



Two Echinostome Species, *Pegosomum bubulcum* and *Nephrostomum ramosum* (Digenea: Echinostomatidae), from an Eastern Cattle Egret, *Bubulcus ibis coromandus*, in Republic of Korea

Seongjun Choe¹, Dongmin Lee¹, Hansol Park¹, Hyeong-Kyu Jeon¹, Youngsun Lee², Eunju Kim², Ki-Jeong Na^{2,3}, Keeseon S. Eom^{1,*}

¹Department of Parasitology, Medical Research Institute and Parasite Resource Bank, School of Medicine, Chungbuk National University, Cheongju 28644, Korea; ²The Wildlife Center of Chungbuk, Cheongju 28116, Korea; ³College of Veterinary Medicine, Chungbuk National University, Cheongju 28644, Korea

Abstract: We describe 2 echinostome species recovered from an Eastern cattle egret, *Bubulcus ibis coromandus*, from Cheongju-si (city), Chungcheongbuk-do (province), Korea. Total 72 *Pegosomum bubulcum* specimens were recovered from the bile duct. They were 7,566 × 2,938 μm in average size and had 27 collar spines with vitelline extension from anterior 1/3 level of the esophagus to mid-level of the posterior testis as characteristic features. Total 9 specimens of *Nephrostomum ramosum* were recovered in the small intestines of the bird. They were ribbon-shaped, 11,378 × 2,124 μm in average size, and morphologically variable in some organs, i.e., the number of collar spines (47-50), the shape of ovary and testes, and the extension of vitelline follicles. These morphological variations observed in a single host indicated that these features are not critical for the classification of *Nephrostomum* species and thus were reconsidered taxonomically as synonym of *N. ramosum*. This study is the first report documenting and describing both flukes and their associated genera in Korea.

Key words: *Pegosomum bubulcum*, *Nephrostomum ramosum*, *Bubulcus ibis coromandus*, echinostome, parasite, cattle egret

INTRODUCTION

The cattle egret, *Bubulcus ibis* (Family Ardeidae) is a piscivorous bird that is characterized by dull orange feathers on the chest and head regions during the breeding season. It exhibits a cosmopolitan distribution and feeds on various preys such as fish, insects, and frogs. The Eastern cattle egret, *Bubulcus ibis coromandus*, has a wide distribution ranging from Eastern Pakistan to Eastern Australia [1]. In the 1960s, the Eastern cattle egret was rarely observed in Korea, and almost all of its populations were observed only in the southern part of the Korean peninsula [1,2]. Today, however, it is commonly observed throughout Korea near rice fields, grass fields, and wetlands. It migrates to Korea only in the summer season (mid-April to

late September) [1]. Since the Eastern cattle egret, like other herons and egrets, has indiscriminate food habits, it is considered a host for many trematode parasites.

Cattle egret carries *Pegosomum* sp., *Apharyngostrigea ardeolina*, *Metorchis ozakii*, and *Stephanoprora ozakii* in Japan [3-6], and *Apharyngostrigea ramai*, *Nephrostomum bicolanum*, *Pegosomum egretti*, and *Uroproctepisthmium taiwanense* in Taiwan [7]. However, the helminth parasites of this avian species have never been reported previously in Korea. During the investigation for the avian helminth fauna of Korea, 2 unrecorded echinostome species, *Pegosomum bubulcum* and *Nephrostomum ramosum*, were recovered from an Eastern cattle egret, *B. ibis coromandus*. Therefore, we described the morphological and faunistic characteristics of these 2 echinostomes, and compared deeply with those of previous studies.

MATERIALS AND METHODS

A male *B. ibis coromandus* (WCC No.: 20140205) was found in June 2014 in the town of Cheongju-si (city), Chungcheong-

•Received 19 January 2016, revised 15 May 2016, accepted 8 June 2016.

*Corresponding author (kseom@chungbuk.ac.kr)

© 2016, Korean Society for Parasitology and Tropical Medicine

This is an Open Access article distributed under the terms of the Creative Commons Attribution Non-Commercial License (<http://creativecommons.org/licenses/by-nc/4.0>) which permits unrestricted non-commercial use, distribution, and reproduction in any medium, provided the original work is properly cited.

buk-do (province), Korea, near its breeding colony. The egret was rescued and treated by the Wildlife Center of Chungbuk (WCC), but it died soon after. It was then transferred to the Parasite Resource Bank of Korea from the WCC for parasitological examination. In the autopsy of egret, 1 species of trematode was recovered in the biliary system and other trematode species was collected in the small intestines. The collected flukes were rinsed in phosphate buffer several times and placed with 10% buffered formalin under a cover glass with slight pressure for microscopic observation after staining with Semichon's acetocarmine. The various structures of the worms were observed under a light microscope and measured.

Abbreviations used in Tables and descriptions are as follows: BL: body length; BW: maximum body width; CL: collar length; CW: collar width; NCS: number of collar spine; OSL: oral

sucker length; OSW: oral sucker width; PL: prepharynx length; PHL: pharynx length; PHW: pharynx width; ESL: esophagus length; CSL: cirrus sac length; CSW: cirrus sac width; SVL: seminal vesicle length; SVW: seminal vesicle width; VSL: ventral sucker length; VSW: ventral sucker width; OVL: ovary length; OVW: ovary width; MEL: Mehlis' gland length; MEW: Mehlis' gland width; ATL: anterior testis length; ATW: anterior testis width; PTL: posterior testis length; PTW: posterior testis width; EL: egg length; EW: egg width; FO: forebody length (distance from the anterior end to the anterior margin of the ventral sucker); U: uterine field length (distance from the anterior margin of the ovary to the posterior margin of the ventral sucker); and T: post-testicular field length. The size and proportions of the standard structures relative to the body length were calculated as described by Kostadinova [8]. Each propor-

Table 1. Morphometric comparison of *Pegosomum bubulcum* with other *Pegosomum* species

Data source Species Location	Present study <i>Pegosomum bubulcum</i> Korea	Tubangui and Masilungan (1935) <i>P. bubulcum</i> Philippines	MacCallum (1918) <i>P. herodiae</i> Indonesia
Host	<i>Bubulcus ibis coromandus</i>	<i>B. i. coromandus</i>	<i>Ardea modesta</i>
BL	6,593-8,635 (7,566)	5,000-6,600	8,000
BW	2,804-3,321 (2,938)	1,700-2,300	3,000
CL			
CW	343-461 (381)	340-360	450
OSL			
OSW	88-157 (127)	140-150	
PL	0-20 (5)		
PHL	225-274 (245)	260-280	150 ^a
PHW	147-176 (161)	150-170	100 ^a
ESL	833-1,538 (1,043)	1,240-1,460	
CSL	1,568-2,257 (1,856) ^b	320-470	1,000 ^{a,b}
CSW		500-600	
VSL	686-843 (776)		1,000 ^a
VSW	763-892 (817)	680-830	
OVL	333-470 (408)	190-420	500 ^a
OVW	421-774 (574)	280-300	
MEL	500-886 (644)		
MEW	813-1,470 (1,058)		
ATL	886-1,402 (1,156)	380-410	
ATW	2,042-2,411 (2,163)	1,180-1,520	1,400
PTL	983-1,402 (1,153)	570-820	1,000
PTW	1,476-1,820 (1,661)	570-1,200	600
EL	108-121 (115)	115-123	70 ^a
EW	66-74 (71)	73-77	40 ^a
FO	2,066-3,050 (2,378)		
U	221-467 (366)		
T	910-1,328 (1,111)		
BW/BL	36-43 (39)%	34%	38%
FO/BL	29-35 (31)%	37%	24%
U/BL	3-6 (5)%	1%	5%
T/BL	12-19 (15)%	18%	14%

Measurements are in μm .

^aEstimated by present authors.

^bProjected cirrus sac length.

