the JAK/STAT signalling axis and specifically inhibited STAT3 signalling, suppressing the macrophage inflammatory response and release of paracrine mediators driving myocyte hypertrophy.⁴¹ See Figure 3C.

4.5 Heart failure

An ongoing inflammatory response is associated with chronic HF. HF patients have elevated levels of circulating pro-inflammatory cytokines including IL-6, IL-1 β , and TNF α , and recent analysis of the results of the CANTOS phase III clinical trial revealed that use of the anti-IL-1 β monoclonal antibody canakinumab (Ilaris), reduced hospitalizations and mortality related to HF in MI-patients.⁹⁶ However, other phase III trials aimed at modulating HF with anti-inflammatory interventions have had less successful outcomes, suggesting that inflammatory responses may only drive HF in certain patient cohorts.⁹⁷ In DCM patients, CD4⁺ T cell over-activation and proliferation is associated with down-regulation of miR-451a-5p and increased expression of its target Myc,⁹⁸ marking the importance of T cell responses and their regulation by miRNAs in the development of HF. Immunosenescence is also related to the development of HF associated with aging. Ageing-associated miRNAs, such as miR-181 family members—which are important positive regulators of Bcell lymphopoiesis,²⁰ are reduced in immune cells of aged patients⁴⁵ and miR-34a-5p which inhibits B cell maturation and demonstrates enhanced expression in leucocytes with aging^{99,100} may underlie these effects.⁴⁶ Targeting these molecules may mitigate aging-associated HF progression by restoring lymphocyte function. See Figure 3D.

4.6 Vasculitis

Vasculitis covers a highly heterogeneous group of multi-system autoimmune, inflammatory conditions that affect blood vessels. Although rare, complications of vasculitides can include systemic emboli, thrombosis, aneurysms, sepsis, vessel occlusion, and major end-organ damage. In Wegener's granulomatosis—the most common form of anti-neutrophil cytoplasmic autoantibody-associated vasculitis, accumulation of effector CD4⁺ T cells and a T-helper profile skewed towards IL-17 and IL-23

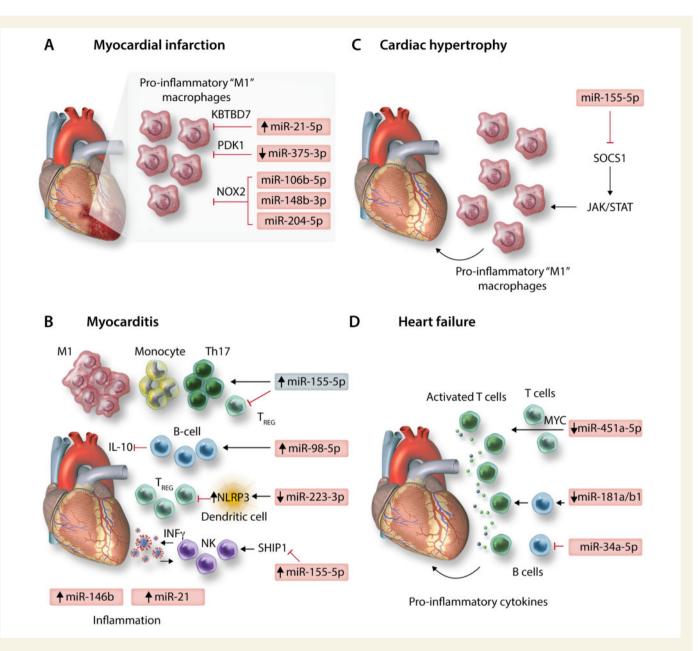


Figure 3 MicroRNAs involved in immune-regulation of diseases of the heart: depicting (A) myocardial infarction, (B) myocarditis, (C) cardiac hypertrophy, and (D) heart failure.

producing Th₁₇ cells^{101–103} alongside a reduction in T_{REGS} suppressive capacity^{104,105} is thought to critically regulate disease pathogenesis. In this setting, aberrantly up-regulated expression of miR-142-3p may underlie defective T_{REGS} suppressive activity, potentially driving pathogenic effector T cell responses. miRNA expression profiling of a circulating memory-like population of T_{REGS} found to be enriched within the CD4⁺ T cell pool of patients relative to healthy controls revealed miR-142-3p to be significantly up-regulated alongside decreased expression of the miR-142-3p target adenylate cyclase 9 (ADCY9) and a corresponding reduction in intracellular cyclic adenosine monophosphate (cAMP)¹⁰⁶— a central mediator of T_{REGS} activity alongside aberrant changes in T_{REGS} miRNA expression has also been reported in Kawasaki's disease (KD)—an acute form of systemic vasculitis, which can affect the coronary arteries. The proportion of circulating T_{REGS} and T_{REGS} expression

of markers of maturation and suppressive function were significantly lower in KD patients (aged 3–54 months), while miR-155-5p and miR-21-5p expression was reduced and miR-31-5p expression was increased significantly.¹⁰⁷ Putatively, these miRNA expression changes during KD lead to reduced proportion and activity of T_{REGS} through de-repression of the miR-155-5p target SOCS1-inhibiting STAT5 signalling critical for T_{REGS} homoeostasis and function, and restricted expression of the T_{REGS} master transcription factor FOXP3.^{107,108}

5. Immune cell-derived EVs and miRNAs—implications for CVD pathogenesis

EVs are a heterogeneous group of lipid-bilayer encapsulated particles, naturally shed into the cellular microenvironment from the cell

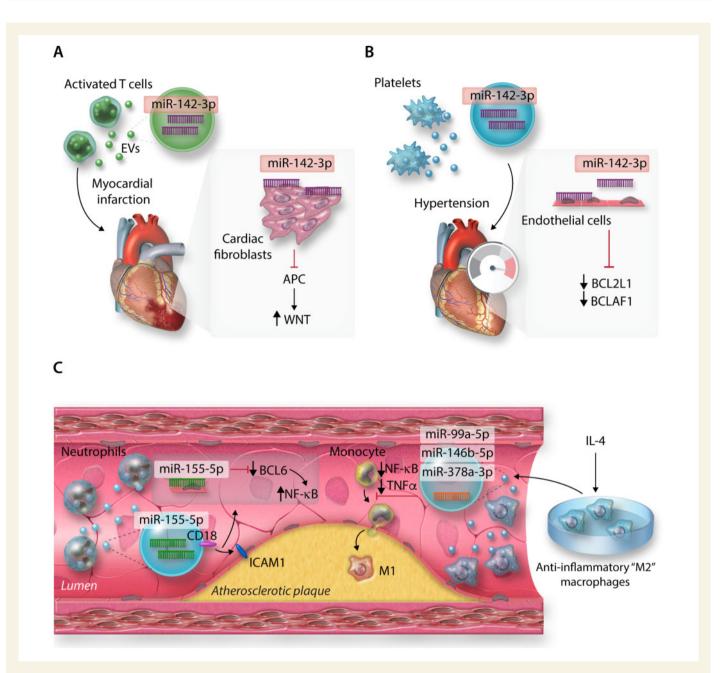


Figure 4 MicroRNAs from immune cell-derived extracellular vesicles (EVs) and their targets in (A) cardiac fibroblasts post-MI, (B) Endothelial cells (ECs) in hypertension, (C) Atherosclerotic plaques.

membrane. Once considered merely a disposal route for unwanted cellular components and debris, it is now appreciated that although this may be the case for a subset of specialized EVs (apoptotic bodies), EVs (including exosomes and microvesicles) can also be packed with numerous forms of viable cargo derived from their parental cells, including intracellular proteins, DNA, and multiple RNA species, such as miRNAs.¹⁰⁹ Following uptake by recipient cells, molecular cargo delivered within these packages can exert important biological functions, including post-transcriptional gene regulation by non-host cell-derived miRNAs, forming a critical component of intercellular communication and regulatory networks.

In the context of CVD, cardiovascular non-immune cells can release EVs loaded with miRNAs to influence intercellular communication and

regulation of other cell types, affecting multiple facets of their responses including cell migration and tissue influx, and inflammation.^{110,111} However, immune cells also utilize these pathways to modulate the surrounding cardiovascular tissue environment¹¹² (*Figure 4*). For instance, EVs derived from activated CD4⁺ T cells may be packaged with miR-142-3p.^{113–116} When delivered into mouse hearts following experimental MI, T-cell-derived EVs worsened post-MI cardiac dysfunction via the repression of the miR-142-3p target, adenomatous polyposis coli in cardiac fibroblasts, stimulating up-regulation of WNT-signalling, and profibrotic pathways¹¹⁶ (*Figure 4A*). Platelet-derived EVs, thought to be responsible for the majority of circulating EVs,¹¹⁷ may also play an important role in CVD pathogenesis via miR-142-3p delivery to cardiovascular cell types. Previously, it has been demonstrated that miR-142-3p plays

critical roles in generation of platelets, via the regulation of a network of actin cytoskeletal targets with critical roles to play during megakaryopoiesis.¹¹⁸ However, as for T cells, platelet-derived miR-142-3p can extend its regulatory actions beyond merely cell-endogenous effects; in the context of hypertension, ECs may take-up platelet-derived EVs, resulting in abnormal EC proliferation¹¹⁹ and apoptosis¹²⁰ via regulation of BCL2-like1¹¹⁹ and BCL2-associated transcription factor 1 by platelet-derived miR-142-3p¹²⁰ (*Figure 4B*).

