



Review Research Advances of Beneficial Microbiota Associated with Crop Plants

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Abstract: Plants are associated with hundreds of thousands of microbes that are present outside on the surfaces or colonizing inside plant organs, such as leaves and roots. Plant-associated microbiota plays a vital role in regulating various biological processes and affects a wide range of traits involved in plant growth and development, as well as plant responses to adverse environmental conditions. An increasing number of studies have illustrated the important role of microbiota in crop plant growth and environmental stress resistance, which overall assists agricultural sustainability. Beneficial bacteria and fungi have been isolated and applied, which show potential applications in the improvement of agricultural technologies, as well as plant growth promotion and stress resistance, which all lead to enhanced crop yields. The symbioses of arbuscular mycorrhizal fungi, rhizobia and Frankia species with their host plants have been intensively studied to provide mechanistic insights into the mutual beneficial relationship of plant-microbe interactions. With the advances in second generation sequencing and omic technologies, a number of important mechanisms underlying plant-microbe interactions have been unraveled. However, the associations of microbes with their host plants are more complicated than expected, and many questions remain without proper answers. These include the influence of microbiota on the allelochemical effect caused by one plant upon another via the production of chemical compounds, or how the monoculture of crops influences their rhizosphere microbial community and diversity, which in turn affects the crop growth and responses to environmental stresses. In this review, first, we systematically illustrate the impacts of beneficial microbiota, particularly beneficial bacteria and fungi on crop plant growth and development and, then, discuss the correlations between the beneficial microbiota and their host plants. Finally, we provide some perspectives for future studies on plant-microbe interactions.

Keywords: endosphere; microbiota; phyllosphere; plant-microbe interaction; rhizosphere

1. Introduction

A microbiota comprises all microbes, including viruses, bacteria, archaea, protozoa, and fungi that are present in a special environment, which is usually investigated together with its host and surrounding environment such as plant, animal gut, and soil, whereas a microbiome includes all the living microorganisms (viruses, bacteria, archaea, and lower and higher eukaryotes), their genomic sequences, and the environmental conditions surrounding the entire habitat [1–3]. Microbes are highly diverse and abundantly present in the Earth's ecosystem, and can survive in extreme environments [1,2]. Microbes are closely associated with their living environment, and environmental changes can alter different microbial species [1,4,5]. Thus, different crop plant species can selectively assemble various microbes in their rhizosphere, phyllosphere, and endosphere [6,7]. Studies have revealed that the microbiota associated with plants is crucial in determining plant performance and health, because certain beneficial microbes improve plant growth and resistance to stresses [8–12].

Crop plants which are cultivated on a large scale in fields for human consumption, livestock feeding, profits, or as raw materials for industrial products are often referred to as agronomic crops [13,14]. With the industrialization and development of human society, improvement of plant productivity and product quality in a natural environment remains a concern. The technologies for crop cultivation have also changed with the use of machines, chemical fertilizers, and pesticides, and thus the composition of microbiota associated with plants has been affected [12,15,16]. Increasing evidence has shown that plant-associated microbiota plays important roles in plant growth and development and is able to provide protection for plants against invading pathogens and various abiotic stresses [17,18]. Moreover, studies have proven that different assemblies of microbiota occur in association with different members of the plant kingdom [19–21]. One recent report revealed that the nitrogen (N) absorptions by both the rice (Oryza sativa) indica and japonica species are associated with the NRT1.1B gene, encoding a rice nitrate transporter and sensor, which is responsible for root-associated microbiota composition and N usage in field-grown rice [22]. Community structures of rhizomicrobiomes associated with four traditional Chinese medicinal herbs (Mentha haplocalyx, Perilla frutescens, Glycyrrhiza uralensis, and Astragalus membranaceus) presented significantly different bacterial and fungal communities after culturing in the same soil and air conditions [20].

The advance of next generation sequencing (NGS) technologies has presented and promoted an effective method for studying the plant–microbe interactions by facilitating the investigations of whole plant-associated microbial communities [23–25]. Numerous research groups have focused on studies of microbial communities using the advanced sequencing technologies. For instance, using NGS technologies, Perez-Jaramillo et al. reported a link between the rhizosphere microbiota compositions of domesticated and wild common bean (*Phaseolus vulgaris*) with their genotypic and root phenotypic traits [26], while Shenton et al. performed a study to examine the effects of *O. rufipogon* (wild rice) and *O. sativa* (cultivated rice) on the rhizosphere bacterial community compositions [27].

Studies have reported that certain species of bacteria and fungi are closely associated with their host plants [28,29]. Various genera of beneficial microbes associated with crop plants have been studied for their symbiotic formation with the host plants in recent years [30,31]. Previous reports have detailed the importance of microbiota on their host plants; however, only a few reviews have been available on this important topic in a systemic manner [17,18,32]. Thus, in this review, we have made an effort to systemically illustrate the latest research advances of beneficial microbiota associated with crop plants and make suggestions for the research trends on the plant and microbiota relationships.

2. Beneficial Microorganisms for Crop Plants

2.1. Plant Growth-Promoting Bacteria

Plant growth-promoting bacteria (PGPBs) occupy a large proportion of plant microbes and are able to promote plant growth under various environmental conditions in different ways. PGPBs such as beneficial plant-symbiotic bacteria, including rhizobia and *Frankia* spp., form a symbiotic structure in plant roots [30,31] (Figure 1a). The host plants of rhizobia and *Frankia* spp. are leguminous and non-leguminous plants (e.g., actinorhizal plants), respectively [30]. The symbiosis of rhizobia with legumes is a host-specific association [33], and the rhizobia from each genus are well known to nodulate a specific host legume to generate nodules that fix atmospheric nitrogen (N₂) [33]. Among the rhizobia,

the *Rhizobium* spp. are often associated with chickpea (*Cicer arietinum*) and common bean, whereas the *Bradyrhizobium* spp. are largely found to nodulate cowpea (*Vigna unguiculata*) and soybean (*Glycine max*) plants [34]. For instance, *B. japonicum* fixes the N₂ to promote plant growth, and thus improves soybean production and reduces the fertilizer requirement in the fields [35]. *Rhizobium*-legume association is used as a biofertilizer in several countries due to the effective of N₂ fixation of rhizobia [36]. It is well known that inoculation of rhizobia also helps to save the N source for cereal production, thus, reducing the application of N fertilizers which in turn conserves the environment [31,37–40]. Interestingly, Wu et al. indicated that inoculation of rice with one species of rhizobia, *Sinorhizobium meliloti*, helped rice growth by elevating the expression of genes that function to enhance cell expansion and accelerate cell division [41]. Furthermore, another study revealed that the alliance of arbuscular mycorrhizal (*Glomus mosseae*) and rhizobial (*R. leguminosarum*) symbioses alleviated damage to clover (*Trifolium repens*) by root hemiparasitic *Pedicularis* species [42].



Figure 1. (a) Beneficial microbes and microbiota for crop plants. Beneficial microbiota assembles in rhizosphere, phyllosphere, and endosphere. Arbuscular mycorrhizal fungi (AMFs), *Frankia* spp. and *Rhizobium* spp. act as symbionts for plant roots; (b) Future research trends of microbiota and microbiomes associated with crop plants. PGPBs, plant growth-promoting bacteria; PGPFs, plant growth-promoting fungi.

The N₂-fixing *Frankia*, one genus belonging to the actinobacteria group, form the actinorhizal nodules with actinorhizal plants, and promote the growth and abiotic resistance for several woody plants, such as *Alnus rubra*, *Hippophae tibetana*, *Elaeagnus angustifolia*, and *Casuarina glauca* [30,43–46]. Root nodule formation in plants by *Frankia* spp. is a host plant-specific process, and the *Frankia* spp. can be clustered into three clusters that include some identified species, namely (1) *Frankia alni*, *F. casuarinae*, and *F. torreyi*; (2) *F. coriariae* and *F. datiscae*; and (3) *F. elaeagni* and *F. irregularis*, which represent the associated *Alnus*, *Dryas* and *Elaeagnus* hosts, respectively [47–49]. Ngom et al. determined that the *Frankia* spp. in Cluster 1 can also promote the growth of *Casuarina glauca* through symbiosis [50]. Several reports revealed that *Alnus*, *Hippophae*, and *Elaeagnus* plants can resist drought and salt stress with the help of specific *Frankia* strains [51,52]. On the basis of the results of genome analyses, *Frankia* strains possess some shared or specific genes for promoting plant growth, as well as genes related to N₂ fixation [49]. Further studies are required to understand the underlying mechanisms, as well as the clustering of the *Frankia* species based on their specific symbiotic relationship with the host plants [52–54].

Other types of PGPBs, either residing in the rhizosphere or phyllosphere or endosphere, can also promote plant growth and stress resistance [5]. These PGPBs have some beneficial traits, such as the ability to produce indole-3-acetic acid (IAA), 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase and siderophores, as well as to solubilize phosphate [55–57]. Furthermore, some PGPBs also compete with certain pathogenic bacteria or fungi for nutrients or niches, and thus inhibit the spread of pathogens and reduce damage to plants [58]. Some PGPB species of *Bacillus* and *Pseudomonas* genera have been studied for their potential applications in sustainable agriculture development [59–61]. For instance, PGPBs belonging to *Bacillus* and *Pseudomonas*, as well as *Azospirillum* and *Azotobacter* genera were

shown to improve alkaloid contents in the medicinal plant *Withania somnifera* in a two-consecutive-year experiment [62], while *P. putida* and *P. fluorescens* were proven to improve tropane alkaloid production in *Hyoscyamus niger* plants under water scarcity [63]. In this context, Maggini et al. proposed *Echinacea purpurea* as a new model plant to study crosstalk between medicinal plants and bacterial endophytes, with the aim to discover bioactive compounds [64]. Their suggestion was made based on the finding that the secondary metabolite levels in non-infected and infected plants were different, indicating that the bacterial infection modulated the biosyntheses of several secondary metabolites [64]. Additionally, the transcript levels of a gene involved in the alkamide metabolic pathway were higher in the roots of infected *E. purpurea* than in that of the non-infected control [64]. Several secondary metabolites produced by PGPBs can also be used to control pathogens and salinity [60,65]. Several studies have also reported that PGPBs lead to induced systemic resistance (ISR) in plants, which helps the plants to resist the pathogens [66–68] (Figure 1b).

