

Research Article

Cloning, Expression, and Characterization of a Novel Thermophilic Monofunctional Catalase from *Geobacillus* sp. CHB1

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Catalases are widely used in many scientific areas. A catalase gene (*Kat*) from *Geobacillus* sp. CHB1 encoding a monofunctional catalase was cloned and recombinantly expressed in *Escherichia coli* (*E. coli*), which was the first time to clone and express this type of catalase of genus *Geobacillus* strains as far as we know. This *Kat* gene was 1,467 bp in length and encoded a catalase with 488 amino acid residuals, which is only 81% similar to the previously studied *Bacillus* sp. catalase in terms of amino acid sequence. Recombinant catalase was highly soluble in *E. coli* and made up 30% of the total *E. coli* protein. Fermentation broth of the recombinant *E. coli* showed a high catalase activity level up to 35,831 U/mL which was only lower than recombinant *Bacillus* sp. WSHDZ-01 among the reported catalase production strains. The purified recombinant catalase had a specific activity of 40,526 U/mg and K_m of 51.1 mM. The optimal reaction temperature of this recombinant enzyme was 60°C to 70°C, and it exhibited high activity over a wide range of reaction temperatures, ranging from 10°C to 90°C. The enzyme retained 94.7% of its residual activity after incubation at 60°C for 1 hour. High yield and excellent thermophilic properties are valuable features for this catalase in industrial applications.

1. Introduction

Catalases (EC1.11.1.6) are a class of enzymes that specifically catalyze the decomposition of H_2O_2 to H_2O and O_2 [1]. H_2O_2 has been applied to sterilization and bleaching processes in the medical, food, textile, and paper-making industries [2, 3]. Residual H_2O_2 in products or by-products is harmful to human health and environment. Thus, the removal of residual H_2O_2 is necessary in textile production processes, food health, pollution prevention, and other fields [2]. Catalase is an ideal choice for removing H_2O_2 due to its efficiency and lack of secondary pollution compared with chemical decomposition methods [4].

Catalase can be divided into four classes: monofunctional heme catalases, catalase-peroxidases, non-heme catalases, and minor catalases [5]. Catalases are widely present in animals, plants, fungi, most aerobic bacteria, and some anaerobic bacteria [5]. Commercial catalase produced by animals liver, plant tissues, and microbial fermentation is mainly mesophilic catalase, but many industrial applications need

thermostable catalase; for example, textile bleaching temperature is always up to 60°C [6], so thermophilic catalase is more comparable on that condition. There have been some thermophilic catalases reported [7–10], but they were either catalase-peroxidase or Mn-dependent catalase. Thermophilic monofunctional heme catalases are rarely reported as we know.

High yielding strains are also necessary for enzyme production in fermentation processes. Recombinant expression is a practical method to increase the yield of a target gene. Currently, *Pichia pastoris* [11], *Hansenula polymorpha* [12], *Lactococcus lactis* [13], *Bacillus subtilis* (*B. subtilis*) [14, 15], and *Escherichia coli* (*E. coli*) [9] are used as host cells to produce recombinant catalase, but those recombinant catalases were mainly mesophilic, successfully recombinant expression precedents of thermophilic catalases which were few. The advantages of *E. coli* systems, such as their convenience, high yields, and ease of purification promote their wide application in genetic engineering. But many thermophilic enzymes cannot be overproduced in active forms in mesophilic host [8].

For example, though *Thermus thermophilus* catalase could solubly be expressed in *E. coli*, the recombinant catalase was absolutely inactive [8]; recombinant *E. coli* with *Bacillus stearothermophilus* catalase-peroxidase gene also showed a comparatively lower catalase level (1055.3 U/mL) [16]. So high soluble expression with an active form in *E. coli* is important for thermophilic catalase.

Previously, we screened thermophilic bacteria strains isolated from different high-temperature compost samples and *Geobacillus* sp. CHB1 was found to be a high catalase production strain. So, in this work, we cloned a novel *Kat* gene encoding a monofunctional catalase from thermophiles *Geobacillus* sp. CHB1 and recombinant expressed this gene in *E. coli* in a highly soluble form. The recombinant catalase was also purified and characterized.

2. Materials and Methods

2.1. Materials, Bacterial Strains, Plasmids, and Medium. *E. coli* BL21 (DE3) was cultured in our laboratory, and the expression vector pEASY-E2 was purchased from Beijing TransGen Biotech. The *Geobacillus* sp. CHB1 was isolated in Fuzhou, China. LB medium (10 g/L tryptone, 5 g/L yeast extract, 10 g/L NaCl, pH 7.0) was used in culturing *E. coli* BL21 (DE3). Auto-induction medium ZYM-5052 [17] was used for inducing the expression of recombinant catalase. The medium for culturing the *Geobacillus* sp. CHB1 consisted of soya peptone 0.9%, yeast extract 0.5%, NaCl 0.1%, K₂HPO₄ 0.1%, and KH₂PO₄ 0.075%, at pH 7.2.

2.2. Expression Vector Construction. *Geobacillus* sp. CHB1 was incubated at 60°C for 18 h at 180 r/min; then the genome was extracted according to the method of Zhou et al. [18]. The *Kat* gene was amplified using primers CatalaseF: (5'-GCA-GATACAAAAAGCTCACAAAC-3') and CatalaseR: (5'-TGCGTTTGTAAATCACATCGTCCG-3'). Polymerase chain reaction (PCR) was performed with ExTaq DNA polymerase (TaKaRa, Dalian, China) under the following conditions: 95°C initial denaturation for 5 min, followed by 32 cycles of 40 s at 94°C, 40 s at 55°C, and 1 min 30 s at 72°C. The PCR product was purified using a PCR purification kit (Omega Bio-Tek, Inc., USA) and sequenced by Sangon Biotech Co., Ltd. (Shanghai, China).