As EVs are secreted into the extracellular environment and may be carried in the circulation, this also raises the possibility that immune cell types not overtly present in cardiovascular tissues in the context of inflammation, may significantly contribute to CVD progress. For example, neutrophils are not typically enriched within atherosclerotic plagues, but may still influence disease pathogenesis through the provision of integrin beta-2 (CD18)⁺ EVs containing neutrophil-derived miR-155-5p (among other miRs), which targets BCL-6 to promote pro-inflammatory NF-kB signalling in ECs at atheroprone arterial regions in a murine model of atherosclerosis¹²¹ (Figure 4C). Additionally, the therapeutic potential of utilizing immune cell-derived EVs to deliberately target pathways involved in CVD pathogenesis has recently become a point of interest.^{112,122} For example, EVs derived from macrophages polarized towards antiinflammatory, 'M2' phenotypes by IL-4, enriched with miR-99a-5p, miR-146b-5p, and miR-378a-3p may restrict plaque development and proinflammatory polarization of circulating monocytes and macrophages in the Apoe^{-/-} model of murine atherosclerosis, via down-regulation of NFkB and pro-inflammatory TNF- α production¹²³ (Figure 4C).

However, it is important to note that, despite the large number of studies that support a functional capacity for EV-delivered miRNAs to regulate critical cellular pathways, there persists a significant degree of controversy and debate surrounding these claims.^{124–126} For instance, the majority of EVs may actually hold very few copies of individual miRNAs, and the miRNA containing EVs represent only a very small fraction of the total circulating miRNA in plasma.¹²⁵ As such, work in the EV field has been greatly criticized for not, in general, considering the physiological importance of establishing the stoichiometric requirements between the delivered miRNAs and the predicted targets of relevance, in order for a functional impact upon target expression levels to be seen. It is unlikely that EVs carrying very few copies of a specific miRNA will functionally impact the expression levels of a high abundance target in another cell type, unless these EVs are delivered and taken up by target cells in sufficiently large quantities to do so. Additionally, few, if any standardized methods have been agreed on by the field for conducting these studies.

6. T_{REGS} immunotherapy and miRNAs: therapeutic potential of miRNA regulated pathways for the treatment of CVD

There has been significant investment in research to target and/or utilize T_{REGS} for treatment of conditions with underlying immunological dysfunctions. This has included graft-versus-host disease (GvHD),¹²⁷ autoimmune conditions, such as Crohn's disease and diabetes,^{128,129} and in the context of solid organ transplantation wherein induction of longterm tolerance to the allograft is a major clinical goal.¹²⁹ As it has become clear that T_{REGS} can serve many cardioprotective and vasoprotective roles during CVD, and accumulated evidence also implicates

Table | Meanings of abbreviations used in this review

Abbreviation	Meaning
AT	Adipose tissue
CAD	Coronary artery disease
CVD	Cardiovascular disease
DCM	Dilated cardiomyopathy
EAM	Experimental autoimmune myocarditis
EAT	Epicardial adipose tissue
ECs	Endothelial cells
EVs	Extracellular vesicles
FC	Foam cells
HF	Heart failure
LAEAT	Left atrial epicardial adipose tissue
MI	Myocardial infarction
Th	CD4 ⁺ helper T cell
T _{REGS}	CD4 ⁺ regulatory T cells

dysregulated T_{REGS} numbers and/or suppressive functions in CVD pathogenesis, $^{51,89,90,104-107,130-132}_{\rm I}$ immunotherapy delivered by exogenously delivered T_{REGS} or through targeted T_{REGS} -specific therapies might be a novel therapeutic option. $^{133-135}_{\rm I}$

Many studies have identified specific miRNAs, which critically regulate development, differentiation, and the suppressive actions of the spectrum of T_{REGS} subsets identified—from thymus-derived 'natural' T_{REGS} $(tT_{REGS} \text{ or } nT_{REGS})$ to T_{REGS} induced from naïve T cell precursors in the periphery (pT_{REGS}) or differentiated in vitro (iT_{REGS}), in addition to unique T_{REGS} from specialized niches, such as those found in umbilical cord blood T_{REGS} (Table 2). Of the miRNAs, which regulate T_{REGS}, miR-142 isoforms particularly appear to regulate many facets of T_{REGS} biology (Table 2 and Figure 5). A central mediator of T_{REGS} suppression of effector T cells, cAMP production is critically regulated on several molecular levels by miR-142 isoforms in T_{REGS}. Maintenance of ADCY9 expression through the specific down-regulation of miR-142-3p in T_{REGS} constitutes one mechanism of ensuring cAMP production and T_{REGS} suppressive activity.^{106,141} In addition, our group have also recently demonstrated that a major role for the miR-142-5p isoform in T_{REGS} is to ensure complete suppression of the potent cAMP hydrolase, phosphodiesterase 3B (PDE3B).²⁶ In the absence of T_{REGS}-specific miR-142-restriction of PDE3B, a severe breakdown in peripheral immune tolerance occurs, resulting in multi-organ tissue damage and widespread immune cell activation in mice.²⁶ Conversely, we also recently identified miR-142-3p as a critical regulator of TGF- β mediated T_{REGS} development through the regulation of TGFBR1, and established that deletion of the Mir142 locus promotes indefinite tolerance to cardiac allografts in a major MHCmismatched murine model of heterotopic heart transplantation.²⁷

However, what these findings and much of the collective information regarding miRNA regulation of T_{REGS} mean in the context of CVD largely remains to be investigated. Indeed, despite good evidence for protective functions of T_{REGS} during CVD, studies that characterize the potential for T_{REGS} therapy in major forms of CVD, such as atherosclerosis, are currently lacking–as recently outlined by Albany *et al.*¹³⁵ The concept of using miRNA mimics and/or synthetic miRNA inhibitory molecules (i.e. antagomiRs) as therapeutic agents is highly topical.^{151,152} Candidates in early development aim to target a range of conditions including hepatitis C infection (Mirvirsen, RG-101), cancer (MesomiR,

Table 2 Summar	y of miRNAs a	Table 2 Summary of miRNAs and identified targets involved		in regulation of T _{REGS} biology		
MicroRNA	Organism studied	T _{REGS} subset investigated	ldentified target/s	Altered microRNA expression (↑↓)	Effect of altered microRNA expression on T_{REGS}	Ref.
miR-15a-5p/16-5p	Mm	CB T _{REGS}	FOXP3	←	Forced overexpression suppresses FOXP3 and reduces cellular suppressive functions.	136
miR-17-5p	Mm	iT _{regs} , pT _{regs}	EOS, IRF4, SATB1	÷	IL-6 promotes miR-17-5p, which suppresses identified targets, compromising FOXP3 function and leading to pro-inflammatory cytokine expression and	137
miR-24-3p	Hs	рТ _{REGS}	FOXP3	\rightarrow	toss of suppressive activity. miR-24-3p is naturally lowly expressed in T _{REGS} . Enforced expression reduces FOXP3 expression	138
miR-31-5p	Hs, Mm	CB/nT _{REGS} , pT _{REGS}	FOXP3	<i>←</i>	miR-31-5p expression is low in T _{REGS} —enforced expression reduces FOXP3 expression and TREGS differentiation.	108,139
miR-125a-5p	Мт	рТ _{REGS} , iT _{REGS}	STAT3, IFNG, IL13	\rightarrow	Loss of miR-125a-5p reduces T_{ReGS} and promotes inflammatory responses, worsening colitis, and EAE, miR-125a stabilizes T_{ReGS} homoeostasis.	140
miR-142-3p	Mm	pT _{REGS} , iT _{REGS}	TGFBR1	\rightarrow	Deletion of Mir142 in T cells prevented MHC-mismatched heart and skin allo-	27
	Hs, Mm Uc. M.::	рТ _{REGS} , iT _{REGS} т	TET2 ADCV8	$\rightarrow \leftarrow$	graft rejection by augmenting intragraft T _{REGS} via up-regulation of TGFBR1.	28 106,141
	Hs, Mm	P I REGS iT _{REGS}	KDM6A	_ →	Decreasing miner 142-54 up-1 eguated up ing paner eauc ister aucommuner. restores T _{REGs} induction and stability during disease.	142
					Raised miR-142-3p inhibits T _{REGS} suppressive activity via reduced ADCY9 and	
					Exerciting synutesis. miR-142-3p knockdown led to increased FOXP3 expression and suppressive function via up-regulation of KDM6A and promotion of BCL-2 via demeth- vlation of H3K77me3	
miR-142-5p	Mm	pTregs	PDE3B	\rightarrow	T _{REGS} specific loss of <i>Mir</i> 142 resulted in loss of peripheral tolerance, wide- spread systemic autoimmunity, and inflammation due to reduction in cAMP	26
miR-146a-5p	Mm	PT _{REGS}	STAT1	\rightarrow	and loss of suppressive function. Loss of miR-146a-5p in T _{REGS} leads to breakdown in tolerogenic mechanisms and promotes Th.	143,144
miR-146b-5p	Hs	tT _{REGS}	TRAF6	\rightarrow	Antagomir-knockdown of miR-146b-5p promotes tT _{REGS} function and inhibi- tory function in GvHD	145
miR-155-5p	Mm	PT _{REGS}	socs1	\rightarrow	FOXP3 promotes T _{REGS} miR-155-5p expression. Genetic loss of miR-155 sup- pressed STAT5 signalline and reduced T _{REGS} numbers and homoeostasis.	6
miR-181a/b-5p	Mm Mm	tT _{REGS} iT _{REGS}	CTLA4 SMAD7	$\rightarrow \leftarrow$	Loss of miR-181a/b-5p restricts thymic T _{REGS} development but promotes peripheral TREGS suppressive activity.	146 147
					Transfected miR-181a-5p mimics enhance FOXP3 expression and $T_{\mathtt{REGS}}$ differentiation.	
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	Organism studied	T _{REGS} subset investigated	ldentifie target/s	Altered microRNA expression (↑↓)	Effect of altered microRNA expression on T _{REGS}	Ref.
miR-202-5p	Hs	miR-202-5p Hs pTREGS iTREGS	MATN2	÷	During allergic rhinitis, up-regulated miR-202-5p inhibits T_{REGS} development and function.	148
miR-210-3p	Hs	PT _{REGS}	FOXP3	\rightarrow	miR-210-3p is naturally lowly expressed in T _{REGS} . Enforced expression reduces FOXP3 expression.	138
miR-340-5p	Mm	pT _{REGS}	IL4	\rightarrow	CD226 deficiency in TREGS promoted Th2, by reducing miR-340-5p, promot- ing T _{REGS} IL-4 release.	149
miR-1224-5p	Mim	PTREGS	FOXP3	\rightarrow	AhR signalling attenuates pertussis toxin-induced systemic inflammation by suppressing miR-1224-5p, enhancing FOXP3 expression, and promoting Trenses	150
miR-4281-3p	Hs	IT _{REGS}	FOXP3	←	Intranuclear localization and binding of miR-4281-3p to FOXP3 promotor TATA-box enhanced FOXP3 expression, T _{REGS} differentiation, stability, and function.	ω