Table 2. Morphometric comparison of *Nephrostomum ramosum* with other *Nephrostomum* species

Data Source	Present study	Various authors ^a	Tubangui (1933)	Travassos (1922)	Ukoi (1967)	Viguera (1944)	Ibáñez (1966)	Kasimov, Vaidova and Falaulaev (1958)	Kanyakarte (1969)	Gupta and Mehrotra (1970)	Agarwal (1968)	Gupta and Singh (1986)	Gupta (1983)	Khan and Ghazi (2011)	Kalkbaid and Bliques (1972)	
Species Location	<i>N. ramosum</i> Korea	<i>N. ramosum</i> Africa and Europe	<i>N. bimaculatum</i> Philippines	<i>N. jirai</i> Brazil	<i>N. legonum</i> Ghana	<i>N. robustum</i> Cuba	<i>N. sinchirocai</i> Peru	<i>N. skrjabini</i> Azerbaijan	<i>N. reticulatum</i> India	<i>N. chandragensis</i> India	<i>N. dolfusii</i> India	<i>N. guptai</i> India	<i>N. udajpurensis</i> India	<i>N. odelalensis</i> Pakistan	<i>N. dubashi</i> Pakistan	
Host	<i>Bubulcus ibis</i> coromandus	<i>Ardea</i> sp., <i>Bubulcus ibis</i> and <i>Egretta gazetta</i>	<i>Bubulcus ibis</i>	<i>Syrigma sibilatrix</i>	<i>Bubulcus ibis</i>	<i>Columbus dominicus dominicus</i>	<i>Egretta thula</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	<i>Bubulcus ibis</i>	Unknown
BL	7,947-13,579 (11,378)	8,000-18,000	8,100-15,150	15,000	9,680-12,650 (11,330)	6,500	16,670	7,000-10,500	11,820-13,270 (12,030)	12,285-16,477	8,440-13,360	10,410	8,200-9,000	7,800-8,820	9,050-10,900	
BW	1,796-2,411 (2,124)	2,000-3,400	1,400-2,400	2,200	2,320-2,700 (2,530)	1,800	2,930	2,300-3,000	2,770	2,457-2,730	1,950-2,900	2,060	2,000-2,240	1,280-1,310	2,090-2,500	
CL	541-738 (619)	880	1,080-1,400	630	610-750 (670)	533	533	1,150-1,460	45	NA	1,070-1,400	46-47	540-560	40-48 ^b	520-550	
CW	910-1,255 (1,111)	1,000-1,700	1,080-1,400	1,400	1,270-1,380 (1,330)	50	26	1,150-1,460	45	NA	46-47	47	1,100-1,200	40-48 ^b	860-860	
NCS	47-50	47-51	47	48	50	50	26	47	45	NA	46-47	47	46	40-48 ^b	27-34	
OSL	176-265 (225)	200-430	260-280	270	170-230 (190)	230	333	146-203	273-455	273-455	194-286	280-330	280-330	120-200	250	
OSW	245-343 (294)	280-360	280-360	270	200-380 (320)	230	270	140-200 (170)	318-364	318-364	208-286	340-360	340-360	130-220	280-380	
PL	29-147 (78)								182	52	52	0	100-170	100-170		
PHL	235-314 (266)	240-310	240	260	290-330 (300)	340	230-315	230-315	280	200-250	195-520	130	250-300	110-230	490-500	
PHW	157-255 (206)	160-230	220-240	200-240	170-230 (200)	245	532	209-377	220	150-300	208-260	120	200-300	100-200		
ESL	216-430 (381)	170-185	320-560	660-730	340	340	532	209-377	270-340 (300)	408-455	390-576	330	320-420	300-370		
CSL	363-578 (462)	360-560	360-560	510-520	720	720	524	524	520-560	320-960	320-960	380	380-390	430-530		
CSW	284-412 (325)	300-380	340-360	330	330	330	333-503	333-503	360-440	260-400	260-400	210	280-300	400-500		
SVL									240	256-490	256-490	350	210-220			
SWW									240	96-260	96-260	110	150-170			
VSL	1,068-1,343 (1,266)	1,250-1,560	1,040-1,340	1,400	1,270-1,470 (1,370)	1,045	1,800	945-1,407	1,520	1,456-1,638	1,200-1,380	1,200	1,160-1,200	660-1,260	1,150-1,180	
VSW	862-1,245 (1,083)	1,140-1,610	1,100-1,440	1,200	1,130-1,350 (1,270)	1,127	1,670	1,005-1,509	540	880-1,190	880-1,190	1,000	1,000-1,140	440-1,140	900-910	
OVL	176-480 (385)	240-360	240-540	640	300-470 (370)	240	370	377-752	280-340 (310)	455	240-440	390	300-340	230-270		
OWW	294-608 (492)	640-730	300-640	400	470-650 (530)	325	583	377-628	560-700 (630)	728-773	440-680	470	540-560	240-260		
MEL	382-529 (465)				330-610 (480)	583	583	330-610 (480)	819	819						
MEW	431-725 (580)				470-610 (570)	583	583	470-610 (570)	864	864						
ATL	402-1,068 (654)	620-690	460-1,440	1,200-1,300	500-760 (640)	820	570	985	680-1,280 (930)	1,047-1,192 ^c	480-1,150	490	650-710	630-650	800-820	
ATW	441-588 (524)	640	320-660	660-710	290-470 (410)	370	333	721-817	720-960 (830)	546-682	490-680	490	520-600	320-350	460-480	
PTL	617-1,078 (895)	620-690	1,000-1,100	1,000-1,100	500-720 (630)	420	420	630-1,048	940-1,000 (970)	910	482-1,000	590	530-880	580-630		
PTW	402-617 (516)	480	660	660	320-520 (420)	333	333	524-691	800	455-701	490-680	420	440-510	320-340		
EL	92-102 (98)	90-119	96-104	113-120	88-102	74	113	92-104	87-94 (90)	88-90	91-100	80-100	60-110	90-110	120-160	
EW	47-67 (53)	50-68	60-62	63-75	41-48	52	66	52-62	54-63 (58)	38-49	32-52	50	40-50	46-59 ^d	60-80	
FO	910-1,279 (1,144)		1,040-1,540						1,456-1,638			870				
U	1,378-4,748 (3,112)															
T	2,411-3,567 (3,137)		2,980-3,960 (3,590)									2970				
BW/BL ^e	16-23 (19%)	27%	21%	18%	21%	24%	15%	30%	20%	16%	26-27%	20%	28%	16%	23%	
FO/BL ^e	9-12 (10%)	8%	11%	10%	9%	18%	10%	9%	10%	9%	11-15%	9%	12%	8%	12%	
U/BL ^e	18-35 (26%)	24%	27%	31%	33%	7%	25%	23%	24%	36%	24-26%	28%	25%	27%	27%	
T/BL ^e	22-31 (28%)	35%	28%	23%	33%	22%	28%	27%	33%	25%	26-29%	29%	29%	25%	29%	

Measurements are in µm.
^aCombined measurements of Copher (1910), Mendheim (1943), and Skriabin and Baschkirova (1956) [12, 13, 22].
^bEstimated by present authors.
^cEstimated from the drawings of literatures. Measurements shown in µm.
^dEstimated from the drawings of literatures. Measurements shown in µm.

tion for previously described species was estimated from each drawing of the species. Measurements of the present specimens and those of previously described fluke species that resemble our specimens are shown in Tables 1 and 2.

RESULTS

Recovery of worms

Six parasite species were identified during parasitological examinations of the host. One was the echinostome *Pegosomum bubulcum* and 72 specimens of this species were collected from

the ball-like bulge in the biliary system. These flukes were pinkish red. Two other echinostome species were found in the small intestine of the host, namely, *Nephrostomum ramosum* (n=9) and *Echinochasmus japonicus* (n=2). Other than trematodes, *Desportesius invaginatus*, *Eustrongylides excisus*, and an unidentified nematode species were also recovered in the proventriculus.

