

The evaluation of the oxidative stress for patients receiving neoadjuvant chemoradiotherapy for locally advanced rectal cancer

Serbanescu GL^{* **}, Gruia MI^{***}, Bara M^{* **}, Anghel RM^{* **}

^{*}Prof. Dr. Al.Trestioreanu" Institute of Oncology, Bucharest, Romania

^{***}"Carol Davila" University of Medicine and Pharmacy, Bucharest, Romania

^{***}Research Department, "Prof. Dr. Al. Trestioreanu" Institute of Oncology, Bucharest, Romania

Correspondence to: Luiza Serbanescu, MD, PhD student, Assistant Professor,
Clinical Department No. 8, "Carol Davila" University of Medicine and Pharmacy, Bucharest,
"Prof. Dr. Al. Trestioreanu" Institute of Oncology
252 Fundeni Street, Code: 22328, Bucharest, Romania,
Mobile phone: +40723 666 279, E-mail: luizaserbanescu@yahoo.com

Received: September 29th, 2016 – Accepted: December 17th, 2016

Abstract

Hypothesis: Nowadays, rectal cancer is an important healthcare challenge that affects many thousands of people each year worldwide, being diagnosed especially after the age of 50 years.

Objective: This study attempted to evaluate the oxidative stress in patients with rectal cancer.

Methods and results: 30 patients from the "Prof. Dr. Al. Trestioreanu" Institute of Oncology in Bucharest were treated with neoadjuvant radiochemotherapy during 2014 and 2016 and were included in the clinical study. Blood samples were obtained in dynamics during the treatment. From the blood samples, the serum was separated and used to identify the biochemical oxidative stress parameters.

Results: Regarding the determination of lipid peroxides, albumin thiols, the cuprum oxidase activity of ceruloplasmin, the values registered in the dynamic of the treatment highlighted their increase to a maximum at the treatment's endpoint due to an important oxidative stress. Regarding the serum values for total antioxidants, the results pointed out the activation of the natural protection systems, which in time were overwhelmed, due to the installed oxidative stress.

Conclusion: Part of the cytotoxic effect of radiotherapy was due to the production of oxidative stress. The cell was constantly exposed to the cytotoxic action of the reactive oxygen species. The obtained results indicated the dual relation to which the tumoral cell exposed itself and the installed oxidative stress, respectively, the oxidative stress being a cause or a consequence of the malign transformation.

Abbreviations

CT = computed tomography, MRI = magnetic resonance imaging, ESMO = European Society for Medical Oncology, ECOG = performance status scale

Keywords: rectal cancer, radiotherapy, oxidative stress

Introduction

In terms of incidence, colorectal cancer is the third most common cancer in men (10.0% of the total) and the second in women (9.2% of the total) worldwide, with an estimated 1,36 million new cases in both sexes in 2012 [1]. 694,000 deaths in both sexes from colorectal cancer were estimated worldwide in 2012, accounting for 8.5% of the total deaths caused by cancer [1]. In Europe, the incidence of rectal cancer represents 35% of all colorectal cancers (15-25/100000) [2]. In Romania, colorectal cancer is the second malignant cause of death after lung cancer [1].

Surgical intervention is the mainstay of the treatment of rectal cancers and, in 60% of the earliest stages, provides cure [3,4]. Associated to surgery, pelvic radiation represents a standard procedure in the management of locally advanced rectal cancers [2,5]. In combination with 5-FU based

chemotherapy, radiotherapy increases the sphincter preservation rate in the neoadjuvant setting and decreases local relapse rates [6].

The antineoplastic effect of radiotherapy is due to the production of oxidative stress and can result through two mechanisms: it leads to the damaging of the cellular macromolecules (lipids, proteins, DNA) and indirectly by determining abnormal signaling at cell level [7].

Materials and Methods

30 patients who were treated in "Prof. Dr. Al. Trestioreanu" Institute of Oncology in Bucharest were

enrolled in this study between January 2014 and September 2016.

The inclusion criteria were: age over 18 years, histopathological confirmation of rectal cancer, ECOG ≤ 2 performance status, no distant metastasis, no prior pelvic radiation, informed consent signed by all the patients and approved by the Ethics Committee of the Institute of Oncology. Pre-therapeutic evaluation consisted of an initial physical and rectal examination, complete blood count, liver and renal function assessment, CEA level, chest X-ray/ CT and abdominal and pelvic CT/ MRI.