CB, cord blood; Hs, Homo sopiens; i/n/p/t T_{REGS}, inducible/natural/peripheral/thymic regulatory T cells; Mm, Mus musculus.

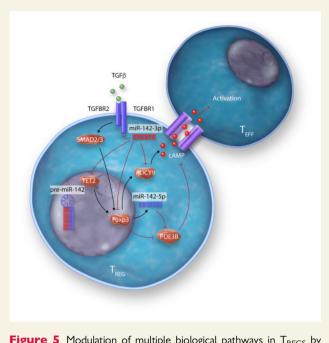


Figure 5 Modulation of multiple biological pathways in T_{REGS} by miR-142 isoforms.

MRG-106), and ischaemia (MRG-110), with promising results in phase I and II clinical trials. Furthermore, pre-clinical data using antagomiRtreated T_{REGS} in humanized mouse models of GvHD indicate that ex vivo miRNA modulation of T_{REGS} is a promising avenue to enhance T_{REGS} stability and suppressive activity during adoptive cell transfer immunotherapy.¹⁴⁵ The prospect that targeted miRNA manipulation of T_{REGS} might overcome some of the predicted limitations of adoptive transfer T_{REGS} immunotherapy,¹²⁹ such as stabilization of specific regulatory phenotypes or better control over trafficking and migration to desired sites, is particularly enticing both for CVD and other conditions. In addition, these points also raise the possibility that correction of dysfunctional T_{REGS} in specific forms of CVD might also be achieved therapeutically by targeting aberrantly expressed miRNAs in ex vivo expanded populations of the patient's cells. This may be particularly important when considering that, although interventions targeted to cardiovascular resident or infiltrating cell populations may be desirable for treatment of CVD, offtarget, systemic suppression of immunity and inflammatory responses may theoretically result in unwanted general and/or opportunistic infections and other undesirable outcomes-despite any benefits to CVD treatment.

7. Concluding remarks

Attempts to characterize and better understand how immune system responses are regulated by miRNAs is a highly topical avenue of research. Unravelling the dynamics of these regulatory pathways and their functions in specific immune and inflammatory cell types is key to their therapeutic potential. With regards to CVD, it is critical to examine the dynamics of immune system regulation by miRNAs in each specific disease context, rather than make generalized inferences. Against this background, for certain well-known immuno-miRNAs, such as miR-155-5p, there is a robust body of literature, which implicates dysregulated expression in CVD pathogenesis. In our opinion, this molecule represents an obvious therapeutic candidate target of interest in several forms of CVD. However, for other immuno-miRNAs, there are notable gaps in the literature surrounding immune system regulation specifically in CVD. For example, the mature isoforms of miR-142 demonstrably influence key immune system developmental and cellular signalling pathways, and circulating levels are altered in CVD,^{153–155} but they have rarely been directly investigated in this setting.

As discussed in this review, among leucocytes, macrophages and CD4⁺ T cell responses are particularly important in CVD aetiology and progression. Targeting miRNAs and miRNA-regulated signalling networks in these cells-particularly those which regulate multiple aspects of immune cell biology, may prove to be especially useful in immunotherapeutic settings. However, how best to target these molecules in an effective, safe, yet cell type-specific manner is still unclear. The use of modified synthetic anti-sense oligonucleotides, such as antagomirs may be associated with off-target effects, particularly at higher therapeutic doses in vivo. Additionally, such effects could be relatively long-lasting, given the enhanced stability of these molecules. Packaging technologies. such as modified polymeric NPs may overcome some of the challenges the field faces, for instance, allowing for more controlled delivery or release of therapeutic molecules to target cells.¹⁵⁶ As we have outlined, an alternative to this cell-targeted approach may be to turn to immunotherapy using miRNA modified T_{REGs} to suppress dysregulated inflammation and immune responses underlying CVD pathogenesis. This area warrants further investigation and significant research effort.

Finally, recent advances in single-cell sequencing and analytical technologies allow for a depth in the resolution of data concerning rare cardiac immune cell populations, previously not accessible to researchers. For instance, it has only recently come to light that a population of tissue-resident, helper-like innate immune cells (ILCs) displaying a predominantly group 2-commited profile (ILC2) exists within heart tissue in mice and humans. These cells may play important roles during CVD including myocarditis, cardiac ischaemia, and pericarditis, ^{157–159} while populations observed in para-aortic AT and lymph nodes may also serve vital roles in directing progression of atherosclerosis.¹⁶⁰ However, these cells have primarily been studied in mice thus far and the relevance to human CVD remains to be established. Although still in its infancy, our understanding of how miRNAs regulate development, homeostasis, and activity of ILCs is gradually being unravelled.^{161–163} Future studies should aim to identify how these molecules regulate these newly identified populations, both during health and disease. This may potentially reveal exciting new avenues of therapeutic promise for patients suffering from CVD.

Conflict of interest: A.M.S. is a consulting editor for Cardiovascular Research. G.M.L. has filed a patent regarding modulation of PDE3B by miR-142.