2.2. Plant Growth-Promoting Fungi

Plant growth-promoting fungi (PGPFs) have gained immense attention as biofertilizers due to their role in maintaining plant quality and quantity and their environment-friendly relationship [69] (Figure 1a). Advancements in the development of PGPFs for crop cultivation have been achieved in Salvia miltiorrhiza, grapevine (Vitis vinifera), and lettuce (Lactuca sativa) [69–72]. For instance, Trichoderma spp., Ganoderma spp., and yeasts (Saccharomyces spp.) have been used as PGPFs in several studies under both normal growth and environmental stress conditions [70,73–79]. For example, Jaroszuk-Scisel et al. reported that a Trichoderma strain, isolated from a healthy rye (Secale cereale) rhizosphere with the ability to produce auxin, gibberellins, and ACC deaminase in vitro, and to inhibit the growth of *Fusarium* spp., could colonize the roots of wheat (*Triticum aestivum*) seedlings and enhance stem growth [73]. Another study revealed that *T. asperellum* could induce peroxidase and polyphenol oxidase, as well as the cell wall degrading enzymes chitinase and β -1,3-glucanase in lettuce plants to defend the infected plants against the leaf spot disease [70]. Several isolated killer yeast S. cerevisiae strains were shown to serve as PGPFs for controlling *Colletotrichum gloeosporioides* in grape planting [80]. Cell wall isolated from yeast (Rhodosporidium paludigenum) could induce disease resistance against Penicillium expansum in pear (*Pyrus pyrifolia*) fruits by inducing the activities of defense-associated enzymes such as β -1,3-glucanase and chitinase, and by upregulating expression of pathogenesis-related (PR) genes in plants [75].

Arbuscular mycorrhizal fungi (AMFs), equal to the term "Glomeromycota" at the fungal phylum level, can benefit plants as PGPFs during nutrient absorption and stress resistance by acting as symbionts to their hosts [81,82] (Figure 1a). Being one group of obligate endomycorrhizas that has occurred for nearly 400 million years, AMFs also play a key role in liverworts (Marchantiophyta), hornworts (Ceratophyllum demersum) and lycophytes by helping them adapt to the land during the early years [83]. Spagnoletti et al. reported that AMFs promote soybean production by helping soybean plants resist arsenic (As) contaminated soils [82], while Cely et al. indicated that inoculation of *Rhizophagus clarus* increased the soybean production under field conditions [84]. Moreover, AMFs improved rice production by reducing the total content of As, and improving the nitrogen/carbon (N:C) ratio in rice grains [85,86]. Furthermore, one study reported that selenium (Se) \times AMF interactions promoted both soybean and forage grass (Urochloa decumbens) for Se absorption, which improved the quality of the two plants used as animal feed [87]. Rhizoglomus intraradices, one model species of AMFs, help rice, soybean, ginseng (*Panax ginseng*), and other crops resist drought and cold stresses and promote nutrient absorption [10,88,89]. R. intraradices also promoted rice plants to resist blast disease (caused by Magnaporthe oryzae spores), and the transcriptome data revealed that R. intraradices-inoculated rice plants exhibited significantly higher expression levels of genes related to the auxin and salicylic acid signaling pathways than the noninoculated rice plants in response to *M. oryzae* [11].

2.3. Biocontrol Agents

The biocontrol agents (BCAs), including various beneficial AMFs, PGPBs, and PGPFs have been investigated by several scientists in the last 30 years [11,90–93]. For example, various species of *Pseudomonas, Saccharomyces, Streptomyces,* and *Trichoderma* genera are the most effective BCAs that help plants acquire nutrients and control the plant pathogens [71,91,94]. These species possess a number of antagonistic mechanisms, including competition for nutrients and space with pathogenic fungi or bacteria, producing antifungal and bacterial compounds for resisting pathogenic fungi and bacteria, and direct parasitism [71,91,94]. In addition, these microbes can trigger plant resistance by strengthening the cell wall and enhancing the physiological and biochemical responses of plants to stresses [95].

3. Why Plants Need Beneficial Microbiota

As discussed above, a myriad of microbes populated on plant surfaces or inside the plants can considerably affect plant growth, nutrient uptake, and resistance to environmental stresses [11,90–93]. Therefore, gaining deep insights into the mechanisms underlying the evolution, compositions, and functions of plant microbiomes would open new avenues to enhance crop health and yield, ensuring global food security [96]. Wei et al. proposed a framework for plant breeding, which would enable plants to obtain economically novel phenotypes by altering plants' genomic information along with plant-associated microbiota [97]. In the breeding strategy of plants, microbes are considered to be one of the direct targets subjected to selection for achieving a desirable plant phenotype [97].

Bai et al. reported different bacterial communities in the rhizophere and phyllosphere [98]. Roots secrete some organic C and N sources into the rhizosphere, which help in enrichment and assembly of the soil microbes in the rhizosphere [99–101]. A number of studies have revealed that the correlation between microbiota and related metabolic pathways in plants mutually affects the growth and responses of plants to environmental stresses [21,102,103]. Considering various compounds secreted by different plants, a different rhizosphere microbiota is associated with a different plant species to support their performance and stress resistance [20,104,105]. The beneficial microbiota, containing AMFs, PGPBs, and PGPFs, also affects the metabolism of various substances in plants [10,106,107]. This mutual relationship between microbiota and plants effectively supports plant performance and resistance to adverse environmental conditions [8–11] (Figure 1b).

Soil represents an extremely rich microbial reservoir on the Earth [108]. Soil is the origin of the rhizosphere microbiota, and a driver of microbial community formation [109,110]. The diversity and abundance of a microbiota in soil are about $(1~10) \times 10^9$ CFU bacteria/g soil and $(1~10) \times 10^5$ colony-forming unit (CFU) fungi/g soil in normal soils [111,112]. Rhizosphere is the soil surrounding the plant roots within 2 to 5 mm, in which numerous microbes assemble [25,113,114]. The rhizosphere of the crop plants acts as the major ecological environment for establishment of plant–microbe interactions. The mutual interactions involve colonization of numerous microbes growing in the inner parts or on the surface of roots, and the microbiota result in associative, symbiotic, and even parasitic collaborations among the plants and the associated microflora [115,116]. Soil, as the natural medium for bacteria and fungi, provides C and N sources needed by these microbes [117,118]; however, these C and N sources are normally provided by plants, further illustrating the mutual relationship between plants and microbes. Furthermore, the soil characteristics differ due to the flora grown on it, the microbes in the soil, and soil management by humans [19,20,119].

Whereas the use of chemicals, such as insecticides, herbicides and pesticides, hampers the quality of plant products, and thus adversely affects human health [120,121], extracts and metabolites of PGPBs and PGPFs effectively protect plants from various biotic and abiotic adversities [60,77,122]. Thus, if the microbiota containing PGPBs and PGPFs is effectively used, the problems regarding crop production could be solved in an efficient and environment-friendly manner. Endophytic microbiota residing inside the roots, stems, and leaves of plants help to maintain plant health and help plants resist adverse conditions [123]. It has been reported that plants selectively construct the community of

their endophytic microbiota, which helps in upregulating expression levels of stress resistance-related genes in plants [123].

In addition, plants need soil microbiota for degrading their residues [124]. Plant residues are abundantly found in the soil [124,125]. Although chemical or physical methods can help degrade the residues, the natural degradation is carried out by soil microbes [125]. The microbiota assembles and is enriched in the plant residues and further degrades them into macro- or micromolecules, which can serve as soil organic matter [125]. For instance, the utilization of plant residues and enrichment of soil microbiota could improve the organic matter content and composition of the soils in northeast China [126,127]. Improved soil organic matter properties protect the soil from destruction and maintain its nutrient content, which would make the soil more suitable for crop production [128,129]. In addition, when plants are grown under biotic or abiotic stress conditions, such as in pathogenic fungi-contaminated, heavy metal-contaminated, or alkaline soils, the plant-associated microbiota helps them resist stresses by mediating plant abscisic acid (ABA) levels, plant jasmonic acid (JA) levels, or Bradford reactive soil protein protection [11,130,131].

4. Influences of the Community Compositions of Rhizosphere, Phyllosphere, and Endosphere Microbiota on Growth and Performance of Crop Plants

A few advances have been made to illustrate the diversity and community structure of the microbes associated with crop plants, and their relationship with the changes in plant metabolites and gene expression [21,102,132]. Community compositions of the rhizosphere, phyllosphere, or endosphere microbiota associated with plants indicate the health or nutrient conditions of the plants [21,115,116,133,134]. On the one hand, the highly enriched pathogenic fungi in the rhizosphere indicate stress conditions that can adversely affect the growth of crop plants [10,135]. On the other hand, the beneficial rhizospheric microbes improve plant growth, nutrient absorption, and development based on the mechanisms, such as organic matter mineralization, disease containment against soil-borne pathogens, N₂ fixation, potassium (K) and phosphate solubilization, and IAA and ACC deaminase production [115,116]. Thus, the soil/plant/microbial relationship must be appropriately maintained for sustainable agricultural practices [115,116]. Phyllospheric microbes, consisting of mostly bacteria and fungi, can act as (i) beneficial mutualists that improve plant growth and resistance to environmental stresses; (ii) commensals that use the leaf habitat for their own growth, development, and reproduction; or (iii) antagonistic pathogens [136–138] (Figure 1a). A study of 14 phylogenetically diverse plant species grown under controlled greenhouse conditions showed high presentation of *Bacillus* and Stenotrophomonas genera in their phyllosphere microbial communities, which showed antagonistic potential toward Botrytis cinerea [134], a foliar pathogen that causes gray mold disease in more than 200 dicot crop plants [139].

Endophytic bacteria and fungi, inhabiting within the plant tissues, play crucial roles in plant growth, fitness, development and protection without causing any evident damage to the host plants [140,141]. These endophytic bacteria and fungi reside on intercellular spaces in the plants for a certain period of their life cycle, and obtain carbohydrates, amino acids, and inorganic nutrients from their hosts [142]. Some endophytic bacteria or fungi also produce IAA, soluble phosphate, and siderophores, which can promote the host plant growth [143]. For instance, Borah et al. studied the endophytic microbes of cultivated rice (*O. sativa*) and wild rice (*O. rufipogon*) plants and reported their diversity in terms of their functional characteristics related to multiple traits associated with promotion of plant growth and development [144]. Murphy et al. screened fungal endophytes from wild barley (*Hordeum murinum*) for testing the beneficial and promoting traits in cultivated barley (*H. vulgare*) and found that some fungi could promote the growth of cultivated barley [145].

