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CKJ REVIEW The metabolomic quest for a biomarker in chronic kidney disease

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ABSTRACT

Chronic kidney disease (CKD) is a growing burden on people and on healthcare for which the diagnostics are niether disease-specific nor indicative of progression. Biomarkers are sought to enable clinicians to offer more appropriate patient-centred treatments, which could come to fruition by using a metabolomics approach. This mini-review highlights the current literature of metabolomics and CKD, and suggests additional factors that need to be considered in this quest for a biomarker, namely the diet and the gut microbiome, for more meaningful advances to be made.

Keywords: biomarkers, chronic kidney disease, diet, metabolomics

THE PROBLEM OF CHRONIC KIDNEY DISEASE

Globally, chronic kidney disease (CKD) has increased by 36.9% between 1990 and 2013 with increases in CKD due to diabetes by 106.5%, hypertension by 29.4% and other causes by 58.8% [1]. Global CKD prevalence is increasing with different rates between countries, ethnicities and sexes, reflecting health inequalities, and even within these categories there are differences with respect to CKD aetiology [1–3].

Although estimated glomerular filtration rate (eGFR), albuminuria and serum creatinine form part of the assessment along with clinical context and data [4, 5], there are limitations with the current diagnostic criteria. Estimation of GFR and creatinine is based on the Chronic Kidney Disease Epidemiology Collaboration creatinine equation, which requires a correction factor for sex and those of African-Caribbean or African background [5] and is dependent on muscle mass; therefore, those who embody extremes of muscle mass such as bodybuilders, amputees and those with sarcopenia or other muscle-wasting disorders may have exaggerated and erroneous results. Kidney biopsies are also used as diagnostic tools but are invasive, and require skilled professionals and resources to undertake [6].

Therefore, it would be beneficial to investigate other diagnostic measures to aid in further understanding CKD inception, progression and prognosis, to offer more suitable treatment options to patients and to advance and improve therapeutics [7]. Indeed, in 2016, the International Society of Nephrology identified key strategic points to enhance kidney-related research, of which diagnostic methods and CKD progression were highlighted [6].

As CKD is a condition of various aetiologies with complex networks of inter- and intra-molecular signalling, studies on CKD could utilize the '-omics' approaches (Figure 1): genomics, transcriptomics, proteomics and metabolomics, which should enable the clinician and researcher to have a better understanding of the interconnecting genetic and molecular networks in CKD by how the disease affects different body systems and responses to stimuli such as diet, medication and the microbiome [8]. This mini-review will focus on contemporary human studies of CKD utilizing a metabolomics approach published between 2016 and 2017. The research strategy for this involved reviewing relevant

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FIGURE 1: Overview of '-omics' approaches.

literature using PubMed, Google Scholar and Plymouth University's Library Primo databases for articles in English with the following key words: 'CKD', 'dialysis', 'metabolomics', 'biomarker' used in various combinations. The bibliographies of identified articles with these key words were searched for additional references. See Table 1 for a summary of included studies.

THE METABOLOMIC APPROACH

For CKD, metabolomics may offer the best '-omics' approach as this involves examining the whole-body system by highlighting changes in metabolites from cellular processes evident in bodily fluids [19, 20] demonstrating the phenotype of the disease. It is through metabolomics that a biomarker, or a panel of biomarkers, may be identified to ameliorate diagnosis and elucidate progression in those with CKD [7]. A new prognostic biomarker in CKD would not only be beneficial in enhancing patient-centred care and treatment management but also in elucidating the mechanisms by which the disease progresses and how effective treatment is by monitoring the rate of change of the identified biomarker(s) [6].

The metabolomics approach has been applied to the study of various kidney diseases [21] but the discovery and implementation into clinical practice of specific disease biomarkers remains elusive. The science of metabolomics has greatly advanced due to the progress in technological developments in recent years, with better instrumentation and the ability to store, analyse and share data with the concomitant development of bioinformatics and computational platforms [20, 22, 23].

For metabolomics to be fruitful, metabolites need to be quantified. Biofluids and tissue samples can be used for metabolomic analysis with technologies such as nuclear magnetic resonance (NMR) spectroscopy and mass spectrometry (MS) coupled with a preparatory chromatographic separation step of the sample such as capillary electrophoresis MS (CE-MS), liquid chromatography MS (LC-MS) or gas-chromatography (GC-MS) (Table 2). NMR is an analytical technique that uses magnetic fields to yield molecular information. MS is a method that measures the mass-to-charge ratio of an intact ion and tandem MS (MS/MS) can be used to measure selected isolated ions that are then fragmented, and the mass-to-charge ratio of each fragment is measured and used for analysis [28]. Both methods require bioinformatic analysis for the data to be interpretable and meaningful. These methods provide information on identification and quantification of metabolites present in the sample. Additional factors such as sample preparation, sample matrix, and carryover effects should be considered when analysing and interpreting the data [24, 29]. For a more comprehensive review and analysis of metabolomic techniques and methodology consult references [24-29].

This progress in technology has enabled the identification of endogenous and exogenous metabolites as potential disease biomarkers, which could place personalized and precision patient-centred medicine within reach [30]. Metabolomics can be used to identify metabolites from a range of samples and for CKD the most pertinent are blood and urine [31], with the dialysate fluid also offering potential benefits.

Blood

As the current diagnostics for CKD are not indicative of disease progression, Rhee et al. [9] investigated progression of CKD within a CKD cohort stratifying by stable and rapid decline based on eGFR slope. This study incorporated a mix of ethnic backgrounds and CKD aetiologies reflecting the phenotype of CKD, and suggested lower levels of threonine, methionine and arginine as potential biomarkers of renal dysfunction by analysing plasma samples processed by LC-MS. In a study by Kimura et al. [10] the authors aimed to identify prognostic biomarkers for CKD progression and mortality in participants with CKD Stages 3-5 over a 4-year period. Plasma samples were processed using CE and LC-MS from which 16 metabolites were identified with MasterHands software. These 16 metabolites were identified as intrinsic to variable metabolic pathways including nucleotides, glycolysis and amino acids, with others unidentified. Medical history was noted including CKD aetiology and presence of comorbidities, and some medications were listed but glycaemic agents were not. Furthermore, no changes in nutritional status, dietary intake or weight were reported; therefore, it is unknown whether the identified metabolites could arise from CKD or be derived from the diet, or gut microbiome as variation has been shown to occur both intra- and interindividually in the blood metabolome largely due to dietary influences [32-34].