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References

- Virani SS, Alonso A, Benjamin EJ, Bittencourt MS, Callaway CW, Carson AP, Chamberlain AM, Chang AR, Cheng S, Delling FN, Djousse L, Elkind MSV, Ferguson JF, Fornage M, Khan SS, Kissela BM, Knutson KL, Kwan TW, Lackland DT, Lewis TT, Lichtman JH, Longenecker CT, Loop MS, Lutsey PL, Martin SS, Matsushita K, Moran AE, Mussolino ME, Perak AM, Rosamond WD, Roth GA, Sampson UKA, Satou GM, Schroeder EB, Shah SH, Shay CM, Spartano NL, Stokes A, Tirschwell DL, VanWagner LB, Tsao CW; On behalf of the American Heart Association Council on Epidemiology and Prevention Statistics Committee and Stroke Statistics Subcommittee. Heart disease and stroke statistics—2020 update: a report from the American Heart Association. *Circulation* 2020;**141**:e139–e596.
- Swirski FK, Nahrendorf M. Cardioimmunology: the immune system in cardiac homeostasis and disease. Nat Rev Immunol 2018;18:733–744.
- Martini E, Kunderfranco P, Peano C, Carullo P, Cremonesi M, Schorn T, Carriero R, Termanini A, Colombo FS, Jachetti E, Panico C, Faggian G, Fumero A, Torracca L, Molgora M, Cibella J, Pagiatakis C, Brummelman J, Alvisi G, Mazza EMC, Colombo MP, Lugli E, Condorelli G, Kallikourdis M. Single-cell sequencing of mouse heart immune infiltrate in pressure overload-driven heart failure reveals extent of immune activation. *Circulation* 2019;**140**:2089–2107.
- Strassheim D, Dempsey EC, Gerasimovskaya E, Stenmark K, Karoor V. Role of inflammatory cell subtypes in heart failure. *J Immunol Res* 2019;2019:1–9.
- Mehta A, Baltimore D. MicroRNAs as regulatory elements in immune system logic. Nat Rev Immunol 2016;16:279–294.
- Liu H, Lei C, He Q, Pan Z, Xiao D, Tao Y. Nuclear functions of mammalian MicroRNAs in gene regulation, immunity and cancer. *Mol Cancer* 2018;17:14.
- Wei Y, Li L, Wang D, Zhang CY, Zen K. Importin 8 regulates the transport of mature microRNAs into the cell nucleus. *J Biol Chem* 2014;289:10270–10275.
- Zhang Y, Liu W, Chen Y, Liu J, Wu K, Su L, Zhang W, Jiang Y, Zhang X, Zhang Y, Liu C, Tao L, Liu B, Zhang H. A cellular microRNA facilitates regulatory T lymphocyte development by targeting the FOXP3 promoter TATA-box motif. *J Immunol* 2018;**200**:1053–1063.
- 9. Berezikov E, Chung W-J, Willis J, Cuppen E, Lai EC. Mammalian mirtron genes. *Mol Cell* 2007;**28**:328–336.
- Ruby JG, Jan CH, Bartel DP. Intronic microRNA precursors that bypass Drosha processing. Nature 2007;448:83–86.
- 11. Stavast CJ, Erkeland SJ. The non-canonical aspects of microRNAs: many roads to gene regulation. *Cells* 2019;**8**:1465.
- O'Connell RM, Rao DS, Chaudhuri AA, Baltimore D. Physiological and pathological roles for microRNAs in the immune system. Nat Rev Immunol 2010;10:111–122.
- Zampetaki A, Willeit P, Drozdov I, Kiechl S, Mayr M. Profiling of circulating microRNAs: from single biomarkers to re-wired networks. *Cardiovasc Res* 2012;93: 555–562.
- Mogilyansky E, Rigoutsos I. The miR-17/92 cluster: a comprehensive update on its genomics, genetics, functions and increasingly important and numerous roles in health and disease. *Cell Death Differ* 2013;20:1603–1614.
- Kuo G, Wu CY, Yang HY. MiR-17-92 cluster and immunity. J Formos Med Assoc 2019;118:2–6.
- Sheedy FJ. Turning 21: induction of miR-21 as a key switch in the inflammatory response. Front Immunol 2015;6:1–9.
- Medina PP, Nolde M, Slack FJ. OncomiR addiction in an in vivo model of microRNA-21-induced pre-B-cell lymphoma. *Nature* 2010;467:86–90.
- 18. Feng YH, Tsao CJ. Emerging role of microRNA-21 in cancer (Review). *Biomed Rep* 2016;**5**:395–402.
- Kim C, Hu B, Jadhav RR, Jin J, Zhang H, Cavanagh MM, Akondy RS, Ahmed R, Weyand CM, Goronzy JJ. Activation of miR-21-regulated pathways in immune aging selects against signatures characteristic of memory T cells. *Cell Rep* 2018;25: 2148–2162.E5.
- Chen C-Z. MicroRNAs modulate hematopoietic lineage differentiation. Science 2004;303:83–86.
- 21. Landgraf P, Rusu M, Sheridan R, Sewer A, Iovino N, Aravin A, Pfeffer S, Rice A, Kamphorst AO, Landthaler M, Lin C, Socci ND, Hermida L, Fulci V, Chiaretti S, Foà R, Schliwka J, Fuchs U, Novosel A, Müller R-U, Schermer B, Bissels U, Inman J, Phan Q, Chien M, Weir DB, Choksi R, De Vita G, Frezzetti D, Trompeter H-I, Hornung V, Teng G, Hartmann G, Palkovits M, Di Lauro R, Wernet P, Macino G, Rogler CE, Nagle JW, Ju J, Papavasiliou FN, Benzing T, Lichter P, Tam W, Brownstein MJ, Bosio A, Borkhardt A, Russo JJ, Sander C, Zavolan M, Tuschl T. A mammalian microRNA expression atlas based on small RNA library sequencing. *Cell* 2007;**129**:1401–1414.
- Sharma S. Immunomodulation: a definitive role of microRNA-142. Dev Comp Immunol 2017;77:150–156.

- Nimmo R, Ciau-Uitz A, Ruiz-Herguido C, Soneji S, Bigas A, Patient R, Enver T. MiR-142-3p controls the specification of definitive hemangioblasts during ontogeny. *Dev Cell* 2013;26:237–249.
- Kramer NJ, Wang W-L, Reyes EY, Kumar B, Chen C-C, Ramakrishna C, Cantin EM, Vonderfecht SL, Taganov KD, Chau N, Boldin MP. Altered lymphopoiesis and immunodeficiency in miR-142 null mice. *Blood* 2015;**125**:3720–3730.
- Rivkin N, Chapnik E, Mildner A, Barshtein G, Porat Z, Kartvelishvily E, Dadosh T, Birger Y, Amir G, Yedgar S, Izraeli S, Jung S, Hornstein E. Erythrocyte survival is controlled by microRNA-142. *Haematologica* 2017;**102**:676–685.
- Anandagoda N, Willis JCD, Hertweck A, Roberts LB, Jackson I, Refik Gökmen M, Jenner RG, Howard JK, Lord GM. MicroRNA-142–mediated repression of phosphodiesterase 3B critically regulates peripheral immune tolerance. *J Clin Invest* 2019; 129:1257–1271.
- Anandagoda N, Roberts LB, Willis JCD, Sarathchandra P, Xiao F, Jackson I, Hertweck A, Kapoor P, Jenner RG, Howard JK, Lord GM. Dominant regulation of long-term allograft survival is mediated by microRNA-142. *Am J Transplant* 2020;**20**: 2715–2727.
- Scherm MG, Serr I, Zahm AM, Schug J, Bellusci S, Manfredini R, Salb VK, Gerlach K, Weigmann B, Ziegler AG, Kaestner KH, Daniel C. miRNA142-3p targets Tet2 and impairs Treg differentiation and stability in models of type 1 diabetes. *Nat Commun* 2019;**10**:15.
- Suzuki HI, Young RA, Sharp PA. Super-enhancer-mediated RNA processing revealed by integrative microRNA network analysis. *Cell* 2017;**168**:1000–1014.E15.
- Skårn M, Barøy T, Stratford EW, Myklebost O. Epigenetic regulation and functional characterization of microRNA-142 in mesenchymal cells. *PLoS One* 2013;8:e79231.
- Razak SRA, Baba Y, Nakauchi H, Otsu M, Watanabe S. DNA methylation is involved in the expression of miR-142-3p in fibroblasts and induced pluripotent stem cells. *Stem Cells Int* 2014;2014:1–8.
- Saba R, Sorensen DL, Booth SA. MicroRNA-146a: a dominant, negative regulator of the innate immune response. *Front Immunol* 2014;5:1–11.
- Rusca N, Monticelli S. MiR-146a in immunity and disease. Mol Biol Int 2011;2011: 1–7.
- Yang L, Boldin MP, Yu Y, Liu CS, Ea CK, Ramakrishnan P, Taganov KD, Zhao JL, Baltimore D. miR-146a controls the resolution of T cell responses in mice. J Exp Med 2012;209:1655–1670.
- Taganov KD, Boldin MP, Chang KJ, Baltimore D. NF-kB-dependent induction of microRNA miR-146, an inhibitor targeted to signaling proteins of innate immune responses. *Proc Natl Acad Sci USA* 2006;**103**:12481–12486.
- Rodriguez A, Vigorito E, Clare S, Warren MV, Couttet P, Soond DR, Dongen S, van Grocock RJ, Das PP, Miska EA, Vetrie D, Okkenhaug K, Enright AJ, Dougan G, Turner M, Bradley A. Requirement of bic/microRNA-155 for normal immune function. *Science* 2007;**316**:608–611.
- Vigorito E, Perks KL, Abreu-Goodger C, Bunting S, Xiang Z, Kohlhaas S, Das PP, Miska EA, Rodriguez A, Bradley A, Smith KGC, Rada C, Enright AJ, Toellner KM, MacLennan ICM, Turner M. microRNA-155 regulates the generation of immunoglobulin class-switched plasma cells. *Immunity* 2007;27:847–859.
- Alivernini S, Gremese E, McSharry C, Tolusso B, Ferraccioli G, McInnes IB, Kurowska-Stolarska M. MicroRNA-155-at the critical interface of innate and adaptive immunity in arthritis. *Front Immunol* 2018;8:1932.
- Wang D, Tang M, Zong P, Liu H, Zhang T, Liu Y, Zhao Y. MiRNA-155 regulates the Th17/Treg ratio by targeting SOCS1 in severe acute pancreatitis. *Front Physiol* 2018; 9:1–10.
- Lu L-F, Thai T-H, Calado DP, Chaudhry A, Kubo M, Tanaka K, Loeb GB, Lee H, Yoshimura A, Rajewsky K, Rudensky AY. Foxp3-dependent microRNA155 confers competitive fitness to regulatory T cells by targeting SOCS1 protein. *Immunity* 2009;**30**:80–91.
- 41. Heymans S, Corsten MF, Verhesen W, Carai P, Leeuwen REW, Van Custers K, Peters T, Hazebroek M, Stöger L, Wijnands E, Janssen BJ, Creemers EE, Pinto YM, Grimm D, Schürmann N, Vigorito E, Thum T, Stassen F, Yin X, Mayr M, Windt LJ, De Lutgens E, Wouters K, Winther MPJ, De Zacchigna S, Giacca M, Bilsen M, Van Papageorgiou AP, Schroen B. Macrophage microRNA-155 promotes cardiac hypertrophy and failure. *Circulation* 2013;**128**:1420–1432.
- Williams A, Henao-Mejia J, Harman CCD, Flavell RA. miR-181 and metabolic regulation in the immune system. *Cold Spring Harb Symp Quant Biol* 2013;**78**:223–230.
- Henao-Mejia J, Williams A, Goff LA, Staron M, Licona-Limón P, Kaech SM, Nakayama M, Rinn JL, Flavell RA. The microRNA miR-181 is a critical cellular metabolic rheostat essential for NKT cell ontogenesis and lymphocyte development and homeostasis. *Immunity* 2013;**38**:984–997.
- 44. Su R, Lin HS, Zhang XH, Yin XL, Ning HM, Liu B, Zhai PF, Gong JN, Shen C, Song L, Chen J, Wang F, Zhao HL, Ma YN, Yu J, Zhang JW. MiR-181 family: regulators of myeloid differentiation and acute myeloid leukemia as well as potential therapeutic targets. *Oncogene* 2015;**34**:3226–3239.
- Ye Z, Li G, Kim C, Hu B, Jadhav RR, Weyand CM, Goronzy JJ. Regulation of miR-181a expression in T cell aging. Nat Commun 2018;9:11.
- Seeger T, Haffez F, Fischer A, Koehl U, Leistner DM, Seeger FH, Boon RA, Zeiher AM, Dimmeler S. Immunosenescence-associated microRNAs in age and heart failure. *Eur J Heart Fail* 2013;15:385–393.