Homology search of gene and amino acid sequence was carried out at BLAST server (<http://blast.ncbi.nlm.nih.gov>). Program blastp was used to analyze homology of the amino acid sequences; nonredundant protein sequences (Nr) database and Swiss Prot database were both used for blastp program. Sequences with high similarity in Swiss Prot database were selected to construct phylogenetic tree and do multiple alignment. MEGA 4.0.2 was used to construct the phylogenetic tree. Multiple alignment of the amino acid sequences was carried out using DNAMAN V6.0.3.99.

The purified fragment was ligated with pEASY-E2 using the described protocol of the kit. The ligated product was transformed into the competent *E. coli* cell strain Trans-T1 (TransGen Biotech, Beijing, China). A positive clone was

selected using the T7 promoter primer (5'-TAATAC-GACTCACTATAGGG-3') and CatalaseR (5'-TGCGTT-TGTAATCACATCGTCCG-3') and then cultured with LB medium containing 100 µg/mL of ampicillin. The recombinant vector pEASY-E2-*Kat* was extracted using a Plasmid Mini Kit (Omega Bio-Tek, Inc., USA) and then transformed into competent *E. coli* BL21 (DE3) cells. A positive clone for BL21/pEASY-E2-*Kat* was selected using PCR as described above.

2.3. Inducing Expression and Purification of Recombinant Catalase. Positive clones were cultured in 5 mL of LB medium containing 100 µg/mL of ampicillin at 37°C at 220 rpm for 12 h. Next, 1 mL of the above culture was inoculated into 50 mL of ZYM-5052 medium and was cultured at 30°C and 220 rpm for 16 h to induce the expression of the enzyme. To confirm expression, 8 mL of the induced culture was centrifuged at 10,000 ×g for 2 min and then suspended with 3 mL PBS (50 mM, pH 7.0). The suspended culture was subjected to ultrasonication at 400 W until clear. Then, 200 µL of ultrasonicated sample was centrifuged at 12,000 ×g for 5 min at 4°C, and the supernatant was transferred to a new centrifuge tube. The supernatant and sediment sample were both subjected to 12% SDS-PAGE to detect the expression of the recombinant enzyme; a sample containing an empty vector was used as a blank control. Electrophoresis parameters were set as follows: 30 V for 30 min, followed by 80 V until the end. The coomassie brilliant blue R250 protocol [23] was used to dye the gel.

2.4. Recombinant Strain Enzyme Production Curve. First, 1 mL of seed liquid was inoculated into shake flasks containing 50 mL of ZYM-5052 medium, and the flasks were placed in shaking tables at 220 rpm and 30°C. Every 4 h, samples were collected and analyzed for catalytic activity. The samples were ultrasonicated before activity measurement collection. Enzyme production curves were then obtained.

2.5. Purification of Recombinant Catalase. First, 100 mL of the induced recombinant *E. coli* was centrifuged at 10,000 ×g for 5 min and washed once with Buffer A (50 mM NaH₂PO₄, 300 mM NaCl, pH 8.0). After centrifugation, the recombinant *E. coli* pellet was resuspended with 3 mL of Buffer A. The resuspended sample was ultrasonicated until clear. The ultrasonicated sample was centrifuged, and the supernatant was subjected to a Nickel-iminodiacetic acid (Ni-IDA) column. Then, 10 volumes of Buffer A (50 mM NaH₂PO₄, 300 mM NaCl, pH 8.0) were used to wash proteins that were non-specifically bound to the column. Then, 10 volumes of Buffer B (50 mM NaH₂PO₄, 300 mM NaCl, 20 mM imidazole, pH 8.0) were used to remove other proteins. Finally, 5 volumes of Buffer C (50 mM NaH₂PO₄, 300 mM NaCl, 100 mM imidazole, pH 8.0) and 5 volumes of Buffer D (50 mM NaH₂PO₄, 300 mM NaCl, 500 mM imidazole, pH 8.0) were applied to elute the recombinant catalase. The purity of catalase was confirmed by SDS-PAGE. The purified catalase was placed into a dialysis bag and dialyzed against Buffer A to remove the high concentrations of imidazole. Buffer A was replaced every several hours until the imidazole was removed. The quantity

of purified catalase was assayed using the Bradford method [24].

2.6. Recombinant Catalase Characteristics. A catalase activity assay was performed according to the method described by Beers Jr. and Sizer [25]. Purified recombinant catalase was diluted several times with PBS (20 mM, pH 7.0) to a suitable concentration, and 2.7 mL of 30 mM H_2O_2 (diluted with 20 mM PBS, pH 7.0) was then immediately added to 300 μ L of diluted enzyme. The absorbance changes at 240 nm were measured every 5 s for 1 min. 1 U of activity was defined as the amount of enzyme to decompose 1 μ mol of H_2O_2 per min. Each measurement was performed at least three times.

To assess the optimal reaction temperature, 30 mM H_2O_2 was preincubated at different temperatures from 10°C to 90°C (in 10°C increment), and then activities at different temperatures were measured with above preincubated H_2O_2 .

To assess enzyme thermostability, diluted enzyme was incubated at 40, 50, 60, 70, 80, and 90°C for 30 min and 1 hour. After the incubation time, the heated enzyme was immediately transferred onto ice. Residual activities were measured with method described above.

The optimum pH was assayed with 30 mM H_2O_2 prepared in the following 50 mM buffers: sodium citrate 50 mM (pH 4–6), sodium phosphate 50 mM (pH 6–8), and glycine-NaOH 50 mM (pH 9–11). Activities at different pH values were measured as above in least three replicates.

