

## CRITICAL REVIEW

# Drug-resistant epilepsy: Drug target hypothesis and beyond the receptors

Daniel Fonseca-Barriendos<sup>1</sup>  | Christian Lizette Frías-Soria<sup>1</sup> | Daniel Pérez-Pérez<sup>2</sup>  | Rosenda Gómez-López<sup>3</sup> | Dasiel O. Borroto Escuela<sup>4</sup> | Luisa Rocha<sup>1</sup> 

<sup>1</sup>Pharmacobiology Department, Center for Research and Advanced Studies, México City, México

<sup>2</sup>Plan of Combined Studies in Medicine (PECEM), Faculty of Medicine, UNAM, México City, Mexico

<sup>3</sup>Escuela Nacional de Medicina y Homeopatía, Instituto Politécnico Nacional, Mexico City, México

<sup>4</sup>Biomedicum, Karolinska Institutet, Stockholm, Sweden

## Correspondence

Luisa Rocha, Biomedicum, Karolinska Institutet, Stockholm, Sweden.  
Email: [lrocha@cinvestav.mx](mailto:lrocha@cinvestav.mx)

## Funding information

Consejo Nacional de Ciencia y Tecnología, Grant/Award Number: A3-S-26782

## Abstract

Epilepsy is a chronic neurological disorder that affects more than 50 million people worldwide. Despite a recent introduction of antiseizure drugs for the treatment of epileptic seizures, one-third of these patients suffer from drug-resistant epilepsy (DRE). The therapeutic target hypothesis is a cited theory to explain DRE. According to the target hypothesis, the failure to achieve seizure freedom leads to alteration of the structure and/or function of the antiseizure medication (ASM) target. However, this hypothesis fails to explain why patients with DRE do not respond to antiseizure medications of different targets. This review presents different conditions, such as epigenetic mechanisms and protein-protein interactions that may result in alterations of diverse drug targets using different mechanisms. These novel conditions represent new targets to control DRE.

## KEYWORDS

antiseizure medications, drug-resistant, epigenetics, epilepsy, mosaics, oligomers, receptors, target hypothesis

## 1 | INTRODUCTION

Epilepsy is a common chronic neurological disorder that affects more than 50 million people worldwide.<sup>1</sup> Antiseizure drug that serves as the first line of treatment is used to control seizures. However, one-third of patients with epilepsy suffer from drug-resistant epilepsy (DRE) because they fail to achieve control of seizures despite the appropriate treatment schemes used (in monotherapy or various combinations).<sup>2</sup>

According to the target hypothesis that explains the drug-resistance phenotype in epilepsy, failure to control epileptic activity by antiseizure medication (ASM) results in losing therapeutic efficacy as a consequence of

alterations in the structure and/or function of their targets<sup>3</sup> (Figure 1). In this regard, adopting different therapeutic targets in patients with DRE is crucial, which include alterations in voltage-gated sodium channels (VGSCs) and  $\gamma$ -aminobutyric acid (GABA) receptors.

Some studies confirm that resected brain tissue obtained from patients with drug-resistant temporal lobe epilepsy (DR-TLE) who have undergone surgery show reduced sensitivity to carbamazepine, a drug that inhibits VGSC.<sup>4,5</sup> Experimental models have not yet reproduced this condition in DRE. However, studies in models of acute seizures and epilepsy have demonstrated induced alterations in VGSC similar to those detected in brain tissue of patients with the DRE.<sup>6-8</sup>

This is an open access article under the terms of the [Creative Commons Attribution](https://creativecommons.org/licenses/by/4.0/) License, which permits use, distribution and reproduction in any medium, provided the original work is properly cited.

© 2021 The Authors. *Epilepsia Open* published by Wiley Periodicals LLC on behalf of International League Against Epilepsy

Status epilepticus induced in rats led to increase in spontaneous limbic seizures and window current of VGSC of dentate granule cells. The outcome is associated with a  $\beta 2$  subunit (days 1 and 3 after seizures) and  $\beta 1$  subunit (days 5 and 30 after seizures) reduced expression in VGSC.<sup>8</sup> From the results, sodium currents were enhanced, leading to aberrant burst discharges.<sup>9,10</sup> In addition, rats submitted to *status epilepticus* show impaired ASMs efficacy in VGSC, such as phenytoin and lamotrigine. This is more evident when spontaneous seizures are established in animals with a high frequency of ictal events.<sup>11</sup>

GABA<sub>A</sub> receptor is an important target for several ASMs.<sup>12</sup> In the hippocampal tissue of patients with DR-TLE, a decrease in the expression and rearrangement of GABA<sub>A</sub> receptors subunits has been reported.<sup>13</sup> Positron emission tomography is an effective neuroimaging method that has increasingly been used for epileptic focus *in vivo* analysis owing to a reduction of GABA<sub>A</sub> receptor binding and decrease in [<sup>11</sup>C]-flumazenil binding.<sup>14-17</sup> However, in the cortex of patients with DR-TLE, *in vitro* techniques revealed increased [<sup>3</sup>H]-flunitrazepam binding.<sup>18</sup> These findings suggest that the changes in GABA<sub>A</sub> receptors are region-specific.

Alterations in expression and/or stoichiometry of  $\alpha$  and  $\beta$  subunits of GABA<sub>A</sub> receptors lead to the loss of efficacy of benzodiazepines and barbiturates in DRE. Studies on animals with spontaneous recurrent seizures previously submitted to *status epilepticus*<sup>19</sup> or electrical stimulation<sup>20</sup> indicate that 40% of the experimental subjects fail to respond to phenobarbital, which is an agonist of GABA<sub>A</sub> receptor. The issue is associated with decreased expression of GABA<sub>A</sub>  $\alpha 1-3,5$  and  $\beta 2/3$  receptor subunits in the hippocampus up to 10 weeks after the insult.<sup>21</sup> An interesting finding is the increased expression of the  $\alpha 4$  subunits accompanied by a decrease in the  $\alpha 1$  subunit (1-4 months after pilocarpine-induced *status epilepticus*), a condition that leads to the lower effectiveness of benzodiazepines.<sup>22</sup> The increased mRNA expression of  $\alpha 4$  and  $\gamma 2$  subunits is reported in the cortex of patients with TLE-DR.<sup>18</sup> Thus, the  $\alpha 4\beta 2$  subtype of GABA<sub>A</sub> receptors are nonsensitive to benzodiazepine, revealing low brain expression.<sup>23</sup>

Alterations of ASM targets in patients with DRE and experimental models explain the pharmaco-resistance to individual drugs, that is, drug resistance to GABAergic drugs or VGSC blockers. However, patients with DRE normally fail to respond to diverse ASMs with dissimilar mechanisms, indicating simultaneous changes in different molecular targets. Thus, DRE is associated with a pathological condition that simultaneously modifies diverse targets with different mechanisms. In the subsequent sections of this manuscript, evidence explains epigenetic changes and protein-protein interactions changes of therapeutic targets with different mechanisms in DRE.

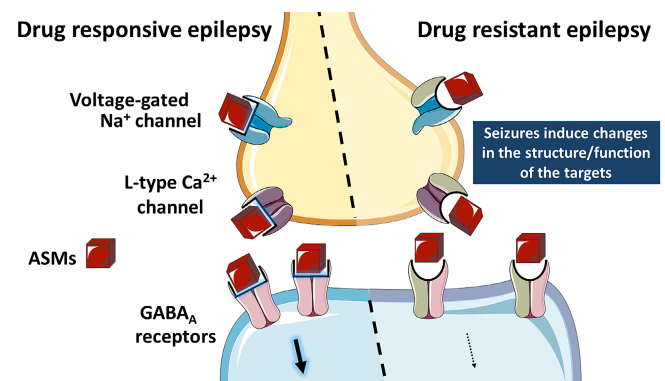
### Key points

- Patients with DRE show simultaneous lack of effects of ASMs with different mechanisms.
- Epigenetic changes can lead to simultaneous changes in the expression of different proteins.
- The receptor mosaics expression explains the loss of diverse ASMs efficacy.