Lee et al. [11] attempted to identify prognostic biomarkers from blood serum comparing CKD patients with and without diabetes, versus a healthy control group using NMR spectroscopy analysis. Participants were placed in groups based on eGFR and diabetes diagnosis before enrolment, but the authors did not test participants in this study, which is a limitation as participants may have developed diabetes but have not yet been diagnosed. This study highlighted differences between healthy controls and the CKD groups with increases in

Table 1. Summary of metabolomic studies included in this mini-review

Proposed metabolite biomarkers	Study population group	Metabolomic platform	Biological matrix for metabolomic analysis	Study outcome	Bibliographic reference
Uric acid, glucuronate, 4-hydroxymandelate, 3-methyladipate/pime- late, cytosine and homo- gentisate were higher in cases than in controls; threonine, methionine, phenylalanine and argi- nine were lower in cases than in controls	200 rapidly declining eGFR, and 200 stable eGFR	LC-MS	Plasma	CKD progression	Rhee et al. [9]
Isethionate, saccharate, TMAO, 4-oxopentanoate, cytidine, gluconate, glu- curonate, guanidinosuc- cinate, 2-hydroxyisobutyrate, uridine, 5-oxoproline, pimelate, N-acetylneura- minate, 3-methylhisti- dine, citramalate, nhthalate	112 participants with CKD Stages 3–5 not on dialysis at start of study	CE-MS	Plasma	Composite: predic- tive value for CKD progression to ESRF, requiring RRT, all-cause death	Kimura et al. [10]
TMAO, creatinine, urea, glu- cose, higher in CKD than healthy controls; argi- nine, leucine, valine, glu- tamine, tyrosine, pyruvate, citrate, acetate and formate decreased in CKD compared with healthy group	291 pre-dialysis CKD patients with/with- out type 2 diabetes and 56 healthy controls	NMR	Serum	Progression of CKD	Lee et al. [11]
G-Glycosyltryptophan, pseudouridine, O-sulfo- tyrosine, N-acetylthreo- nine, N-acetylserine, N6-carbamoylthreonyla- denosine, N6-acetyllysine	158 patients with type 1 diabetes, protein- uria and CKD Stage 3	GC-MS and LC-MS	Serum	eGFR decline and progression to ESRF	Niewczas et al. [12]
4-Hydroxyphenylacetate, phenylacetylglutamine, hippurate and prolyl- hydroxyproline	Discovery cohort of 141 CKD patients on dialysis and an in- dependent replica- tion cohort of 180 CKD patients on dialysis	GC/LC-MS/MS	Plasma	Uraemic metabolites and impaired ex- ecutive function	Tamura et al. [13]
Kynurenine and its metabo- lites (quinolinic acid, kynurenic acid, xanthur- enic acid) and indoxyl sulphate	27 CKD patients	LC-MS/MS	Serum	Kidney function, tryptophan me- tabolism, markers for inflammation and oxidative stress, psychologi- cal/cognitive function	Karu et al. [14]
Citrulline, dimethylamine, proline, acetoacetate, alphaketoisovaleric acid, valine, isobutyrate, D-Palmitylcarnitine, histidine and N-methylnicotinamide	15 patients with bi- opsy-proven FSG	NMR	Urine	Pathogenic path- ways and molecu- lar changes in FSG disease progression	Kalantari et al. [15]
Urinary excretion rate of 27 metabolites and plasma	First cohort: 22 non- diabetic CKD Stages	GC-MS	Plasma and urine	Metabolic pathway analysis of CKD	Hallan et al. [16]

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Table 1. Continued

Proposed metabolite biomarkers	Study population group	Metabolomic platform	Biological matrix for metabolomic analysis	Study outcome	Bibliographic reference
concentration of 33 metabolites differed sig- nificantly in CKD patients versus controls. Citric acid cycle was the most significantly affected	3–4 and 10 healthy adults. Second co- hort: 45 non-dia- betic CKD patients and 15 controls. Additional 155 patients from the European Renal cDNA Bank cohort and 31 kidney biop- sies from healthy kidney transplant donors				
Significant differences in concentration of 214 metabolites between healthy control and ESRF patients' pre-dialysis plasma (126 increased and 88 reduced in ESRF group). Pre-dialysis ver- sus post-dialysis showed significant changes in 362 metabolites—including as yet unidentified metabolites	80 ESRF haemodialysis patients and 80 healthy controls	LC-MS	Plasma and dialysate	Metabolic profile of ESRF patients on dialysis	Zhang et al. [17]
TMAO and choline	80 controls and 179 CKD Stages 3–5 patients	LC-MS/MS	Plasma	TMAO, inflamma- tion and mortality in CKD patients	Missailidis et al. [18]

FSG, focal segmental glomerulonephritis; RRT, renal replacement therapy.