- Yuan X, Berg N, Lee JW, Le TT, Neudecker V, Jing N, Eltzschig H. MicroRNA miR-223 as regulator of innate immunity. J Leukoc Biol 2018;104:515–524.
- Roffel MP, Bracke KR, Heijink IH, Maes T. miR-223: a key regulator in the innate immune response in asthma and COPD. Front Med 2020;7:1–16.
- Guzik TJ, Hoch NE, Brown KA, McCann LA, Rahman A, Dikalov S, Goronzy J, Weyand C, Harrison DG. Role of the T cell in the genesis of angiotensin II-induced hypertension and vascular dysfunction. J Exp Med 2007;204:2449–2460.
- Nosalski R, Siedlinski M, Denby L, McGinnigle E, Nowak M, Cat AND, Medina-Ruiz L, Cantini M, Skiba D, Wilk G, Osmenda G, Rodor J, Salmeron-Sanchez M, Graham G, Maffia P, Graham D, Baker AH, Guzik TJ. T-cell-derived miRNA-214 mediates perivascular fibrosis in hypertension. *Circ Res* 2020;**126**:988–1003.
- Li X, Cai W, Xi W, Sun W, Shen W, Wei T, Chen X, Sun L, Zhou H, Sun Y, Chen W, Gao P, Wang H, Li Q. MicroRNA-31 regulates immunosuppression in Ang II (Aangiotensin II)-induced hypertension by targeting Ppp6C (protein phosphatase 6c). *Hypertension* 2019;**73**:E14–E24.
- 52. Marí-Alexandre J, Barceló-Molina M, Sanz-Sánchez J, Molina P, Sancho J, Abellán Y, Santaolaria-Ayora ML, Giner J, Martínez-Dolz L, Estelles A, Braza-Boils A, Zorio E. Thickness and an altered miRNA expression in the epicardial adipose tissue is associated with coronary heart disease in sudden death victims. *Rev Española Cardiol (English Ed)* 2019;**72**:30–39.
- Pan Y, Hui X, Chong Hoo RL, Ye D, Cheung Chan CY, Feng T, Wang Y, Ling Lam KS, Xu A. Adipocyte-secreted exosomal microRNA-34a inhibits M2 macrophage polarization to promote obesity-induced adipose inflammation. *J Clin Invest* 2019; 129:834–849.
- 54. Vacca M, Di Eusanio M, Cariello M, Graziano G, D'Amore S, Petridis FD, D'orazio A, Salvatore L, Tamburro A, Folesani G, Rutigliano D, Pellegrini F, Sabbà C, Palasciano G, Di Bartolomeo R, Moschetta A. Integrative miRNA and whole-genome analyses of epicardial adipose tissue in patients with coronary atherosclerosis. *Cardiovasc Res* 2016;**109**:228–239.
- 55. Tran K, Van Majka J, Sanghai S, Sardana M, Lessard D, Milstone Z, Tanriverdi K, Freedman JE, Fitzgibbons TP, McManus D. Micro-RNAs are related to epicardial adipose tissue in participants with atrial fibrillation: data from the MiRhythm study. *Front Cardiovasc Med* 2019;6:1–8.
- Apostolakis S, Vogiatzi K, Amanatidou V, Spandidos DA. Interleukin 8 and cardiovascular disease. *Cardiovasc Res* 2009;84:353–360.
- 57. Wei Y, Zhu M, Schober A. Macrophage microRNAs as therapeutic targets for atherosclerosis, metabolic syndrome, and cancer. *Int J Mol Sci* 2018;**19**:1–20.
- Zheng CG, Chen BY, Sun RH, Mou XZ, Han F, Li Q, Huang HJ, Liu JQ, Tu YX. miR-133b downregulation reduces vulnerable plaque formation in mice with AS through inhibiting macrophage immune responses. *Mol Ther Nucleic Acids* 2019;16: 745–757.
- Nus M, Martínez-Poveda B, MacGrogan D, Chevre R, D'Amato G, Sbroggio M, Rodríguez C, Martínez-González J, Andrés V, Hidalgo A, de la Pompa JL. Endothelial Jag1-RBPJ signalling promotes inflammatory leucocyte recruitment and atherosclerosis. *Cardiovasc Res* 2016;**112**:568–580.
- Fukuda D, Aikawa E, Swirski FK, Novobrantseva TI, Kotelianski V, Gorgun CZ, Chudnovskiy A, Yamazaki H, Croce K, Weissleder R, Aster JC, Hotamisligil GS, Yagita H, Aikawa M. Notch ligand delta-like 4 blockade attenuates atherosclerosis and metabolic disorders. *Proc Natl Acad Sci USA* 2012;**109**:E1868–E1877.
- Liu ZJ, Tan Y, Beecham GW, Seo DM, Tian R, Li Y, Vazquez-Padron RI, Pericak-Vance M, Vance JM, Goldschmidt-Clermont PJ, Livingstone AS, Velazquez OC. Notch activation induces endothelial cell senescence and pro-inflammatory response: implication of Notch signaling in atherosclerosis. *Atherosclerosis* 2012;**225**: 296–303.
- Chen J, Xu L, Hu Q, Yang S, Zhang B, Jiang H. MiR-17-5p as circulating biomarkers for the severity of coronary atherosclerosis in coronary artery disease. *Int J Cardiol* 2015;**197**:123–124.
- Tan L, Liu L, Jiang Z, Hao X. Inhibition of microRNA-17-5p reduces the inflammation and lipid accumulation, and up-regulates ATP-binding cassette transporterA1 in atherosclerosis. J Pharmacol Sci 2019;139:280–288.
- Feinberg MW, Moore KJ. MicroRNA regulation of atherosclerosis. Circ Res 2016; 118:703–720.
- 65. Horie T, Ono K, Horiguchi M, Nishi H, Nakamura T, Nagao K, Kinoshita M, Kuwabara Y, Marusawa H, Iwanaga Y, Hasegawa K, Yokode M, Kimura T, Kita T. MicroRNA-33 encoded by an intron of sterol regulatory element-binding protein 2 (Srebp2) regulates HDL in vivo. *Proc Natl Acad Sci USA* 2010;**107**: 17321–17326.
- Najafi-Shoushtari SH, Kristo F, Li Y, Shioda T, Cohen DE, Gerszten RE, Naar AM. MicroRNA-33 and the SREBP host genes cooperate to control cholesterol homeostasis. *Science* 2010;**328**:1566–1569.
- 67. Karunakaran D, Thrush AB, Nguyen M-A, Richards L, Geoffrion M, Singaravelu R, Ramphos E, Shangari P, Ouimet M, Pezacki JP, Moore KJ, Perisic L, Maegdefessel L, Hedin U, Harper M-E, Rayner KJ. Macrophage mitochondrial energy status regulates cholesterol efflux and is enhanced by anti-miR33 in atherosclerosis. *Circ Res* 2015; **117**:266–278.
- Sorci-Thomas MG, Thomas MJ. Microdomains, inflammation, and atherosclerosis. Circ Res 2016;118:679–691.
- 69. Meng X, Yin J, Yu X, Guo Y. MicroRNA-205-5p promotes unstable atherosclerotic plaque formation in vivo. *Cardiovasc Drugs Ther* 2020;**34**:25–39.