## 2 | EPIGENETIC REGULATION

Epigenetics is the series of mechanisms that regulate gene function or expression that are heritable and do not entail a change in DNA sequence.<sup>24,25</sup> Histone modifications, DNA methylation, noncoding RNAs, and restrictive element-1 silencing transcription factor/neuron-restrictive silencing factor (REST/NRSF) are among the epigenetic involved in health and disease.<sup>26,27</sup>

Histone tails and globular domains are the targets of several posttranslational modifications, which include methylation, acetylation, ubiquitination, ADP-ribosylation, sumoylation, and phosphorylation.<sup>28</sup> The alterations are associated with active transcription (euchromatin modifications) or inactive regions (heterochromatin modifications).<sup>28</sup> Previous studies support that epilepsy and seizures induce histone modifications. Brain tissue obtained from patients with DR-TLE shows



**FIGURE 1** Schematic representation of the antiseizure medication (ASM) target hypothesis. In drug-responsive epilepsy (DRE) (left side), the ASMs interact with their targets and exert their pharmacological effect. Drug target hypothesis indicates that seizures induce molecular changes alter the structure and/or function of the drug targets. This condition results in the lack of sensitivity to ASMs leading to DRE. Thus, patients with DRE are refractory to several drugs with different mechanisms, which is not explained by the drug target hypothesis. The figure was created using illustrations from Servier Medical Art

upregulation of class 1 histone deacetylase (HDAC) 2, a condition associated with a decrease in histone acetylation and a decrease in gene expression.<sup>29</sup> Similar results were found in experimental models of seizures. Several histone deacetylases from class 1<sup>29,30</sup> and class 2<sup>31,32</sup> are upregulated during the *status epilepticus*-induced epileptogenesis. However, the class 2 member HDAC4 downmodulates the gene expression of GABA<sub>A</sub>  $\alpha$ 1 subunit,<sup>31</sup> which is benzodiazepine-sensitive<sup>33</sup> and the glutamatergic  $\alpha$ -amino-3-hydroxy-5-methyl-4-isoxazolepropionic acid (AMPA) receptor subunit GluA2.<sup>34</sup> The GluA2 subunit regulates calcium permeability of AMPA receptors,<sup>35</sup> and expression changes of this subunit is associated with many neurodevelopmental disorders,<sup>36</sup> tetramerizations,<sup>37</sup> and neuronal plasticity.<sup>38</sup> These findings suggest that histone deacetylation following epileptic seizures is associated with simultaneous changes in two different targets: (a) reduced gene expression of  $\alpha$ 1 subunit of GABA<sub>A</sub> benzodiazepine sensitive receptors, (b) downregulation of GluA2 subunit of AMPA receptors, a condition that enhances glutamate neurotransmission, aberrant plasticity, and neurotoxicity.<sup>39</sup>

DNA methylation is an epigenetic process of gene silencing mediated by the attachment of methyl groups to cytosine residues, located in promoter regions.<sup>40</sup>

Genome-wide methylation changes have been evaluated in the brain tissue of patients with DR-TLE and in epileptic experimental models. A previous study by Miller-Delaney<sup>41</sup> indicated that hippocampal resected tissue of patients with DR-TLE comprised 119 hypermethylated genes and 27 hypomethylated genes, which were compared with the methylation autopsies profile without neurological disorders. Gene ontology revealed that genes with differential methylation were correlated to developmental processes and cellular differentiation, connected to neuronal remodeling and maturation. In addition, neuron-expressed genes showing hypermethylation in patients with DR-TLE were associated with subunits of voltage-dependent calcium channels, voltage-gated potassium channels, and potassium, inwardly rectifying channels. A major finding of that study was that, despite the intrinsic variability associated with human tissue, less than 150 genes were differentially methylated, which suggested that most of the DNA methylation found in the hippocampus of patients with DR-TLE was static and disease-specific.<sup>41</sup> However, the temporal neocortex of patients with DR-TLE presents a high expression of DNA methyltransferase 1 (Dnmt1) and Dnmt3a, involved in DNA methylation.<sup>42</sup>

In rats with chronic epilepsy, whole-genome DNA methylation profiling showed increased DNA methylation in the dorsal hippocampus. However, this analysis fails to show significant changes in the gene expression of the

DNA methyltransferases Dnmt1, Dnmt3a, and Dnmt3b in rats with chronic epilepsy.<sup>43</sup> The results were in contrast with findings obtained from previous studies that focused on the cortex of patients with TLE.<sup>42</sup> In addition, diverse methylation patterns were detected in hippocampal tissue based on various experimental models of epilepsy (electrical amygdala stimulation, traumatic brain injury, or pilocarpine-induced *status epilepticus*), ranging from 1121 to 2741 altered regions and hyper- or hypomethylation in coding and noncoding sequences.<sup>44</sup> The present study revealed a broad pathophysiological difference between models. According to the previous information, it is suggested that DNA methylation in epilepsy is disease-specific that depends on different situations, such as the etiology and chronicity of the disorder, the stage of the disease, the rate of hippocampal sclerosis,<sup>41,45</sup> and the previous history of prolonged febrile seizures.<sup>46</sup>

In addition, DNA methylation affects several genes, thereby altering protein expression, signaling pathways, neuronal plasticity, inflammation, and immune response, amongst others. Moreover, changes in the transcriptome of different proteins induced by DNA methylation as a consequence of seizures and epilepsy may underlie the resistance of several ASMs.<sup>47,48</sup>

Noncoding RNA is the class of RNA transcripts that are not translated into proteins, but modulate gene expression in a sequence-specific manner.<sup>49</sup> Thus, noncoding RNA modulates 30% of all human genes as each noncoding RNA can regulate hundreds of genes.<sup>50</sup> In hippocampal tissue of patients with DR-TLE, approximately 18 microRNA (<200 nt) and 4 long noncoding (>200 nt) have been found differentially expressed. In addition, 13 microRNA were methylation-sensitive, in which DNA methylation exerts strong control of microRNA in DR-TLE.<sup>41</sup> Overexpression of miRNA-134 is in the hippocampal tissue of patients with DR-TLE and mice sacrificed 3 months after *status epilepticus*.<sup>51</sup> Previous studies show that miRNA-134 induces cell damage, has proconvulsant effects, and augments spontaneous seizure activity.<sup>51</sup> Moreover, microRNA-134 is negatively associated with synaptic growth and remodeling,<sup>52</sup> mediated by Limk1 inhibition, which induces dendritic spine growth<sup>53</sup> and inhibits palmitoyltransferase DHHC9 that regulate protein trafficking<sup>54,55</sup> to the synaptic membrane in GABAergic neurons.<sup>56</sup> In addition, microRNA-155 is upregulated in the hippocampal tissue of patients with DR-TLE, a condition that correlates with seizure frequency and postsurgical outcome.<sup>57</sup> The results from an experimental model showed that microRNA-155 was upregulated up to 60 days after *status epilepticus*, which was when spontaneous seizures were established. In addition, microRNA-155 provokes cell damage, leads to proconvulsant effects, and increases oxidative stress.<sup>57</sup> Interestingly, higher microRNA-155

expression (2.45-fold) was detected in patients who experienced postoperative seizures, but not in patients without seizures. Computational analysis revealed that microRNA-155 downregulated the  $\alpha 1$  subunit of VGSC expression.<sup>58</sup> Alterations in this subunit are associated with epileptic pathologies<sup>59</sup> based on persistent currents, which are shifts in the voltage dependence of activation<sup>60</sup> and decrease the activity of GABAergic neurons.<sup>61</sup> These findings demonstrate that epilepsy alters microRNA-134 and 155 expressions and modifies its progression. In patients with DRE, the absence of response to different ASMs is due to a simultaneous impairment of GABAergic synapse, increasing oxidative stress and altered VGSC as a consequence of microRNAs changes.