Table 2. Platforms for metabolomic analysis with possible advantages and disadvantages [24-27]

Platform	Advantages	Disadvantages	
CE-MS	Small sample volume	Migration time variability	
	High separation efficiency	Poor concentration sensitivity	
	High resolution	Low sample loading capacity	
LC-MS	Detects a large pool of metabolites	Destructive of sample	
	High sensitivity	Time-consuming	
	High resolution	Sample preparation required	
GC-MS	Wide dynamic range	Requires thermal stability	
	High resolution	Destructive of sample	
	High sensitivity	Sample preparation required	
NMR	Minimal sample preparation	Low resolution	
	Non-destructive of the sample	Low sensitivity	
	High reproducibility	Expensive	

trimethylamine-N-oxide (TMAO), creatinine, urea and glucose that correlated with CKD progression. It was also shown that levels of arginine, leucine, valine, glutamine, tyrosine, pyruvate, citrate, acetate and formate decreased in CKD patients compared with the healthy group. However, these metabolites are not specific to kidney disease and may be influenced by a myriad of other factors such as age, diet, nutritional status, medication, nutritional supplements and other diseases not accounted for in this study. Indeed, this study did not exclude those with other CKD-associated conditions, namely hypertension and immune-related, subsequently, the results should be interpreted with caution. In another study on diabetes, Niewczas *et al.* [12] monitored patients with CKD Stage 3 and type 1 diabetes for a median of 11 years. Serum samples were analysed by Metabolon Inc. using GC-MS and LC-MS, and seven metabolites were identified that correlated with CKD progression. This study obtained repeat blood serum for metabolomic analysis, and urine samples for protein and renal function markers, which is a strength when investigating the progression of CKD. However, this study did not report on medication use and may obfuscate the study results because an improvement in medication treatment regimens and i:S

patient adherence to glycaemic, hypertensive and dyslipidaemic agents may slow the decline in kidney function, and hence delay CKD progression [35].

Tamura et al. [13] sought to elucidate the impact of uraemic metabolites on executive function in a cohort of dialvsis patients by using GC/LC-MS/MS in pre-dialysis plasma samples and analysis performed by Metabolon Inc. Four metabolites were associated with impaired executive function in those with CKD: 4-hydroxyphenylacetate, phenylacetylglutamine, hippurate and prolyl-hydroxyproline. However, these metabolites can be derived from the diet and gut microbial metabolism, which this study did not investigate [36-39]. Furthermore, cognitive impairment could result from other confounding factors in this study such as age, frailty, hypertension, incidence of neurological disorders and cardiovascular factors [40-46] rather than the metabolites identified. In a similar study into cognitive decline, Karu et al. [14] identified kynurenine and its metabolites (quinolinic acid, kynurenic acid, xanthurenic acid), and indoxyl sulphate as being greatly elevated in CKD patients, especially in those with cognitive impairment, compared with healthy controls. The proposed mechanism for this is that tryptophan is involved in the synthesis of the indoxyl sulphate (uraemic toxin) via colonic microbes [47, 48] and can affect brain activity through the kynurenine pathway. However, the same confounding factors are attributable to this study as were for Tamura et al. [13], and hypertension was documented in 78% of CKD participants in Karu et al. [14]; therefore, the cognitive decline may be as a result of hypertension or exacerbated by the co-presence of accumulating uraemic toxins and hypertension.

Urine

Metabolomic urinary analysis in CKD could be useful as it is non-invasive, easily obtained and provides a global state of physiological function. However, caution should be employed as there is evidence to suggest that urinary metabolites fluctuate throughout the day suggesting vigilance should be taken when interpreting results from such studies [49, 50]. Kalantari et al. [15] collected urine samples over 24 h from 15 patients with focal segmental glomerulosclerosis to identify 10 metabolites using NMR spectroscopy and ProMetab software, that were deemed to be prognostic when compared with kidney biopsy results. This study implemented a diet on its participants for 24 h prior to collecting urine as a mitigating measure to control for dietary influences on the urinary metabolome. However, urine samples were collected in 2011 but no information is given on how the samples were stored nor when the samples were processed, which could limit the reliability of these results [50, 51]. Hallan et al. [16] used GC-MS and MetaboAnalyst 3.0 software on urinary samples in non-diabetic CKD patients showing decreased excretion of citric acid cycle metabolites corroborated with analysis of kidney biopsies, showing a reduction in gene expression for citric acid cycle enzymes. These findings, however, could be accounted for by considering the nutritional status and dietary intake of these participants, which this study did not do.

Dialysate

The only currently identified study that applied metabolomics to the effect of haemodialysis on the metabolome and dialysate effluent was by Zhang *et al.* [17]. In this study, end-stage renal failure (ESRF) patients receiving dialysis were compared with matched heathy controls with samples collected from blood plasma and dialysate effluent at regular timings during the dialvsis process but on a single occasion. The metabolome in the plasma samples was compared with both groups using ultra performance LC-MS and MetaboAnalyst 3.0 software, which showed that the haemodialysis process not only removed, as expected, uraemic products (TMAO, indoxyl sulphate, p-cresol sulphate, p-cresol glucuronide, uric acid and hippuric acid), fluid and excessive electrolytes, but a plethora of metabolitesmainly amino acids (arginine, glutamine, alanine and phenylalanine) and lipids, which the authors concluded may be the cause of increased mortality within the CKD population. These changes in metabolites were also identified in the dialysate effluent when measured at the corresponding time intervals. However, the authors did not include information on the medical comorbities or medications, CKD aetiology of the disease group, or whether those in this group had any residual kidney function. Although for the control group hypertension, cardiovascular disease and diabetes were exclusion criteria, no indication is given of diabetes prevalence within the ESRF group, which could have implications for interpreting the study's results.

DIET AND THE MICROBIOME—THE MISSING LINKS?

Diet and nutritional status

Changes in amino acid metabolism are widely seen in those with CKD and on dialysis [9, 10, 17, 52, 53] but whether this is due to CKD progression, other diseases or concomitant with poor nutritional status and dietary protein intake remains elusive, compounded by the fact that very few studies that include an assessment of nutritional status or dietary intake. Diet is an important factor that should be assessed as those who display malnutrition and protein-energy malnutrition have worse outcomes and early mortality in CKD and on dialysis [54–56]. Utilizing a subjective global assessment (SGA) tool will enable the clinician and researcher to understand and appreciate whether the metabolites identified result from nutritional status, dietary intake or from disease [57–59].