Kinetic parameters were assayed by calculating the activity of the enzyme under different concentrations of H_2O_2 ranging from 2.5 to 25 mM in 20 mM PBS (pH 7.0) at 30°C. K_m was obtained by double-reciprocal plots according to Lineweaver and Burk [26].

3. Results and Discussion

3.1. Kat Sequence. The PCR product (Figure 1) and sequencing results showed that the total length of the *Kat* gene was 1,467 bp, which encoded a protein with 488 amino acids. Nucleotide sequence was submitted to GenBank, and GenBank accession number KP202252 was assigned. blastp was run in the nonredundant protein sequences of NCBI database for homologs of the amino acid sequence. The amino sequence had higher similarity with the catalase sequences of other *Geobacillus* spp.; however, all of these highly similar *Kat* genes were submitted in the form of sequenced genomes, and none had been previously cloned or expressed. Among all studied catalase sequences obtained from Swiss Prot database, this catalase was most similar to that of a *Bacillus* sp. [27], and its identity was 80%. Phylogenetic tree (Figure 2) based on the amino acid sequences similarity showed this catalase shared closest phylogenetic relationship with catalase of *Bacillus subtilis* and *Lactobacillus sakei* in Swiss Prot database. Multiple alignment of the amino acid sequence showed this catalase has seven typically conserved amino acid residuals for heme binding (Figure 3). All of the reported thermostable catalases [28, 29] from *Geobacillus stearothermophilus* were different from this catalase herein because they belonged to the second group of catalase family (catalase-peroxidase) and were encoded by *Per* gene, but catalase in

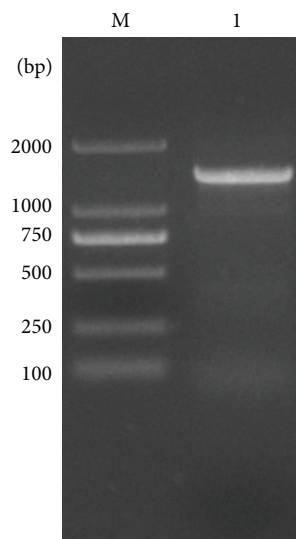


FIGURE 1: PCR product showing a 1,452-length band that corresponds to *Kat* from a *Geobacillus* sp. CHB1. Lanes: M, DNA marker; Lane 1, PCR product.

this paper belonged to the first group (monofunctional heme catalase) which was encoded by gene *Kat* [5]. Therefore, to our knowledge this study was the first to clone and express this type of *Geobacillus* spp. *Kat* gene.

3.2. Construction of Recombinant Expression Vector pEASY-E2-Kat. A positive clone was selected, and its sequence was confirmed by sequencing. The expression vector pEASY-E2-Kat (Figure 4) was transformed into BL21 (DE3) cells. Then, SDS-PAGE was applied to confirm whether the catalase was expressed and to determine the molecular weight of the enzyme. A band was observed between 60 and 70 kDa, and the recombinant catalase was highly soluble (make up to 30% of the total *E coli* protein) with a low inclusion body content (Figure 5). The theoretical molecular weight of the recombinant enzyme herein, calculated using its amino sequence, was 56 kDa, including the 6 \times His tag, which conflicted with the molecular weight of 65 kDa observed via SDS-PAGE. Some studies [30] have found that the 6 \times His tag might result in a recombinant protein with a greater molecular weight; however, the reason for this phenomenon remains unclear. The specific activity of the purified catalase was 40,526 U/mg under optimal conditions (60°C, pH 7.0).

3.3. Production Curve for Recombinant Catalase. Production curves for the recombinant catalase were constructed at 30°C. When cultured at 30°C for 20 h, the total activity of the fermentation broth reached a maximum level of 35,831 U/mL assayed under optimal conditions (Figure 6). This level was much higher than many catalase production strains and only lower than recombinant *Bacillus* sp. WSHDZ-01 (Table 1) as we know. High production ensured further application of this catalase.