REST/NRSF is a protein that acts as a transcriptional repressor. REST binds to RE-1, a 17-33 bp sequence found in the DNA, which regulates the expression of approximately 1800 genes.<sup>62</sup> Once REST binds to RE-1, it prevents the expression of hundreds of genes and augments the expression of other genes.<sup>63,64</sup> Hippocampus of patients with DR-TLE shows REST mRNA overexpression, a finding that positively correlated with seizure frequency.<sup>65</sup> In addition, REST mRNA overexpression is induced in hippocampal tissue of rats, 24 hours after kainic acid-induced seizures.<sup>66</sup> The evidence suggests that REST overexpression induced by seizure activity alters many target proteins of the transcriptional repressor, which include brain-derived neurotrophic factor (BDNF), AMPA receptor GluR2 subunit, and noncoding RNA.<sup>67</sup> After seizures, REST overexpression may evoke a negative regulatory mechanism and suppress excessive expression of neuronal genes.<sup>66</sup>

Previous studies confirm that failure to achieve seizure freedom in patients with DR-TLE is associated with the overexpression of REST, which affects multiple target genes, such as expression of VGSC,<sup>68</sup> calcium channels,<sup>69</sup> Kv channels,<sup>70</sup> chloride transporter,<sup>71</sup> N-methyl-D-aspartate (NMDA) receptor subunits,<sup>72</sup>  $\mu$  opioid receptor,<sup>73</sup> dense-core vesicles,<sup>74</sup> and other pre- and postsynaptic proteins.

### 3 | EPIGENETIC CHANGES INDUCED BY ANTISEIZURE MEDICATIONS

Many ASMs induce epigenetic alterations.<sup>75</sup> Phenobarbital is associated with a decreased expression of methyltransferases<sup>76</sup> and increased gene expression. Valproic acid is a histone deacetylase inhibitor<sup>77</sup> that increases the miRNA-134 expression in patients with bipolar mania.<sup>78</sup> Carbamazepine inhibits histone deacetylases<sup>79</sup> and associates with GABA<sub>A</sub> gene silencing.<sup>80</sup> Topiramate is a histone deacetylase inhibitor<sup>81</sup> that inhibits ethanol-induced

methyltransferase overexpression.<sup>82</sup> In addition, lacosamide inhibits histone deacetylase.<sup>83</sup>

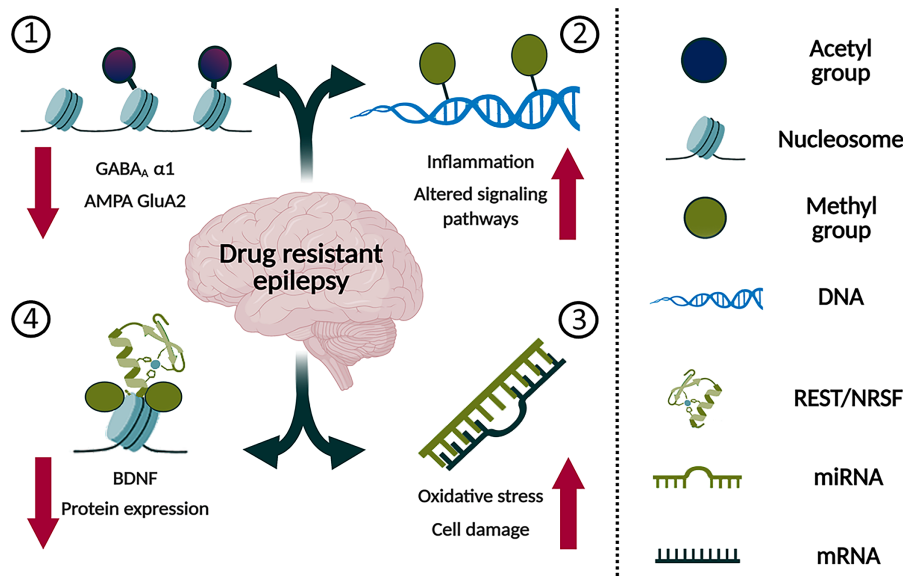
The findings raise issues about whether ASM-induced epigenetic changes aggravate alterations produced by ictogenic activity and contribute to the DRE. Concerning this issue, HDAC inhibitors, such as valproic acid, augment the expression and activity of the multidrug resistance protein 1 transporter in human brain endothelial cells.<sup>84</sup> The effect facilitates the drug-resistant phenotype of patients with epilepsy.<sup>85</sup> On the other hand, specific epigenetic changes were identified in patients with TLE, explaining their resistance to levetiracetam.<sup>86</sup> Thus, future research is crucial to determine if epigenetic effects induced by ASMs modify epilepsy. Further research is also required to investigate if the expression of the drug-resistant phenotype depends on clinical factors (dose and duration of the treatment, age of the patient, the condition of the disease, etc). The knowledge derived will help to identify patients with epilepsy susceptible to develop drug-resistant phenotype when receiving specific ASMs (Figure 2) and will help to design novel epigenetic strategies to control DRE.

### 4 | OLIGOMERIC RECEPTOR COMPLEXES AS THERAPEUTIC TARGETS IN DRUG-RESISTANT EPILEPSY

Some cells require external stimuli to survive. The stimuli are different molecules that include neurotransmitters, hormones, ions, and metabolites, among others. The cell's plasma membrane contains proteins that act as receptors and ion channels used to respond to external signals. Several conditions control the signaling of receptors such as the expression of G proteins, phosphorylation of intracellular residues that terminates receptor-effector coupling, activation of G protein-coupled receptor kinases, receptor internalization via clathrin-dependent endocytosis, etcetera.<sup>87,88</sup> On the other hand, sustained receptor activation may result in changes in sensitivity, conformation, uncoupling of the effector molecules, and internalization. Several of these receptor regulation mechanisms are based on the drug-resistant phenotype, and their evaluation has a significant therapeutic implication in designing new strategies to control DRE.

At present, receptors offer physical interaction, representing a cross-talk mechanism, with relevance to health and disease.<sup>89</sup> Approximately one-third of all receptors/ion channels in the cell are oligomeric complexes, that is, supramolecular assemblies of two or more associated subunits.<sup>90</sup> The protein assemblies consist of the same (homo-oligomers) or different protein subunits (hetero-oligomers).<sup>91</sup> For example, G protein-coupled





**FIGURE 2** Different epigenetic mechanisms and the drug target hypothesis of drug-resistant epilepsy. (1) Histone deacetylation downmodulates the gene expression of the benzodiazepine sensitive GABA<sub>A</sub>  $\alpha$ 1 subunit and the calcium impermeable AMPA receptor subunit GluA2. (2) Differential DNA methylation has been reported in thousands of coding and noncoding regions, including those associated with protein expression, signaling pathways, inflammation, among others. (3) Noncoding microRNAs overexpression has been associated to cell damage, oxidative stress, and proconvulsant effects, impairing the GABAergic synapse and increasing sodium currents. (4) REST/NRSF overexpression acts as a transcriptional repressor of several target genes including BDNF, ionic channels, glutamatergic receptors, transporters as well as some pre- and postsynaptic proteins. The figure was created using illustrations from BioRender.com

receptors (GPCRs) serve as monomers and homomers ( $\beta$ 2-adrenergic receptors,<sup>92</sup> the metabotropic glutamate receptor 5,<sup>93</sup> dopamine D2 receptor,<sup>94</sup>) and heteromers (GABA<sub>B</sub> hetero-oligomeric complex<sup>95</sup>), in which allosteric receptor-receptor interactions modulate the functions of the participating GPCR protomers. In addition, GPCRs form heteroreceptor complexes with ionotropic receptors (D1R-NMDA)<sup>96</sup> and receptor tyrosine kinases (FGFR1-5-HT<sub>1A</sub>R,<sup>97</sup> A2AR-FGFR1<sup>98</sup>), thereby modulating their functions. Moreover, adaptor proteins interact with receptor protomers and modulate their interactions.