Consideration should also be made of dietary regimes as these can have influence over the metabolome and microbiome composition [60-62], such as differences between vegans, vegetarians, pescatarians and carnivores. In Wu et al. [63], healthy vegans consumed more carbohydrates, but less protein and fat, than healthy omnivores, resulting in a 25% difference between the identified metabolites of omnivores and vegans of which lipid and amino acid metabolites were significantly elevated in omnivores and the metabolites often associated with CKD hippurate, catechol sulphate and 3-hydroxyhippurate were increased in vegans compared with omnivores. It is, therefore, necessary to account for differences in dietary intake when assessing the metabolites identified in CKD patients as what could have been considered to be a potential biomarker may be derived from or greatly influenced by factors other than kidney disease. Furthermore, it would also be informative to collect multiple samples across time-points for metabolomic analysis of dietary intake to understand how the metabolome changes especially for amino acids [32, 64, 65] in CKD patients. Indeed, current suggested dietary protein requirements for CKD patients are contentious and vary globally from 0.55 g/kg to 1 g/kg [66-71] with greater requirements for those on dialysis, 1.1-1.4 g/kg [58, 72], which may impact on the levels of amino acids and uraemic toxins seen in these metabolomic CKD studies.

Microbiome

Combining the study of diet and the microbiome in CKD studies with metabolomics would enable elucidation of these complex and interwoven relationships, especially for TMAO and uraemic toxins [73, 74]. Phenylacetylglutamine is associated with levels of p-cresyl sulphate and indoxyl sulphate in CKD patients not yet on dialysis and is considered to be a risk factor for cardiovascular disease and mortality [39], but whether it is as a result of gut microbiome dysbiosis or due to impaired renal function is yet to be elucidated. TMAO is derived from the gut microbiota and L-carnitine and choline precursors derived from dietary intake of meat and eggs, and p-cresyl sulphate and sulphate are uraemic toxins derived from the metabolism of amino acids by commensal gut microbiota, consequently greatly influenced by dietary intake [62, 75], and TMAO is implicated in greater mortality amongst those with CKD concomitant with progressing impaired renal function [76] and increased cardiovascular events [77]. Missailidis et al. [18] assessed plasma samples from those with CKD Stages 3-5 from various aetiologies, comorbidities and nutritional status using SGA, but lacked an assessment of dietary intake. TMAO increased as CKD stage progressed and was associated with greater mortality in a 5-year follow-up. However, the nutritional status score also increased with progressive CKD, which may have a more negative impact on mortality than the presence of TMAO. CKD patients had higher TMAO levels than controls, which continued to increase as renal function declined. When a study participant received a kidney transplant, TMAO levels decreased and nutritional status improved. It was demonstrated that CKD patients with the highest TMAO levels had a significantly lower survival rate, which the authors deemed to suggest that high levels of TMAO predicted reduced 5-year survival. Although this study did consider nutritional status by utilizing the SGA tool examining weight loss, anorexia and vomiting, muscle wasting, oedema and loss of fat mass, it did not consider dietary intake or the microbiome, which can both have an impact on TMAO levels [62, 73–75]. Furthermore, it could have collected faecal samples to assess the gut microbiome and its influence on TMAO levels [31]. Stubbs et al. [78] also assessed the impact of TMAO on CKD and demonstrated the beneficial effect of transplant on decreased levels of TMAO compared with pre-transplant, therefore suggesting that increased levels of TMAO are a consequence of decreased renal function and urinary excretion.

It would have been advantageous to assess levels of TMAO in conjunction with an assessment of dietary intake for a more comprehensive investigation of the relationship between TMAO, CKD, diet and the microbiome [79]. Stubbs et al. [78] did not comment on the effect of diet and Missailidis et al. [18] concluded that dietary changes could not explain the normalized levels of TMAO after kidney transplant; however, it is not documented whether the participants receiving the transplant were urinating [80, 81] as this would allow TMAO levels to decrease due to it being excreted in the urine [82]. It remains unclear if TMAO can be used as a biomarker in CKD and cardiovascular dysfunction as it may just be a marker of poor renal clearance or poor nutritional status; therefore, TMAO should be monitored in those with CKD along with an assessment of dietary intake and gut microbiome to further elucidate this mechanism and potential biomarker.

Very few studies have been identified that incorporate the study of the metabolome with dietary and microbiome considerations. Pallister *et al.* [38] identified that metabolites are influenced by microbiome diversity, particularly hippurate, which

was associated with intakes of coffee, fruit and wholegrains; and p-cresol sulphate and phenylacetylglutamine from the putrefaction of undigested dietary proteins by colonic bacteria. Furthermore, Lees et al. [36] suggested that hippurate excretion is associated with co-excretion of metabolic intermediates, especially citrate, succinate and 2-oxoglutarate, and has been associated with a range of conditions besides kidney disease including liver disease, hypertension, diabetes, atherosclerosis and psychiatric disorders, but is also dependent on intestinal microbiota diversity. Diversity and abundance of human microbiome varies widely even among healthy subjects and important factors such as diet need to be considered due to its effect on microbiome composition and metabolism, and wider effects on health status and disease [20, 83-85]. Furthermore, dietary advice given to those with CKD and on dialysis may negatively impact the microbiome of the kidney-gut axis due to reducing the ability to produce beneficial short-chain fatty acids [86, 87] as fruit and vegetable consumption is rationed to prevent electrolyte derangement [57, 86, 88]. Short-chain fatty acids are thought to be implicated in CKD through their deleterious depletion and consequential effect on increasing oxidative stress, fibrosis and the immune response [86, 89]. Utilizing faecal samples to assess the microbiome may offer further insights in the pathological progression of CKD [31] and provide potential dysbiotic targets for treatment of CKD [73].

CONSIDERATIONS FOR FURTHER CKD METABOLOMICS

Single biomarker

As studies incorporate metabolomics into their methodology to identify potential biomarkers, it should also be embedded how to evaluate the clinical usefulness of these biomarkers, to elucidate to what degree they can be used in clinical care, drug development and therapy, and, ultimately, point-of-care testing devices [30, 90]. A single biomarker may be elusive, but a panel of biomarkers based on ratios of identified altered metabolites may offer potential benefits [47] such as glutamate:glutamine, which may indicate nervous system disorders and energy dysmetabolism in uraemic patients, and tryptophan:kynurenine, which may indicate immune responses and increased atherosclerosis risk in uraemic patients [53].