The treatment scheme consisted in concomitant radiochemotherapy due to a locally advanced tumor (T3 or T4) or lymph node involvement suspicion (N+), according to ESMO international guidelines. All the patients underwent external beam radiation therapy up to a total dose of 50-54 Gy, in daily 180-200 cGy fractions, 5 days per week. Chemotherapy schedule was based on Capecitabine 825mg/ m² twice a day, at 30 min after main meals, 5 days per week, during radiotherapy. 6 weeks after the end of the oncological treatment, the patients were evaluated for surgery.

Blood samples were obtained in dynamics during radiotherapy as it follows: at the initiation of the treatment, at the 1/ 3 and 2/ 3 of the total radiation dose and at the end of radiotherapy. From the blood samples (5ml each of

them), the serum was separated by centrifugation and used to identify the biochemical oxidative stress parameters such as:

Malondialdehyde, the final result of the lipid peroxidation reaction, was determined by using the Carboneau method. It had normal levels between 0-2 $\mu\text{mol}/100\text{ ml}$ serum;

Serum ceruloplasmin is an acute phase protein. The liver is the most important site where the synthesis of ceruloplasmin takes place, but it can also occur extrahepatically. The Ravin method was used for its measurement, which was based on its p-phenylenediamine oxidase activity. Normal values were registered between 80 and 120 U.I.;

Albumin thiol compounds, part of the serum antioxidants, were determined with the Albini method. This test is based on the capacity of the SH groups to develop a colored complex with the acid 5,5-dithio-bis-2-nitrobenzoic. Normal levels were encountered among 370 and 450 $\mu\text{mol}/\text{l}$.

The serum's capacity to reduce iron through the redox colorimetric test with reductants was used to determine the total antioxidants. Normal values were between 0,9 and 1,4 $\mu\text{mol}/\text{l}$.

Results

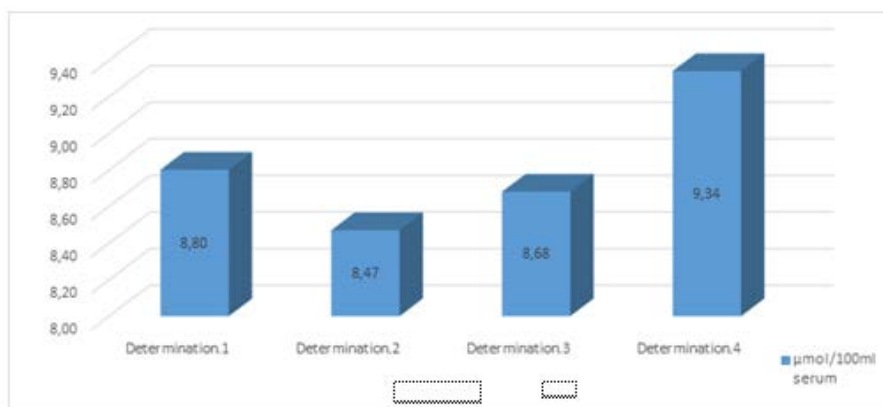


Fig. 1 Determination of the lipid peroxidation reaction ($\mu\text{mol}/100\text{ ml}$ serum)

It is known that the lipids, which are components of the cell membrane, represent the primary target of the oxidative attack [8]. The results showed an overproduction of oxidants, for example free radicals. Therefore, the lipid peroxidation reaction started and presented the maximum values at the end of

radiotherapy, due to the installation of the oxidative stress. The result of the second determination was smaller than the initial one, probably, due to antioxidant endogenous systems that were activated as an adaptive mechanism.

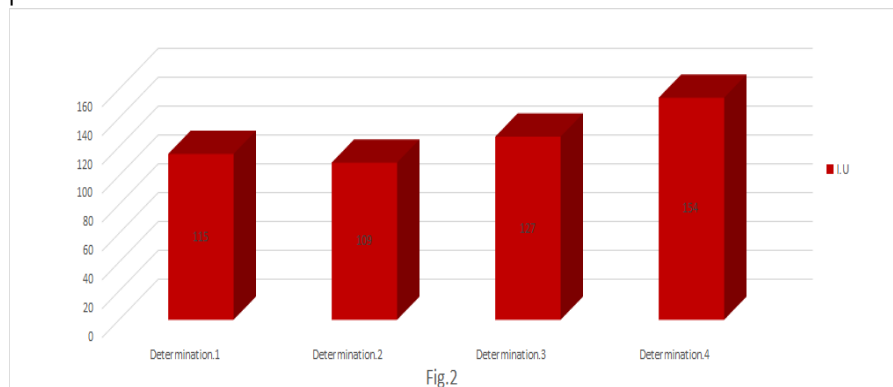


Fig. 2 Determination of ceruloplasmin (I.U.)