Structural-functional molecular studies show that hetero-oligomerization between mutant and wild-type  $\alpha$  subunits of the VGSC leads to the channel gating probability impairment.<sup>99,100</sup> In this channel, the  $\beta$ 3 subunit in the VGSC hetero-oligomer reduces the carbamazepine action.<sup>101</sup> The hetero-dimerization of Q2 and Q3 subunits of the voltage-gated potassium channel increases the sensitivity of the channel for retigabine.<sup>102</sup> In addition, mutations in the GABA<sub>A</sub> receptor complex modify the function of the receptors and increase the possibility of forming hetero-oligomers or homo-oligomers, thereby facilitating the seizure activity.<sup>103</sup>

However, the expression of some oligomers depends on environmental conditions (pH, temperature, or drug concentration).<sup>104</sup> The oligomeric complexes' organization and their allosteric protomer-protomer interactions

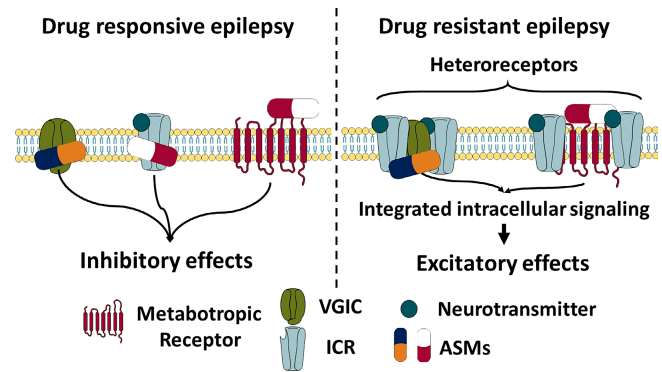
are reciprocal, highly dynamic, substantially altering the signaling, trafficking, recognition, and pharmacology of the participating protomers. The pattern of changes is unique for each heteroreceptor complex and favors antagonistic or facilitation interaction. Integration of signals to the plasma membrane by homo and heteroreceptor oligomers is crucial. This is based on the hypothesis that learning and memory at the molecular level occur through the reorganization of homo and heteroreceptor complexes in the postsynaptic membrane. Typically, homo and heteroreceptor complexes balance each other, and their disbalance provokes diseases.<sup>105</sup> In patients with DRE, the waxing-and-waning (or fluctuating) pattern in the ASM efficacy<sup>106</sup> could be associated with the dynamic formation of oligomers.<sup>91</sup> Future studies are needed to support this hypothesis.

In certain conditions, the receptors and/or ion channels establish physical interactions at a short distance (<40 nm), leading to mosaics (higher-order oligomers).<sup>107-109</sup> Heteroreceptors are mosaics established with receptors of different pharmacological properties. However, homoreceptors consist of the mosaic that contains different subtypes of the same receptor.<sup>107,110-113</sup> Some examples of heteroreceptors are established between  $\kappa$  and  $\delta$ -opioid receptors<sup>112</sup> and the adenosine A1 and dopamine D1 receptors. For example, A2A receptors are activated in homoreceptor complexes.<sup>114</sup> Thus, the receptor activation in the mosaics alters the

function (horizontal molecular network) or the intracellular transductional signals (vertical molecular network) of other receptors.<sup>115,116</sup> From this, the formation of receptor mosaics explains some clinical features that cannot be explained by the activation of isolated receptors.<sup>117</sup> Based on the expression of NMDA-D2 heteroreceptor in the striatum, the activation of D2 receptors can turn glutamatergic-induced long-term potentiation into long-term depression.<sup>118</sup> In Parkinson's disease, motor alterations are associated with the expression of heteroreceptors of dopamine receptors with adenosine, neurotensin, or different subtype of dopamine receptors.<sup>119</sup> The different heteroreceptors (serotonergic and metabotropic glutamate receptors, neurotensin and dopamine receptors, adenosine and dopamine receptors) are associated with the pathophysiology of psychosis and schizophrenia.<sup>120-122</sup> The dopamine D2-adenosine A2A heteroreceptors lead to the acute locomotor changes and sensitization induced by cocaine.<sup>123</sup> In addition, the cocaine-induced psychostimulant is associated with heteroreceptor complex formation between dopamine D2 receptors and NMDA receptor NR2B subunits in the neostriatum.<sup>124</sup> In patients suffering from depression, the neurotrophic and antidepressant effects induced by serotonin agonists are partially mediated by the activation of fibroblast growth factor 1 and 5-hydroxytryptamine 1A heteroreceptor complexes.<sup>125</sup>

At present, information regarding the expression of the receptor mosaics formation associated with epilepsy and DRE is lacking. However, the expression of homo- and heterooligomers complexes and receptor mosaics explain the resistance to several ASMs with different mechanisms of action showed by patients with DRE (Figure 3). Thus, patients with DRE have receptor mosaics that are formed by different ASM targets, including the neurotransmitter receptors or voltage-gated channels. Abnormal interaction within the mosaics could modify their sensitivity to diverse ASMs, thereby explaining the multidrug resistance phenotype. Everitt<sup>126</sup> hypothesized that a pathological memory, called drug memory, is associated with drug addiction. They also proposed that its molecular basis could lead to novel drug abuse user therapies. Our hypothesis on learning and memory molecular basis<sup>127</sup> is in line with the Everitt hypothesis. We suggest that drug memories can be produced through a reorganization of the homo and heterooligomeric complexes in synapses and extra-synaptic regions.

Future studies should determine if the DRE condition is associated with the dynamic expression of receptor mosaics. In addition, the abnormal receptor-receptor interactions in DRE opens the novel therapeutic approaches<sup>117</sup> depending on the heteroreceptor or mosaic expression.<sup>128,129</sup>



**FIGURE 3** Possible influence of heteroreceptor formation in drug-resistant epilepsy. In drug-responsive epilepsy, the antiseizure medications (ASMs) induce inhibitory effects, thus lowering seizure frequency or intensity. On the other hand, the heteroreceptor formation simultaneously changes the intracellular signaling of several receptors with different mechanisms and induces excitatory effects, instead of inhibition. This situation may lead to the drug resistant phenotype of patients with epilepsy. The figure was created using illustrations from Servier Medical Art

## 5 | CONCLUSION

The DRE target hypothesis indicated that the loss of therapeutic efficacy of ASMs was due to alterations in the molecular target. However, patients with DRE suffer from the loss of therapeutic efficacy of ASMs with dissimilar mechanisms. Other mechanisms can induce simultaneous changes in different molecular targets. In this review, information obtained from the literature suggests that altered gene expression due to epigenetic regulation and receptor mosaics formation is involved in the loss of several targets efficacy.

An important issue is to consider how the brain of patients with epilepsy is organized and the concept of the epileptic focus. The surgical strategy focuses to reduce seizure activity through resection of the epileptogenic region (the brain area sufficient for the generation of spontaneous seizures) and the distant irritative zone (characterized by spontaneous interictal spike activity, and involved in the seizure propagation).<sup>130,131</sup> Eventually, the irritative zone zone serves as a control condition, compared with the epileptogenic region of the same patient. However, studies indicate that both regions present significant changes, suggesting an increased excitatory influence in the epileptic focus and decreased inhibitory neurotransmission in the irritative zone.<sup>132</sup> Indeed, the irritative zone increases vascularity, microlesions, and marked activation of mitogen-activated protein kinase and cAMP-response element-binding protein (MAPK/CREB, a signaling pathway involved in synaptic plasticity).<sup>133</sup> Previous studies that focused on the human brain with epilepsy revealed a high expression

of long noncoding RNA (lncRNAs) genes involved in synaptic plasticity, a situation that correlated with the interictal spiking activity.<sup>134</sup> From this information and considering that lncRNAs involve in gene regulation,<sup>135</sup> the epileptogenic region and irritative zone present different patterns of gene and protein expression. Thus, the conclusions about changes in targets obtained from the evaluation of epileptogenic region vs irritative zone could lead to misconceptions. Future studies should identify changes in targets and mechanisms in the different zones of the epileptogenic region.

## ACKNOWLEDGMENT

This study was supported by the National Council for Science and Technology (CONACyT), grant A3-S-26782, scholarships 489736 (DFB), 704471 (CLFS), 261481 (DPP), and 734205 (RGL).