Samples

Results from metabolomic studies are not always reproducible due to differences in patient demographics, samples used, methodology and computational analysis [20, 23, 91-94]. Studies should report on when samples are taken and what presampling checks have been done to limit variability, as well as on the time between sample acquisition and sample processing, because this may increase the possibility of metabolite degradation and yielding false-positive results [50, 93, 95-99]. Each patient is an individual, and each has their own individual metabolic phenotype that is subject to dynamic daily changes due to diet and diet-microbiome interactions [32-34, 49]. It is problematic when studies rely on a single sample from which to extrapolate prognostic markers with hindsight, and studies seeking to investigate prognostic questions should have at least two measurements over the study time period in order to monitor dynamic changes and allow for more meaningful interpretation of the data [90].

The other '-omics'

Current limitations with metabolomics in CKD studies stem from the inability to identify all metabolites in the metabolome and concomitant lack of overlap of metabolite coverage in comparable studies and validation [100]. Corroborating identified metabolites with other physiological functions would make the results more robust and may offer great benefits in ascertaining phenotypical data for an individual patient [16, 101], which will probably become even more useful with advances in technology and the ability to have wider coverage. Genetics coupled with metabolomics could provide valuable information on an individual's metabolic profile, coined as metabotype, which has been demonstrated in genome-wide association studies (GWAS) through the identification of genetic variation and its effect on metabolic functionality [19, 102-105]. GWAS data sets can be utilized by researchers to enrich the study of metabolic dysregulation and be applied to metabolomic studies of CKD, but such studies are currently lacking [106].

Advancement of metabolomics, and the wider '-omics' family, requires a collaborative effort to share and store metabolite data such as the CKDdb database [100]. Once the technology is available, it could conceivably progress to readily available point-of-care devices such as lateral flow devices, dipsticks, breath testing and wearable technology utilizing biosensors and chemometric-based analyses [8] to monitor for disease inception, progression and prognosis, with additional benefits arising from measuring dietary and microbiome influences [107].

CONCLUSIONS

This mini-review has highlighted the current need for better diagnostic and prognostic markers for CKD. Further studies on CKD should utilize the metabolomic approach, but also examine the diets and microbiome of the individual participants with CKD. Multiple samples should be taken over a pre-determined time period and assessed for changes in the metabolome and cross-referenced with the CKD phenotype. Studies should also stratify patients based on their ethnicity, sex and CKD aetiology, and perform further analysis based on nutritional status, dietary intake and on the microbiome particularly to elucidate the interconnectedness of amino acid metabolism, uraemic toxins, dietary factors and the gut microbiome. This approach would strengthen the research output on CKD fostering greater understanding of how metabolites change, and through what influences, so that biomarkers for CKD inception, prognosis and prognostics may be identified.

CONFLICT OF INTEREST STATEMENT

None declared.

REFERENCES

- Abubakar II, Tillmann T, Banerjee A. Global, regional, and national age-sex specific all-cause and cause-specific mortality for 240 causes of death, 1990-2013: a systematic analysis for the Global Burden of Disease Study 2013. Lancet 2015; 385: 117–171
- Hill NR, Fatoba ST, Oke JL *et al*. Global prevalence of chronic kidney disease – a systematic review and meta-analysis. PLoS ONE 2016; 11: e0158765

- Carrero J-J, Hecking M, Ulasi I et al. Chronic kidney disease, gender, and access to care: a global perspective. Semin Nephrol 2017; 37: 296–308
- Kellum JA, Lameire N, Aspelin P et al. Kidney Disease: Improving Global Outcomes (KDIGO) acute kidney injury work group. KDIGO clinical practice guideline for acute kidney injury. Kidney Int Suppl 2013; 2: 1–138
- NICE. Chronic Kidney Disease in Adults: Assessment and Management | Clinical Guideline [CG182] 2015. https://www. nice.org.uk/guidance/cg182 (28 November 2017, date last accessed)
- Levin A, Tonelli M, Bonventre J et al. Global kidney health 2017 and beyond: a roadmap for closing gaps in care, research, and policy. Lancet 2017; 390: 1888–1917
- Hocher B, Adamski J. Metabolomics for clinical use and research in chronic kidney disease. Nat Rev Nephrol 2017; 13: 269
- Trivedi DK, Hollywood KA, Goodacre R. Metabolomics for the masses: the future of metabolomics in a personalized world. New Horiz Transl Med 2017; 3: 294–305
- Rhee EP, Clish CB, Wenger J et al. Metabolomics of chronic kidney disease progression: a case-control analysis in the chronic renal insufficiency cohort study. Am J Nephrol 2016; 43: 366–374
- Kimura T, Yasuda K, Yamamoto R et al. Identification of biomarkers for development of end-stage kidney disease in chronic kidney disease by metabolomic profiling. Sci Rep 2016; 6: 26138
- Lee J, Choi J-Y, Kwon Y-K et al. Changes in serum metabolites with the stage of chronic kidney disease: comparison of diabetes and non-diabetes. Clin Chim Acta 2016; 459: 123–131
- Niewczas MA, Mathew AV, Croall S et al. Circulating modified metabolites and a risk of esrd in patients with type 1 diabetes and chronic kidney disease. Diabetes Care 2017; 40: 383–390
- Tamura MK, Chertow GM, Depner TA et al. Metabolic profiling of impaired cognitive function in patients receiving dialysis. J Am Soc Nephrol 2016; 27: 3780–3787
- Karu N, McKercher C, Nichols DS et al. Tryptophan metabolism, its relation to inflammation and stress markers and association with psychological and cognitive functioning: Tasmanian Chronic Kidney Disease pilot study. BMC Nephrol 2016; 17: 171
- Kalantari S, Nafar M, Samavat S et al. NMR-based metabolomics exploring urinary biomarkers correlated with proteinuria in focal segmental glomerulosclerosis: a pilot study: urine metabolomics based on NMR. Magn Reson Chem 2016; 54: 821–826
- Hallan S, Afkarian M, Zelnick LR et al. Metabolomics and gene expression analysis reveal down-regulation of the citric acid (TCA) cycle in non-diabetic CKD patients. EBioMedicine 2017; 26: 68–77
- Zhang Z-H, Mao J-R, Chen H et al. Removal of uremic retention products by hemodialysis is coupled with indiscriminate loss of vital metabolites. Clin Biochem 2017; 50: 1078–1086
- Missailidis C, Hällqvist J, Qureshi AR et al. Serum trimethylamine-n-oxide is strongly related to renal function and predicts outcome in chronic kidney disease. PLoS ONE 2016; 11: e0141738
- Atzler D, Schwedhelm E, Zeller T. Integrated genomics and metabolomics in nephrology. Nephrol Dial Transplant 2014; 29: 1467–1474

