



The continuous long reads (CLR) were assembled *de novo* with the PacBio SMRT analysis software (7.0.0.63985) for the CBS11 genome and SMRT analysis software (6.0.0.47841) for CBS12, using the HGAP4 protocol (SMRTLink v6.0.0). A subset of reads were subjected to BLAST analyses to ensure quality control and authenticity of the genome assembly. Contig end trimming and circularization with Circlator (v1.5.5) were performed, followed by polishing with Quiver by the Canadian Center for Computational Genomics (C3G). Once polished, sequences were annotated with Prokka (v1.12) (4) referencing a genus-specific genome (NCBI number [ASM78391v2](#)) produced by the University of Maryland School of Medicine and Institute for Genome Science (IGS). In all tool applications, default parameters were used except where otherwise noted. The CBS11 genome sequence resolved into 2 contigs, namely, a 5,352,713-bp contig and a 139,857-bp contig, representing a chromosome and a plasmid, respectively, with a G+C content of 59.39%, that combined included 4,950 genes, 22 rRNAs, 101 tRNAs, and 1 noncoding RNA. The CBS12 genome sequence resolved into 1 5,642,069-bp contig ascribed to the sole chromosome containing 5,204 genes, 22 rRNAs, 93 tRNAs, and 1 noncoding RNA.

Genome sequence analyses of CBS11 and CBS12 indicated that these two isolates are not clones and therefore are unique strains. Furthermore, a majority of the pAM01 plasmid sequence shares high identity (>90%) with *S. marcescens* plasmid PWNM146 (GenBank accession number [LT575492](#)) (5). Further genomic analyses are ongoing to understand the ability of these two *S. marcescens* strains to survive and thrive in PCs.

**Data availability.** This genome project has been deposited in GenBank under accession numbers [CP053927](#) (chromosome of CBS11), [CP053926](#) (plasmid pAM01), and [CP053925](#) (chromosome of CBS12). Accession numbers for raw reads for the CBS11 chromosome and plasmid pAM01 and for the CBS12 chromosome are [PRJNA634532](#) and [PRJNA634544](#), respectively.

## ACKNOWLEDGMENTS

This work was funded by a University of Manitoba grant (URGP number 49703) and by a The Dr. Paul H. T. Thorlakson Foundation grant (University of Manitoba) awarded to A.K.C.B. The Canadian Center for Computational Genomics (C3G) is a Genomics Technology Platform (GTP) supported by the Canadian Government through Genome Canada.

## REFERENCES

1. Greco-Stewart VS, Brown EE, Parr C, Kalab M, Jacobs MR, Yomtovian RA, Ramirez-Arcos SM. 2012. *Serratia marcescens* strains implicated in adverse transfusion reactions form biofilms in platelet concentrates and demonstrate reduced detection by automated culture. *Vox Sanguinis* 102: 212–220. <https://doi.org/10.1111/j.1423-0410.2011.01550.x>.
2. Schrezenmeier H, Walther-Wenke G, Müller TH, Weinauer F, Younis A, Holland-Letz T, Geis G, Asmus J, Bauerfeind U, Burkhart J, Deitenbeck R, Förstmann E, Gebauer W, Höchsmann B, Karakassopoulos A, Liebscher U-M, Sänger W, Schmidt M, Schunter F, Sireis W, Seifried E. 2007. Bacterial contamination of platelet concentrates: results of a prospective multicenter study comparing pooled whole blood-derived platelets and apheresis platelets. *Transfusion* 47:644–652. <https://doi.org/10.1111/j.1537-2995.2007.01166.x>.
3. Wilson-Nieuwenhuis JST, Dempsey-Hibbert N, Liauw CM, Whitehead KA. 2017. Surface modification of platelet concentrate bags to reduce biofilm formation and transfusion sepsis. *Colloids Surf B Biointerfaces* 160:126–135. <https://doi.org/10.1016/j.colsurfb.2017.09.019>.
4. Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
5. Vicente CSL, Nascimento FX, Barbosa P, Ke H-M, Tsai IJ, Hirao T, Cock PJA, Kikuchi T, Hasegawa K, Mota M. 2016. Evidence for an opportunistic and endophytic lifestyle of the *Bursaphelenchus xylophilus*-associated bacteria *Serratia marcescens* PWN146 isolated from wilting *Pinus pinaster*. *Microb Ecol* 72:669–681. <https://doi.org/10.1007/s00248-016-0820-y>.