## Check for updates

## RESEARCH HIGHLIGHT OPEN

## Supramolecular attack particle: the way cytotoxic T lymphocytes kill target cells

Jiaqi Liu<sup>1</sup>, Yanqi Ye<sup>2</sup> and Lulu Cai 100

Signal Transduction and Targeted Therapy (2020)5:210

; https://doi.org/10.1038/s41392-020-00319-z

A very recent study by Bálint et al. published in *Science* deciphered in detail how cytotoxic T lymphocytes (CTLs) kill target cells. They found that perforin and pellet enzyme—the cytotoxic substances in CTLs—combined and assembled into supramolecular attack particles (SMAPs). The SMAPs were subsequently released toward the cell membrane of the target cell. The shell of SMAP was rich in glycoproteins that enabled its stability to maintain the cell-killing activity for hours, in which the thrombospondin-1 (TSP-1) was critical (Fig. 1).

There are many types of T cells, among which the CTLs play a major role in destroying virus-infected cells and cancerous cells.<sup>2</sup> The killing mechanism mediated by CTLs has been studied extensively before and there are two main mechanisms. One is degranulation induced upon the target cell recognition. The CTL identifies the target cell, forms an immunological synapse (IS) and releases perforin (Prf1) and soluble granzymes (Gzmb) from secretory lysosomes (SLs). Prf1 creates holes in the membrane of target cells through which Gzmb enters to initiate various apoptosis pathways.<sup>3,4</sup> Tamzalit et al. found that even in the absence of perforin, the CTL could initiate necrosis and kill target cells by its mechanical movement.<sup>5</sup> The other mechanism is by activating the self-apoptosis process of the cells. CTLs can recognize the death receptor Fas (FasL) expressed on target cells to induce programmed cell death. Despite the ongoing studies, it remains unclear on the details of killing mechanism and its relation to the secretion of cytotoxic molecules Prf1

To track how perforin and granzyme work when CTLs kill the targets, the authors labeled granular enzymes in CTLs and glycoproteins in SLs with different fluorescein in this work. After in contact with the CTLs, the target cells showed intense double-positive puncta on the surface. These labeled multiprotein structures were defined as SMAPs. With a supported lipid bilayers (SLB) to simulate the target cell, the authors investigated the kinetics of SMAPs release. The SLs in the CTLs rapidly accumulated at the IS after the CTLs were incubated on the SLB. It was rapidly followed by the appearance of Gzmb puncta in the SLB and the attachment of SMAPs to the surface of the simulated target cell.

After CTL removal, Prf1<sup>+</sup> and Gzmb<sup>+</sup> SMAPs were found to remain attached on the SLB. The authors then tested the stability of SMAPs after being released. After 20 minutes of CTL removal, the SMAPs still existed in the IS and maintained the activity of killing the target cell. The authors further investigated the SMAPs captured on SLB by mass spectrometry. More than 285 proteins were present in SMAPs, including peptides from Prf1 and Gzmb. This result was also confirmed by SDS-PAGE and immunoblotting. Among these proteins, TSP-1 caught the authors' attention due to its signature Ca<sup>2+</sup> binding repeats, which corresponded to the well-established Ca<sup>2+</sup> dependent steps in CTL mediated killing. After knocking out TSP-1 by 60%, the CTLs killing efficacy reduced by 30%. These results indicated that the TSP-1 component in SMAP played an important role in the process of CTL killing target cells.

Furthermore, they investigated SMAPs from a microscopic perspective. Each IS contained about 27 SMAPs with a diameter of about 120 nm. TSP-1 was found to be co-localized with Prf1 and Gzmb, either in the secreted SMAPs or in the CTLs. It was likely that molecules such as Prf1 and Gzmb were assembled into SMAPs and waited to be released in the secretory lysosome of CTLs. The distribution of TSP-1 in SMAP was also consistent with the glycoprotein shell, which turned out to be a component of the SMAP glycoprotein shell. All these results indicated the specific construction of SMAP—a mixed particle with a diameter of about 120 nm consisting of a dense shell with TSP-1, and a core structure containing Prf1, Gzmb, and other cytotoxic substances.

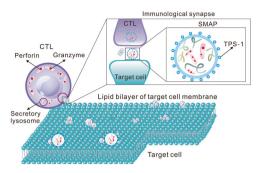
In summary, the study by Bálint and coworkers innovatively illustrated how CTLs attack the target cells by secreting SMAPs containing TSP-1 and deadly chemicals such as Prf1 and Gzmb. They revealed the details and mechanisms that CTLs kill target cells, further enhancing our understanding of CTLs. Besides, SMAPs contain many immune regulatory factors that may regulate the immunity in the tumor microenvironment. The key protein TSP-1 has a binding site for "don't eat me" signal CD47 to myeloid cells, so that any cell escaped from SMAPs may be culled by phagocytosis. All of these findings provide implications for developing new ideas for cancer immunotherapy.

Received: 30 June 2020 Revised: 18 August 2020 Accepted: 20 August 2020

Published online: 21 September 2020

© The Author(s) 2020 SPRINGER NATURE

<sup>&</sup>lt;sup>1</sup>Personalized Drug Therapy Key Laboratory of Sichuan Province, Department of Pharmacy, Sichuan Provincial People's Hospital, University of Electronic Science and Technology of China, Chengdu 610072, China and <sup>2</sup>Zenomics Inc., California NanoSystems Institute, University of California, Los Angeles, CA 90095, USA Correspondence: Lulu Cai (cailulu@med.uestc.edu.cn)



**Fig. 1** The cytotoxic substances in CTL, perforin, and granzyme, assembled into supramolecular attack particles (SMAPs), which were released toward and remained attached to the target cell membrane after the CTL removal. SMAPs, ~120 nm in diameter, have a dense shell that includes TSP-1 and a core of perforin, granzymes, and other cytotoxic proteins

## **REFERENCES**

- Bálint, Š. et al. Supramolecular attack particles are autonomous killing entities released from cytotoxic T cells. Science 368, 897–901 (2020).
- Jerome, T. & Lieberman, J. Perforin: a key pore-forming protein for immune control of viruses and cancer. Sub-Cell. Biochem. 80, 197–220 (2014).

- 3. Halle, S., Halle, O. & Förster, R. Mechanisms and dynamics of T cell-mediated cytotoxicity in vivo. *Trends Immunol.* **38**, 432–443 (2017).
- Jamie, A. et al. Perforin forms transient pores on the target cell plasma membrane to facilitate rapid access of granzymes during killer cell attack. *Blood* 121, 2659–2668 (2013).
- Fella, T. et al. Interfacial actin protrusions mechanically enhance killing by cytotoxic T cells. Sci. Immunol. 4, eaav5445 (2019).

Open Access This article is licensed under a Creative Commons Attribution 4.0 International License, which permits use, sharing, adaptation, distribution and reproduction in any medium or format, as long as you give appropriate credit to the original author(s) and the source, provide a link to the Creative Commons license, and indicate if changes were made. The images or other third party material in this article are included in the article's Creative Commons license, unless indicated otherwise in a credit line to the material. If material is not included in the article's Creative Commons license and your intended use is not permitted by statutory regulation or exceeds the permitted use, you will need to obtain permission directly from the copyright holder. To view a copy of this license, visit http://creativecommons.org/licenses/by/4.0/.

© The Author(s) 2020