Family Echinostomatidae Looss, 1899

Genus *Pegosomum* Ratz, 1903

Pegosomum bubulcum Tubangui & Masilungan, 1935

Materials examined: 20 of 72 specimens collected from a cat-

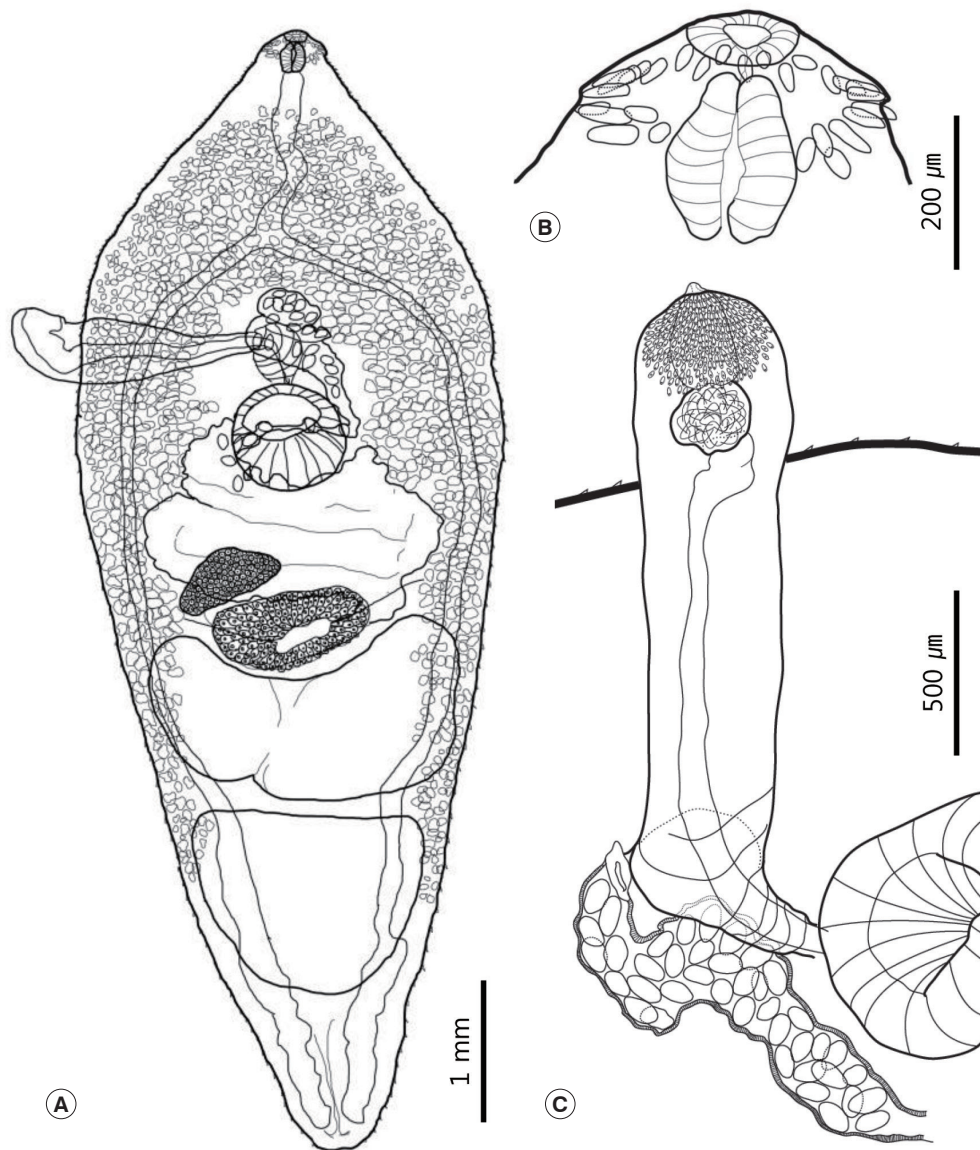


Fig. 1. Drawings of *Pegosomum bubulcum* specimens recovered from a cattle egret, *Bubulcus ibis coromandus*. (A) Whole body. (B) Head collar region. (C) Terminal genitalia with projected cirrus sac.

tle egret, *B. ibis coromandus*, from Cheongju-si (city), Chungcheongbuk-do (province), Korea (GPS: 36°36'56.9"N 127°28'54.9"E). Two of the specimens were deposited in the National Institute of Biological Resources in Korea (NIBR) (NIBR specimen no. KOSPIV0000221885 and KOSPIV0000221886).

The morphological characteristics were described with 20 intact mature adults (Fig. 1). Morphometric measurements of their features are provided in Table 1. Body medium sized, plump, and leaf-like. Maximum width of the body at the level of the ventral sucker (BW/BL=36-43%). Tegument covered with minute spines, which distributed in the whole body and

their density decreased posteriorly (Fig. 1A). Head small with a weakly developed head collar. Oral sucker subterminal to terminal, subspherical, and surrounded by 27 collar spines. Four pairs of angle spines slightly larger than the other spines, 79-92 (86) long and 15-20 (17) wide. Lateral spines 55-69 (63) long and 12-17 (14) wide. Dorsal spines arranged in double rows, 50-60 (53) long and 10-15 (12) wide (Fig. 1B). Prepharynx short. Pharynx strongly muscular, had an elongated oval shape, and larger than the oral sucker (Fig. 1B). Esophagus had minute diverticula that obscured in most specimens. Forebody very long (FO/BL=29-35%). Ventral sucker relative-

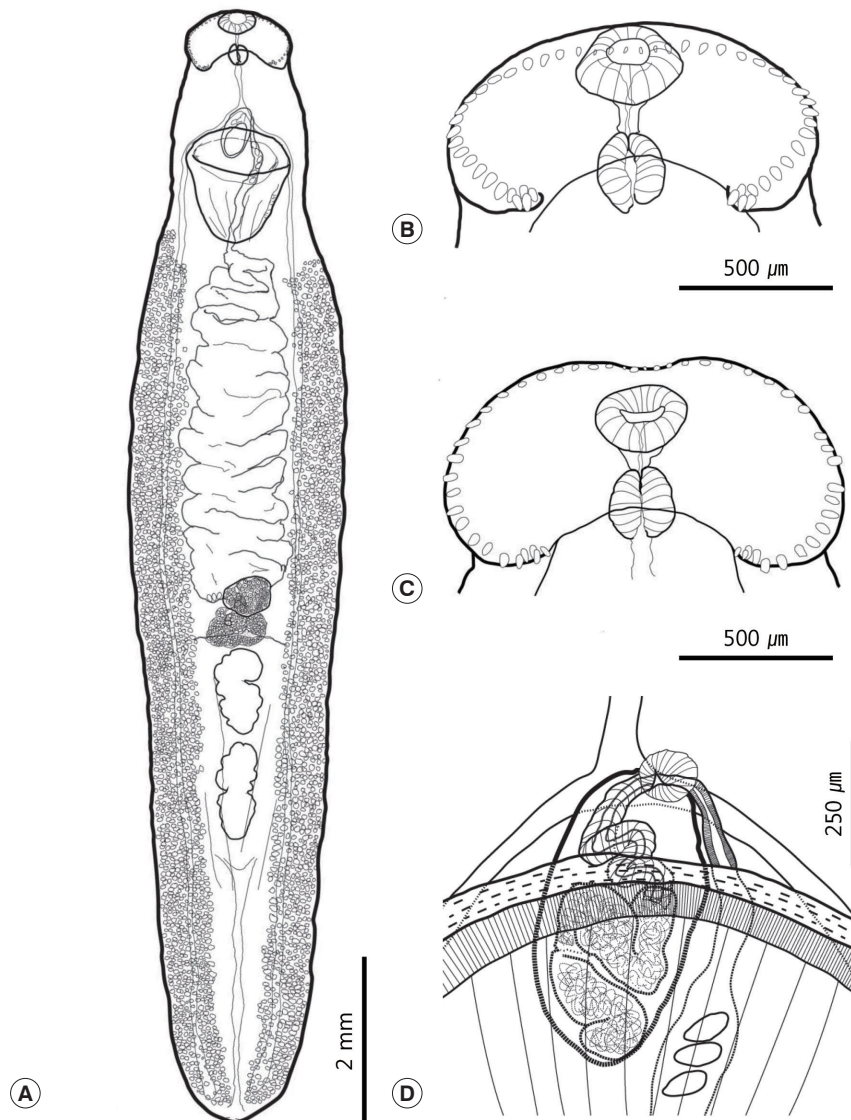


Fig. 2. Drawings of *Nephrostomum ramosum* specimens recovered from a cattle egret, *Bubulcus ibis coromandus*. (A) Whole body. (B) Head region of a specimen whose collar was lifted by the specimen fixation process. (C) Head region of a specimen with a well-positioned collar that made it possible to observe the dorsal recess. (D) Cirrus sac region containing the convoluted seminal vesicle.