- Chen W, Li X, Wang J, Song N, Zhu A, Jia L. miR-378a modulates macrophage phagocytosis and differentiation through targeting CD47-SIRPα axis in atherosclerosis. Scand J Immunol 2019;90:10.
- 71. Ye Z, Ming Yang S, Xia Y, Peng Hu R, Ting Chen S, Li B, Wei Chen S, Li Luo X, Ying Mao L, Li Y, Jin H, Qin C, Hu B. LncRNA MIAT sponges miR-149-5p to inhibit efferocytosis in advanced atherosclerosis through CD47 upregulation. *Cell Death Dis* 2019;**10**:138.
- Thomson DW, Dinger ME. Endogenous microRNA sponges: evidence and controversy. Nat Rev Genet 2016;17:272–283.
- 73. Liu J, Li W, Wang S, Wu Y, Li Z, Wang W, Liu R, Ou J, Zhang C, Wang S. MiR-142-3p attenuates the migration of CD4+T cells through regulating actin cytoskeleton via RAC1 and ROCK2 in arteriosclerosis obliterans. *PLoS One* 2014;**9**:e95514.
- 74. Ji Q, Wei Guo M, Zheng J, Song Mao X, Bo Peng Y, Dong Li S, Nan Liang Z, Shan Dai Z, Yin Mao Y, Zeng Q. Tang Downregulation of T helper cell type 3 in patients with acute coronary syndrome. *Arch Med Res* 2009;40:285–293.
- Methe H, Brunner S, Wiegand D, Nabauer M, Koglin J, Edelman ER. Enhanced Thelper-1 lymphocyte activation patterns in acute coronary syndromes. J Am Coll Cardiol 2005;45:1939–1945.
- Buono C, Binder CJ, Stavrakis G, Witztum JL, Glimcher LH, Lichtman AH. T-bet deficiency reduces atherosclerosis and alters plaque antigen-specific immune responses. Proc Natl Acad Sci USA 2005;102:1596–1601.
- 77. Steiner DF, Thomas MF, Hu JK, Yang Z, Babiarz JE, Allen CDC, Matloubian M, Blelloch R, Ansel KM. MicroRNA-29 regulates T-box transcription factors and interferon-γ production in helper T cells. *Immunity* 2011;35:169–181.
- Ulrich V, Rotllan N, Araldi E, Luciano A, Skroblin P, Abonnenc M, Perrotta P, Yin X, Bauer A, Leslie KL, Zhang P, Aryal B, Montgomery RL, Thum T, Martin K, Suarez Y, Mayr M, Fernandez-Hernando C, Sessa WC. Chronic miR-29 antagonism promotes favorable plaque remodeling in atherosclerotic mice. *EMBO Mol Med* 2016;8: 643–653.
- Peet C, Ivetic A, Bromage DI, Shah AM. Cardiac monocytes and macrophages after myocardial infarction. *Cardiovasc Res* 2020;**116**:1101–1112.
- Yang L, Wang B, Zhou Q, Wang Y, Liu X, Liu Z, Zhan Z. MicroRNA-21 prevents excessive inflammation and cardiac dysfunction after myocardial infarction through targeting KBTBD7. *Cell Death Dis* 2018;9:1–14.
- Bejerano T, Etzion S, Elyagon S, Etzion Y, Cohen S. Nanoparticle delivery of miRNA-21 mimic to cardiac macrophages improves myocardial remodeling after myocardial infarction. *Nano Lett* 2018;**18**:5885–5891.
- 82. Garikipati VNS, Verma SK, Jolardarashi D, Cheng Z, Ibetti J, Cimini M, Tang Y, Khan M, Yue Y, Benedict C, Nickoloff E, Truongcao MM, Gao E, Krishnamurthy P, Goukassian DA, Koch WJ, Kishore R. Therapeutic inhibition of miR-375 attenuates post-myocardial infarction inflammatory response and left ventricular dysfunction via PDK-1-AKT signalling axis. *Cardiovasc Res* 2017;**113**:938–949.
- Krijnen PAJ, Meischl C, Hack CE, Meijer CJLM, Visser CA, Roos D, Niessen HWM. Increased Nox2 expression in human cardiomyocytes after acute myocardial infarction. J Clin Pathol 2003;56:194–199.
- 84. Wu Q, Allouch A, Paoletti A, Leteur C, Mirjolet C, Martins I, Voisin L, Law F, Dakhli H, Mintet E, Thoreau M, Muradova Z, Gauthier M, Caron O, Milliat F, Ojcius DM, Rosselli F, Solary E, Modjtahedi N, Deutsch E, Perfettini JL. NOX2-dependent ATM kinase activation dictates pro-inflammatory macrophage phenotype and improves effectiveness to radiation therapy. *Cell Death Differ* 2017;**24**:1632–1644.
- Looi YH, Grieve DJ, Siva A, Walker SJ, Anilkumar N, Cave AC, Marber M, Monaghan MJ, Shah AM. Involvement of Nox2 NADPH oxidase in adverse cardiac remodeling after myocardial infarction. *Hypertension* 2008;**51**:319–325.
- Yang J, Brown ME, Zhang H, Martinez M, Zhao Z, Bhutani S, Yin S, Trac D, Xi JJ, Davis ME. High-throughput screening identifies microRNAs that target Nox2 and improve function after acute myocardial infarction. *Am J Physiol Hear Circ Physiol* 2017;**312**:H1002–H1012.
- Chen X, Dong S, Zhang N, Chen L, Li MG, Yang PC, Song J. MicroRNA-98 plays a critical role in experimental myocarditis. *Int J Cardiol* 2017;229:75–81.
- Chen L, Hou X, Zhang M, Zheng Y, Zheng X, Yang Q, Li J, Gu N, Zhang M, Sun Y, Wu J, Yu B. MicroRNA-223-3p modulates dendritic cell function and ameliorates experimental autoimmune myocarditis by targeting the NLRP3 inflammasome. *Mol Immunol* 2020;**117**:73–83.
- Wang J, Han B. Dysregulated CD4+ T cells and microRNAs in myocarditis. Front. Immunol 2020;11:539–554.
- Yan L, Hu F, Yan X, Wei Y, Ma W, Wang Y, Lu S, Wang Z. Inhibition of microRNA-155 ameliorates experimental autoimmune myocarditis by modulating Th17/Treg immune response. J Mol Med 2016;94:1063–1079.
- 91. Corsten MF, Papageorgiou A, Verhesen W, Carai P, Lindow M, Obad S, Summer G, Coort SLM, Hazebroek M, Leeuwen R, Van Gijbels MJJ, Wijnands E, Biessen EAL, Winther MPJ, De Stassen FRM, Carmeliet P, Kauppinen S, Schroen B, Heymans S. MicroRNA profiling identifies microRNA-155 as an adverse mediator of cardiac injury and dysfunction during acute viral myocarditis. *Circ Res* 2012;**111**:415–425.
- Zhang Y, Zhang M, Li X, Tang Z, Wang X, Zhong M, Suo Q, Zhang Y, Lv K. Silencing microRNA-155 attenuates cardiac injury and dysfunction in viral myocarditis via promotion of M2 phenotype polarization of macrophages. Sci Rep 2016;6:13.
- Ong S, Ligons DL, Barin JG, Wu L, Talor MV, Diny N, Fontes JA, Gebremariam E, Kass DA, Rose NR, Čiháková D. Natural killer cells limit cardiac inflammation and fibrosis by halting eosinophil infiltration. Am J Pathol 2015;185:847–861.