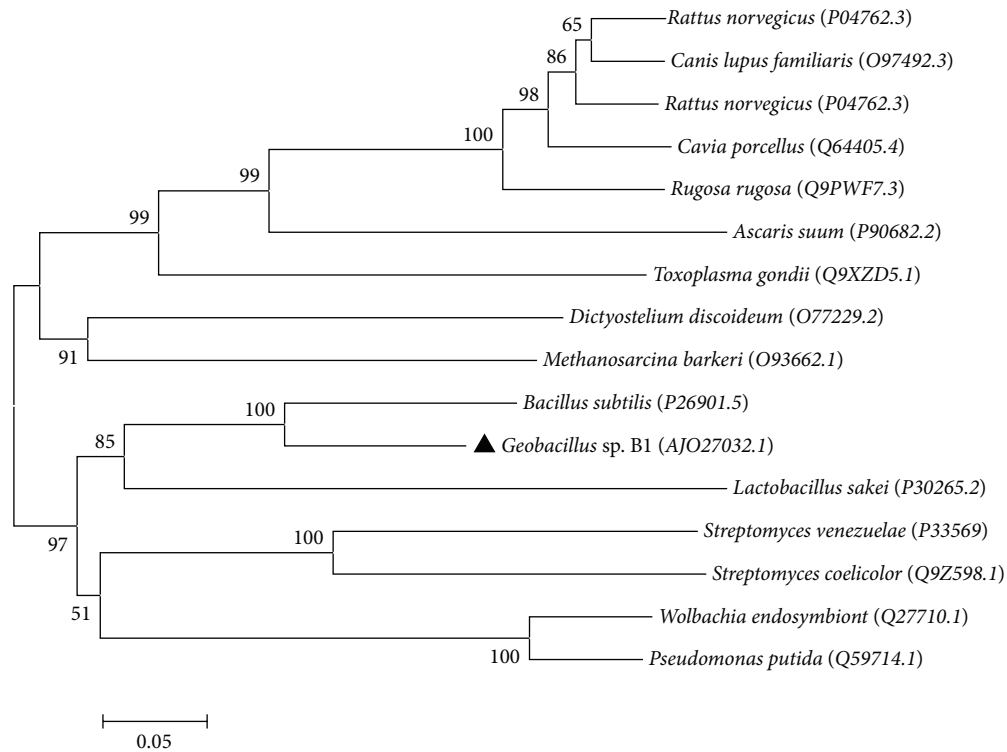


FIGURE 2: Phylogenetic tree of CHB1 catalase amino acid showing the relationship with other strains on catalase amino acid sequence. Protein sequences were selected by running blastp program with CHB1 catalase sequence in Swiss Prot database; accession numbers in brackets were the corresponding accession numbers of the strains in Swiss Prot database. Phylogenetic tree was constructed with MEGA 4.0.2 with the method Neighbor-Joining. Test of inferred phylogeny was Bootstrap for 1000 replications.

TABLE 1: Catalase production levels of recombinant *E. coli* BL21 and other strains.

Strains	Activity (U/mL)	Reference
Recombinant <i>E. coli</i> BL21 (DE3)	35,831	This study
Recombinant <i>Bacillus</i> sp. WSHDZ-01	39,117	[15]
<i>Bacillus</i> sp. WSHDZ-01	28,990	[19]
<i>Exiguobacterium oxidotolerans</i> T-2-2T	22,000	[20]
<i>Serratia marcescens</i> SYBC08	20,353	[21]
<i>Rhizobium radiobacter</i> strain 2-1	17,035	[22]
<i>M. luteus</i>	6920	[22]

3.4. Purified Recombinant Catalase Characteristics

3.4.1. Effect of Temperature and pH on This Catalase. Temperature is a major factor in the application of many enzymes. The activity and stability of our purified catalase at different temperatures were assayed. The catalase in this study showed a high activity over a wide range of temperatures, from 10°C to 90°C (Figure 7(a)), and showed maximum activity at 60°C to 70°C. The temperature range was wider and optimal temperature was higher than many reported catalases, such as *Serratia marcescens* SYBC08 at temperatures ranging from 0°C to

70°C with optimal temperature of 20°C [31]; *Psychrobacter piscatorii* T-3 from 10°C to 60°C with optimal temperature of 45°C [32]; and *Bacillus altitudinis* SYBC hb4 from 20°C to 40°C with optimal temperature of 30°C [33]. This catalase was stable at temperatures $\leq 60^\circ\text{C}$ (Figure 7(b)) and when incubated at 70°C for 1h, the enzyme maintained >70% residual activity; however, when the incubation temperature was >80°C, the residual activity was <10% after incubation for 1h. Although this enzyme was inactive at $\geq 80^\circ\text{C}$, it still maintained high activity when added to reaction systems at 80°C and 90°C. Thermophilic catalase isolated from thermophilic *Thermoascus aurantiacus* [34] can retain 100% residual activity after incubation at 60°C for 1h, which was similar to that of CHB1 catalase. This thermophilic property was much better than the properties of other catalases, such as those from *Psychrobacter piscatorii* (*P. piscatorii*) (65°C, 15 min, 20%) [32, 35], *Vibrio salmonicida* (60°C, 20 min, 0%) [36], *Vibrio rumoiensis* S-1T (65°C, 10 min, 0%) [37], and *Halomonas* sp. SK1 (55°C, 30 min, 0%) [38]. These properties enable the enzyme to be applied in both low and high-temperature processes in industry. The optimal pH of the enzyme was 6-7, and the enzyme retained >50% of its activity between pH 5 and 9 (Figure 8).