Furthermore, a few studies have reported that both artificial management of plant growth environment and conditions, and plant domestication affect the community structure of rhizosphere and endosphere microbiomes [27,119,146,147]. Wild plant species preserve more diversity in associated microbiota for their survival against stress resistance than the domesticated crops. A recent study

reported that common wild rice (*O. rufipogon*) grown in wildland of lower soil nutrients showed higher rhizobacterial diversity than the cultivated rice grown under cultivated field conditions with higher soil nutrient [28]. Martín-Robles et al. discussed the impacts of domestication on the arbuscular mycorrhizal symbioses of 14 crop species under different available phosphorus conditions, and reported that in response to increased available phosphorus, domestication decreased the AMF colonization in domesticated plants as compared with their wild progenitors [148]. Thus, more studies revealing the influence of plant domestication on the compositions of microbiomes are warranted, and the rhizosphere microbiota in wild and cultivated species grown in the field conditions are essentially required to study the correlation of microbiota and host plants.

A plant can release various chemical compounds into the environment that impose either direct or indirect allelochemical effect on another plant [149] (Figure 1b). In addition, studies have revealed that the production of allelochemicals by plants significantly alters the structure of associated microbiomes [150–152]. However, the influence of microbiota on the allelochemical production in plants remains unclear. Moreover, the relationships of specific allelochemicals with a particular microbiota remain unillustrated. A recent study by Li et al. concluded that replantation of Sanqi ginseng (*P. notoginseng*) is not recommended mainly due to the accumulation of soil-borne pathogens and allelochemicals (e.g., ginsenosides) [153]. Li et al. also demonstrated that reductive soil disinfestation approach could effectively alleviate the replantation failure of Sanqi ginseng with allelochemical degradation and pathogen suppression [153] (Table 1). However, the reason behind the continuous cropping obstacle on the community compositions of microbiomes remains the task of future research.

5. Applications of Individual Microbes for Improvement of Crop Performance and Soil Ameliorations

Various beneficial microorganisms have been commercialized [154]; however, their efficacy has not always been consistent in terms of benefiting crop plants in fields. Nevertheless, combining beneficial microbes for application to crop plants has been shown to be more effective in improving plant performance. The PGPBs and PGPFs have been applied in agricultural practices for 30 years. For instance, applications of the AMFs have been known to improve the yield of cotton (Gossypium *hirsutum*) and soybean, as well as soybean resistance to the *Macrophomina phaseolina* pathogen and stress associated with As [82,84]. AMFs also promote N transfer from soil to plants as indicated by a study of Tian et al., in carrot (*Daucus carota*) [155]. The authors also reported that *R. intraradices*-mediated N transfer successively induced the expression of N metabolism-related genes in the intraradical and extraradical mycelia, as well as in the host plants [155]. The interactive effects of the AMF R. irregularis and the PGPB P. putida synergistically improved the growth and defence of the wheat host plants against pathogens [156]. With the global warming, drought has been one of the major limiting conditions for crop production [157]. Studies have shown that AMFs enhanced rice and wheat in resisting drought by improving the plant physiological index [158,159]. Application of PGPBs such as Azospirillum, Azotobacter, Pseudomonas, and Bacillus species improved the alkaloid contents in two medicinal plants W. somnifera and H. niger under limited water conditions [62,63]. With regards to the endosymbiotic PGPBs, Chandrakala et al. screened and isolated one *Rhizobium* species from rice rhizosphere, which could solubilize silicate and promote plant growth potential of the rice plants [160]. B. japonicum has been shown to help soybean to resist a high content of As in the soil [161], whereas S. fredii could effectively nodulate numerous different legumes, thereby promoting growth of the host plants [162]. Vemulapally et al. screened and isolated *Frankia* spp. from the nodules of *Casuarina* spp., one of which (F. casuarinae CcI3 strain) could form nodules in roots of C. equisetifolia seedlings after its inoculation to the plants and promote plant growth [163]. Forchetti et al. demonstrated that certain bacterial species can produce JA and ABA [4]. However, whether the contents of these hormones in plants can be influenced directly by the associated microbes remains unclear. Nevertheless, hormone-producing bacteria or fungi are not always beneficial; for example, Gibberella fujikuroi can produce gibberellin and cause rice bakanae disease [164] (Table 1). Thus, studies on the mutual effects of microbes and host

plants on hormone metabolism in both plants and microbes in the context of plant and microbiota relationships have been novel and interesting, and thus this topic would deserve more attention from the research community.

Microbe-mediated remediation of environmental contaminants requires more research for heavy metal-polluted soils and saline-sodic soils (Figure 1b). With the industrialization, environmental contaminants, such as heavy metal, household garbage, and excessive N and phosphate in the soil or water, have seriously affected the quality of humans' and plants' life. Mitigation of heavy metal contamination in soil by fungal bioremediation has proceeded for several years [165,166], and studies for exploring heavy metal-polluted soils for agriculture by cultivating stress-tolerant plants with the help of microbes are also in process [167]. Cornejo et al. revealed that AMFs assisted Oenothera picensis in resisting copper stress and helped the bioremediation of copper-polluted soil [130]. Saline-sodic soils have been studied for several years to improve their soil characteristics for crop cultivation with higher productivity [19,119]. Chemical and biological ameliorations of saline-sodic soils are the most effective ways; however, both methods affect the compositions of soil microbiomes. For example, Luo et al. revealed that ameliorations of saline-sodic soils by using chemicals or plant planting affected the soil microbiota [19,168], while Shi et al. found various responses of microbial communities and enzyme activities to various chemical and biological amendments in the saline-alkaline soils [119] (Table 1). Research on plant-associated microbiota to improve the soil characteristics is further warranted to facilitate microbiota applications in different aspects of agriculture and life, and more studies considering microbiota applications along with plant cultivation for soil ameliorations are required.

As discussed earlier, beneficial microbes can also improve the useful secondary metabolites in medicinal plants, demonstrating the wide application potential of microbes and microbiota to various types of crops. Application of a microbial consortium of various PBGBs, including *Azospirillum, Azotobacter, Pseudomonas*, and *Bacillus* spp., improved the levels of alkaloids, which have pharmacological properties [62,63], in *W. somnifera* and *H. niger* [62,63]. Furthermore, Huang et al. advocated *S. miltiorrhiza*, which is a herb widely used in traditional Chinese medicine, as a model species for investigating how different microbes can interact with medicinal plants to alter the production of phytochemical compounds [169]. Additionally, it is well known that *Taxus chinensis* can produce taxol, an anticancer metabolite [170], and its endophytic fungi *Paraconiothyrium* spp. can also produce the same anticancer metabolite [171–173]. However, it is not clear whether the content of taxol in *T. chinensis* could be improved by its endophytic fungi, as well as the relationship of *T. chinensis* and associated microbiota in terms of metabolite production has also not yet been determined. Thus, the effect of microbiota on the metabolic pathways of their host plant deserves more studies in future (Figure 1b).

Research Aspects	Microbes/ Microbiomes	Beneficial Traits	References
Rhizobia/ <i>Frankia</i> in colonization with host plants	Rhizobia/ Frankia spp.	<i>Rhizobium</i> spp. promoted the growth of rice seedlings and solubilized silicate;	[160]
		Bradyrhizobium japonicum helped soybean in resisting arsenic contamination in soil	[161]
		<i>Frankia casuarinae</i> Ccl3 formed nodules and promoted the growth of <i>Casuarina equisetifolia</i>	[163]

Table 1. Several representative applications of microbes in crop plants. ACC, 1-aminocyclopropane-1carboxylic acid; AMFs, arbuscular mycorrhizal fungi; IAA, indole-3-acetic acid; JA, jasmonic acid; ABA, abscisic acid; PGPBs, plant growth-promoting bacteria; PGPFs, plant growth-promoting fungi.

Research Aspects	Microbes/ Microbiomes	Beneficial Traits	References
Positive effects of mutualistic symbioses with fungi on stress resistance	AMFs	<i>Rhizoglomus intraradices</i> reduced the relative abundance of pathogenic fungi in plant soil	[10]
		AMFs promote rice and wheat in resisting drought	[158,159]
PGPB/PGPF applications in improving plant growth, productivity, and resistance	PGPBs	Productions of IAA, ACC deaminases, JA and ABA	[55–57]
	PGPFs	<i>Trichoderma</i> spp. and yeasts help plant in resisting diseases	[70,73–79]
Allelochemical effects/soil ameliorations	Microbiomes in soil	Reductive soil disinfestation can reduce the allelochemical effect in soil.	[153]
		Soil amendments associated with changes in the compositions of soil microbiomes	[19,119]
Effects of microbiota on the production of secondary metabolites in plants	PGPBs/AMFs/ Paraconiothyr-ium	AMFs improve the content of gesenosides in <i>P. ginseng</i> planting	[10]
		Endophytic fungi <i>Paraconiothyrium</i> spp. in <i>Taxus</i> chinensis can produce taxol	[171–173]
		Azospirillum, Azotobacter, Pseudomonas and Bacillus improved the alkaloid content in Withania somnifera	[62]
		<i>P. putida</i> and <i>P. fluorescens</i> were proved to improve tropane alkaloid production of <i>Hyoscyamus niger</i> under water deficit stress	[63]

Table 1. Cont.

6. Conclusions and Future Perspectives

Microbiota plays important roles in plant growth, development, productivity, and resistance to environmental stresses. An increasing body of research has been focusing on the associations and relationships between microbiota and host plants. Although some progresses have been made in recent years, applications of microbes and microbiota to improve crop productivity still require more research. Both theoretical research and applied research of microbes and microbiota in connection with plants remain a major task for the scientific community. In the future, AMFs, PGPBs, and PGPF should be applied and utilized more effectively in agriculture. The signaling pathways of AMFs, and rhizobia and *Frankia* spp. with their host plants have been studied for a long time but are still under debate in plant science. Various genes in microbes related to hormones such as strigolactones and nutrient uptake similar to nitrogen uptake should be studied to uncover the mutual connection with the host plants.

In the future, microbiota research using both basic and applied research approaches in connection with crop plants should be focused on the following: (1) Mutualistic symbioses of the fungal and bacterial microbes with the host plants; (2) plant diseases caused by the environmental microbiota; (3) mechanistic studies and explorations of PGPBs and PGPFs for the improvement of plant growth, productivity and resistance, and yield quality; (4) allelochemical effects between plants and microbiota; and (5) mechanisms underlying the degradation of plant residues by the microbiota for acceleration of practical use in this field. Future research on plant and microbiota associations and relationships could play an essential role in ensuring food security in response to global climate change.