- Trotta R, Chen L, Ciarlariello D, Josyula S, Mao C, Costinean S, Yu L, Butchar JP, Tridandapani S, Croce CM, Caligiuri MA. miR-155 regulates IFN-γ production in natural killer cells. *Blood* 2012;**119**:3478–3485.
- Loebel M, Holzhauser L, Hartwig JA, Shukla PC, Savvatis K, Jenke A, Gast M, Escher F, Becker SC, Bauer S, Stroux A, Beling A, Kespohl M, Pinkert S, Fechner H, Kuehl U, Lassner D, Poller W, Schultheiss HP, Zeller T, Blankenberg S, Papageorgiou AP, Heymans S, Landmesser U, Scheibenbogen C, Skurk C. The forkhead transcription factor Foxo3 negatively regulates natural killer cell function and viral clearance in myocarditis. *Eur Heart J* 2018;**39**:876–887.
- Everett BM, Cornel JH, Lainscak M, Anker SD, Abbate A, Thuren T, Libby P, Glynn RJ, Ridker PM. Anti-inflammatory therapy with canakinumab for the prevention of hospitalization for heart failure. *Circulation* 2019;**139**:1289–1299.
- Adamo L, Rocha-Resende C, Prabhu SD, Mann DL. Reappraising the role of inflammation in heart failure. Nat Rev Cardiol 2020;17:269–285.
- 98. Zeng Z, Wang K, Li Y, Xia N, Nie S, Lv B, Zhang M, Tu X, Li Q, Tang T, Cheng X. Down-regulation of microRNA-451a facilitates the activation and proliferation of CD4+ T cells by targeting Myc in patients with dilated cardiomyopathy. *J Biol Chem* 2017;**292**:6004–6013.
- Owczarz M, Budzinska M, Domaszewska-Szostek A, Borkowska J, Polosak J, Gewartowska M, Slusarczyk P, Puzianowska-Kuznicka M. miR-34a and miR-9 are overexpressed and SIRT genes are downregulated in peripheral blood mononuclear cells of aging humans. *Exp Biol Med (Maywood)* 2017;**242**:1453–1461.
- Rao DS, O'Connell RM, Chaudhuri AA, Garcia-Flores Y, Geiger TL, Baltimore D. MicroRNA-34a perturbs B lymphocyte development by repressing the forkhead box transcription factor Foxp1. *Immunity* 2010;33:48–59.
- 101. Nogueira E, Hamour S, Sawant D, Henderson S, Mansfield N, Chavele KM, Pusey CD, Salama AD. Serum IL-17 and IL-23 levels and autoantigen-specific Th17 cells are elevated in patients with ANCA-associated vasculitis. *Nephrol Dial Transplant* 2010;**25**:2209–2217.
- Abdulahad WH, Stegeman CA, Limburg PC, Kallenberg CGM. Skewed distribution of Th17 lymphocytes in patients with Wegener's granulomatosis in remission. *Arthritis Rheum* 2008;58:2196–2205.
- Abdulahad WH, Geld YM, Van Der Stegeman CA, Kallenberg CGM. Persistent expansion of CD4+ effector memory T cells in Wegener's granulomatosis. *Kidney Int* 2006;**70**:938–947.
- 104. Abdulahad WH, Stegeman CA, Geld YM, Van Der Doornbos-Van Der Meer B, Limburg PC, Kallenberg CGM. Functional defect of circulating regulatory CD4+ T cells in patients with Wegener's granulomatosis in remission. Arthritis Rheum 2007; 56:2080–2091.
- 105. Morgan MD, Day CJ, Piper KP, Khan N, Harper L, Moss PA, Savage COS. Patients with Wegener's granulomatosis demonstrate a relative deficiency and functional impairment of T-regulatory cells. *Immunology* 2010;**130**:64–73.
- 106. Dekkema GJ, Bijma T, Jellema PG, A Van Den B, Kroesen BJ, Stegeman CA, Heeringa P, Abdulahad WH, Sanders JS. Increased miR-142-3p expression might explain reduced regulatory T cell function in granulomatosis with polyangiitis. *Front Immunol* 2019;**10**:1–12.
- Ni FF, Li CR, Li Q, Xia Y, Wang GB, Yang J. Regulatory T cell microRNA expression changes in children with acute Kawasaki disease. *Clin Exp Immunol* 2014;**178**: 384–393.
- 108. Rouas R, Fayyad-Kazan H, El Zein N, Lewalle P, Rothé F, Simion A, Akl H, Mourtada M, El Rifai M, Burny A, Romero P, Martiat P, Badran B. Human natural Treg microRNA signature: role of microRNA-31 and microRNA-21 in FOXP3 expression. *Eur J Immunol* 2009;**39**:1608–1618.
- Kalra H, Drummen GPC, Mathivanan S. Focus on extracellular vesicles: introducing the next small big thing. *Int J Mol Sci* 2016;**17**:1–30.
- Chang YJ, Li YS, Wu CC, Wang KC, Huang TC, Chen Z, Chien S. Extracellular microRNA-92a mediates endothelial cell-macrophage communication. *ATVB* 2019; 39:2492–2504.
- 111. Zhao J, Li X, Hu J, Chen F, Qiao S, Sun X, Gao L, Xie J, Xu B. Mesenchymal stromal cell-derived exosomes attenuate myocardial ischaemia-reperfusion injury through miR-182-regulated macrophage polarization. *Cardiovasc Res* 2019;**115**: 1205–1216.
- Wu R, Gao W, Yao K, Ge J. Roles of exosomes derived from immune cells in cardiovascular diseases. Front Immunol 2019;10:1–16.
- 113. Cortes-Troncoso J, Jang S-I, Perez P, Hidalgo J, Ikeuchi T, Greenwell-Wild T, Warner BM, Moutsopoulos NM, Alevizos I, Moutsopoulos NM, Alevizos I. T cell exosome-derived miR-142-3p impairs glandular cell function in Sjögren's syndrome. JCI Insight 2020;5:1–14.
- 114. Dewi IS, Celik S, Karlsson A, Hollander Z, Lam K, McManus JW, Tebbutt S, Ng R, Keown P, McMaster R, McManus B, Öhman J, Gidlöf O. Exosomal miR-142-3p is increased during cardiac allograft rejection and augments vascular permeability through down-regulation of endothelial RAB11FIP2 expression. *Cardiovasc Res* 2017; **113**:440–452.
- 115. Guay C, Kruit JK, Rome S, Menoud V, Mulder NL, Jurdzinski A, Mancarella F, Sebastiani G, Donda A, Gonzalez BJ, Jandus C, Bouzakri K, Pinget M, Boitard C, Romero P, Dotta F, Regazzi R. Lymphocyte-derived exosomal microRNAs promote pancreatic β cell death and may contribute to type 1 diabetes development. *Cell Metab* 2019;**29**:348–361.E6.

- 116. Cai L, Chao G, Li W, Zhu J, Li F, Qi B, Wei Y, Chen S, Zhou G, Lu X, Xu J, Wu X, Fan G, Li J, Liu S. Activated CD4+ T cells-derived exosomal miR-142-3p boosts post-ischemic ventricular remodeling by activating myofibroblast. *Aging (Albany NY)* 2020;**12**:7380–7396.
- 117. Italiano JE, Mairuhu AT, Flaumenhaft R. Clinical relevance of microparticles from platelets and megakaryocytes. *Curr Opin Hematol* 2010;**17**:578–584.
- 118. Chapnik E, Rivkin N, Mildner A, Beck G, Pasvolsky R, Metzl-Raz E, Birger Y, Amir G, Tirosh I, Porat Z, Israel L, Lellouche E, Michaeli S, Lellouche JP, Izraeli S, Jung S, Hornstein E. MiR-142 orchestrates a network of actin cytoskeleton regulators during megakaryopoiesis. *Elife* 2014;**3**:1–22.
- Bao H, Yao Q-P, Huang K, Chen X-H, Han Y, Jiang Z-L, Gao L-Z, Qi Y-X. Plateletderived miR-142-3p induces apoptosis of endothelial cells in hypertension. *Cell Mol Biol (Noisy-le-Grand)* 2017;**63**:3.
- 120. Bao H, Chen YX, Huang K, Zhuang F, Bao M, Han Y, Chen XH, Shi Q, Yao QP, Qi YX. Platelet-derived microparticles promote endothelial cell proliferation in hypertension via miR-142-3p. *Faseb J* 2018;**32**:3912–3923.
- 121. Gomez I, Ward B, Souilhol C, Recarti C, Ariaans M, Johnston J, Burnett A, Mahmoud M, Luong LA, West L, Long M, Parry S, Woods R, Hulston C, Benedikter B, Niespolo C, Bazaz R, Francis S, Kiss-Toth E, Zandvoort M, van Schober A, Hellewell P, Evans PC, Ridger V. Neutrophil microvesicles drive atherosclerosis by delivering miR-155 to atheroprone endothelium. *Nat Commun* 2020;**11**:18.
- 122. Tikhomirov R, Reilly-O'Donnell B, Catapano F, Faggian G, Gorelik J, Martelli F, Emanueli C. Exosomes: from potential culprits to new therapeutic promise in the setting of cardiac fibrosis. *Cells* 2020;**9**:592.
- 123. Bouchareychas L, Duong P, Covarrubias S, Alsop E, Phu TA, Chung A, Gomes M, Wong D, Meechoovet B, Capili A, Yamamoto R, Nakauchi H, McManus MT, Carpenter S, Van K-JK, Raffai RL. Macrophage exosomes resolve atherosclerosis by regulating hematopoiesis and inflammation via microRNA cargo. *Cell Rep* 2020;**32**: 107881.
- Tkach M, Théry C. Communication by extracellular vesicles: where we are and where we need to go. *Cell* 2016;**164**:1226–1232.
- 125. Chevillet JR, Kang Q, Ruf IK, Briggs HA, Vojtech LN, Hughes SM, Cheng HH, Arroyo JD, Meredith EK, Gallichotte EN, Pogosova-Agadjanyan EL, Morrissey C, Stirewalt DL, Hladik F, Yu EY, Higano CS, Tewari M. Quantitative and stoichiometric analysis of the microRNA content of exosomes. *Proc Natl Acad Sci USA* 2014; **111**:14888–14893.
- Kesidou D, Martins C, da PA, Windt LJ, de BM, Beqqali A, Baker AH. Extracellular vesicle miRNAs in the promotion of cardiac neovascularisation. *Front Physiol* 2020; 11:1–25.
- Elias S, Rudensky AY. Therapeutic use of regulatory T cells for graft-versus-host disease. Br J Haematol 2019;187:25–38.
- Zhang X, Olsen N, Zheng SG. The progress and prospect of regulatory T cells in autoimmune diseases. J Autoimmun 2020;111:102461.
- Romano M, Fanelli G, Albany CJ, Giganti G, Lombardi G. Past, present, and future of regulatory T cell therapy in transplantation and autoimmunity. *Front Immunol* 2019;**10**:1–14.
- 130. Bansal SS, Ismahil MA, Goel M, Zhou G, Rokosh G, Hamid T, Prabhu SD. Dysfunctional and proinflammatory regulatory T-lymphocytes are essential for adverse cardiac remodeling in ischemic cardiomyopathy. *Circulation* 2019;**139**: 206–221.
- 131. Bansal SS, Ismahil MA, Goel M, Patel B, Hamid T, Rokosh G, Prabhu SD. Activated T lymphocytes are essential drivers of pathological remodeling in ischemic heart failure. *Circ Hear Fail* 2017;**10**:139–148.
- 132. Emmerson A, Trevelin SC, Mongue-Din H, Becker PD, Ortiz C, Smyth LA, Peng Q, Elgueta R, Sawyer G, Ivetic A, Lechler RI, Lombardi G, Shah AM. Nox2 in regulatory T cells promotes angiotensin II–induced cardiovascular remodeling. J Clin Invest 2018;**128**:3088–3101.
- Foks AC, Lichtman AH, Kuiper J. Treating atherosclerosis with regulatory T cells. Arterioscler Thromb Vasc Biol 2015;35:280–287.
- 134. Ou Han-xiao, Guo Bing-bing, Liu Qi, Yang Zhen, Feng Wen-jie and Mo Zhongcheng Regulatory T cells as a new therapeutic target for atherosclerosis. Acta Pharmacol Sin 2018;39:1249–1258.
- 135. Albany CJ, Trevelin SC, Giganti G, Lombardi G, Scottà C. Getting to the heart of the matter: the role of regulatory T-cells (T_{REGS}) in cardiovascular disease (CVD) and atherosclerosis. *Front Immunol* 2019;**10**:1–11.
- 136. Liu X, Robinson SN, Setoyama T, Tung SS, D'Abundo L, Shah MY, Yang H, Yvon E, Shah N, Yang H, Konopleva M, Garcia-Manero G, McNiece I, Rezvani K, Calin GA, Shpall EJ, Parmar S. FOXP3 is a direct target of miR15a/16 in umbilical cord blood regulatory T cells. *Bone Marrow Transplant* 2014;**49**:793–799.
- 137. Yang HY, Barbi J, Wu CY, Zheng Y, Vignali PDA, Wu X, Tao JH, Park BV, Bandara S, Novack L, Ni X, Yang X, Chang KY, Wu RC, Zhang J, Yang CW, Pardoll DM, Li H, Pan F. MicroRNA-17 modulates regulatory T cell function by targeting coregulators of the Foxp3 transcription factor. *Immunity* 2016;45:83–93.
- 138. Fayyad-Kazan H, Rouas R, Fayyad-Kazan M, Badran R, Zein N, El Lewalle P, Najar M, Hamade E, Jebbawi F, Merimi M, Romero P, Burny A, Badran B, Martiat P. MicroRNA profile of circulating CD4-positive regulatory T cells in human adults and impact of differentially expressed microRNAs on expression of two genes essential to their function. J Biol Chem 2012;287:9910–9922.