ly small, muscular, and spherical. It located behind the intestinal bifurcation. Intestinal caeca lay along both sides of the body and ran to the posterior end.

The cirrus sac projected between the intestinal bifurcation and the ventral sucker. It longer than half of the maximum width of the body. It had a long balloon shape and armed with minute spines on its distal end. Long vas deferens connected with the spherical to oval seminal vesicle. The cirrus surrounded by prostate glands (Fig. 1C). Two large tandem testes located at the posterior half of the body. Anterior testis had a horse-shoe shape, while the posterior testis shaped as either a blunt-ended triangle or a trapezoid. The post-testicular field was short ($T/BL = 12-19\%$).

Ovary small, round to oval shaped, and located slightly to the dextral side of the body. Mehlis' gland located at the median between the ovary and the anterior testis. It fitted almost exactly in the concave region of the anterior margin of the anterior testis. Vitelline follicles distributed slightly behind the pharynx to the level of the posterior testis. Vitelline reservoir located at the center of Mehlis' gland. Vitelline field had a reverse U-shape. The relative length of the uterine field very short ($U/BL = 3-6\%$). Uterus slightly overlapped with Mehlis' gland and the ovary. Genital pore located anterior to the cirrus sac. The eggs yellowish brown. 108-121 (115) long and 66-74 (71) wide.

Genus *Nephrostomum* Dietz, 1909

Nephrostomum ramosum Sonsino, 1895

Materials examined: Six of 9 specimens collected from a *B. ibis coromandus* from Cheongju-si (city), Chungcheongbuk-do (province), Korea (GPS: 36°36'56.9"N 127°28'54.9"E). Two of the specimens were deposited in the NIBR (NIBR specimen numbers: KOSPIV0000221887 and KOSPIV0000221888).

The morphological characteristics were described with 6 adult specimens (Fig. 2). Morphometric measurements of their features are provided in Table 2. While all 6 worms had similar morphological characteristics, variation was also observed. Body large, elongated, and ribbon shaped. Maximum width of the body at the level of the mid-uterus region ($BW/BL = 16-23\%$). Tegumental spines absent (Fig. 2A). Head collar well developed, longer than wide, and reniform. Collar exhibited a dorsal recess and the ventral notch very wide. Ventral ridges well developed. Collar spines small and numbered 47-50. Angle spines 4-6 pairs, comprised a double row, 40-60 (53) long and 15-32 (25) wide. The other spines arranged in a continuous row. Lateral spines 30-64 (52) long and 20-32 (25) wide. Dorsal spines relatively smaller than other spines, being 12-35 (26) long and 10-15 (13) wide. Prepharynx short. Oral sucker subterminal and located at the median region of the collar. It small and spherical. Prepharynx short. Pharynx small (somewhat similar to the oral sucker in size) and had a slight-

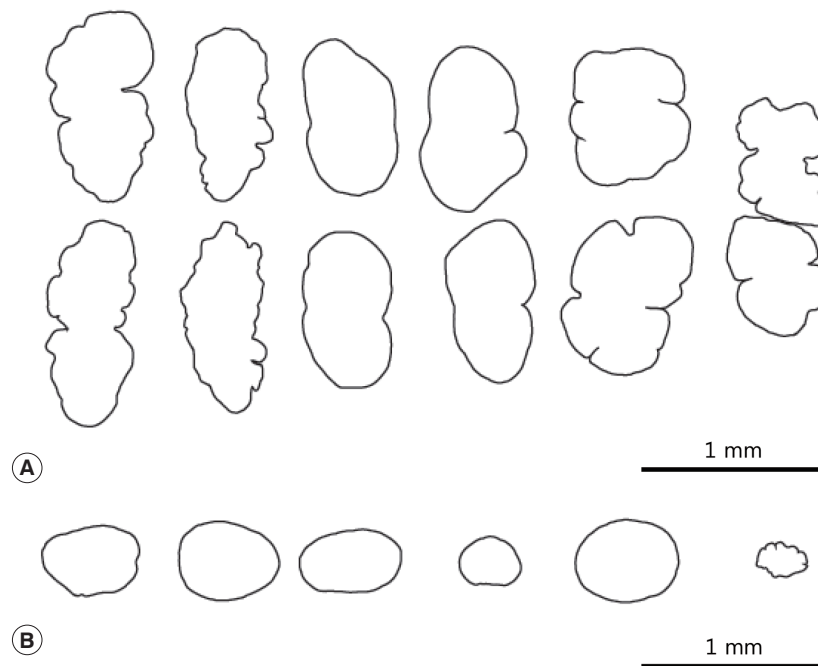


Fig. 3. Morphological variation of the testes (A) and the ovary (B) of *Nephrostomum ramosum*.

ly elongated oval shape (Fig. 2B, C). Forebody short (FO/BL=9-12%). Ventral sucker well developed, muscular, and cup-shaped. Intestinal bifurcation located just anterior to the ventral sucker. Intestinal caecum reached the posterior extremity along the median region of the vitelline field. Excretory vesicle narrow and had several branches.

Cirrus sac oval and relatively small. It located anterodorsally of the ventral sucker. It composed of a long convoluted seminal vesicle that connected to convoluted cirrus (Fig. 2D). There were 2 tandem testes located 3 quarters of the way down the body. They either had an elongated oval shape, were constricted medially, or irregularly lobed (Fig. 3A). Post-testicular field very long (T/BL=22-31%). Ovary generally round and small, but also showed variations in shape and size (Fig. 3B). It located slightly posterior to the equatorial region of the body. Mehlis' gland lay behind the ovary. The vitelline reservoir located at the center of Mehlis' gland. Vitelline follicles very small. They extended anteriorly to the mid-level of the ventral sucker or slightly behind the ventral sucker, and extended posteriorly to the posterior end along each side of the body. Uterine field length long (U/BL=18-35%). Uterus coiled and filled with numerous eggs. Eggs yellowish brown, 92-102 (98) long and 47-67 (53) wide.

DISCUSSION

Among 3 echinostome species recovered in the present study, 2 species, namely, *Pegosomum bubulcum* and *Nephrostomum ramosum*, are the new members of Korean trematode fauna. Choe et al. [9] recently reported 3 echinostome species, i. e., *Patagifer bilobus*, *Petasiger neocomense* and *Saakotrema metatestis*, and they also described another echinostome species, *Chaunocephalus ferox*, from the oriental white stork *Ciconia boyciana* (in press). By the Choe et al. [9], total 14 echinostome species including *E. japonicus* had been reported in Korea before their studies. However, they erroneously omitted an echinostome species, *Hypoderaeum conoideum* from their list [9,10]. Accordingly, a total of 21 echinostome species including 2 by the present study are to be listed in the Korean trematode fauna.