- Beger RD, Dunn W, Schmidt MA et al. Metabolomics enables precision medicine: 'A White Paper, Community Perspective'. Metabolomics 2016; 12: 149
- 21. Weiss RH, Kim K. Metabolomics in the study of kidney diseases. Nat Rev Nephrol 2011; 8: 22–33
- Cisek K, Krochmal M, Klein J et al. The application of multiomics and systems biology to identify therapeutic targets in chronic kidney disease. Nephrol Dial Transplant 2016; 31: 2003–2011
- Meier R, Ruttkies C, Treutler H et al. Bioinformatics can boost metabolomics research. J Biotechnol 2017; 261: 137–141
- Aretz I, Meierhofer D. Advantages and pitfalls of mass spectrometry based metabolome profiling in systems biology. Int J Mol Sci 2016; 17: 632
- Lindon JC. Overview of NMR-based metabonomics. In: Lindon JC, Tranter GE, Koppenaal DW (eds). Encyclopedia of Spectroscopy and Spectrometry, 3rd edn. Oxford: Academic Press, 2017, 517–526
- Nassar AF, Wu T, Nassar SF et al. UPLC–MS for metabolomics: a giant step forward in support of pharmaceutical research. Drug Discov Today 2017; 22: 463–470
- Ramautar R, Somsen GW, de Jong GJ. CE–MS for metabolomics: developments and applications in the period 2014– 2016. Electrophoresis 2017; 38: 190–202
- Patti GJ, Yanes O, Siuzdak G. Metabolomics: the apogee of the omics trilogy: Innovation. Nat Rev Mol Cell Biol 2012; 13: 263–269
- Gowda GAN, Djukovic D. Overview of mass spectrometrybased metabolomics: opportunities and challenges. In: Raftery D (ed). Mass Spectrometry in Metabolomics. New York, NY: Springer, 2014, 3–12
- Wishart DS. Emerging applications of metabolomics in drug discovery and precision medicine. Nat Rev Drug Discov 2016; 15: 473
- Barrios C, Spector TD, Menni C. Blood, urine and faecal metabolite profiles in the study of adult renal disease. Arch Biochem Biophys 2016; 589: 81–92
- 32. Kim K, Mall C, Taylor SL et al. Mealtime, temporal, and daily variability of the human urinary and plasma metabolomes in a tightly controlled environment. PLoS ONE 2014; 9: e86223
- Dunn WB, Lin W, Broadhurst D et al. Molecular phenotyping of a UK population: defining the human serum metabolome. Metabolomics 2015; 11: 9–26
- Chaleckis R, Murakami I, Takada J et al. Individual variability in human blood metabolites identifies age-related differences. Proc Natl Acad Sci USA 2016; 113: 4252–4259
- Aitken GR, Roderick PJ, Fraser S et al. Change in prevalence of chronic kidney disease in England over time: comparison of nationally representative cross-sectional surveys from 2003 to 2010. BMJ Open 2014; 4: e005480. doi: 10.1136/bmjopen-2014-005480
- Lees HJ, Swann JR, Wilson ID et al. Hippurate: the natural history of a mammalian-microbial cometabolite. J Proteome Res 2013; 12: 1527–1546
- Vázquez-Fresno R, Llorach R, Perera A et al. Clinical phenotype clustering in cardiovascular risk patients for the identification of responsive metabotypes after red wine polyphenol intake. J Nutr Biochem 2016; 28: 114–120
- Pallister T, Jackson MA, Martin TC et al. Hippurate as a metabolomic marker of gut microbiome diversity: modulation by diet and relationship to metabolic syndrome. Sci Rep 2017; 7: 13670. doi: 10.1038/s41598-017-13722-4