## CONFLICT OF INTEREST

No authors disclose conflict of interest. The authors confirm that they have read the Journal's position regarding the ethical publication and affirm that this report is consistent with those guidelines.

## ORCID

Daniel Fonseca-Barriendos  <https://orcid.org/0000-0002-7884-1593>

Daniel Pérez-Pérez  <https://orcid.org/0000-0002-9712-2543>

Luisa Rocha  <https://orcid.org/0000-0003-4495-9427>

## REFERENCES

1. WHO. Epilepsy. World Heal. Organ. 2019.
2. Kwan P, Arzimanoglou A, Berg AT, Brodie MJ, Allen Hauser W, Mathern G, et al Definition of drug resistant epilepsy: Consensus proposal by the ad hoc Task Force of the ILAE Commission on Therapeutic Strategies. *Epilepsia*. 2010;51:1069–77.
3. Remy S, Beck H. Molecular and cellular mechanisms of pharmacoresistance in epilepsy. *Brain*. 2006;129:18–35.
4. Jandová K, Päsler D, Antonio LL, Raue C, Ji S, Njunting M, et al Carbamazepine-resistance in the epileptic dentate gyrus of human hippocampal slices. *Brain*. 2006;129:3290–306.
5. Remy S, Gabriel S, Urban BW, Dietrich D, Lehmann TN, Elger CE, et al A novel mechanism underlying drug resistance in chronic epilepsy. *Ann Neurol*. 2003;53:469–79.
6. Bartolomei F, Gastaldi M, Massacrier A, Planells R, Nicolas S, Cau P. Changes in the mRNAs encoding subtypes I, II and III sodium channel alpha subunits following kainate-induced seizures in rat brain. *J Neurocytol*. 1997;26:667–78.
7. Gastaldi M, Robaglia-Schlupp A, Massacrier A, Planells R, Cau P. mRNA coding for voltage-gated sodium channel  $\beta 2$  subunit in rat central nervous system: Cellular distribution and changes following kainate-induced seizures. *Neurosci Lett*. 1998;249:53–6.
8. Ellerkmann RK, Remy S, Chen J, Sochivko D, Elger CE, Urban BW, et al Molecular and functional changes in voltage-dependent Na<sup>+</sup> channels following pilocarpine-induced status epilepticus in rat dentate granule cells. *Neuroscience*. 2003;119:323–33.
9. Bouza AA, Isom LL. Voltage-gated sodium channel  $\beta$  subunits and their related diseases. *Handb Exp Pharmacol*. 2018;246:423–50.
10. Azouz R, Jensen MS, Yaari Y. Ionic basis of spike afterdepolarization and burst generation in adult rat hippocampal CA1 pyramidal cells. *J Physiol*. 1996;492:211–23.
11. Remy S, Urban BW, Elger CE, Beck H. Anticonvulsant pharmacology of voltage-gated Na<sup>+</sup> channels in hippocampal neurons of control and chronically epileptic rats. *Eur J Neurosci*. 2003;17:2648–58.
12. Greenfield LJ Jr. Molecular mechanisms of antiseizure drug activity at GABA<sub>A</sub> receptors. *Seizure*. 2013;22:589–600.
13. Loup F, Wieser HG, Yonekawa Y, Aguzzi A, Fritschy JM. Selective alterations in GABA(A) receptor subtypes in human temporal lobe epilepsy. *J Neurosci*. 2000;20:5401–19.
14. Koepp MJ, Hammers A, Labbe C, Woermann FG, Brooks DJ, Duncan JS. 11C-flumazenil PET in patients with refractory temporal lobe epilepsy and normal MRI. *Neurology*. 2000;54:332–332.
15. Lamusuo S, Pitkänen A, Jutila L, Ylinen A, Partanen K, Kälviäinen R, et al [11 C]Flumazenil binding in the medial temporal lobe in patients with temporal lobe epilepsy: correlation with hippocampal MR volumetry, T2 relaxometry, and neuropathology. *Neurology*. 2000;54:2252–60.
16. Ryvlin P, Bouvard S, Le Bars D, De Lamérie G, Grégoire M, Kahane P, et al Clinical utility of flumazenil-PET versus [18F] fluorodeoxyglucose-PET and MRI in refractory partial epilepsy. A prospective study in 100 patients. *Brain*. 1998;121(Pt 1):2067–81.
17. Bouvard S, Costes N, Bonnefoi F, Lavenne F, Mauguière F, Delforge J, et al Seizure-related short-term plasticity of benzodiazepine receptors in partial epilepsy: a [11C]flumazenil-PET study. *Brain*. 2005;128:1330–43.
18. Rocha L, Alonso-Vanegas M, Martínez-Juarez IE, Orozco-Suárez S, Escalante-Santiago D, Feria-Romero IA, et al Gabaergic alterations in neocortex of patients with pharmacoresistant temporal lobe epilepsy can explain the comorbidity of anxiety and depression: The potential impact of clinical factors. *Front Cell Neurosci*. 2015;8:1–10.
19. Bankstahl M, Bankstahl JP, Löscher W. Inter-individual variation in the anticonvulsant effect of phenobarbital in the pilocarpine rat model of temporal lobe epilepsy. *Exp Neurol*. 2012;234:70–84.
20. Brandt C, Volk HA, Löscher W. Striking differences in individual anticonvulsant response to phenobarbital in rats with spontaneous seizures after status epilepticus. *Epilepsia*. 2004;45:1488–97.
21. Bethmann K, Fritschy JM, Brandt C, Löscher W. Antiepileptic drug resistant rats differ from drug responsive rats in GABA<sub>A</sub> receptor subunit expression in a model of temporal lobe epilepsy. *Neurobiol Dis*. 2008;31:169–87.
22. Brooks-Kayal AR, Shumate MD, Jin H, Rikhter TY, Coulter DA. Selective changes in single cell GABA(A) receptor subunit expression and function in temporal lobe epilepsy. *Nat Med*. 1998;4:1166–72.
23. Sieghart W, Sperk G. Subunit composition, distribution and function of GABA. *Curr Top Med Chem*. 2002;2:795–816.