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Abbreviations

ABA	abscisic acid
ACC	1-aminocyclopropane-1-carboxylic acid
AMFs	Arbuscular mycorrhizal fungi
BCAs	biocontrol agents
PGPBs	plant growth-promoting bacteria
PGPFs	Plant growth-promoting fungi
JA	jasmonic acid
IAA	indole-3-acetic acid

References

- 1. Gibbons, S.M.; Gilbert, J.A. Microbial diversity—Exploration of natural ecosystems and microbiomes. *Curr. Opin. Genet. Dev.* **2015**, *35*, 66–72. [CrossRef] [PubMed]
- 2. Pedrós-Alió, C.; Manrubia, S. The vast unknown microbial biosphere. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 6585. [CrossRef] [PubMed]
- 3. Marchesi, J.R.; Ravel, J. The vocabulary of microbiome research: A proposal. *Microbiome* **2015**, *3*, 31. [CrossRef] [PubMed]
- 4. Forchetti, G.; Masciarelli, O.; Alemano, S.; Alvarez, D.; Abdala, G. Endophytic bacteria in sunflower (*Helianthus annuus* L.): isolation, characterization, and production of jasmonates and abscisic acid in culture medium. *Appl. Microbiol. Biot.* **2007**, *76*, 1145–1152. [CrossRef]
- Orozco-Mosqueda, M.d.C.; Rocha-Granados, M.d.C.; Glick, B.R.; Santoyo, G. Microbiome engineering to improve biocontrol and plant growth-promoting mechanisms. *Microbiol. Res.* 2018, 208, 25–31. [CrossRef]
- Fitzpatrick, C.R.; Copeland, J.; Wang, P.W.; Guttman, D.S.; Kotanen, P.M.; Johnson, M.T.J. Assembly and ecological function of the root microbiome across angiosperm plant species. *Proc. Natl. Acad. Sci. USA* 2018, 115, 1157. [CrossRef]
- 7. Zgadzaj, R.; Garrido-Oter, R.; Jensen, D.B.; Koprivova, A.; Schulze-Lefert, P.; Radutoiu, S. Root nodule symbiosis in *Lotus japonicus* drives the establishment of distinctive rhizosphere, root, and nodule bacterial communities. *Proc. Natl. Acad. Sci. USA* **2016**, *113*, 7996. [CrossRef]
- Ferentinos, K.P. Deep learning models for plant disease detection and diagnosis. *Comput. Electron. Agric.* 2018, 145, 311–318. [CrossRef]
- 9. Nejat, N.; Rookes, J.; Mantri, N.L.; Cahill, D.M. Plant-pathogen interactions: Toward development of next-generation disease-resistant plants. *Crit. Rev. Biotechnol.* **2017**, *37*, 229–237. [CrossRef]
- 10. Tian, L.; Shi, S.; Ma, L.; Zhou, X.; Luo, S.; Zhang, J.; Lu, B.; Tian, C. The effect of *Glomus intraradices* on the physiological properties of *Panax ginseng* and on rhizospheric microbial diversity. *J. Ginseng Res.* **2019**, *43*, 77–85. [CrossRef]
- 11. Tian, L.; Chang, C.; Ma, L.; Nasir, F.; Zhang, J.; Li, W.; Tran, L.S.P.; Tian, C. Comparative study of the mycorrhizal root transcriptomes of wild and cultivated rice in response to the pathogen *Magnaporthe oryzae*. *Rice* **2019**, *12*, 35. [CrossRef]
- 12. Chaparro, J.M.; Sheflin, A.M.; Manter, D.K.; Vivanco, J.M. Manipulating the soil microbiome to increase soil health and plant fertility. *Biol. Fert. Soils* **2012**, *48*, 489–499. [CrossRef]
- 13. Fageria, N.K. The Use of Nutrients in Crop Plants; Fageria, N.K., Ed.; CRC Press: New York, NY, USA, 2016.
- 14. Byrt, C.S.; Munns, R.; Burton, R.A.; Gilliham, M.; Wege, S. Root cell wall solutions for crop plants in saline soils. *Plant Sci.* **2018**, *269*, 47–55. [CrossRef] [PubMed]
- Bhatt, P.; Nailwal, T.K. Chapter 11—Crop Improvement through Microbial Technology: A Step toward Sustainable Agriculture. In *Crop Improvement through Microbial Biotechnology*; Prasad, R., Gill, S.S., Tuteja, N., Eds.; Elsevier: New Delhi, India, 2018; pp. 245–253.
- 16. Altieri, M.A.; Nicholls, C.I.; Montalba, R. Technological approaches to sustainable agriculture at a crossroads: an agroecological perspective. *Sustainability* **2017**, *9*, 349. [CrossRef]

- 17. Müller, D.B.; Vogel, C.; Bai, Y.; Vorholt, J.A. The plant microbiota: systems-level insights and perspectives. *Annu. Rev. Genet.* **2016**, *50*, 211–234. [CrossRef]
- Hussain, S.S.; Mehnaz, S.; Siddique, K.H.M. Harnessing the Plant Microbiome for Improved Abiotic Stress Tolerance. In *Plant Microbiome: Stress Response*; Egamberdieva, D., Ahmad, P., Eds.; Springer: Singapore, 2018; pp. 21–43.
- Luo, S.; Tian, L.; Chang, C.; Wang, S.; Zhang, J.; Zhou, X.; Li, X.; Tran, L.S.P.; Tian, C. Grass and maize vegetation systems restore saline-sodic soils in the Songnen Plain of northeast China. *Land Degrad. Dev.* 2018, 29, 1107–1119. [CrossRef]
- Shi, S.; Tian, L.; Ma, L.; Tian, C. Community structure of rhizomicrobiomes in four medicinal herbs and its implication on growth management. *Microbiology* 2018, *87*, 425–436. [CrossRef]
- Shi, S.; Tian, L.; Nasir, F.; Li, X.; Li, W.; Tran, L.S.P.; Tian, C. Impact of domestication on the evolution of rhizomicrobiome of rice in response to the presence of *Magnaporthe oryzae*. *Plant Physiol. Biochem.* 2018, 132, 156–165. [CrossRef] [PubMed]
- Zhang, J.; Liu, Y.X.; Zhang, N.; Hu, B.; Jin, T.; Xu, H.; Qin, Y.; Yan, P.; Zhang, X.; Guo, X.; et al. NRT1.1B is associated with root microbiota composition and nitrogen use in field-grown rice. *Nat. Biotechnol.* 2019, 37, 676–684. [CrossRef]
- 23. Schlaeppi, K.; Bulgarelli, D. The plant microbiome at work. *Mol. Plant Microbe Interact.* **2014**, *28*, 212–217. [CrossRef]
- 24. Nasir, F.; Tian, L.; Shi, S.; Chang, C.; Ma, L.; Gao, Y.; Tian, C. Strigolactones positively regulate defense against *Magnaporthe oryzae* in rice (*Oryza sativa*). *Plant Physiol. Biochem.* **2019**, *142*, 106–116. [CrossRef] [PubMed]
- 25. Tian, L.; Shi, S.; Ma, L.; Tran, L.S.P.; Tian, C. Community structures of the rhizomicrobiomes of cultivated and wild soybeans in their continuous cropping. *Microbiol. Res.* **2020**, *232*, 126390. [CrossRef] [PubMed]
- Perez-Jaramillo, J.E.; Carrion, V.J.; Bosse, M.; Ferrao, L.F.V.; de Hollander, M.; Garcia, A.A.F.; Ramirez, C.A.; Mendes, R.; Raaijmakers, J.M. Linking rhizosphere microbiome composition of wild and domesticated *Phaseolus vulgaris* to genotypic and root phenotypic traits. *ISME J.* 2017, *11*, 2244–2257. [CrossRef] [PubMed]
- 27. Shenton, M.; Iwamoto, C.; Kurata, N.; Ikeo, K. Effect of wild and cultivated rice genotypes on rhizosphere bacterial community composition. *Rice* **2016**, *9*, 42–53. [CrossRef]
- 28. Tian, L.; Zhou, X.; Ma, L.; Xu, S.; Nasir, F.; Tian, C. Root-associated bacterial diversities of *Oryza rufipogon* and *Oryza sativa* and their influencing environmental factors. *Arch. Microbiol.* **2017**, *199*, 563–571. [CrossRef]
- 29. Yeoh, Y.K.; Dennis, P.G.; Paungfoo-Lonhienne, C.; Weber, L.; Brackin, R.; Ragan, M.A.; Schmidt, S.; Hugenholtz, P. Evolutionary conservation of a core root microbiome across plant phyla along a tropical soil chronosequence. *Nat. Commun.* **2017**, *8*, 215. [CrossRef]
- 30. Cissoko, M.; Hocher, V.; Gherbi, H.; Gully, D.; Carré-Mlouka, A.; Sane, S.; Pignoly, S.; Champion, A.; Ngom, M.; Pujic, P.; et al. Actinorhizal signaling molecules: *Frankia* root hair deforming factor shares properties with NIN inducing factor. *Front. Plant Sci.* **2018**, *9*, 1494. [CrossRef]
- Poole, P.; Ramachandran, V.; Terpolilli, J. Rhizobia: from saprophytes to endosymbionts. *Nat. Rev. Microbiol.* 2018, 16, 291–303. [CrossRef]
- 32. Escudero-Martinez, C.; Bulgarelli, D. Tracing the evolutionary routes of plant–microbiota interactions. *Curr. Opin. Microbiol.* **2019**, *49*, 34–40. [CrossRef]
- 33. Batista, L.; Irisarri, P.; Rebuffo, M.; Cuitiño, M.J.; Sanjuán, J.; Monza, J. Nodulation competitiveness as a requisite for improved rhizobial inoculants of *Trifolium pratense*. *Biol. Fert. Soils* **2015**, *51*, 11–20. [CrossRef]
- Koskey, G.; Mburu, S.W.; Kimiti, J.M.; Ombori, O.; Maingi, J.M.; Njeru, E.M. Genetic characterization and diversity of *Rhizobium* isolated from root nodules of mid-altitude climbing bean (*Phaseolus vulgaris* L.) varieties. *Front. Microbiol.* 2018, 9, 968. [CrossRef] [PubMed]
- 35. Ronner, E.; Franke, A.C.; Vanlauwe, B.; Dianda, M.; Edeh, E.; Ukem, B.; Bala, A.; van Heerwaarden, J.; Giller, K.E. Understanding variability in soybean yield and response to P-fertilizer and rhizobium inoculants on farmers' fields in northern Nigeria. *Field Crop Res.* **2016**, *186*, 133–145. [CrossRef]
- 36. Mia, M.B.; Shamsuddin, Z. *Rhizobium* as a crop enhancer and biofertilizer for increased cereal production. *Afr. J. Biotechnol.* **2010**, *9*, 6001–6009.
- Crespo-Rivas, J.C.; Navarro-Gomez, P.; Alias-Villegas, C.; Shi, J.; Zhen, T.; Niu, Y.; Cuellar, V.; Moreno, J.; Cubo, T.; Vinardell, J.M.; et al. *Sinorhizobium fredii* HH103 RirA is required for oxidative stress resistance and efficient symbiosis with soybean. *Int. J. Mol. Sci.* 2019, 20, 787. [CrossRef] [PubMed]