3.4.2. Kinetic Parameters of This Catalase. The enzyme kinetics of the recombinant catalase were assayed using

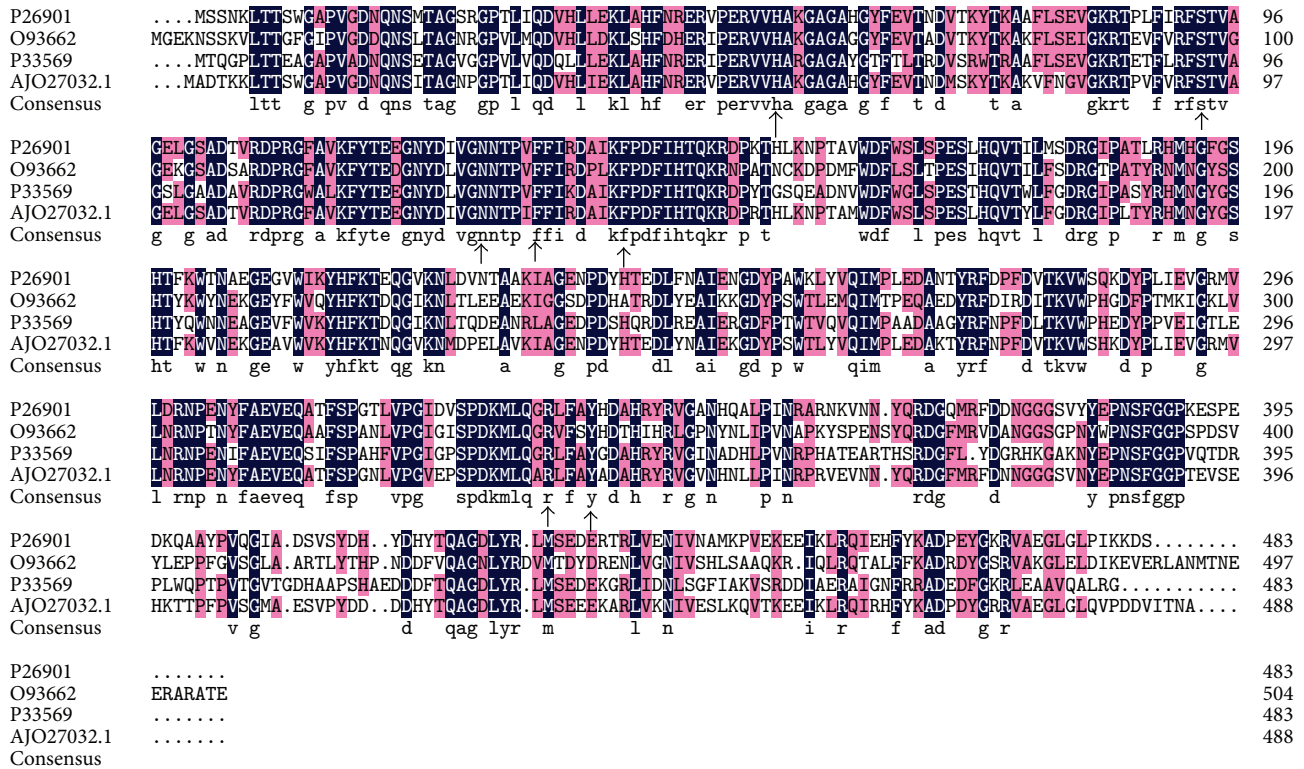


FIGURE 3: Multiple alignment of amino acid sequences for CHB1 catalase and catalase from Swiss Prot database. The sequences were those from *Bacillus subtilis* subsp. str. 168 (Swiss Prot number P26901), *Methanosarcina barkeri* str. Fusaro (Swiss Prot number O93662), *Streptomyces venezuelae* ATCC 10712 (Swiss Prot number P33569), and *Geobacillus* sp. CHB1 (GenBank number AJO27032.1). Identical amino acid residuals were shaded in black and conserved residuals were shaded in gray. Arrows showed the conserved residuals of home binding pocket of this kind of catalase.

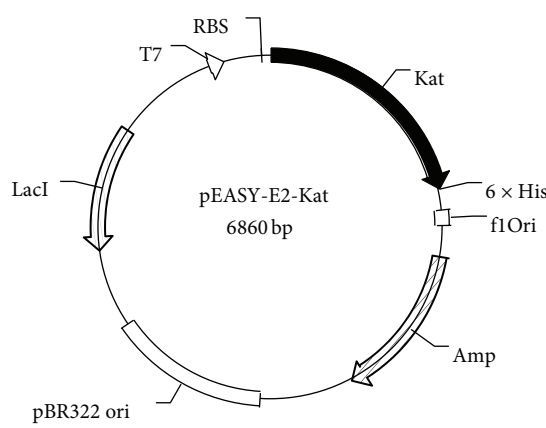


FIGURE 4: Recombinant expression construction of vector pEASY-E2-kat. A schematic diagram of recombinant expression vector pEASY-E2-kat. The kat gene encoding catalase was inserted into an expression vector with upstream RBS and T7 promoters and downstream 6xHis.

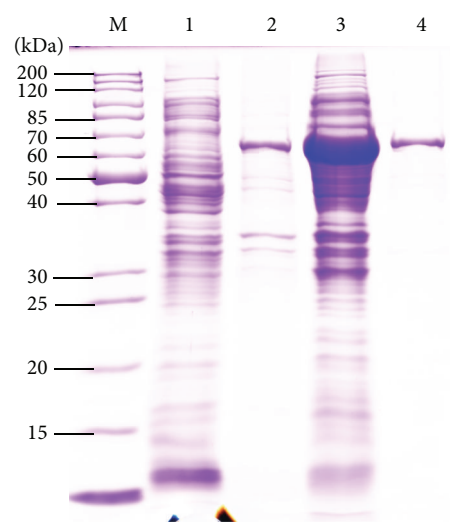


FIGURE 5: SDS-PAGE analyses of recombinant catalase expression and the purified enzyme. Lanes: M, protein marker; Lane 1, total bacterial protein of BL21 (DE3) cells with empty vector pEASY-E2; Lane 2, insoluble bacterial proteins of BL21 (DE3) with vector pEASY-E2-kat after induced expression; Lane 3, soluble bacterial proteins of BL21 (DE3) with vector pEASY-E2-kat after induced expression; Lane 4, SDS-PAGE analysis of purified catalase using a Ni-IDA column.

different H₂O₂ concentrations as substrates; a Lineweaver-Burk plot was used to calculate K_m , 51.1 mM, and V_{max} , 151.5 mol/min-mg (Figure 9), which is similar to previously

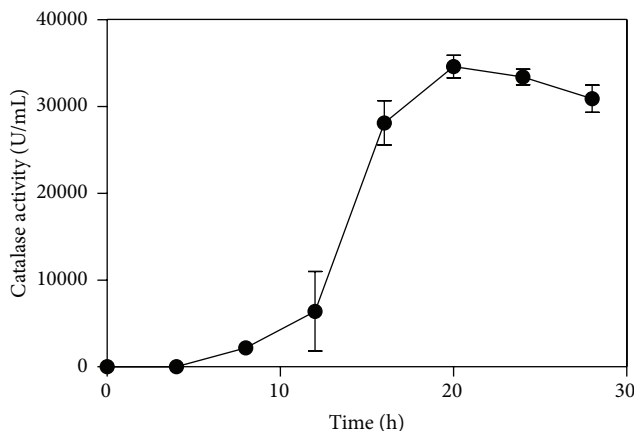


FIGURE 6: Catalase production curve of the recombinant *E. coli* BL21 (DE3).