- Poesen R, Claes K, Evenepoel P et al. Microbiota-derived phenylacetylglutamine associates with overall mortality and cardiovascular disease in patients with CKD. J Am Soc Nephrol 2016; 27: 3479–3487
- Tamura M, Covinsky KE, Chertow GM et al. Functional status of elderly adults before and after initiation of dialysis. N Engl J Med 2009; 361: 1539–1547
- 41. Tamura MK, Meyer JB, Saxena AB et al. Prevalence and significance of stroke symptoms among patients receiving maintenance dialysis. *Neurology* 2012; 79: 981–987
- 42. Bugnicourt J-M, Godefroy O, Chillon J-M et al. Cognitive disorders and dementia in CKD: the neglected kidney-brain axis. J Am Soc Nephrol 2013; 24: 353–363
- McAdams-DeMarco MA, Tan J, Salter ML. Frailty and cognitive function in incident hemodialysis patients. Clin J Am Soc Nephrol 2015; 10: 2181–2189
- 44. Kallenberg MH, Kleinveld HA, Dekker FW. Functional and cognitive impairment, frailty, and adverse health outcomes in older patients reaching esrd–a systematic review. Clin J Am Soc Nephrol 2016; 11: 1624–1639
- Iyasere O, Okai D, Brown E. Cognitive function and advanced kidney disease: longitudinal trends and impact on decision-making. Clin Kidney J 2017; 10: 89–94
- 46. Shirazian S, Grant CD, Aina O. Depression in chronic kidney disease and end-stage renal disease: similarities and differences in diagnosis, epidemiology, and management. Kidney Int Rep 2017; 2: 94–107
- 47. Breit M, Weinberger KM. Metabolic biomarkers for chronic kidney disease. Appl Metabolomics 2016; 589: 62–80
- Leong S, Sirich T. Indoxyl sulfate—review of toxicity and therapeutic strategies. Toxins 2016; 8: 358
- Heinzmann SS, Merrifield CA, Rezzi S et al. Stability and robustness of human metabolic phenotypes in response to sequential food challenges. J Proteome Res 2012; 11: 643–655
- Budde K, Gök Ö-N, Pietzner M et al. Quality assurance in the pre-analytical phase of human urine samples by 1H NMR spectroscopy. Arch Biochem Biophys 2016; 589: 10–17
- Capati A, Ijare OB, Bezabeh T. Diagnostic applications of nuclear magnetic resonance–based urinary metabolomics. Magn Reson Insights 2017; 10: 1178623X17694346. https://www.ncbi.nlm.nih.gov/pmc/articles/PMC5428226/ (13 December 2017)
- 52. Duranton F, Lundin U, Gayrard N et al. Plasma and urinary amino acid metabolomic profiling in patients with different levels of kidney function. Clin J Am Soc Nephrol 2014; 9: 37–45
- 53. Liu S, Wang L, Hu C et al. Plasma metabolomics profiling of maintenance hemodialysis based on capillary electrophoresis—time of flight mass spectrometry. Sci Rep 2017; 7: 8150. doi: 10.1038/s41598-017-08327-w
- Magalhães LP, dos Reis LM, Graciolli FG et al. Predictive factors of one-year mortality in a cohort of patients undergoing urgent-start hemodialysis. PLoS ONE 2017; 12: e0167895
- Sabatino A, Regolisti G, Karupaiah T et al. Protein-energy wasting and nutritional supplementation in patients with end-stage renal disease on hemodialysis. Clin Nutr 2017; 36: 663–671
- 56. Zha Y, Qian Q. Protein nutrition and malnutrition in CKD and ESRD. Nutrients 2017; 9: 208
- Wright M, Jones C. Renal association clinical practice guideline on nutrition in CKD. Nephron Clin Pract 2011; 118: c153-c164
- Naylor HL, Jackson H, Walker GH et al. British Dietetic Association evidence-based guidelines for the protein requirements of adults undergoing maintenance

haemodialysis or peritoneal dialysis. J Hum Nutr Diet 2013; 26: 315–328

- 59. Santin F, Rodrigues J, Brito FB et al. Performance of subjective global assessment and malnutrition inflammation score for monitoring the nutritional status of older adults on hemodialysis. Clin Nutr 2018; 37: 604–611
- 60. Poesen R, Mutsaers HAM, Windey K et al. The influence of dietary protein intake on mammalian tryptophan and phenolic metabolites. PLoS ONE 2015; 10: e0140820
- 61. Schmidt JA, Rinaldi S, Scalbert A *et al.* Plasma concentrations and intakes of amino acids in male meat-eaters, fisheaters, vegetarians and vegans: a cross-sectional analysis in the EPIC-Oxford cohort. *Eur J Clin Nutr* 2016; 70: 306–312
- Cheung W, Keski-Rahkonen P, Assi N et al. A metabolomic study of biomarkers of meat and fish intake. Am J Clin Nutr 2017; 105: 600–608
- Wu GD, Compher C, Chen EZ et al. Comparative metabolomics in vegans and omnivores reveal constraints on dietdependent gut microbiota metabolite production. Gut 2016; 65: 63–72
- Andersen M-BS, Rinnan Å, Manach C et al. Untargeted metabolomics as a screening tool for estimating compliance to a dietary pattern. J Proteome Res 2014; 13: 1405–1418
- Scalbert A, Brennan L, Manach C et al. The food metabolome: a window over dietary exposure. Am J Clin Nutr 2014; 99: 1286–1308
- Fouque D, Laville M. Low protein diets for chronic kidney disease in non diabetic adults. In: The Cochrane Collaboration (ed). Cochrane Database of Systematic Reviews. Chichester: John Wiley & Sons, Ltd, 2009; doi: 10.1002/14651858.CD001892.pub3
- Kovesdy CP, Kopple JD, Kalantar-Zadeh K. Management of protein-energy wasting in non-dialysis-dependent chronic kidney disease: reconciling low protein intake with nutritional therapy. Am J Clin Nutr 2013; 97: 1163–1177
- Piccoli GB, Vigotti FN, Leone F et al. Low-protein diets in CKD: how can we achieve them? A narrative, pragmatic review. Clin Kidney J 2015; 8: 61–70
- Riccio E, Di Nuzzi A, Pisani A. Nutritional treatment in chronic kidney disease: the concept of nephroprotection. *Clin Exp Nephrol* 2015; 19: 161–167
- Bellizzi V, Cupisti A, Locatelli F et al. Low-protein diets for chronic kidney disease patients: the Italian experience. BMC Nephrol 2016; 17:77
- Di Iorio B, Di Micco L, Marzocco S et al. Very low-protein diet (VLPD) reduces metabolic acidosis in subjects with chronic kidney disease: the "nutritional light signal" of the renal acid load. Nutrients 2017; 9: 69
- Cano N, Fiaccadori E, Tesinsky P et al. ESPEN guidelines on enteral nutrition: adult renal failure. Clin Nutr 2006; 25: 295–310
- Zhang LS, Davies SS. Microbial metabolism of dietary components to bioactive metabolites: opportunities for new therapeutic interventions. *Genome Med* 2016; 8: 46. http:// genomemedicine.biomedcentral.com/articles/10.1186/ s13073-016-0296-x (28 November 2017, date last accessed)
- 74. Fernandez-Prado RE, Perez-Gomez M, Gracia-Iguacel C et al. Nutrients turned into toxins: microbiota modulation of nutrient properties in chronic kidney disease. Nutrients 2017; 9:489
- 75. Schmedes M, Aadland EK, Sundekilde UK et al. Lean-seafood intake decreases urinary markers of mitochondrial lipid and energy metabolism in healthy subjects:

Metabolomics results from a randomized crossover intervention study. Mol Nutr Food Res 2016; 60: 1661–1672