- Miao, J.; Zhang, N.; Liu, H.; Wang, H.; Zhong, Z.; Zhu, J. Soil commensal rhizobia promote *Rhizobium* etli nodulation efficiency through CinR-mediated quorum sensing. *Arch. Microbiol.* 2018, 200, 685–694. [CrossRef]
- 39. Siczek, A.; Lipiec, J. Impact of faba bean-seed rhizobial inoculation on microbial activity in the rhizosphere soil during growing season. *Int. J. Mol. Sci.* **2016**, *17*, 784. [CrossRef]
- 40. Ozturk, A.; Caglar, O.; Sahin, F. Yield response of wheat and barley to inoculation of plant growth promoting rhizobacteria at various levels of nitrogen fertilization. *J. Plant Nut. Soil Sci.* **2003**, *166*, 262–266. [CrossRef]
- Wu, Q.; Peng, X.; Yang, M.; Zhang, W.; Dazzo, F.B.; Uphoff, N.; Jing, Y.; Shen, S. Rhizobia promote the growth of rice shoots by targeting cell signaling, division and expansion. *Plant Mol. Biol.* 2018, 97, 507–523. [CrossRef]
- 42. Sui, X.L.; Zhang, T.; Tian, Y.Q.; Xue, R.J.; Li, A.R. A neglected alliance in battles against parasitic plants: arbuscular mycorrhizal and rhizobial symbioses alleviate damage to a legume host by root hemiparasitic *Pedicularis* species. *New Phytol.* **2019**, *221*, 470–481. [CrossRef]
- 43. Van Nguyen, T.; Pawlowski, K. *Frankia* and Actinorhizal Plants: Symbiotic Nitrogen Fixation. In *Rhizotrophs: Plant Growth Promotion to Bioremediation;* Mehnaz, S., Ed.; Springer: Singapore, 2017; pp. 237–261.
- 44. Berry, A.M.; McIntyre, L.; McCully, M.E. Fine structure of root hair infection leading to nodulation in the *Frankia–Alnus* symbiosis. *Can. J. Bot.* **1986**, *64*, 292–305. [CrossRef]
- 45. Tian, C.; He, X.; Zhong, Y.; Chen, J. Effects of VA mycorrhizae and *Frankia* dual inoculation on growth and nitrogen fixation of *Hippophae tibetana*. *For. Ecol. Manag.* **2002**, *170*, 307–312. [CrossRef]
- 46. Gtari, M.; Brusetti, L.; Skander, G.; Mora, D.; Boudabous, A.; Daffonchio, D. Isolation of *Elaeagnus*-compatible *Frankia* from soils collected in Tunisia. *FEMS Microbiol. Lett.* **2006**, 234, 349–355. [CrossRef]
- 47. Samant, S.; Dawson, J.O.; Hahn, D. Growth responses of introduced *Frankia* strains to edaphic factors. *Plant Soil* **2016**, 400, 123–132. [CrossRef]
- Wolters, D.J.; Van Dijk, C.; Zoetendal, E.G.; Akkermans, A.D.L. Phylogenetic characterization of ineffective *Frankia* in *Alnus glutinosa* (L.) Gaertn. nodules from wetland soil inoculants. *Mol. Ecol.* 1997, *6*, 971–981. [CrossRef] [PubMed]
- 49. Nouioui, I.; Cortés-albayay, C.; Carro, L.; Castro, J.F.; Gtari, M.; Ghodhbane-Gtari, F.; Klenk, H.P.; Tisa, L.S.; Sangal, V.; Goodfellow, M. Genomic insights into plant-growth-promoting potentialities of the genus *Frankia*. *Front. Microbiol.* **2019**, *10*. [CrossRef]
- 50. Ngom, M.; Diagne, N.; Laplaze, L.; Champion, A.; Sy, M.O. Symbiotic ability of diverse *Frankia* strains on *Casuarina glauca* plants in hydroponic conditions. *Symbiosis* **2016**, *70*, 79–86. [CrossRef]
- 51. Lee, J.T.; Tsai, S.M. The nitrogen-fixing *Frankia* significantly increases growth, uprooting resistance and root tensile strength of *Alnus formosana*. *Afr. J. Biotechnol.* **2018**, *17*, 213–225.
- 52. Ngom, M.; Oshone, R.; Diagne, N.; Cissoko, M.; Svistoonoff, S.; Tisa, L.S.; Laplaze, L.; Sy, M.O.; Champion, A. Tolerance to environmental stress by the nitrogen-fixing actinobacterium *Frankia* and its role in actinorhizal plants adaptation. *Symbiosis* **2016**, *70*, 17–29. [CrossRef]
- 53. Rey, T.; Dumas, B. Plenty is no plague: *Streptomyces* symbiosis with crops. *Trends Plant Sci.* **2017**, *22*, 30–37. [CrossRef]
- Numan, M.; Bashir, S.; Khan, Y.; Mumtaz, R.; Shinwari, Z.K.; Khan, A.L.; Khan, A.; Al-Harrasi, A. Plant growth promoting bacteria as an alternative strategy for salt tolerance in plants: A review. *Microbiol. Res.* 2018, 209, 21–32. [CrossRef]
- 55. Misra, S.; Dixit, V.K.; Khan, M.H.; Kumar Mishra, S.; Dviwedi, G.; Yadav, S.; Lehri, A.; Singh Chauhan, P. Exploitation of agro-climatic environment for selection of 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase producing salt tolerant indigenous plant growth promoting rhizobacteria. *Microbiol. Res.* 2017, 205, 25–34. [CrossRef]
- 56. Patil, N.; Inchanalkar, M.; Desai, D.; Landge, V.; Bhole, B.D. Screening of 1-aminocyclopropane-1-carboxylic acid (ACC) deaminase producing multifunctional plant growth promoting rhizobacteria from onion (*Allium cepa*) rhizosphere. *Int. J. Curr. Microbiol. Appl. Sci.* **2016**, *5*, 118–127. [CrossRef]
- Jogaiah, S.; Kurjogi, M.; Govind, S.R.; Huntrike, S.S.; Basappa, V.A.; Tran, L.S.P. Isolation and evaluation of proteolytic actinomycete isolates as novel inducers of pearl millet downy mildew disease protection. *Sci. Rep. UK* 2016, *6*, 30789. [CrossRef]
- 58. Hassan, M.K.; McInroy, J.A.; Kloepper, J.W. The interactions of rhizodeposits with plant growth-promoting rhizobacteria in the rhizosphere: A review. *Agriculture* **2019**, *9*, 142. [CrossRef]

- Santoyo, G.; Orozco-Mosqueda, M.d.C.; Govindappa, M. Mechanisms of biocontrol and plant growth-promoting activity in soil bacterial species of *Bacillus* and *Pseudomonas*: A review. *Biocontrol. Sci. Technol.* 2012, 22, 855–872. [CrossRef]
- 60. Jiang, Y.; Ran, C.; Chen, L.; Yin, W.; Liu, Y.; Chen, C.; Gao, J. Purification and characterization of a novel antifungal flagellin protein from endophyte *Bacillus methylotrophicus* NJ13 against *Ilyonectria robusta*. *Microorganisms* **2019**, *7*, 605. [CrossRef]
- 61. Maiyappan, S.; Amalraj, E.; Santhosh, A.; Peter, A. Isolation, evaluation and formulation of selected microbial consortia for sustainable agriculture. *J. Biofertil. Biopestic.* **2010**, *2*, 2.
- 62. Rajasekar, S.; Elango, R. Effect of microbial consortium on plant growth and improvement of alkaloid content in *Withania somnifera* (Ashwagandha). *Curr. Bot.* **2011**, *2*, 27–30.
- 63. Ghorbanpour, M.; Hatami, M.; Khavazi, K. Role of plant growth promoting rhizobacteria on antioxidant enzyme activities and tropane alkaloid production of *Hyoscyamus niger* under water deficit stress. *Turkish J. Biol.* **2013**, *37*, 350–360. [CrossRef]
- 64. Maggini, V.; Mengoni, A.; Bogani, P.; Firenzuoli, F.; Fani, R. Promoting model systems of microbiota–medicinal plant interactions. *Trends Plant Sci.* 2020. [CrossRef]
- 65. Mishra, J.; Fatima, T.; Arora, N.K. Role of Secondary Metabolites from Plant Growth-Promoting Rhizobacteria in Combating Salinity Stress. In *Plant Microbiome: Stress Response*; Egamberdieva, D., Ahmad, P., Eds.; Springer: Singapore, 2018; pp. 127–163.
- 66. Kannojia, P.; Choudhary, K.K.; Srivastava, A.K.; Singh, A.K. Chapter Four—PGPR Bioelicitors: Induced Systemic Resistance (isr) and Proteomic Perspective on Biocontrol. In *PGPR Amelioration in Sustainable Agriculture*; Singh, A.K., Kumar, A., Singh, P.K., Eds.; Woodhead Publishing: Cambridge, UK, 2019; pp. 67–84.
- 67. Narendra Babu, A.; Jogaiah, S.; Ito, S.I.; Kestur Nagaraj, A.; Tran, L.S.P. Improvement of growth, fruit weight and early blight disease protection of tomato plants by rhizosphere bacteria is correlated with their beneficial traits and induced biosynthesis of antioxidant peroxidase and polyphenol oxidase. *Plant Sci.* **2015**, *231*, 62–73. [CrossRef] [PubMed]
- 68. Berger, B.; Brock, A.K.; Ruppel, S. Nitrogen supply influences plant growth and transcriptional responses induced by *Enterobacter radicincitans* in *Solanum lycopersicum*. *Plant Soil* **2013**, *370*, 641–652. [CrossRef]
- Zhou, L.; Tang, K.; Guo, S. The plant growth-promoting fungus (PGPF) *Alternaria* sp. A13 markedly enhances *Salvia miltiorrhiza* root growth and active ingredient accumulation under greenhouse and field conditions. *Int. J. Mol. Sci.* 2018, *19*, 270. [CrossRef] [PubMed]
- Baiyee, B.; Ito, S.I.; Sunpapao, A. *Trichoderma asperellum* T1 mediated antifungal activity and induced defense response against leaf spot fungi in lettuce (*Lactuca sativa* L.). *Physiol. Mol. Plant Pathol.* 2019, 106, 96–101. [CrossRef]
- 71. El-Sharkawy, H.H.A.; Abo-El-Wafa, T.S.A.; Ibrahim, S.A.A. Biological control agents improve the productivity and induce the resistance against downy mildew of grapevine. *J. Plant Pathol.* **2018**, *100*, 33–42. [CrossRef]
- 72. El-Sharkawy, H.H.A.; Rashad, Y.M.; Ibrahim, S.A. Biocontrol of stem rust disease of wheat using arbuscular mycorrhizal fungi and *Trichoderma* spp. *Physiol. Mol. Plant Pathol.* **2018**, *103*, 84–91. [CrossRef]
- 73. Jaroszuk-Scisel, J.; Tyskiewicz, R.; Nowak, A.; Ozimek, E.; Majewska, M.; Hanaka, A.; Tyskiewicz, K.; Pawlik, A.; Janusz, G. Phytohormones (auxin, gibberellin) and ACC deaminase in vitro synthesized by the mycoparasitic *Trichoderma* DEMTkZ3A0 strain and changes in the level of auxin and plant resistance markers in wheat seedlings inoculated with this strain conidia. *Int. J. Mol. Sci.* **2019**, *20*, 4923. [CrossRef]
- 74. Pretscher, J.; Fischkal, T.; Branscheidt, S.; Jäger, L.; Kahl, S.; Schlander, M.; Thines, E.; Claus, H. Yeasts from different habitats and their potential as biocontrol agents. *Fermentation* **2018**, *4*, 31. [CrossRef]
- 75. Sun, C.; Lin, M.; Fu, D.; Yang, J.; Huang, Y.; Zheng, X.; Yu, T. Yeast cell wall induces disease resistance against *Penicillium expansum* in pear fruit and the possible mechanisms involved. *Food Chem.* 2018, 241, 301–307. [CrossRef]
- Zhang, S.; Xu, B.; Gan, Y. Seed treatment with *Trichoderma longibrachiatum* T6 promotes wheat seedling growth under NaCl stress through activating the enzymatic and nonenzymatic antioxidant defense systems. *Int. J. Mol. Sci.* 2019, 20, 3729. [CrossRef]
- 77. Abdelrahman, M.; Abdel-Motaal, F.; El-Sayed, M.; Jogaiah, S.; Shigyo, M.; Ito, S.I.; Tran, L.S.P. Dissection of *Trichoderma longibrachiatum*-induced defense in onion (*Allium cepa* L.) against *Fusarium oxysporum* f. sp. cepa by target metabolite profiling. *Plant Sci.* 2016, 246, 128–138. [CrossRef]