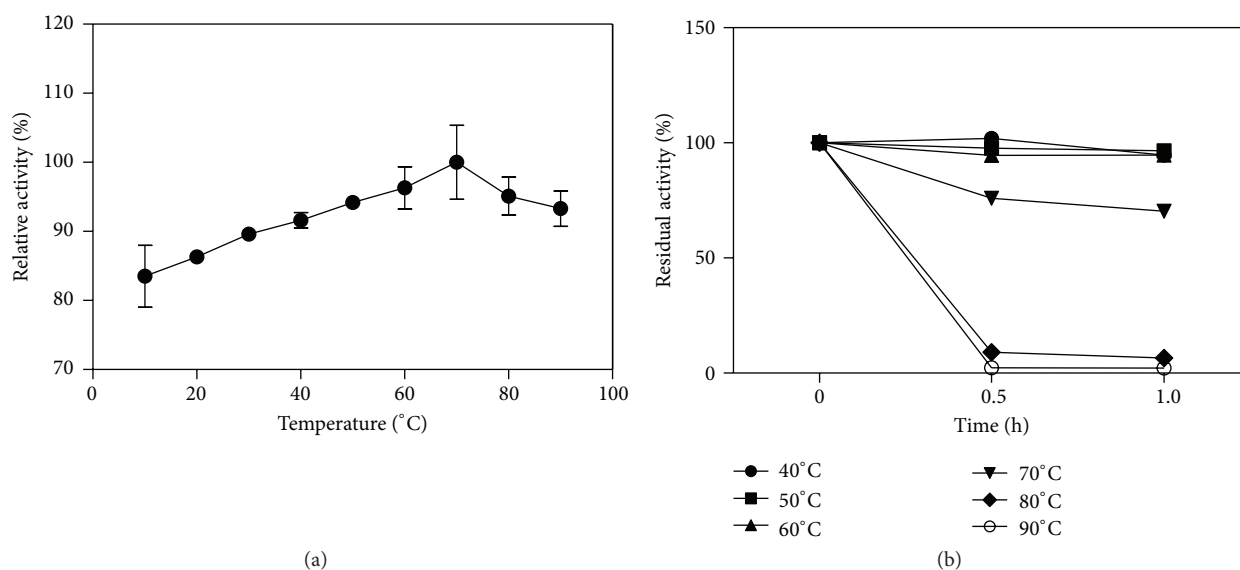


FIGURE 7: Effect of temperature on recombinant catalase activity. (a) Relative activity of the enzyme at 10, 20, 30, 40, 50, 60, 70, 80, and 90°C; (b) temperature stability of the enzyme. Residual activity of the recombinant enzyme by incubation at 40, 50, 60, 70, 80, and 90°C for 0.5 and 1 h.

reported values (52.5 mM [23] and 40.1 mM [2]). Many other catalases from *P. piscatorii* T-3, *Micrococcus luteus*, *Bacteroides fragilis*, *Helicobacter pylori*, *Serratia marcescens*, and *Xanthomonas campestris* have higher K_m values compared to the purified catalase [39]. The relatively low K_m indicates that the enzyme has a high affinity for H_2O_2 and that the enzyme is stable under high H_2O_2 concentrations. K_m is an important characteristic of enzymes and is vital for the assessment of their potential applications.

4. Conclusions

In this paper, we cloned, expressed, purified, and characterized a *Kat* gene of thermophilic bacteria *Geobacillus* sp. CHB1

for the first time. The recombinant enzyme could maintain its stability and showed a high activity over a wide range of temperatures from 10°C to 90°C. Specific activity of the purified recombinant catalase was 40,526 U/mg of protein, and recombinant *E. coli* BL21 strain reached a high level of catalase production to 35,831 U/mL. This enzyme's wide range of reaction temperatures, good thermostability, and high production may be suitable for the textile, paper-making, and other industries.

Competing Interests

The authors declare no conflict of interests.

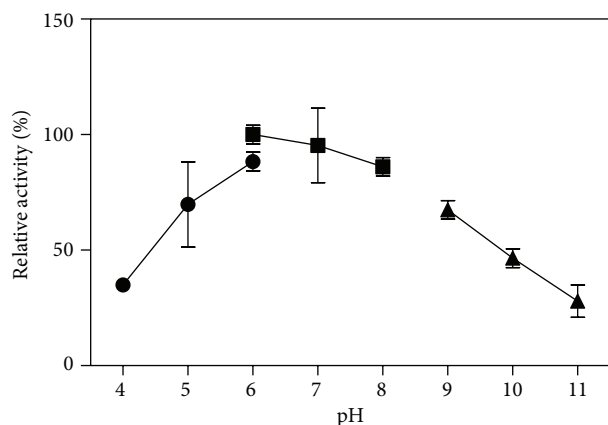


FIGURE 8: Optimal pH of the recombinant catalase. The buffers used for each pH region were 50 mM sodium citrate (pH 4–6), 50 mM sodium phosphate (pH 6–8), and 50 mM glycine-NaOH (pH 9–11). Each pH was assayed at least three times.

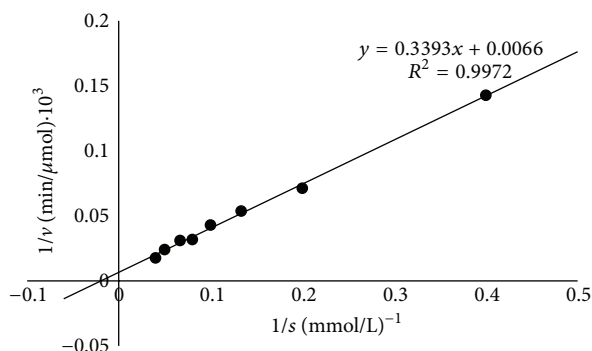


FIGURE 9: A Lineweaver-Burk plot of the recombinant catalase. Enzyme activity was assayed in 50 mM phosphate buffer (pH 7.0) and at 30°C.