24. Holliday R. INTRODUCTION epigenetics: an overview INTRODUCTION AND DEFINITION. *Dev Genet.* 1994;15:453-457.
25. Deans C, Maggert KA. What do you mean, “Epigenetic”? *Genetics.* 2015;199:887–96.
26. Hwang JY, Zukin RS. REST, a master transcriptional regulator in neurodegenerative disease. *Curr Opin Neurobiol.* 2018;48:193–200.
27. Radford EJ. *An Introduction to Epigenetic Mechanisms.* 1st ed. Elsevier Inc., 2018. <https://doi.org/10.1016/bs.pmbts.2018.04.002>
28. Li B, Carey M, Workman JL. The role of Chromatin during transcription. *Cell.* 2007;128:707–19.
29. Huang Y, Zhao F, Wang L, Wang L, Yin H, Zhou C, et al Increased expression of histone deacetylases 2 in temporal lobe epilepsy: A study of epileptic patients and rat models. *Synapse.* 2012;66:151–9.
30. Jagirdar R, Drexel M, Kirchmair E, Tasan RO, Sperk G. Rapid changes in expression of class I and IV histone deacetylases during epileptogenesis in mouse models of temporal lobe epilepsy. *Exp Neurol.* 2015;273:92–104.
31. Zhang Y, Dong HT, Duan L, Niu L, Yuan G-Q, Dai J-Q, et al HDAC4 gene silencing alleviates epilepsy by inhibition of GABA in a rat model. *Neuropsychiatr Dis Treat.* 2019;15:405–16.
32. Jagirdar R, Drexel M, Bukovac A, Tasan RO, Sperk G. Expression of class II histone deacetylases in two mouse models of temporal lobe epilepsy. *J Neurochem.* 2016;136:717–30.
33. Smith GB. *The GABA receptors.* Humana Press, 2007. <https://doi.org/10.1007/978-1-59745-465-0>
34. Huang Y, Doherty JJ, Dingleline R. Altered histone acetylation at glutamate receptor 2 and brain-derived neurotrophic factor genes is an early event triggered by status epilepticus. *J Neurosci.* 2002;22:8422–8.
35. Wright A, Vissel B. The essential role of AMPA receptor GluA2 subunit RNA editing in the normal and diseased brain. *Front Mol Neurosci.* 2012;5:1–13.
36. Salpietro V, Dixon CL, Guo H, Bello OD, Vandrovcova J, Efthymiou S, et al AMPA receptor GluA2 subunit defects are a cause of neurodevelopmental disorders. *Nat Commun.* 2019;10:3094. <https://doi.org/10.1038/s41467-019-10910-w>
37. Greger IH, Khatri L, Kong X, Ziff EB. Is mediated by Q / R editing. *Neuron.* 2003;40:763–74.
38. Wiltgen BJ, Royle GA, Gray EE, Abdipranoto A, Thangthaeng N, Jacobs N, et al A role for calcium-permeable AMPA receptors in synaptic plasticity and learning. *PLoS One.* 2010;5:e12818. <https://doi.org/10.1371/journal.pone.0012818>
39. Pellegrini-Giampietro DE, Gorter JA, Bennett MVL, Zukin RS. The GluR2 (GluR-B) hypothesis: Ca<sup>2+</sup>-permeable AMPA receptors in neurological disorders. *Trends Neurosci.* 1997;20:464–70.
40. Egger G, Liang G, Aparicio A, Jones PA. Epigenetics in human disease and prospects for epigenetic therapy. *Nature.* 2004;429:457–63.
41. Miller-Delaney SFC, Bryan K, Das S, McKiernan RC, Bray IM, Reynolds JP, et al Differential DNA methylation profiles of coding and non-coding genes define hippocampal sclerosis in human temporal lobe epilepsy. *Brain.* 2015;138:616–31.
42. Zhu Q, Wang L, Zhang Y, Zhao F-H, Luo J, Xiao Z, et al Increased expression of DNA methyltransferase 1 and 3a in human temporal lobe epilepsy. *J Mol Neurosci.* 2012;46:420–6.
43. Kobow K, Kaspi A, Harikrishnan KN, Kiese K, Ziemann M, Khurana I, et al Deep sequencing reveals increased DNA methylation in chronic rat epilepsy. *Acta Neuropathol.* 2013;126:741–56.
44. Debski KJ, Pitkanen A, Puhakka N, Bot AM, Khurana I, Harikrishnan KN, et al Etiology matters-genomic DNA methylation patterns in three rat models of acquired epilepsy. *Sci Rep.* 2016;6:1–14.
45. Kobow K, Ziemann M, Kaipananickal H, Khurana I, Mühlebner A, Feucht M, et al Genomic DNA methylation distinguishes subtypes of human focal cortical dysplasia. *Epilepsia.* 2019;60:1091–103.
46. De Nijs L, Choe K, Steinbusch H, Schijns OEMG, Dings J, van den Hove DLA, et al DNA methyltransferase isoforms expression in the temporal lobe of epilepsy patients with a history of febrile seizures. *Clin Epigenetics.* 2019;11:1–14.
47. Kobow K, Blümcke I. Epigenetics in epilepsy. *Neurosci Lett.* 2017;667:40–6.
48. Xiao W, Liu C, Zhong K, Ning S, Hou R, Deng NA, et al CpG methylation signature defines human temporal lobe epilepsy and predicts drug-resistant. *CNS Neurosci Ther.* 2020;26:1021–30.
49. Patil VS, Zhou R, Rana TM. Gene regulation by noncoding RNAs Veena. *Crit Rev Biochem Mol Biol.* 2014;25:289–313.
50. Chen K, Rajewsky N. The evolution of gene regulation by transcription factors and microRNAs. *Nat Rev Genet.* 2007;8:93–103.
51. Jimenez-Mateos EM, Engel T, Merino-Serrais P, McKiernan RC, Tanaka K, Mouri G, et al Silencing microRNA-134 produces neuroprotective and prolonged seizure-suppressive effects. *Nat Med.* 2012;18:1087–94.
52. Brennan GP, Henshall DC. MicroRNAs as regulators of brain function and targets for treatment of epilepsy. *Nat Rev Neurol.* 2020;16:506–19.
53. Schratt GM, Tuebing F, Nigh EA, Kane CG, Sabatini ME, Kiebler M, et al A brain-specific microRNA regulates dendritic spine development. *Nature.* 2006;439:283–9.
54. Matt L, Kim K, Chowdhury D, Hell JW. Role of palmitoylation of postsynaptic proteins in promoting synaptic plasticity. *Front Mol Neurosci.* 2019;12:1–19.
55. Park M, Penick EC, Edwards JG, Kauer JA, Ehlers MD. Recycling endosomes supply AMPA receptors for LTP. *Science (80-).* 2004;305:1972–5.
56. Chai S, Cambonne XA, Eichhorn SW, Goodman RH. MicroRNA-134 activity in somatostatin interneurons regulates H-Ras localization by repressing the palmitoylation enzyme, DHHC9. *Proc Natl Acad Sci USA.* 2013;110:17898–903.
57. Huang L-G, Zou J, Lu Q-C. Silencing rno-miR-155-5p in rat temporal lobe epilepsy model reduces pathophysiological features and cell apoptosis by activating Sestrin-3. *Brain Res.* 2018;1689:109–22.
58. Zhang Z, Wang Z, Zhang B, Liu Y. Downregulation of microRNA-155 by preoperative administration of valproic acid prevents postoperative seizures by upregulating SCN1A. *Mol Med Rep.* 2018;17:1375–81.
59. Catterall WA, Kalume F, Oakley JC. NaV1.1 channels and epilepsy. *J Physiol.* 2010;588:1849–59.