- 78. Jogaiah, S.; Abdelrahman, M.; Tran, L.S.P.; Shin-ichi, I. Characterization of rhizosphere fungi that mediate resistance in tomato against bacterial wilt disease. *J. Exp. Bot.* **2013**, *64*, 3829–3842. [CrossRef]
- 79. Jogaiah, S.; Shetty, H.S.; Ito, S.I.; Tran, L.S.P. Enhancement of downy mildew disease resistance in pearl millet by the G_app7 bioactive compound produced by *Ganoderma applanatum*. *Plant Physiol. Biochem.* **2016**, *105*, 109–117. [CrossRef]
- 80. Liu, Z.; Du, S.; Ren, Y.; Liu, Y. Biocontrol ability of killer yeasts (*Saccharomyces cerevisiae*) isolated from wine against *Colletotrichum gloeosporioides* on grape. *J. Basic Microb.* **2018**, *58*, 60–67. [CrossRef]
- 81. Stürmer, S.L.; Bever, J.D.; Morton, J.B. Biogeography of arbuscular mycorrhizal fungi (Glomeromycota): a phylogenetic perspective on species distribution patterns. *Mycorrhiza* **2018**, *28*, 587–603. [CrossRef]
- Spagnoletti, F.; Carmona, M.; Gómez, N.E.T.; Chiocchio, V.; Lavado, R.S. Arbuscular mycorrhiza reduces the negative effects of *M. phaseolina* on soybean plants in arsenic-contaminated soils. *Appl. Soil Ecol.* 2017, 121, 41–47. [CrossRef]
- 83. MacLean, A.M.; Bravo, A.; Harrison, M.J. Plant signaling and metabolic pathways enabling arbuscular mycorrhizal symbiosis. *Plant Cell* **2017**, *29*, 2319–2335. [CrossRef]
- 84. Cely, M.V.T.; de Oliveira, A.G.; de Freitas, V.F.; de Luca, M.B.; Barazetti, A.R.; dos Santos, I.M.O.; Gionco, B.; Garcia, G.V.; Prete, C.E.C.; Andrade, G. Inoculant of arbuscular mycorrhizal fungi (*Rhizophagus clarus*) increase yield of soybean and cotton under field conditions. *Front. Microbiol.* **2016**, *7*, 720. [CrossRef]
- 85. Li, H.; Chen, X.W.; Wong, M.H. Arbuscular mycorrhizal fungi reduced the ratios of inorganic/organic arsenic in rice grains. *Chemosphere* **2016**, *145*, 224–230. [CrossRef]
- Zhang, X.; Wang, L.; Ma, F.; Yang, J.; Su, M. Effects of arbuscular mycorrhizal fungi inoculation on carbon and nitrogen distribution and grain yield and nutritional quality in rice (*Oryza sativa* L.). *J. Sci. Food Agric.* 2017, 97, 2919–2925. [CrossRef]
- 87. Bamberg, S.M.; Ramos, S.J.; Carneiro, M.A.C.; Siqueira, J.O. Effects of selenium (Se) application and arbuscular mycorrhizal (AMF) inoculation on soybean ('*Glycine max*') and forage grass ('*Urochloa decumbens*') development in oxisol. *Aust. J. Crop Sci.* **2019**, *13*, 380. [CrossRef]
- Liu, Z.; Li, Y.; Ma, L.; Wei, H.; Zhang, J.; He, X.; Tian, C. Coordinated regulation of arbuscular mycorrhizal fungi and soybean MAPK pathway genes improved mycorrhizal soybean drought tolerance. *Mol. Plant Microbe Interact.* 2014, 28, 408–419. [CrossRef]
- Jafari, M.; Yari, M.; Ghabooli, M.; Sepehri, M.; Ghasemi, E.; Jonker, A. Inoculation and co-inoculation of alfalfa seedlings with root growth promoting microorganisms (*Piriformospora indica, Glomus intraradices* and *Sinorhizobium meliloti*) affect molecular structures, nutrient profiles and availability of hay for ruminants. *Anim. Nutr.* 2018, 4, 90–99. [CrossRef]
- Gramisci, B.R.; Lutz, M.C.; Lopes, C.A.; Sangorrín, M.P. Enhancing the efficacy of yeast biocontrol agents against postharvest pathogens through nutrient profiling and the use of other additives. *Biol. Control* 2018, 121, 151–158. [CrossRef]
- 91. Chandra, S.; Singh, B.K. *Trichoderma* spp.: As potential bio-control agents (BCAs) against fungal plant pathogens. *Indian J. Life Sci.* **2016**, *5*, 105.
- 92. Elsen, A.; Gervacio, D.; Swennen, R.; De Waele, D. AMF-induced biocontrol against plant parasitic nematodes in *Musa* sp.: A systemic effect. *Mycorrhiza* **2008**, *18*, 251–256. [CrossRef]
- 93. Han, J.; Sun, L.; Dong, X.; Cai, Z.; Sun, X.; Yang, H.; Wang, Y.; Song, W. Characterization of a novel plant growth-promoting bacteria strain *Delftia tsuruhatensis* HR4 both as a diazotroph and a potential biocontrol agent against various plant pathogens. *Syst. Appl. Microbiol.* **2005**, *28*, 66–76. [CrossRef]
- 94. Cabot, C.; Bosch, R.; Martos, S.; Poschenrieder, C.; Perelló, A. Salinity is a prevailing factor for amelioration of wheat blast by biocontrol agents. *Biol. Control.* **2018**, *125*, 81–89. [CrossRef]
- 95. Siahmoshteh, F.; Siciliano, I.; Banani, H.; Hamidi-Esfahani, Z.; Razzaghi-Abyaneh, M.; Gullino, M.L.; Spadaro, D. Efficacy of *Bacillus subtilis* and *Bacillus amyloliquefaciens* in the control of *Aspergillus parasiticus* growth and aflatoxins production on pistachio. *Int. J. Food Microbiol.* **2017**, *254*, 47–53. [CrossRef]
- 96. Xie, K.; Guo, L.; Bai, Y.; Liu, W.; Yan, J.; Bucher, M. Microbiomics and plant health: an interdisciplinary and international workshop on the plant microbiome. *Mol. Plant* **2019**, *12*, 1–3. [CrossRef]
- 97. Wei, Z.; Jousset, A. Plant breeding goes microbial. Trends Plant Sci. 2017, 22, 555–558. [CrossRef]
- 98. Bai, Y.; Müller, D.B.; Srinivas, G.; Garrido-Oter, R.; Potthoff, E.; Rott, M.; Dombrowski, N.; Münch, P.C.; Spaepen, S.; Remus-Emsermann, M.; et al. Functional overlap of the *Arabidopsis* leaf and root microbiota. *Nature* 2015, 528, 364–369. [CrossRef]