Acknowledgments

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References

- [1] H.-S. Ko, H. Fujiwara, Y. Yokoyama et al., "Inducible production of alcohol oxidase and catalase in a pectin medium by *Thermoascus aurantiacus* IFO 31693," *Journal of Bioscience and Bioengineering*, vol. 99, no. 3, pp. 290–292, 2005.
- [2] H.-W. Zeng, Y.-J. Cai, X.-R. Liao, S.-L. Qian, F. Zhang, and D.-B. Zhang, "Optimization of catalase production and purification and characterization of a novel cold-adapted Cat-2 from mesophilic bacterium *Serratia marcescens* SYBC-01," *Annals of Microbiology*, vol. 60, no. 4, pp. 701–708, 2010.
- [3] H. D. Rowe, "Biotechnology in the textile/clothing industry—a review," *Journal of Consumer Studies & Home Economics*, vol. 23, no. 1, pp. 53–61, 1999.
- [4] X. Fu, W. Wang, J. Hao, X. Zhu, and M. Sun, "Purification and characterization of catalase from marine bacterium *Acinetobacter* sp. YS0810," *BioMed Research International*, vol. 2014, Article ID 409626, 7 pages, 2014.
- [5] B. S. Soodh, B. S. Kauldhar, and M. Puri, "Recent insights into microbial catalases: isolation, production and purification," *Biotechnology Advances*, vol. 32, no. 8, pp. 1429–1447, 2014.
- [6] M. Spiro and W. P. Griffith, "The mechanism of hydrogen peroxide bleaching," *Textile Chemist and Colorist*, vol. 29, no. 11, pp. 12–13, 1997.
- [7] M. Kagawa, N. Murakoshi, Y. Nishikawa et al., "Purification and cloning of a thermostable manganese catalase from a thermophilic bacterium," *Archives of Biochemistry and Biophysics*, vol. 362, no. 2, pp. 346–355, 1999.
- [8] A. Hidalgo, L. Betancor, R. Moreno et al., "*Thermus thermophilus* as a cell factory for the production of a thermophilic Mn-dependent catalase which fails to be synthesized in an active form in *Escherichia coli*," *Applied and Environmental Microbiology*, vol. 70, no. 7, pp. 3839–3844, 2004.
- [9] S. Ebara and Y. Shigemori, "Alkali-tolerant high-activity catalase from a thermophilic bacterium and its overexpression in *Escherichia coli*," *Protein Expression and Purification*, vol. 57, no. 2, pp. 255–260, 2008.
- [10] M. Gudelj, G. O. Fruhwirth, A. Paar et al., "A catalase-peroxidase from a newly isolated thermoalkaliphilic *Bacillus* sp. with potential for the treatment of textile bleaching effluents," *Extremophiles*, vol. 5, no. 6, pp. 423–429, 2001.
- [11] X.-L. Shi, M.-Q. Feng, J. Shi, Z.-H. Shi, J. Zhong, and P. Zhou, "High-level expression and purification of recombinant human catalase in *Pichia pastoris*," *Protein Expression and Purification*, vol. 54, no. 1, pp. 24–29, 2007.
- [12] G. Gellissen, M. Piontek, U. Dahlems et al., "Recombinant *Hansenula polymorpha* as a biocatalyst: coexpression of the spinach glycolate oxidase (GO) and the *S. cerevisiae* catalase T (CTT1) gene," *Applied Microbiology and Biotechnology*, vol. 46, no. 1, pp. 46–54, 1996.
- [13] T. Rochat, A. Miyoshi, J. J. Gratadoux et al., "High-level resistance to oxidative stress in *Lactococcus lactis* conferred by *Bacillus subtilis* catalase KatE," *Microbiology*, vol. 151, no. 9, pp. 3011–3018, 2005.
- [14] X. Shi, M. Feng, Y. Zhao, X. Guo, and P. Zhou, "Overexpression, purification and characterization of a recombinant secretory catalase from *Bacillus subtilis*," *Biotechnology Letters*, vol. 30, no. 1, pp. 181–186, 2008.
- [15] S. Xu, Y. Guo, G. Du, J. Zhou, and J. Chen, "Self-cloning significantly enhances the production of catalase in *Bacillus subtilis* WSHDZ-01," *Applied Biochemistry and Biotechnology*, vol. 173, no. 8, pp. 2152–2162, 2014.
- [16] H. Luo, Y. Zhou, Y. H. Chang, L. Xiong, and L. Z. Liu, "Rapid gene cloning, overexpression and characterization of a thermophilic catalase in *E. coli*," *Advanced Materials Research*, vol. 365, pp. 367–374, 2012.
- [17] F. W. Studier, "Protein production by auto-induction in high density shaking cultures," *Protein Expression and Purification*, vol. 41, no. 1, pp. 207–234, 2005.
- [18] J. Zhou, M. A. Bruns, and J. M. Tiedje, "DNA recovery from soils of diverse composition," *Applied and Environmental Microbiology*, vol. 62, no. 2, pp. 316–322, 1996.
- [19] Y. Deng, Z. Hua, Z. Zhao, D. Yao, G. Du, and J. Chen, "Effect of nitrogen sources on catalase production by *Bacillus subtilis* WSHDZ-01," *Chinese Journal of Applied and Environmental Biology*, vol. 14, no. 4, pp. 544–547, 2008.