- Alrafas HR, Busbee PB, Nagarkatti M. Resveratrol downregulates miR-31 to promote T regulatory cells during prevention of TNBS-induced colitis. *Mol Nutr Food Res* 2020;64:1–11.
- 140. Pan W, Zhu S, Dai D, Liu Z, Li D, Li B, Gagliani N, Zheng Y, Tang Y, Weirauch MT, Chen X, Zhu W, Wang Y, Chen B, Qian Y, Chen Y, Fang J, Herbst R, Richman L, Jallal B, Harley JB, Flavell RA, Yao Y, Shen N. MiR-125a targets effector programs to stabilize Treg-mediated immune homeostasis. *Nat Commun* 2015;**6**:1–12.
- 141. Huang B, Zhao J, Lei Z, Shen S, Li D, Shen GX, Zhang GM, Feng ZH. miR-142-3p restricts cAMP production in CD4+CD25-T cells and CD4+CD25+ TREG cells by targeting AC9 mRNA. *EMBO Rep* 2009;**10**:180–185.
- 142. Gao J, Gu J, Pan X, Gan X, Ju Z, Zhang S, Xia Y, Lu L, Wang X. Blockade of miR-142-3p promotes anti-apoptotic and suppressive function by inducing KDM6Amediated H3K27me3 demethylation in induced regulatory T cells. *Cell Death Dis* 2019;**10**:332.
- 143. Wang X, Cao J, Yu Y, Ma B, Gao C, Lu J, Lin Y, Li P, Qi F. Role of microRNA 146a in regulating regulatory T cell function to ameliorate acute cardiac rejection in mice. *Transplant Proc* 2019;**51**:901–912.
- 144. Lu L-F, Boldin MP, Chaudhry A, Lin L-L, Taganov KD, Hanada T, Yoshimura A, Baltimore D, Rudensky AY. Function of miR-146a in controlling treg cell-mediated regulation of Th1 responses. *Cell* 2010;**142**:914–929.
- 145. Lu Y, Hippen KL, Lemire AL, Gu J, Wang W, Ni X, Ranganathan P, Levine BL, Riley JL, June CH, Turka LA, Munn DH, Garzon R, Lu L, Blazar BR. MiR-146b antagomir-treated human T_{REGS} acquire increased GVHD inhibitory potency. *Blood* 2016;**128**: 1424–1435.
- 146. Łyszkiewicz M, Winter SJ, Witzlau K, Föhse L, Brownlie R, Puchałka J, Verheyden NA, Kunze-Schumacher H, Imelmann E, Blume J, Raha S, Sekiya T, Yoshimura A, Frueh JT, Ullrich E, Huehn J, Weiss S, Gutierrez MG, Prinz I, Zamoyska R, Zietara N, Krueger A. mir-181A/B-1 controls thymic selection of treg cells and tunes their suppressive capacity. *PLoS Biol* 2019;**17**:e2006716.
- 147. Ghorbani S, Talebi F, Chan WF, Masoumi F, Vojgani M, Power C, Noorbakhsh F. MicroRNA-181 variants regulate T cell phenotype in the context of autoimmune neuroinflammation. *Front Immunol* 2017;8:1–14.
- Wang L, Yang X, Li W, Song X, Kang S. MiR-202-5p/MATN2 are associated with regulatory T-cells differentiation and function in allergic rhinitis. *Hum Cell Springer Japan* 2019;**32**:411–417.
- 149. Mu Y, Zhang J, Liu Y, Ma J, Jiang D, Zhang X, Yi X, Cheng K, Shen S, Yang Y, Zhuang R, Zhang Y. CD226 deficiency on regulatory T cells aggravates renal fibrosis via up-regulation of Th2 cytokines through miR-340. *J Leukoc Biol* 2020;**107**:573–587.
- 150. Al-Ghezi ZZ, Singh N, Mehrpouya-Bahrami P, Busbee PB, Nagarkatti M, Nagarkatti PS. AhR activation by TCDD (2,3,7,8-tetrachlorodibenzo-p-dioxin) attenuates pertussis toxin-induced inflammatory responses by differential regulation of T_{REGS} and Th17 cells through specific targeting by microRNA. *Front Microbiol* 2019;**10**:1–14.

- Hanna J, Hossain GS, Kocerha J. The potential for microRNA therapeutics and clinical research. Front Genet 2019;10:1–6.
- 152. Bonneau E, Neveu B, Kostantin E, Tsongalis GJ, De Guire V. How close are miRNAs from clinical practice? A perspective on the diagnostic and therapeutic market. *Electron J Int Fed Clin Chem Lab Med* 2019;**30**:114–127.
- Nair N, Kumar S, Gongora E, Gupta S. Circulating miRNA as novel markers for diastolic dysfunction. *Mol Cell Biochem* 2013;**376**:33–40.
- 154. Jiao M, You HZ, Yang XY, Yuan H, Li YL, Liu WX, Jin M, Du J. Circulating microRNA signature for the diagnosis of childhood dilated cardiomyopathy. *Sci Rep* 2018;8:1–9.
- 155. Ellis KL, Cameron VA, Troughton RW, Frampton CM, Ellmers LJ, Richards AM. Circulating microRNAs as candidate markers to distinguish heart failure in breathless patients. *Eur J Heart Fail* 2013;**15**:1138–1147.
- Lee SWL, Paoletti C, Campisi M, Osaki T, Adriani G, Kamm RD, Mattu C, Chiono V. MicroRNA delivery through nanoparticles. J Control Release 2019;313:80–95.
- 157. Bracamonte-Baran W, Chen G, Hou X, Talor MV, Choi HS, Davogustto G, Taegtmeyer H, Sung J, Hackam DJ, Nauen D, Čiháková D. Non-cytotoxic cardiac innate lymphoid cells are a resident and quiescent type 2-commited population. *Front Immunol* 2019;**10**:1–18.
- 158. Deng Y, Wu S, Yang Y, Meng M, Chen X, Chen S, Li L, Gao Y, Cai Y, Imani S, Chen B, Li S, Deng Y, Li X. Unique phenotypes of heart resident type 2 innate lymphoid cells. *Front Immunol* 2020;**11**:1–13.
- 159. Choi HS, Won T, Hou X, Chen G, Bracamonte-Baran W, Talor MV, Jurčová I, Szárszoi O, Čurnova L, Stříž I, Hooper JE, Melenovský V, Čiháková D. Innate lymphoid cells play a pathogenic role in pericarditis. *Cell Rep* 2020;**30**:2989–3003.E6.
- 160. Newland SA, Mohanta S, Clément M, Taleb S, Walker JA, Nus M, Sage AP, Yin C, Hu D, Kitt LL, Finigan AJ, Rodewald HR, Binder CJ, McKenzie ANJ, Habenicht AJ, Mallat Z. Type-2 innate lymphoid cells control the development of atherosclerosis in mice. *Nat Commun* 2017;8:11.
- 161. Berrien-Elliott MM, Sun Y, Neal C, Ireland A, Trissal MC, Sullivan RP, Wagner JA, Leong JW, Wong P, Mah-Som AY, Wong TN, Schappe T, Keppel CR, Cortez VS, Stamatiades EG, Li MO, Colonna M, Link DC, French AR, Cooper MA, Wang W, Le Boldin MP, Reddy P, Fehniger TA. MicroRNA-142 is critical for the homeostasis and function of type 1 innate lymphoid cells. *Immunity* 2019;**51**:479–490.E6.
- 162. Pelosi A, Alicata C, Tumino N, Ingegnere T, Loiacono F, Mingari MC, Moretta L, Vacca P. An anti-inflammatory microRNA signature distinguishes group 3 innate lymphoid cells from natural killer cells in human decidua. *Front Immunol* 2020;**1**:1–15.
- 163. Ramos-Ram P, Johansson K, Malmh C, Madeleine R. MicroRNA-155 is a critical regulator of type 2 innate lymphoid cells and IL-33 signaling in experimental models of allergic airway inflammation. J Allergy Clin Immunol 2017;**139**:1007–1016.E9.