60. Ragsdale DS. How do mutant Nav1.1 sodium channels cause epilepsy? *Brain Res Rev.* 2008;58:149–59.
61. Escayg A, Goldin AL. Sodium channel SCN1A and epilepsy: Mutations and mechanisms. *Epilepsia.* 2010;51:1650–8.
62. Johnson DS, Mortazavi A, Myers RM, Wold B. Genome-wide mapping of in vivo protein-DNA interactions. *Science (80-).* 2007;316:1497–502.
63. Garcia-Manteiga JM, D'alessandro R, Meldolesi J. News about the role of the transcription factor REST in neurons: From physiology to pathology. *Int J Mol Sci.* 2020;21:235. <https://doi.org/10.3390/ijms21010235>
64. Ooi L, Wood IC. Chromatin crosstalk in development and disease: Lessons from REST. *Nat Rev Genet.* 2007;8:544–54.
65. Navarrete-Modesto V, Orozco-Suárez S, Alonso-Vanegas M, Feria-Romero IA, Rocha L. REST/NRSF transcription factor is overexpressed in hippocampus of patients with drug-resistant mesial temporal lobe epilepsy. *Epilepsy Behav.* 2019;94:118–23.
66. Palm K, Belluardo N, Metsis M, Timmusk T. Neuronal expression of zinc finger transcription factor REST/NRSF/XBR gene. *J Neurosci.* 1998;18:1280–96.
67. Roopra A, Huang Y, Dingleline R. Neurological disease: listening to gene silencers. *Mol Interv.* 2001;1:219–28.
68. Pozzi D, Lignani G, Ferrea E, Contestabile A, Paonessa F, D'Alessandro R, et al REST/NRSF-mediated intrinsic homeostasis protects neuronal networks from hyperexcitability. *EMBO J.* 2013;32:2994–3007.
69. Ariano P, Zamburini P, D'Alessandro R, Meldolesi J, Lovisolo D. Differential repression by the transcription factor REST/NRSF of the various Ca<sup>2+</sup> signalling mechanisms in pheochromocytoma PC12 cells. *Cell Calcium.* 2010;47:360–8.
70. Uchida H, Sasaki K, Ma L, Ueda H. Neuron-restrictive silencer factor causes epigenetic silencing of Kv4.3 gene after peripheral nerve injury. *Neuroscience.* 2010;166:1–4.
71. Uvarov P, Pruunsild P, Timmusk T, Airaksinen MS. Neuronal K<sup>+</sup>/Cl<sup>-</sup> co-transporter (KCC2) transgenes lacking neurone restrictive silencer element recapitulate CNS neurone-specific expression and developmental up-regulation of endogenous KCC2 gene. *J Neurochem.* 2005;95:1144–55.
72. Rodenas-Ruano A, Chávez AE, Cossio MJ, Castillo PE, Zukin RS. REST-dependent epigenetic remodeling promotes the developmental switch in synaptic NMDA receptors. *Nat Neurosci.* 2012;15:1382–90.
73. Chun SK, Cheol KH, Hack SC, Song KY, Law P-Y, Wei L-N, et al Neuron-restrictive Silencer Factor (NRSF) functions as a repressor in neuronal cells to regulate the  $\mu$  opioid receptor gene. *J Biol Chem.* 2004;279:46464–73.
74. D'Alessandro R, Klajn A, Meldolesi J. Expression of dense-core vesicles and of their exocytosis are governed by the repressive transcription factor NRSF/REST. *Ann N Y Acad Sci.* 2009;1152:194–200.
75. Navarrete-Modesto V, Orozco-Suárez S, Feria-Romero IA, Rocha L. The molecular hallmarks of epigenetic effects mediated by antiepileptic drugs. *Epilepsy Res.* 2019;149:53–65.
76. Kong F-C, Ma C-L, Zhong M-K. Epigenetic effects mediated by antiepileptic drugs and their potential application. *Curr Neuropharmacol.* 2019;18:153–66.
77. Göttlicher M, Minucci S, Zhu P, Krämer OH, Schimpf A, Giavara S, et al Valproic acid defines a novel class of HDAC inhibitors inducing differentiation of transformed cells. *EMBO J.* 2001;20:6969–78.
78. Rong H, Liu TB, Yang KJ, Yang HC, Wu DH, Liao CP, et al MicroRNA-134 plasma levels before and after treatment for bipolar mania. *J Psychiatr Res.* 2011;45:92–5.
79. Beutler AS, De LS, Nicol R, Walsh MJ. Carbamazepine is an inhibitor of histone deacetylases. *Life Sci.* 2005;76:3107–15.
80. Houtepen LC, Van Bergen AH, Vinkers CH, Boks MPM. DNA methylation signatures of mood stabilizers and antipsychotics in bipolar disorder. *Epigenomics.* 2016;8:197–208.
81. Eyal S, Yagen B, Sobol E, Altschuler Y, Shmuel M, Bialer M. The activity of antiepileptic drugs as histone deacetylase inhibitors. *Epilepsia.* 2004;45:737–44.
82. Echeverry-Alzate V, Giné E, Bühler KM, Calleja-Conde J, Olmos P, Gorriti MA, et al Effects of topiramate on ethanol-cocaine interactions and DNA methyltransferase gene expression in the rat prefrontal cortex. *Br J Pharmacol.* 2014;171:3023–36.
83. Bang SR, Ambavade SD, Jagdale PG, Adkar PP, Waghmare AB, Ambavade PD. Lacosamide reduces HDAC levels in the brain and improves memory: Potential for treatment of Alzheimer's disease. *Pharmacol Biochem Behav.* 2015;134:65–9.
84. You D, Wen X, Gorczyca L, Morris A, Richardson JR, Aleksunes LM. Increased MDR1 transporter expression in human brain endothelial cells through enhanced histone acetylation and activation of aryl hydrocarbon receptor signaling. *Mol Neurobiol.* 2019;56:6986–7002.
85. Lazarowski A, Czornyj L. Potential role of multidrug resistant proteins in refractory epilepsy and antiepileptic drugs interactions. *Drug Metabol Drug Interact.* 2011;26:21–6.
86. Griminger T, Pernhorst K, Surges R, Niehusmann P, Priebe L, Lehe M, et al Levetiracetam resistance: Synaptic signatures & corresponding promoter SNPs in epileptic hippocampi. *Neurobiol Dis.* 2013;60:115–25.
87. Doupnik CA. RGS Redundancy and Implications in GPCR-GIRK Signaling, 1st edn. Elsevier Inc.; 2015. <https://doi.org/10.1016/bs.irn.2015.05.010>
88. Onfroy L, Galandrin S, Pontier SM, Seguelas M-H, N'Guyen DU, Sénard J-M, et al G protein stoichiometry dictates biased agonism through distinct receptor-G protein partitioning. *Sci Rep.* 2017;7:1–14.
89. González-Maeso J. GPCR oligomers in pharmacology and signaling. *Mol Brain.* 2011;4:1–7.
90. Kumari N, Yadav S. Modulation of protein oligomerization: An overview. *Prog Biophys Mol Biol.* 2019;149:99–113.
91. Ali MH, Imperiali B. Protein oligomerization: How and why. *Bioorg Med Chem.* 2005;13:5013–20.
92. Hebert TE, Moffett S, Morello JP, Loisel TP, Bichet DG, Barret C, et al A peptide derived from a  $\beta$  2-adrenergic receptor transmembrane domain inhibits both receptor dimerization and activation. *J Biol Chem.* 1996;271:16384–92.
93. Romano C, Yang WL, O'Malley KL. Metabotropic glutamate receptor 5 is a disulfide-linked dimer. *J Biol Chem.* 1996;271:28612–6.
94. Ng GYK, O'Dowd BF, Lee SP, Chung HT, Brann MR, Seeman P, et al Dopamine D2 receptor dimers and receptor-blocking peptides. *Biochem Biophys Res Commun.* 1996;227:200–4.
95. Kaupmann K, Malitschek B, Schuler V, Heid J, Froestl W, Beck P, et al GABA(B)-receptor subtypes assemble into functional heteromeric complexes. *Nature.* 1998;396:683–7.
96. Lee FJS, Xue S, Pei L, Vukusic B, Chéry N, Wang Y, et al Dual regulation of NMDA receptor functions by direct