The morphological characteristics of specimens expelled from the ball-like bulge on the liver agreed well with those of the genus *Pegosomum* [8]. Ratz [11] erected the genus *Pegosomum* by including 3 species, namely, *P. asperum* (Wright, 1879) Ratz, 1903, *P. saginatum* (Ratz, 1897) Ratz, 1903, and *P. spiniferum* Ratz, 1903. Thereafter, Mendheim [12] recognized 4 species of

Pegosomum as valid species, *P. asperum*, *P. bubulcum* Tubanguui and Masilungan, 1935, *P. saginatum*, and *P. spiniferum*. Subsequently, Skrjabin and Bashkirova [13] recognized 3 additional valid species, namely, *P. herodiae* (McCallum, 1918) Skrjabin and Bashkirova, 1956, *P. petrovi* Kurashvili, 1949, and *P. skrjabini* Shakhtakhtinskaya, 1949. Although *P. asperum* was not mentioned by Skrjabin and Bashkirova, it was recognized as a valid species by Aleksandrova and Podgornova and they considered *P. spiniferum* to be its synonym [14]. In the latest publication regarding the genus *Pegosomum*, Unar et al. [15] recognized 5 additional species, *P. indicum* Saksena, 1960, *P. ixobrychi* Gvozdev, 1960, *P. lucknowensis* Pandey, 1973, *P. egretti* Srivastava, 1957, and *P. munifi* Dharejo, 2006, and they also added *P. garzettae* Ulnar, 2011 as a new species. Therefore, excluding *P. spiniferum* (which may be a synonym of *P. asperum*), 11 species were recently recognized as valid species of *Pegosomum* [15]. However, an earlier review by Aleksandrova suggested that on the basis of several features (body length and width ratio, number of collar spines, presence and shape of intestinal diverticula, and distribution of the vitelline follicles), only 5 species are valid, namely, *P. asperum*, *P. saginatum*, *P. herodiae*, *P. egretti*, and *P. ixobrychi*, and that the remaining species (*P. spiniferum*, *P. skrjabini*, *P. petrovi*, *P. bubulcum*, and *P. ixobrychi pici*) are not valid [16].

The vitelline follicles of our *Pegosomum* specimens differed from some of the previously described species of the genus. They were limited to the middle of the posterior testis, whereas they reach the posterior extremity in *P. asperum*, *P. petrovi*, *P. saginatum*, *P. skrjabini*, *P. spiniferum*, and *P. garzettae* [11-16]. In *P. ixobrychi*, they only extend to the anterior testis and members of the species have diverticula in the esophagus and intestine [17]. *P. indicum*, *P. lucknowensis*, and *P. munifi*, which have all been reported from Indo-Pakistan regions, also differ from our specimens in regard to vitelline distribution. Their vitelline follicles extend to the posterior end of the body [15,18]. Moreover, *P. egretti* has 25 spines, whereas our specimens had 27 [18]. Only *P. bubulcum* and *P. herodiae* are largely consistent with our specimens in terms of morphology [19,20]. Although Aleksandrova [16] considered *P. bubulcum* to be a synonym of *P. herodiae*, we do not agree with this assessment because the descriptions of *P. herodiae* were incomplete, as shown by the absence of details in the original description regarding the number of spines [20]. In addition, *P. herodiae* seems to differ from our specimens and the drawing of *P. bubulcum* in terms of the shape of the testes and their distribution [20]. The drawing of *P. herodiae* shows that the anterior testis is irregularly square and the posterior tes-

tis has a somewhat blunt conical shape. Moreover, both testes were limited to a region between the intestines and did not overlap [20]. In contrast, the testes of our specimens overlapped with the intestines and their shapes were that of a horseshoe and blunt-ended trapezoid, respectively. This suggests that our specimens are not *P. herodiae*. Since the type host of *P. bubulcum* is the cattle egret *B. ibis coromandus* (= *Bubulcus coromandus*), and this was also the host of our specimens [19], we recognize the validity of *P. bubulcum* and consider our specimens to be *P. bubulcum*. Notably, there are some differences between our specimens and *P. bubulcum* in terms of distribution of the vitel-

line follicles surrounding the esophagus, which in our specimens more resembles the true distribution of the vitelline follicles in *P. herodiae* [20]. However, the stained parenchyma of our specimens seemed similar to that shown in the drawing of *P. bubulcum* [19]. It is possible that the apparent distribution of vitelline follicles around the esophagus and post-pharynx region of *P. bubulcum* actually represents stained parenchyma. Finally, this is the first report detailing an armed cirrus and minute diverticula of the esophagus in *P. bubulcum* (Fig. 4A, B).

The flukes we recovered from the small intestines had 47-50 collar spines forming a single continuous row on a reniform

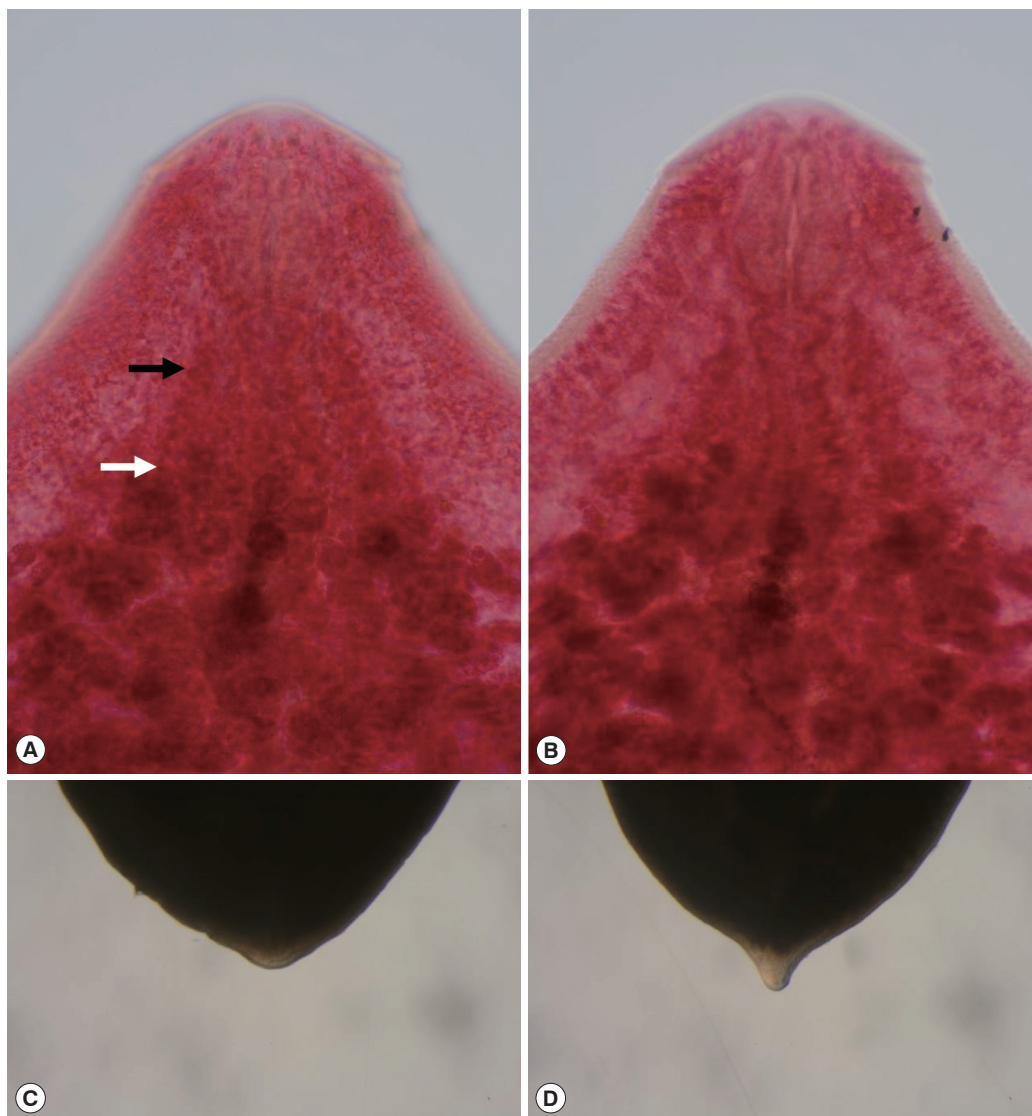


Fig. 4. Photographs of *Pegosomum bubulcum* (A, B) and *Nephrostomum ramosum* (C, D). (A) *P. bubulcum*. Anterior distribution of vitelline follicles. Black arrow: stained parenchyma, White arrow: true distribution of vitelline follicles. (B) Esophagus of *P. bubulcum* showing a minute diverticulum. (C) Blunt posterior end of *N. ramosum*. (D) Posterior end of *N. ramosum* that seems to be similar to that described by Ukoli [25].