- Knee, E.M.; Gong, F.C.; Gao, M.; Teplitski, M.; Jones, A.R.; Foxworthy, A.; Mort, A.J.; Bauer, W.D. Root mucilage from pea and its utilization by rhizosphere bacteria as a sole carbon source. *Mol. Plant Microbe Interact.* 2001, 14, 775–784. [CrossRef]
- 100. Jones, D.L.; Nguyen, C.; Finlay, R.D. Carbon flow in the rhizosphere: carbon trading at the soil–root interface. *Plant Soil* **2009**, *321*, 5–33. [CrossRef]
- 101. Rudrappa, T.; Czymmek, K.J.; Paré, P.W.; Bais, H.P. Root-secreted malic acid recruits beneficial soil bacteria. *Plant Physiol.* **2008**, *148*, 1547. [CrossRef]
- 102. Tian, L.; Shi, S.; Ma, L.; Nasir, F.; Li, X.; Tran, L.S.P.; Tian, C. Co-evolutionary associations between root-associated microbiomes and root transcriptomes in wild and cultivated rice varieties. *Plant Physiol. Biochem.* 2018, 128, 134–141. [CrossRef]
- 103. Ha, S.; Vankova, R.; Yamaguchi-Shinozaki, K.; Shinozaki, K.; Tran, L.S.P. Cytokinins: metabolism and function in plant adaptation to environmental stresses. *Trends Plant Sci.* **2012**, *17*, 172–179. [CrossRef]
- 104. Jalali, J.; Gaudin, P.; Capiaux, H.; Ammar, E.; Lebeau, T. Isolation and screening of indigenous bacteria from phosphogypsum-contaminated soils for their potential in promoting plant growth and trace elements mobilization. *J. Environ. Manag.* 2020, 260, 110063. [CrossRef]
- 105. Goswami, D.; Thakker, J.N.; Dhandhukia, P.C. Portraying mechanics of plant growth promoting rhizobacteria (PGPR): A review. *Cogent Food Agric.* **2016**, *2*, 1127500. [CrossRef]
- French, K.E. Engineering mycorrhizal symbioses to alter plant metabolism and improve crop health. *Front. Microbiol.* 2017, *8*, 1403. [CrossRef]
- 107. Ezawa, T.; Saito, K. How do arbuscular mycorrhizal fungi handle phosphate? New insight into fine-tuning of phosphate metabolism. *New Phytol.* **2018**, *220*, 1116–1121. [CrossRef]
- Gans, J.; Wolinsky, M.; Dunbar, J. Computational improvements reveal great bacterial diversity and high metal toxicity in soil. *Science* 2005, 309, 1387. [CrossRef]
- 109. Berg, G.; Smalla, K. Plant species and soil type cooperatively shape the structure and function of microbial communities in the rhizosphere. *FEMS Microbiol. Ecol.* **2009**, *68*, 1–13. [CrossRef]
- Lundberg, D.S.; Lebeis, S.L.; Paredes, S.H.; Yourstone, S.; Gehring, J.; Malfatti, S.; Tremblay, J.; Engelbrektson, A.; Kunin, V.; Rio, T.G.d.; et al. Defining the core *Arabidopsis thaliana* root microbiome. *Nature* 2012, 488, 86–90. [CrossRef]
- 111. Basha, S.J.; Basavarajappa, R.; Shimalli, G.; Babalad, H. Soil microbial dynamics and enzyme activities as influenced by organic and inorganic nutrient management in vertisols under aerobic rice cultivation. *J. Environ. Biol.* 2017, *38*, 131. [CrossRef]
- 112. Li, R.; Tao, R.; Ling, N.; Chu, G. Chemical, organic and bio-fertilizer management practices effect on soil physicochemical property and antagonistic bacteria abundance of a cotton field: implications for soil biological quality. *Soil Tillage Res.* 2017, 167, 30–38. [CrossRef]
- Wang, X.; Tang, C.; Severi, J.; Butterly, C.R.; Baldock, J.A. Rhizosphere priming effect on soil organic carbon decomposition under plant species differing in soil acidification and root exudation. *New Phytol.* 2016, 211, 864–873. [CrossRef]
- 114. Ma, X.; Zarebanadkouki, M.; Kuzyakov, Y.; Blagodatskaya, E.; Pausch, J.; Razavi, B.S. Spatial patterns of enzyme activities in the rhizosphere: Effects of root hairs and root radius. *Soil Biol. Biochem.* 2018, 118, 69–78. [CrossRef]
- 115. Meena, V.S.; Meena, S.K.; Verma, J.P.; Kumar, A.; Aeron, A.; Mishra, P.K.; Bisht, J.K.; Pattanayak, A.; Naveed, M.; Dotaniya, M.L. Plant beneficial rhizospheric microorganism (PBRM) strategies to improve nutrients use efficiency: A review. *Ecol. Eng.* 2017, 107, 8–32. [CrossRef]
- 116. Zhang, X.; Zhang, R.; Gao, J.; Wang, X.; Fan, F.; Ma, X.; Yin, H.; Zhang, C.; Feng, K.; Deng, Y. Thirty-one years of rice-rice-green manure rotations shape the rhizosphere microbial community and enrich beneficial bacteria. *Soil Biol. Biochem.* 2017, 104, 208–217. [CrossRef]
- 117. Simon, J.; Dannenmann, M.; Pena, R.; Gessler, A.; Rennenberg, H. Nitrogen nutrition of beech forests in a changing climate: Importance of plant-soil-microbe water, carbon, and nitrogen interactions. *Plant Soil* 2017, 418, 89–114. [CrossRef]
- 118. Hestrin, R.; Hammer, E.C.; Mueller, C.W.; Lehmann, J. Synergies between mycorrhizal fungi and soil microbial communities increase plant nitrogen acquisition. *Commun. Biol.* **2019**, *2*, 233. [CrossRef] [PubMed]

- Shi, S.; Tian, L.; Nasir, F.; Bahadur, A.; Batool, A.; Luo, S.; Yang, F.; Wang, Z.; Tian, C. Response of microbial communities and enzyme activities to amendments in saline-alkaline soils. *Appl. Soil Ecol.* 2019, 135, 16–24. [CrossRef]
- 120. Kim, K.H.; Kabir, E.; Jahan, S.A. Exposure to pesticides and the associated human health effects. *Sci. Total Environ.* **2017**, *575*, 525–535. [CrossRef]
- Gomiero, T. Food quality assessment in organic vs. conventional agricultural produce: Findings and issues. *Appl. Soil Ecol.* 2018, 123, 714–728. [CrossRef]
- 122. Tian, L.; Shi, S.; Ji, L.; Nasir, F.; Ma, L.; Tian, C. Effect of the biocontrol bacterium *Bacillus amyloliquefaciens* on the rhizosphere in ginseng plantings. *Int. Microbiol.* **2018**, *21*, 153–162. [CrossRef]
- 123. Siciliano, S.D.; Fortin, N.; Mihoc, A.; Wisse, G.; Labelle, S.; Beaumier, D.; Ouellette, D.; Roy, R.; Whyte, L.G.; Banks, M.K.; et al. Selection of specific endophytic bacterial genotypes by plants in response to soil contamination. *Appl. Environ. Microb.* 2001, 67, 2469–2475. [CrossRef]
- 124. Banfield, C.C.; Pausch, J.; Kuzyakov, Y.; Dippold, M.A. Microbial processing of plant residues in the subsoil –The role of biopores. *Soil Biol. Biochem.* **2018**, 125, 309–318. [CrossRef]
- 125. De León-Lorenzana, A.S.; Delgado-Balbuena, L.; Domínguez-Mendoza, C.A.; Navarro-Noya, Y.E.; Luna-Guido, M.; Dendooven, L. Soil salinity controls relative abundance of specific bacterial groups involved in the decomposition of maize plant residues. *Front. Ecol. Evol.* **2018**, *6*, 51. [CrossRef]
- 126. Lian, T.; Wang, G.; Yu, Z.; Li, Y.; Liu, X.; Jin, J. Carbon input from ¹³C-labelled soybean residues in particulate organic carbon fractions in a Mollisol. *Biol. Fert. Soils* **2016**, *52*, 331–339. [CrossRef]
- 127. Yang, X.; Meng, J.; Lan, Y.; Chen, W.; Yang, T.; Yuan, J.; Liu, S.; Han, J. Effects of maize stover and its biochar on soil CO₂ emissions and labile organic carbon fractions in Northeast China. *Agric. Ecosyst. Environ.* 2017, 240, 24–31. [CrossRef]
- 128. Kantola, I.B.; Masters, M.D.; DeLucia, E.H. Soil particulate organic matter increases under perennial bioenergy crop agriculture. *Soil Biol. Biochem.* 2017, *113*, 184–191. [CrossRef]
- 129. Crews, T.; Rumsey, B. What agriculture can learn from native ecosystems in building soil organic matter: A review. *Sustainability* **2017**, *9*, 578. [CrossRef]
- 130. Cornejo, P.; Meier, S.; García, S.; Ferrol, N.; Durán, P.; Borie, F.; Seguel, A. Contribution of inoculation with arbuscular mycorrhizal fungi to the bioremediation of a copper contaminated soil using *Oenothera picensis*. *J. Soil Sci. Plant Nut.* **2017**, *17*, 14–21.
- 131. Zhou, C.; Li, F.; Xie, Y.; Zhu, L.; Xiao, X.; Ma, Z.; Wang, J. Involvement of abscisic acid in microbe-induced saline-alkaline resistance in plants. *Plant Signal. Behav.* **2017**, *12*, e1367465. [CrossRef]
- Musilova, L.; Ridl, J.; Polivkova, M.; Macek, T.; Uhlik, O. Effects of secondary plant metabolites on microbial populations: changes in community structure and metabolic activity in contaminated environments. *Int. J. Mol. Sci.* 2016, *17*, 1205. [CrossRef]
- 133. Hardoim, P.R.; van Overbeek, L.S.; Elsas, J.D. Properties of bacterial endophytes and their proposed role in plant growth. *Trends Microbiol.* **2008**, *16*, 463–471. [CrossRef]
- 134. Ortega, R.A.; Mahnert, A.; Berg, C.; Müller, H.; Berg, G. The plant is crucial: specific composition and function of the phyllosphere microbiome of indoor ornamentals. *FEMS Microbiol. Ecol.* **2016**, *92*, 173. [CrossRef]
- 135. Liu, D.; Sun, H.; Ma, H. Deciphering microbiome related to rusty roots of *Panax ginseng* and evaluation of antagonists against pathogenic *Ilyonectria*. *Front. Microbiol.* **2019**, *10*, 1350. [CrossRef]
- 136. Stone, B.W.; Weingarten, E.A.; Jackson, C.R. The role of the phyllosphere microbiome in plant health and function. *Annu. Plant Rev. Online* **2018**, *6*, 533–556.
- 137. Vacher, C.; Hampe, A.; Porté, A.J.; Sauer, U.; Compant, S.; Morris, C.E. The phyllosphere: microbial jungle at the plant–climate interface. *Annu. Rev. Ecol. Evol. Syst.* **2016**, *47*, 1–24. [CrossRef]
- 138. Bringel, F.; Couée, I. Pivotal roles of phyllosphere microorganisms at the interface between plant functioning and atmospheric trace gas dynamics. *Front. Microbiol.* **2015**, *6*, 486. [CrossRef]
- AbuQamar, S.; Moustafa, K.; Tran, L.S. Mechanisms and strategies of plant defense against *Botrytis cinerea*. *Crit. Rev. Biotechnol.* 2017, 37, 262–274. [CrossRef]
- 140. Farrar, K.; Bryant, D.; Cope-Selby, N. Understanding and engineering beneficial plant–microbe interactions: plant growth promotion in energy crops. *Plant Biotechnol. J.* **2014**, *12*, 1193–1206. [CrossRef]
- 141. Truyens, S.; Weyens, N.; Cuypers, A.; Vangronsveld, J. Bacterial seed endophytes: genera, vertical transmission and interaction with plants. *Env. Microbiol. Rep.* **2015**, *7*, 40–50. [CrossRef]