- 76. Gruppen EG, Garcia E, Connelly MA et al. TMAO is associated with mortality: impact of modestly impaired renal function. Sci Rep 2017; 7: 13781. doi: 10.1038/ s41598-017-13739-9
- 77. Kim RB, Morse BL, Djurdjev O et al. Advanced chronic kidney disease populations have elevated trimethylamine N-oxide levels associated with increased cardiovascular events. *Kidney Int* 2016; 89: 1144–1152
- Stubbs JR, House JA, Ocque AJ et al. Serum trimethylaminen-oxide is elevated in ckd and correlates with coronary atherosclerosis burden. J Am Soc Nephrol 2016; 27: 305–313
- Aron-Wisnewsky J, Clément K. The gut microbiome, diet, and links to cardiometabolic and chronic disorders. Nat Rev Nephrol 2015; 12: 169
- Khosroshahi HT, Oskui R, Shoja MM et al. Time-dependent variations in urine output after renal transplantation. Transplant Proc 2007; 39: 932–933
- Lai Q, Pretagostini R, Poli L et al. Early urine output predicts graft survival after kidney transplantation. Transplant Proc 2010; 42: 1090–1092
- Moraes C, Fouque D, Amaral ACF et al. Trimethylamine n-oxide from gut microbiota in chronic kidney disease patients: focus on diet. J Ren Nutr 2015; 25: 459–465
- Huttenhower C, Gevers D, Knight R et al. Structure, function and diversity of the healthy human microbiome. Nature 2012; 486: 207–214
- Hall AB, Tolonen AC, Xavier RJ. Human genetic variation and the gut microbiome in disease. Nat Rev Genet 2017; 18: 690–699
- Mendes-Soares H, Chia N. Community metabolic modelling approaches to understanding the gut microbiome: bridging biochemistry and ecology. Free Radic Biol Med 2017; 105: 102–109
- 86. Felizardo RJF, Castoldi A, Andrade-Oliveira V et al. The microbiota and chronic kidney diseases: a double-edged sword. Clin Transl Immunol 2016; 5: e86
- 87. Ríos-Covián D, Ruas-Madiedo P, Margolles A *et al*. Intestinal short chain fatty acids and their link with diet and human health. Front Microbiol 2016; 7: 185
- Mitch WE, Remuzzi G. Diets for patients with chronic kidney disease, should we reconsider? BMC Nephrol 2016; 17: 80. doi: 10.1186/s12882-016-0283-x
- Li L, Ma L, Fu P. Gut microbiota–derived short-chain fatty acids and kidney diseases. Drug Des Devel Ther 2017; 11: 3531–3542
- Ravani P, Parfrey PS, Dicks E et al. Clinical research of kidney diseases II: problems of study design. Nephrol Dial Transplant 2007; 22: 2785–2794
- 91. Everett JR. A new paradigm for known metabolite identification in metabonomics/metabolomics: metabolite identification efficiency. *Comput Struct Biotechnol J* 2015; 13: 131–144
- 92. Darshi M, Van Espen B, Sharma K. Metabolomics in diabetic kidney disease: unraveling the biochemistry of a silent killer. *Am J Nephrol* 2016; 44: 92–103
- 93. Emwas A-H, Roy R, McKay RT et al. Recommendations and standardization of biomarker quantification using nmrbased metabolomics with particular focus on urinary analysis. J Proteome Res 2016; 15: 360–373
- Hajduk J, Matysiak J, Kokot ZJ. Challenges in biomarker discovery with MALDI-TOF MS. Clin Chim Acta 2016; 458: 84–98

- CLINICAL KIDNEY JOURNAL
- Townsend MK, Clish CB, Kraft P. Reproducibility of metabolomic profiles among men and women in 2 large cohort studies. Clin Chem 2013; 59: 1657–1667
- Breier M, Wahl S, Prehn C et al. Targeted metabolomics identifies reliable and stable metabolites in human serum and plasma samples. PLoS ONE 2014; 9: e89728
- 97. Carayol M, Licaj I, Achaintre D et al. Reliability of serum metabolites over a two-year period: a targeted metabolomic approach in fasting and non-fasting samples from EPIC. PLoS ONE 2015; 10: e0135437
- Yin P, Lehmann R, Xu G. Effects of pre-analytical processes on blood samples used in metabolomics studies. Anal Bioanal Chem 2015; 407: 4879–4892
- Jobard E, Trédan O, Postoly D et al. A systematic evaluation of blood serum and plasma pre-analytics for metabolomics cohort studies. Int J Mol Sci 2016; 17: 2035
- 100. Fernandes M, Husi H. Establishment of a integrative multiomics expression database CKDdb in the context of chronic kidney disease (CKD). Sci Rep 2017; 7: 40367
- 101. Chen Dan-Qian, Cao Gang, Chen Hua et al. Gene and protein expressions and metabolomics exhibit activated redox signalling and wnt/β-catenin pathway are associated with

metabolite dysfunction in patients with chronic kidney disease. *Redox* Biol 2017; 12: 505–521

- Illig T, Gieger C, Zhai G et al. A genome-wide perspective of genetic variation in human metabolism. Nat Genet 2010; 42: 137–141
- Suhre K, Shin S-Y, Petersen A-K Human metabolic individuality in biomedical and pharmaceutical research. Nature 2011; 477: 54–60
- 104. Kastenmüller G, Raffler J, Gieger C et al. Genetics of human metabolism: an update. *Hum Mol Genet* 2015; 24: R93–R101
- 105. Raffler J, Friedrich N, Arnold M et al. Genome-wide association study with targeted and non-targeted NMR metabolomics identifies 15 novel loci of urinary human metabolic individuality. PLoS Genet 2015; 11: e1005487
- 106. Köttgen A, Raffler J, Sekula P et al. Genome-wide association studies of metabolite concentrations (mGWAS): relevance for nephrology. Semin Nephrol 2018; 38: 151–174
- 107. Holen T, Norheim F, Gundersen TE et al. Biomarkers for nutrient intake with focus on alternative sampling techniques. Genes Nutr 2016; 11. http://genesandnutrition. biomedcentral.com/articles/10.1186/s12263-016-0527-1 (28 November 2017, date last accessed)