head collar that was composed of a wide ventral notch and a shallow dorsal depression. These characteristics suggest that our specimens belong to the subfamily Nephrostominae Mendheim, 1943 [8,12]. This subfamily includes the genera *Nephrostomum* Dietz, 1909 and *Patagifer* Dietz, 1909. Only 1 species of this subfamily, *Patagifer bilobus*, has been recorded in Korea [9]. However, our specimens lacked the dorsal incision observed in *Patagifer* species and instead had a shallow depression. This suggests that our specimens may belong to the genus *Nephrostomum*.

Among the known species of the genus *Nephrostomum*, our specimens matched well with the descriptions of *N. ramosum* (Sonsino, 1895) Dietz, 1909 by Skrjabin and Bashkirova [13]. Although the morphological characteristics of our specimens varied, we speculate that this reflects morphological variation within the species.

Regarding other possible species in the genus *Nephrostomum*, *N. australe* (Johnston, 1928) Mendheim, 1943 appears to be a valid species since it differs significantly from *N. ramosum* in regard to the number of collar spines [12,21]. However, there is doubt concerning the validity of other reported *Nephrostomum* species. Odhner [22] considered *Nephrostomum garzettae* (MacCallum, 1904) to be a synonym of *N. ramosum*. Moreover, Tubangui [23] believed that *Nephrostomum bicolanum* Tubangui, 1933 differed from *N. ramosum* because it lacked dorsal recesses, while Prudhoe [24] regarded it as a synonym of *N. ramosum*. Although Skrjabin and Bashkirova [7] did not agree with Prudhoe [24], and this opinion was supported by Fischthal and Kuntz [13], our observations support the position of Prudhoe [24]. In general, we found that when the specimens were fixed, the collar lifted up and the dorsal recess overlapped with the head and oral sucker according to our light microscopic observations. This made it difficult to observe the dorsal recess (Fig. 2B). However, in 1 slide specimen that had a well-positioned collar, the dorsal recess was clearly observed (Fig. 2C). Both types of specimens were also observed by Ukoli [25], who also suggested that this variation may reflect the state of the muscles at fixation.

Nephrostomum limai Travassos, 1922, which was collected from *Syrigma sibilatrix* (Temminck, 1824) in South America [26], may also be a synonym of *N. ramosum*. In their key to the species of the genus *Nephrostomum*, Skrjabin and Bashkirova [13] noted that *N. limai* had 38 spines, which is less than the number of spines they reported for *N. ramosum* (47-51). However, this was based on the erroneous key suggested by Mendheim [12]. This error caused confusion among later research-

ers, who continued to cite this erroneous number in their own reports [27,28]. Indeed, Travassos [26] originally noted that *N. limai* had 48 collar spines, which is consistent with the morphological variation of *N. ramosum* reported by Skrjabin and Bashkirova [13]. In addition, the description of *N. limai* and the drawing of the cirrus sac provided by Travassos [26] seem to resemble the morphological attributes of our specimens. Although the origin of *N. ramosum* in the New World is not clear, its history may reflect expansion of the type host, namely, the cattle egret. The cattle egret is regarded as an invasive species in the New World. Since it was already observed in the New World in the late 1800s [29], *N. ramosum* may have been introduced into the New World from Africa by its host at that time. It may then have spread to native hosts in the New World. *N. ramosum* has been found in invasive cattle egrets in both North and South America [30,31].

N. legonum Ukoli, 1967, which was found in *Bubulcus ibis* in Ghana, may also be spreading globally through a similar mechanism. However, this species seems to exhibit only minor differences from *N. ramosum* [13,25], and may in fact be a synonym of *N. ramosum*. This notion is supported by the fact that *N. ramosum* was found in *B. ibis* in the nearby country of Mali, as reported by Fischthal [32]. Ukoli [25] also noted that the tail or a well-marked posterior process also differs from that of other *Nephrostomum* flukes. The feature was denied by Prudhoe and Hussey [33], but it was also observed in some of our specimens (Fig. 4C, D).

It is likely that *Nephrostomum robustum* Viguera, 1944 is a valid species in the New World [34]. Viguera [34] found this species in the Least Grebe *Tachybatus dominicus* (= *Colymbus dominicus*) in Havana, Cuba. Although he did not assess the taxonomy of the species, his drawings suggest that it is not a synonym of *N. ramosum*. The species had all of the key characteristics of the genus *Nephrostomum* and a collar spine number (50) similar to *N. ramosum*; however, it has a posteriorly tapered body whose maximum width is at the ventral sucker region [34]. In contrast, *N. ramosum* has a ribbon-shaped body whose maximum width is located between the uterus and testes regions. In addition, the drawing of *N. robustum* indicates it has a shorter uterine field (7%) than *N. ramosum* (18-35% in the present study). Therefore, we regard *N. robustum* as a valid species.

Another *Nephrostomum* species reported from the New World, *N. sinchirocai* Ibáñez, 1966 is recorded from a snowy egret, *Egretta thula* (= *Leucophoyx thula*) in Peru [35]. Ibáñez [35] considered his specimen a valid species based on the smaller

number of collar spines (n=26) in comparison to other species of the genus *Nephrostomum*. However, the description seems to be based on a specimen of poor condition, and his drawing shows that the angle spines and some of the lateral spines are absent [35]. Loss of spines in echinostome specimens is a common phenomenon when treating specimens in poor condition (personal communication). As a consequence, we believe that the species could be a synonym of *N. ramosum* if we disregard the possibility that it is a genuine morphology described with fresh specimens. *N. skrjabini* is characterized by the presence of tegumental spines, but is also regarded as a synonym of *N. ramosum* [36]. Although we did not observe tegumental spines in our specimens, the fact that Karyakarte [27] observed minute spines on the body surface of juvenile and young adult *N. ramosum* specimens supports the notion that *N. skrjabini* is a young adult of *N. ramosum*. Ukoli [25] also found tegumental spines on the metacercariae of *N. ramosum*. It suggested the species resembles *N. ramosum* and *N. bicolanum*, but differs in the shape of the testes and ovary [36], and therefore *N. skrjabini* could also be a synonym of *N. ramosum*. The shape of the organs also varied in the present study.