- 142. Bacon, C.W.; Hinton, D.M. Bacterial endophytes: The Endophytic Niche, Its Occupants, and Its Utility. In *Plant-Associated Bacteria. Gnanamanickam SS*; Springer: Dordrecht, The Netherlands, 2006; pp. 155–194.
- Zhao, Y.; Xiong, Z.; Wu, G.; Bai, W.; Zhu, Z.; Gao, Y.; Parmar, S.; Sharma, V.K.; Li, H. Fungal endophytic communities of two wild rosa varieties with different powdery mildew susceptibilities. *Front. Microbiol.* 2018, 9, 2462. [CrossRef]
- 144. Borah, M.; Das, S.; Baruah, H.; Boro, R.C.; Barooah, M. Diversity of culturable endophytic bacteria from wild and cultivated rice showed potential plant growth promoting activities. *bioRxiv* **2018**. [CrossRef]
- 145. Murphy, B.R.; Hodkinson, T.R.; Doohan, F.M. Endophytic *Cladosporium* strains from a crop wild relative increase grain yield in barley. *Biol. Environ.* **2018**, *118*, 147–156. [CrossRef]
- 146. Sawers, R.J.H.; Ramirez-Flores, M.R.; Olalde-Portugal, V.; Paszkowski, U. The impact of domestication and crop improvement on arbuscular mycorrhizal symbiosis in cereals: Insights from genetics and genomics. *New Phytol.* 2018, 220, 1135–1140. [CrossRef]
- 147. Majeed, A.; Muhammad, Z.; Ahmad, H. Plant growth promoting bacteria: role in soil improvement, abiotic and biotic stress management of crops. *Plant Cell Rep.* **2018**, *37*, 1599–1609. [CrossRef]
- 148. Martín-Robles, N.; Lehmann, A.; Seco, E.; Aroca, R.; Rillig, M.C.; Milla, R. Impacts of domestication on the arbuscular mycorrhizal symbiosis of 27 crop species. *New Phytol.* **2018**, *218*, 322–334. [CrossRef]
- 149. Chiapusio, G.; Sánchez, A.M.; Reigosa, M.J.; González, L.; Pellissier, F. Do germination indices adequately reflect allelochemical effects on the germination process? *J. Chem. Ecol.* **1997**, *23*, 2445–2453. [CrossRef]
- 150. Vivanco, J.M.; Bais, H.P.; Stermitz, F.R.; Thelen, G.C.; Callaway, R.M. Biogeographical variation in community response to root allelochemistry: novel weapons and exotic invasion. *Ecol. Lett.* 2004, *7*, 285–292. [CrossRef]
- Niro, E.; Marzaioli, R.; De Crescenzo, S.; D'Abrosca, B.; Castaldi, S.; Esposito, A.; Fiorentino, A.; Rutigliano, F.A. Effects of the allelochemical coumarin on plants and soil microbial community. *Soil Biol. Biochem.* 2016, 95, 30–39. [CrossRef]
- 152. Scavo, A.; Abbate, C.; Mauromicale, G. Plant allelochemicals: agronomic, nutritional and ecological relevance in the soil system. *Plant Soil* **2019**, *442*, 23–48. [CrossRef]
- 153. Li, Y.; Wang, B.; Chang, Y.; Yang, Y.; Yao, C.; Huang, X.; Zhang, J.; Cai, Z.; Zhao, J. Reductive soil disinfestation effectively alleviates the replant failure of Sanqi ginseng through allelochemical degradation and pathogen suppression. *Appl. Microbiol. Biot.* **2019**, *103*, 3581–3595. [CrossRef]
- 154. Berg, G. Plant–microbe interactions promoting plant growth and health: perspectives for controlled use of microorganisms in agriculture. *Appl. Microbiol. Biot.* **2009**, *84*, 11–18. [CrossRef]
- 155. Tian, C.; Kasiborski, B.; Koul, R.; Lammers, P.J.; Bucking, H.; Shachar-Hill, Y. Regulation of the nitrogen transfer pathway in the arbuscular mycorrhizal symbiosis: gene characterization and the coordination of expression with nitrogen flux. *Plant Physiol.* **2010**, *153*, 1175–1187. [CrossRef]
- 156. Pérez-de-Luque, A.; Tille, S.; Johnson, I.; Pascual-Pardo, D.; Ton, J.; Cameron, D.D. The interactive effects of arbuscular mycorrhiza and plant growth-promoting rhizobacteria synergistically enhance host plant defences against pathogens. *Sci. Rep. UK* **2017**, *7*, 16409. [CrossRef]
- 157. Senapati, N.; Stratonovitch, P.; Paul, M.J.; Semenov, M.A. Drought tolerance during reproductive development is important for increasing wheat yield potential under climate change in Europe. *J. Exp. Bot.* **2018**, *70*, 2549–2560. [CrossRef]
- 158. Mathur, S.; Tomar, R.S.; Jajoo, A. Arbuscular mycorrhizal fungi (AMF) protects photosynthetic apparatus of wheat under drought stress. *Photosynth. Res.* **2019**, *139*, 227–238. [CrossRef]
- 159. Ruiz-Sánchez, M.; Aroca, R.; Muñoz, Y.; Polón, R.; Ruiz-Lozano, J.M. The arbuscular mycorrhizal symbiosis enhances the photosynthetic efficiency and the antioxidative response of rice plants subjected to drought stress. J. Plant Physiol. 2010, 167, 862–869. [CrossRef]
- Chandrakala, C.; Voleti, S.R.; Bandeppa, S.; Sunil Kumar, N.; Latha, P.C. Silicate solubilization and plant growth promoting potential of *Rhizobium* sp. isolated from rice rhizosphere. *Silicon* 2019, *11*, 2895–2906. [CrossRef]
- 161. Armendariz, A.L.; Talano, M.A.; Olmos Nicotra, M.F.; Escudero, L.; Breser, M.L.; Porporatto, C.; Agostini, E. Impact of double inoculation with *Bradyrhizobium japonicum* E109 and *Azospirillum brasilense* Az39 on soybean plants grown under arsenic stress. *Plant Physiol. Biochem.* 2019, 138, 26–35. [CrossRef]
- 162. Lopez-Baena, F.J.; Ruiz-Sainz, J.E.; Rodriguez-Carvajal, M.A.; Vinardell, J.M. Bacterial molecular signals in the *Sinorhizobium fredii*-soybean symbiosis. *Int. J. Mol. Sci.* **2016**, *17*, 755. [CrossRef]

- 163. Vemulapally, S.; Guerra, T.; Hahn, D. Localization of typical and atypical *Frankia* isolates from *Casuarina* sp. in nodules formed on *Casuarina equisetifolia*. *Plant Soil* **2019**, 435, 385–393. [CrossRef]
- Tudzynski, B. Biosynthesis of gibberellins in *Gibberella fujikuroi*: biomolecular aspects. *Appl. Microbiol. Biot.* 1999, 52, 298–310. [CrossRef]
- Humar, M.; Bokan, M.; Amartey, S.A.; Šentjurc, M.; Kalan, P.; Pohleven, F. Fungal bioremediation of copper, chromium and boron treated wood as studied by electron paramagnetic resonance. *Int. Biodeterior. Biodegrad.* 2004, 53, 25–32. [CrossRef]
- 166. Liu, S.H.; Zeng, G.M.; Niu, Q.Y.; Liu, Y.; Zhou, L.; Jiang, L.H.; Tan, X.F.; Xu, P.; Zhang, C.; Cheng, M. Bioremediation mechanisms of combined pollution of PAHs and heavy metals by bacteria and fungi: A mini review. *Bioresour. Technol.* 2017, 224, 25–33. [CrossRef]
- Sharma, R.; Sharma, K.; Singh, N.; Kumar, A. Rhizosphere biology of aquatic microbes in order to access their bioremediation potential along with different aquatic macrophytes. *Recent Res. Sci. Technol.* 2013, 5, 29–32.
- Luo, S.; Wang, S.; Tian, L.; Shi, S.; Xu, S.; Yang, F.; Li, X.; Wang, Z.; Tian, C. Aggregate-related changes in soil microbial communities under different ameliorant applications in saline-sodic soils. *Geoderma* 2018, 329, 108–117. [CrossRef]
- 169. Huang, W.; Long, C.; Lam, E. Roles of plant-associated microbiota in traditional herbal medicine. *Trends Plant Sci.* **2018**, *23*, 559–562. [CrossRef]
- Liu, Z.; Zheng, X.; Lv, J.; Zhou, X.; Wang, Q.; Wen, X.; Liu, H.; Jiang, J.; Wang, L. Pharmacokinetic synergy from the taxane extract of *Taxus chinensis* improves the bioavailability of paclitaxel. *Phytomedicine* 2015, 22, 573–578. [CrossRef] [PubMed]
- 171. Somjaipeng, S.; Medina, A.; Kwaśna, H.; Ordaz Ortiz, J.; Magan, N. Isolation, identification, and ecology of growth and taxol production by an endophytic strain of *Paraconiothyrium* variabile from English yew trees (*Taxus baccata*). *Fungal Biol. UK* 2015, *119*, 1022–1031. [CrossRef] [PubMed]
- 172. Soliman, S.S.; Greenwood, J.S.; Bombarely, A.; Mueller, L.A.; Tsao, R.; Mosser, D.D.; Raizada, M.N. An endophyte constructs fungicide-containing extracellular barriers for its host plant. *Curr. Biol.* 2015, 25, 2570–2576. [CrossRef] [PubMed]
- 173. Qiao, W.; Ling, F.; Yu, L.; Huang, Y.; Wang, T. Enhancing taxol production in a novel endophytic fungus, *Aspergillus aculeatinus* Tax-6, isolated from *Taxus chinensis* var. *mairei. Fungal Biol. UK* 2017, 121, 1037–1044. [CrossRef]



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