- [20] Y. Hanaoka and I. Yumoto, "Manipulation of culture conditions for extensive extracellular catalase production by *Exiguobacterium oxidotolerans* T-2-2T," *Annals of Microbiology*, vol. 65, pp. 1183–1187, 2014.
- [21] H.-W. Zeng, Y.-J. Cai, X.-R. Liao et al., "*Serratia marcescens* SYBC08 catalase isolated from sludge containing hydrogen peroxide shows increased catalase production by regulation of carbon metabolism," *Engineering in Life Sciences*, vol. 11, no. 1, pp. 37–43, 2011.
- [22] M. Nakayama, T. Nakajima-Kambe, H. Katayama, K. Higuchi, Y. Kawasaki, and R. Fuji, "High catalase production by *Rhizobium radiobacter* strain 2-1," *Journal of Bioscience and Bioengineering*, vol. 106, no. 6, pp. 554–558, 2008.
- [23] V. Neuhoﬀ, N. Arold, D. Taube, and W. Ehrhardt, "Improved staining of proteins in polyacrylamide gels including isoelectric focusing gels with clear background at nanogram sensitivity using Coomassie Brilliant Blue G-250 and R-250," *Electrophoresis*, vol. 9, no. 6, pp. 255–262, 1988.
- [24] M. M. Bradford, "A rapid and sensitive method for the quantitation of microgram quantities of protein utilizing the principle of protein-dye binding," *Analytical Biochemistry*, vol. 72, no. 1-2, pp. 248–254, 1976.
- [25] R. F. Beers Jr. and I. W. Sizer, "A spectrophotometric method for measuring the breakdown of hydrogen peroxide by catalase," *The Journal of Biological Chemistry*, vol. 195, no. 1, pp. 133–140, 1952.
- [26] H. Lineweaver and D. Burk, "The determination of enzyme dissociation constants," *Journal of the American Chemical Society*, vol. 56, no. 3, pp. 658–666, 1934.
- [27] D. K. Bol and R. E. Yasbin, "The isolation, cloning and identification of a vegetative catalase gene from *Bacillus subtilis*," *Gene*, vol. 109, no. 1, pp. 31–37, 1991.
- [28] F. Wang, Z. Wang, W. Shao, J. Liu, C. Xu, and J. Zhuge, "Purification and properties of thermostable catalase in engineered *E. coli*," *Acta microbiologica Sinica*, vol. 42, no. 3, pp. 348–353, 2002.
- [29] S. Loprasert, S. Negoro, and H. Okada, "Cloning, nucleotide sequence, and expression in *Escherichia coli* of the *Bacillus stearothermophilus* peroxidase gene (perA)," *Journal of Bacteriology*, vol. 171, no. 9, pp. 4871–4875, 1989.
- [30] Y. Zhong, X.-H. Zhang, J. Chen et al., "Overexpression, purification, characterization, and pathogenicity of *Vibrio harveyi* hemolysin VHH," *Infection and Immunity*, vol. 74, no. 10, pp. 6001–6005, 2006.
- [31] H.-W. Zeng, Y.-J. Cai, X.-R. Liao, F. Zhang, and D.-B. Zhang, "Production, characterization, cloning and sequence analysis of a monofunctional catalase from *Serratia marcescens* SYBC08," *Journal of Basic Microbiology*, vol. 51, no. 2, pp. 205–214, 2011.
- [32] H. Kimoto, K. Yoshimune, H. Matsuyama, and I. Yumoto, "Characterization of catalase from psychrotolerant *Psychrobacter piscatorii* T-3 exhibiting high catalase activity," *International Journal of Molecular Sciences*, vol. 13, no. 2, pp. 1733–1746, 2012.
- [33] Y. Zhang, X. Li, R. Xi, Z. Guan, Y. Cai, and X. Liao, "Characterization of an acid-stable catalase KatB isolated from *Bacillus altitudinis* SYBC hb4," *Annals of Microbiology*, vol. 66, no. 1, pp. 131–141, 2016.
- [34] H. Wang, Y. Tokusige, H. Shinoyama, T. Fujii, and T. Urakami, "Purification and characterization of a thermostable catalase from culture broth of *Thermoascus aurantiacus*," *Journal of Fermentation and Bioengineering*, vol. 85, no. 2, pp. 169–173, 1998.
- [35] W. Wang, M. Sun, W. Liu, and B. Zhang, "Purification and characterization of a psychrophilic catalase from Antarctic *Bacillus*," *Canadian Journal of Microbiology*, vol. 54, no. 10, pp. 823–828, 2008.
- [36] M. S. Lorentzen, E. Moe, H. M. Jouve, and N. P. Willassen, "Cold adapted features of *Vibrio salmonicida* catalase: characterisation and comparison to the mesophilic counterpart from *Proteus mirabilis*," *Extremophiles*, vol. 10, no. 5, pp. 427–440, 2006.
- [37] I. Yumoto, D. Ichihashi, H. Iwata et al., "Purification and characterization of a catalase from the facultatively psychrophilic bacterium *Vibrio rumoiensis* S-1T exhibiting high catalase activity," *Journal of Bacteriology*, vol. 182, no. 7, pp. 1903–1909, 2000.
- [38] K. Phucharoen, K. Hoshino, Y. Takenaka, and T. Shinozawa, "Purification, characterization, and gene sequencing of a catalase from an alkali- and halo-tolerant bacterium, *Halomonas* sp. SK1," *Bioscience, Biotechnology and Biochemistry*, vol. 66, no. 5, pp. 955–962, 2002.
- [39] J. Switala and P. C. Loewen, "Diversity of properties among catalases," *Archives of Biochemistry and Biophysics*, vol. 401, no. 2, pp. 145–154, 2002.