- protein-protein interactions with the dopamine D1 receptor. *Cell*. 2002;111:219–30.
97. Borroto-Escuela DO, Dupont CM, Li X, Savelli D, Lattanzi D, Srivastava I, et al Disturbances in the FGFR1-5-HT1A heteroreceptor complexes in the raphe-hippocampal 5-HT system develop in a genetic rat model of depression. *Front Cell Neurosci*. 2017;11:1–12.
  98. Flajolet M, Wang Z, Futter M, Shen W, Nuangchamnon N, Bendor J, et al FGF acts as a co-transmitter through adenosine A2A receptor to regulate synaptic plasticity. *Nat Neurosci*. 2008;11:1402–9.
  99. Clatot J, Hoshi M, Wan X, Liu H, Jain A, Shinlapawittayatorn K, et al Voltage-gated sodium channels assemble and gate as dimers. *Nat Commun*. 2017;8:1–14.
  100. Clatot J, Zheng Y, Girardeau A, Liu H, Laurita KR, Marionneau C, et al Mutant voltage-gated Na<sup>+</sup> channels can exert a dominant negative effect through coupled gating. *Am J Physiol Hear Circ Physiol*. 2018;315:H1250–7.
  101. Sokolov MV, Henrich-Noack P, Raynoschek C, Franzén BO, Larsson O, Main M, et al Co-expression of  $\beta$  subunits with the voltage-gated sodium channel NaV1.7: the importance of subunit association and phosphorylation and their effects on channel pharmacology and biophysics. *J Mol Neurosci*. 2018;65:154–66.
  102. Li J, Maghera J, Lamothe SM, Marco EJ, Kurata HT. Heteromeric assembly of truncated neuronal Kv7 channels: Implications for neurologic disease and pharmacotherapy. *Mol Pharmacol*. 2020;98:192–202.
  103. Wang J, Shen D, Xia G, Shen W, Macdonald RL, Xu D, et al Differential protein structural disturbances and suppression of assembly partners produced by nonsense GABRG2 epilepsy mutations: Implications for disease phenotypic heterogeneity. *Sci Rep*. 2016;6:1–16.
  104. Nooren IMA, Thornton JM. Structural characterisation and functional significance of transient protein-protein interactions. *J Mol Biol*. 2003;325:991–1018.
  105. Borroto-Escuela DO, Carlsson J, Ambrogini P, Narváez M, Wydra K, Tarakanov AO, et al Understanding the role of gpcr heteroreceptor complexes in modulating the brain networks in health and disease. *Front Cell Neurosci*. 2017;11:1–20.
  106. Schmidt D, Löscher W. Drug resistance in epilepsy: Putative neurobiologic and clinical mechanisms. *Epilepsia*. 2005;46:858–77.
  107. Fuxe K, Marcellino D, Guidolin D, Woods AS, Agnati L. Brain receptor mosaics and their intramembrane receptor-receptor interactions: molecular integration in transmission and novel targets for drug Development. *JAMS J Acupunct Meridian Stud*. 2009;2:1–25.
  108. López-Cano M, Fernández-Dueñas V, Ciruela F. Proximity ligation assay image analysis protocol: Addressing receptor-receptor interactions. *Methods Mol Biol*. 2019;2040:41–50.
  109. Dale NC, Johnstone EKM, White CW, Pflieger KDG. NanoBRET: The bright future of proximity-based assays. *Front Bioeng Biotechnol*. 2019;7:1–13.
  110. Ginés S, Hillion J, Torvinen M, Le Crom S, Casado V, Canela EI, et al Dopamine D1 and adenosine A1 receptors form functionally interacting heteromeric complexes. *Proc Natl Acad Sci USA*. 2000;97:8606–11.
  111. Fuxe K, Marcellino D, Rivera A, Diaz-Cabiale Z, Filip M, Gago B, et al Receptor-receptor interactions within receptor mosaics. Impact on neuropsychopharmacology. *Brain Res Rev*. 2008;58:415–52.
  112. Jordan BA, Devi LA. G-protein-coupled receptor heterodimerization modulates receptor function. *Nature*. 1999;399:697–700.
  113. Agnati LF, Ferre S, Burioni R, Woods A, Genedani S, Franco R, et al Existence and theoretical aspects of homomeric and heteromeric dopamine receptor complexes and their relevance for neurological diseases. *NeuroMolecular Med*. 2005;7:61–78.
  114. Borroto-Escuela DO, Wydra K, Romero-Fernandez W, Zhou Z, Frankowska M, Filip M, et al A2AR transmembrane 2 peptide administration disrupts the A2AR-A2AR homoreceptor but not the A2AR-D2R heteroreceptor complex: Lack of actions on rodent cocaine self-administration. *Int J Mol Sci*. 2019;20:6100. <https://doi.org/10.3390/ijms20236100>
  115. Agnati LF, Fuxe K, Zoli M, Ogren SO. New vistas on synaptic plasticity: the receptor mosaic hypothesis of the engram. *Med Biol*. 1982;60:183–90.
  116. Agnati LF, Tarakanov AO, Ferré S, Fuxe K, Guidolin D. Receptor-receptor interactions, receptor mosaics, and basic principles of molecular network organization - Possible implications for drug development. *J Mol Neurosci*. 2005;26:193–208.
  117. Fuxe K, Borroto-Escuela DO, Romero-Fernandez W, Palkovits M, Tarakanov AO, Ciruela F, et al Moonlighting proteins and protein-protein interactions as neurotherapeutic targets in the G protein-coupled receptor field. *Neuropsychopharmacology*. 2014;39:131–55.
  118. Higley MJ, Sabatini BL. Competitive regulation of synaptic Ca<sup>2+</sup> influx by D2 dopamine and A2A adenosine receptors. *Nat Neurosci*. 2010;13:958–66.
  119. Borroto-Escuela DO, De La Mora MP, Manger P, Narváez M, Beggiato S, Crespo-Ramírez M, et al Brain dopamine transmission in health and Parkinson's disease: Modulation of synaptic transmission and plasticity through volume transmission and dopamine heteroreceptors. *Front Synaptic Neurosci*. 2018;10:1–24.
  120. González-Maeso J, Ang RL, Yuen T, Chan P, Weisstaub NV, López-Giménez JF, et al Identification of a serotonin/glutamate receptor complex implicated in psychosis. *Nature*. 2008;452:93–7.
  121. Tanganelli S, Antonelli T, Tomasini MC, Beggiato S, Fuxe K, Ferraro L. Relevance of dopamine D2/neurotensin NTS1 and NMDA/neurotensin NTS1 receptor interaction in psychiatric and neurodegenerative disorders. *Curr Med Chem*. 2012;19:304–16.
  122. Wischhof L, Koch M. 5-HT2A and mGlu2/3 receptor interactions: On their relevance to cognitive function and psychosis. *Behav Pharmacol*. 2016;27:1–11.
  123. Filip M, Frankowska M, Zaniewska M, Przegaliński E, Müller CE, Agnati L, et al Involvement of adenosine A2A and dopamine receptors in the locomotor and sensitizing effects of cocaine. *Brain Res*. 2006;1077:67–80.
  124. Liu XY, Chu XP, Mao LM, Wang M, Lan H-X, Li M-H, et al Modulation of D2R-NR2B interactions in response to cocaine. *Neuron*. 2006;52:897–909.
  125. Borroto-Escuela DO, Romero-Fernandez W, Mudó G, Pérez-Alea M, Ciruela F, Tarakanov AO, et al Fibroblast growth factor receptor 1 5-hydroxytryptamine 1A heteroreceptor complexes and their enhancement of hippocampal plasticity. *Biol Psychiatry*. 2012;71:84–91.

126. Everitt BJ. Neural and psychological mechanisms underlying compulsive drug seeking habits and drug memories - indications for novel treatments of addiction. *Eur J Neurosci*. 2014;40:2163–82.
127. Borroto-Escuela DO, Fuxe K. Oligomeric receptor complexes and their allosteric receptor-receptor interactions in the plasma membrane represent a new biological principle for integration of signals in the CNS. *Front Mol Neurosci*. 2019;12:1–17.
128. Fuxe K, Guidolin D, Agnati LF, Borroto-Escuela DO. Dopamine heteroreceptor complexes as therapeutic targets in Parkinson's disease. *Expert Opin Ther Targets*. 2015;19:377–98.
129. Singh SS, Jois SD. Homo- and heterodimerization of proteins in cell signaling: inhibition and drug design, 1st edn. Elsevier Inc.; 2018. <https://doi.org/10.1016/bs.apcsb.2017.08.003>
130. Engel J. Intracerebral recordings: organization of the human epileptogenic region. *J Clin Neurophysiol*. 1993;10:90–8.
131. Oliver A. Surgery of epilepsy: overall procedure. In: Apuzzo MLJ, editor. *Neurosurgical aspects of epilepsy*. American Association of Neurological Surgeons; 1991. pp. 117–48.
132. Ondarza R, Trejo-Martínez D, Corona-Amézcuca R, Briones M, Rocha L. Evaluation of opioid peptide and muscarinic receptors in human epileptogenic neocortex: An autoradiography study. *Epilepsia*. 2002;43:230–4.
133. Dacht F, Bagla S, Keren-Aviram G, Morton A, Balan K, Saadat L, et al Predicting novel histopathological microlesions in human epileptic brain through transcriptional clustering. *Brain*. 2015;138:356–70.
134. Lipovich L, Dacht F, Cai J, Bagla S, Balan K, Jia H, et al Activity-dependent human brain coding/noncoding gene regulatory networks. *Genetics*. 2012;192:1133–48.
135. Statello L, Guo CJ, Chen LL, Huarte M. Gene regulation by long non-coding RNAs and its biological functions. *Nat Rev Mol Cell Biol*. 2021;22:96–118.

**How to cite this article:** Fonseca-Barriendos D, Frías-Soria CL, Pérez-Pérez D, Gómez-López R, Borroto Escuela DO, Rocha L. Drug-resistant epilepsy: Drug target hypothesis and beyond the receptors. *Epilepsia Open*. 2022;7(Suppl. 1):S23–S33. <https://doi.org/10.1002/epi4.12539>