The taxonomic status of *Nephrostomum* species in the Indo-Pakistan region may be more problematic than of species in other regions. To our knowledge, 7 species of *Nephrostomum* flukes have been reported to date from the Indo-Pakistan region. According to Kalikabad and Bilqees [37], *N. dubashi* is only distinguishable from other species by the number of collar spines (27-34). However, *N. reticulatum* Karyakarte, 1969 and *N. chandigarensis* Gupta and Mehrotra, 1970 are already regarded as synonyms of *N. ramosum* [38-40]. The taxonomic status of the other 4 species (*N. dollfusi*, *N. guptai*, *N. udaipurensis*, and *N. oderolalensis*) does not seem to have been discussed previously. We believe that they could be synonyms of *N. ramosum* because they differ only mildly from this species. The authors describing these 4 *Nephrostomum* species all suggested that the distinctive characteristics of each species were the number of collar spines, the shape of the testes and ovary, and the distribution of vitelline follicles. However, these features of the 4 *Nephrostomum* species all fall within the range of variation observed in our specimens, all of which were recovered at the same time from a single host. The number of spines in our specimens ranged from 47 to 50, the ovary and testes had irregular shapes (Fig. 3), and the distribution of vitelline follicles varied from posterior to mid-level or slightly behind of the ventral sucker. Therefore, unless our case in-

volved multiple infections with at least 3 *Nephrostomum* species, it seems quite possible that the remaining 4 *Nephrostomum* species reported from the Indo-Pakistan region are also synonyms of *N. ramosum*. This notion is supported by the fact that their type hosts were *Bubulcus ibis*, which was also the host of our specimens. Moreover, the type locality of each species is not very far from that of other species collected from the Indo-Pakistan region.

N. dollfusi was originally found in *Bubulcus ibis* in the Renital and Melonigaj area, Jabalpur, India, and was initially described as 2 different species, namely, *Paryphostomum dollfusi* Agarwal, 1959 and *P. bubulcusi* Agarwal, 1959 [41]. However, Srivastava [42] regarded *P. bubulcusi* as a synonym of *P. dollfusi*, and Kostadinova and Gibson [43] later placed the species in the genus *Nephrostomum* on the basis of its morphological characteristics, namely, 46-47 spines arranged in a single row, expansion of the vitelline follicles, and an unarmed tegument. We agree with Kostadinova and Gibson. However, since the species does not exhibit any specific differences from *N. ramosum*, it seems likely that this species is also a synonym of *N. ramosum*.

Gupta and Singh [38] suggested a new species, *N. guptai* Gupta and Singh, 1986, on the basis of its slightly lobed ovary, its highly lobed testes, and the distribution of vitelline follicles. However, the ovary was not clearly lobed in their drawing while the other 2 characteristics were observed in some of our specimens. In addition, the description of *N. guptai* was based on only 1 mature and 5 immature specimens [38]. This sample may be too small to adequately reflect the variation of the species. Prior to this study, one of the authors also suggested another new species, *N. udaipurensis* Gupta, 1983, which was found in *Bubulcus ibis* in Udaipur, Rajasthan, India [38]. However, this new species was not mentioned further in their published reports, although they did correctly list the other previously reported species in the genus *Nephrostomum* [39]. Furthermore, the morphological characteristics of *N. udaipurensis* do not differ markedly from those of *N. guptai* [38,39] or *N. ramosum*. Therefore, we consider that both *N. guptai* and *N. udaipurensis* are synonyms of *N. ramosum*.

The description and drawing of the Pakistan species *N. oderolalensis* Khan and Ghazi suggest that the specimens were in poor condition [28]. In addition, their publication had a number of discrepancies: the number of collar spines varied from 40-48 in the abstract, 40-44 in the description, and 40-44 and 40-46 in the remarks [28]. Moreover, we could not detect any marked specific differences between *N. oderolalensis*

and *N. ramosum*. While Khan and Ghazi [28] noted that the number of spines, shape of the ovary, and distribution of vitelline follicles of *N. oderolalensis* differed from those of other *Nephrostomum* species, these features fall in the range of variation shown by *N. ramosum* and our specimens. We therefore regard *N. oderolalensis* as a synonym of *N. ramosum*.

Although we indicated that *N. dubashi* is distinguishable from other species of *Nephrostomum* by the number of collar spines mentioned above, the true taxonomic status of the species does not seem to differ from that of other synonyms of *N. ramosum*. Kaikabad and Bilqees [37] suggested their specimens differed from other species of *Nephrostomum*, mainly by possession of 27-34 collar spines and a poorly developed cirrus sac. However, spines on the ventral lappets were absent in their drawing [37], and this spine absence is likely to have been due to observations of specimens in poor condition as in the case of *N. sinchirocai* [35]. If the absence of spines on the ventral lappets of both *N. dubashi* and *N. sinchirocai* is a genuine morphological character of the species, then this trait needs to be discussed at the genus level. We believe that the absence of spines on ventral lappets and poorly observed cirrus sac of *N. dubashi* were a result of observations of specimens in poor condition, and therefore regard *N. dubashi* as a synonym of *N. ramosum*. Additionally, *N. dubashi* was recorded from an unknown bird in Sindh, Pakistan [37], which was also the type location of another synonymized species, *N. oderolalensis* [28].

In conclusion, we identified *P. bubulcum* and *N. ramosum* from a cattle egret, *B. ibis coromandus*, on the basis of morphological observations. Moreover, comparison of our specimens with previously reported species of the genus *Nephrostomum* led us to the taxonomic opinion that several species of *Nephrostomum*, namely, *N. bicolanum*, *N. changarensis*, *N. dollfusi*, *N. dubashi*, *N. guptai*, *N. legonum*, *N. limai*, *N. oderolaensis*, *N. reticulatum*, *N. sinchirocai*, *N. skjrabini*, and *N. udaipurensis* are possibly synonyms of *N. ramosum*. Among the species that resemble *N. ramosum* morphologically, especially in regard to the number of collar spines (47-51), we only recognized the validity of *N. robustum*. Another species did not exhibit distinct differences regarding morphometric comparisons and estimated proportions of standard structures relative to the body length (Table 2). However, our conclusions are limited by the fact that they are based only on morphological comparisons. Molecular biological comparisons are needed to confirm these conclusions. This report describes the first case of *P. bubulcum* and *N. ramosum* infection of *B. ibis coromandus* in Ko-

rea. This is the first time both flukes and associated genera have been reported as part of the parasite fauna of Korea.

ACKNOWLEDGMENTS

This work was supported by a grant from the National Institute of Biological Resources (NIBR), which is funded by the Ministry of Environment (MOE) of the Republic of Korea (NIBR no. 2013-02-001 and 2014-02-001). One of the authors (Seongjun Choe) was supported by Graduate Program for the Undiscovered Taxa of Korea (NIBR201524202). The parasite materials used in this study were provided by the Parasite Resource Bank of the National Research Resource Center (2012-0000037) of the Republic of Korea. The authors are also grateful to Philippe Vieira Alves, Programa de Pós-Graduação em Biologia Animal, Universidade Federal Rural do Rio de Janeiro, Brazil, for providing useful literature.

CONFLICT OF INTEREST

We have no conflict of interest related to this work.

REFERENCES

1. Park JG. Identification guide to birds of Korea, No. 12, Checklist of organisms in Korea. Nature and Ecology. 2014, pp 1-125.
2. Kim HK. Biology of gray heron and egrets. J Korean Res Inst Bet Liv 1969; 2: 69-81.
3. Murata K, Noda A, Yanai T, Masegi T, Kamegai S. A fatal *Pegosomum* sp. (Trematoda: Echinostomatidae) infection in a wild cattle egret (*Bubulcus ibis*) from Japan. J Zoo Wild Med 1998; 29: 78-80.
4. Uchida A, Uchida K, Kawakami Y, Nagatomo M, Shen M, Ooi H. A helminthological survey of parasites in the waterfowl of Kanagawa Prefecture, Japan. J Jpn Vet Med Assoc 2005; 58: 127-131.
5. Uchida A, Uchida K, Itagaki H. Check list of helminth parasites of Japanese birds. Jpn J Parasitol 1991; 40: 7-85.
6. Yoshino T, Nakamura S, Endoh D, Onuma M, Osa Y, Teraoka H, Kuwana T, Asakawa M. A helminthological survey of four families of waterfowl (Ardeidae, Rallidae, Scolopaciidae and Phalaropodidae) from Hokkaido, Japan. J Yamashita Inst Ornithol 2009; 41: 34-41.
7. Fischthal JH, Kuntz RE. Some digenetic trematodes of birds from Taiwan. Proc Helminthol Soc Wash 1976; 43: 65-79.
8. Kostadinova A. Family Echinostomatida Looss, 1899. In Jones A, Bray RA, Gibson DI eds, Keys to the Trematoda. Volume 2. Wallingford, UK. CAB International and the Natural History Museum. 2005, pp 9-64.

9. Choe S, Lee D, Park H, Oh M, Jeon HK, Lee Y, Na KJ, Kim Y, Lee H, Eom KS. Three echinostome species from wild birds in the Republic of Korea. *Korean J Parasitol* 2014; 52: 513-520.
10. Jang DH. Study on the intestinal helminths of chicken. *J Korean Vet Assoc* 1958; 2(2): 149-155.
11. Ratz I. Un genre nouveau de Fasciolides. *Ann Mus Hungar* 1903; 1: 413-432.
12. Mendheim H. Beiträge zur Systematik und Biologie der Familie Echinostomatidae. *Archive für Naturgeschichte* 1943; 12: 175-302.
13. Skrjabin KI, Bashkirova EY. Family Echinostomatidae Dietz, 1909. In Skrjabin KI ed, *Trematodes of Animals and Man. Osnovy Trematodologii* 12. Izdatel'stvo Akademii Nauk SSSR, Moscow. 1956, pp 53-930.
14. Aleksandrova OV, Podgornova GP. Taxonomical analysis of *Pegosomeum asperum* and *P. saginatum* (Trematoda, Echinostomatidae). *Parazitologiya* 1978; 12: 413-417.
15. Unar MS, Shaikh AA, Khan MM, Channa MA. *Pegosomeum garzettae* n. sp (Digenea: Echinostomatidae) from gallbladder of little egret *Egretta garzetta* (Aves: Ardeidae) of Jamshoro, Sindh, Pakistan. *Parasitology* 2011; 51: 143-148.
16. Aleksandrova OV. Revision of the species composition of the genus *Pegosomeum* (Trematoda, Echinostomatidae). *Zool Zhurnal* 1978; 57: 827-833.
17. Gvosdev EV. New species of fluke *Pegosomeum ixobrychi* sp. nov. from *Ixobrychus minutus* L. *Helminthologia* 1960; 2: 83-86.
18. Chen HT. Fauna Sinica. Platyhelminthes, Trematoda, Digenea (I). Beijing, China. Science Press. 1985, pp 1-697 (in Chinese).
19. Tubangui MA, Masilungan VA. Trematode parasites of Philippine vertebrates, VII. *Phil J Sci* 1935; 58: 435-446.
20. MacCallum GA. Notes on the genus *Telorchis* and other trematodes. *Zoopathologica* 1918; 1: 81-98.
21. Johnston TH. New trematodes from the Australian water-hen *Gallinula tenebrosa*. *Rec S Australian Mus* 1928; 4: 135-142.
22. Odhner T. Nordostafrikanische Trematoden, grösstenteils vom Weissen Nil. I. Fascioliden. Result of the Swedish Zoological Expedition to Egypt and the White Nile 1901 under the direction of LA Jägerskiöld 23A. K.W. Appelberges Boktryckeri, Uppsala, Sweden. 1910, p 1-169.
23. Tubangui MA. Trematode parasites of Philippine vertebrates, VI. Descriptions of new species and classification. *Phil J Sci* 1933; 52: 167-198.
24. Prudhoe SI. On some trematodes from Ceylon. *Mag Nat Hist* 1944; 11: 1-13.
25. Ukoli FMA. On the anatomy, growth and development in the definitive host of *Nephrostomum legonum* n. sp., (Trematoda: Echinostomatidae). *Nigerian J Sci* 1967; 1: 217-231.
26. Travassos L. Informacoes sobre a fauna helminthological de Matto Grosso. Trematoda II. *Mem Inst Oswaldo Cruz* 1938; 33: 461-468.
27. Karyakarte PP. *Nephrostomum reticulatum* sp. N. (Trematoda: Echinostomatidae) from the egret, *Bubulcus ibis* (Linnaeus) in India. *Riv Parassitol* 1969; 30: 91-84.
28. Khan A, Ghazi RR. *Nephrostomum oderolalensis* n. sp. (Trematoda: Digenea) in cattle egret (*Bubulcus ibis* L.) from Sindh, Pakistan. *Int J Biol Biotech* 2011; 8: 487-490.
29. Wetmore A. An early record of the cattle egret in Colombia. *Auk* 1963; 80: 547.
30. Stuart JJ, Dismukes JE, Dixon CF. Endoparasites of the cattle egret (*Bubulcus ibis*) in Alabama. *J Parasitol* 1972; 58: 518.
31. Fischthal JH, Whittaker FH. Two digenetic trematodes from the cattle egret, *Bubulcus ibis* (L.), from Puerto Rico. *J Parasitol* 1977; 63: 491.
32. Fischthal JH. Some digenetic trematodes of birds from Central and West Africa. *Rev Zool Africaine* 1977; 91: 675-680.
33. Prudhoe S, Hussey CG. Some parasitic worms in freshwater fishes and fish-predators from the Transvaal, South Africa. *Zool Afr* 1977; 12: 113-147.
34. Pérez Viguera I. Trematodes de la super-familia Echinostomatoidea, con descripcion de siete especies nuevas de Cuba. *Rev Univ Habana* 1944; 55-57: 221-234.
35. Ibáñez HN. Contribucion al conocimiento de la fauna helmintologica peruana. "*Nephrostomum sinchirocai*" sp. n. (Trematoda, Echinostomatidae). *Rev Brasil Biol* 1966; 26: 93-96.
36. Kasimov GB, Vaidova SM, Felizullaev NA. A new trematode from *Bubulcus ibis*. Papers on Helminthology presented to Academician KI Skrjabin on his 80th birthday 1958; 145-147.
37. Kaikabad SH, Bilqees FM. *Nephrostomum dubashi* n. sp. from a bird of W. Pakistan. In Bilqees FM, Saeed R, Rehana R, Khatoon A, Kaikabad SH eds, *Helminth parasites of some vertebrates chiefly from fishes of West Pakistan*. Agric Res Council Govt of Pakistan, Karachi. 1972, pp 97-99.
38. Gupta PC. *Nephrostomum udaipurensis* n. sp. (family - Echinostomatidae Poche, 1926) from *Bubulcus ibis* (Linn.). *Indian J Parasitol* 1983; 7: 43-45.
39. Gupta PC, Singh RB. Digenetic trematode parasites of fishes and birds. I. On a new trematode *Nephrostomum guptai* n. sp. (family - Echinostomatidae Poche, 1926) from an avian host, *Bubulcus ibis* (Linn.) from Unnao (U.P.). *Indian J Helminthol* 1986; 37: 121-126.
40. Sahay S, Sahay U, Verma DK. On *Nephrostomum ramosum* (Sonsino, 1859) Dietz, 1909 from a new host at Ranchi (Chotanagpur). *Indian J Helminthol* 1987; 39: 12-127.
41. Agarwal SM. Studies on two new species of the genus *Paryphostomum* (Dietz, 1909), (Trematoda: Echinostomatidae) from *Bubulcus ibis*. *Indian J Helminthol* 1959; 10: 19-30.
42. Srivastava CB. The fauna of India and adjacent countries. Platyhelminthes. Vol. I. (Suppl.) Trematoda - Digenea. Zoological Survey of India, Calcutta, 1982, pp 1-163.
43. Kostadinova A, Gibson DI. Redescriptions of two echinostomes from birds in Paraguay, with comments on *Drepanocephalus* Dietz, 1909 and *Paryphostomum* Dietz, 1909 (Digenea: Echinostomatidae). *Sys Parasitol* 2002; 53: 147-158.