To confirm the installation of an important oxidative stress, the determination of the activity of serum ceruloplasmin was introduced in the study.

The increase of the recorded values in the dynamics of radiotherapy was significant and associated

with the induction of an irreversible and intense oxidative stress. The obtained values were in accordance with those from the determination of the lipid peroxidation.

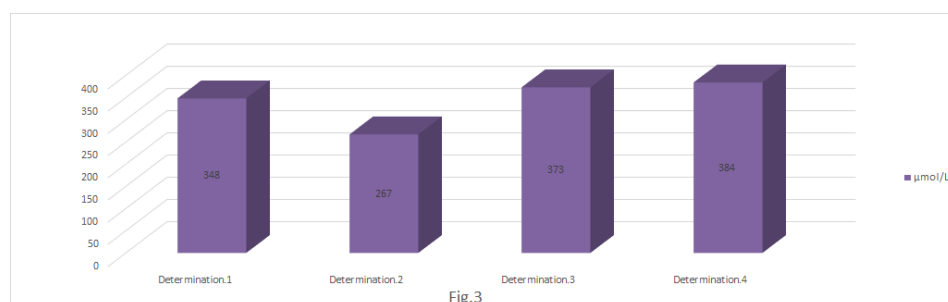


Fig. 3 Determination of thiol compounds (μmol/ l)

The thiol groups were part of the serum antioxidant defense system by having the capacity to cancel the oxidative reactions through the inactivation of the alkoxy and hydroxyl radicals [9]. By own oxidation, the thiol compounds interposed to the aggressive reaction of other free radicals. However, the obtained values were a consequence of the oxidative degradation of the serum albumins. Proteins represented the second target for the

oxidative stress, suggesting the fact that the oxidative stress was a continuous, irreversible process. The profile of the graphic showed a directly proportional relation between the dose of irradiation and the formation of the free thiol compounds. The value of the second determination remained low, apparently unexplainable, but we believed that it was given by the recovery mechanisms.

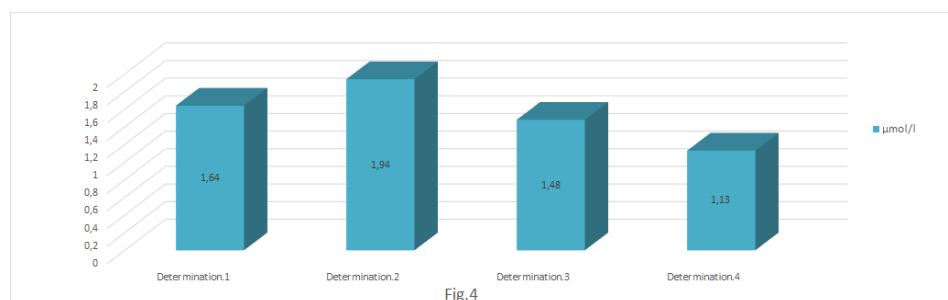


Fig. 4 Determination of total antioxidant level (μmol/ l)

As a response to the installed oxidative stress, initially an increase of the endogenous antioxidant activity, which decreased in time, was recorded. The results pointed out the activation of the natural protection systems, which were overwhelmed in time due to the installed oxidative stress.

Discussion

Many complex molecular and cellular alterations are involved in the process of chemical carcinogenesis [10,11]. The results of a normal cellular metabolism are the reactive species (such as oxygen reactive species), formed especially by the mitochondrial respiratory chain [12,13]. It has been demonstrated that nitric oxide can be generated also in the mitochondrial respiratory chain, but under hypoxic conditions; it can form other reactive species such as malondialdehyde [12]. One of the most important actions of reactive species is to determine damage to DNA, which is not sufficient to produce cancer and they may also play an important role in promoting proliferation, tumor invasion and development of metastases [14]. Once formed, they are rapidly

decomposed by the cellular enzymatic or non-enzymatic systems. The state of oxidative stress appears when these systems are overwhelmed by the in excess formation of the reactive species through an endogenous mechanism or exogenous factors [10,12,15].

Oxidative and antioxidative processes determine an electron transfer, this being the reason for the cellular redox imbalance, which represents a characteristic of cancer cells [15,16].

Even if oxidative stress is involved in the carcinogenesis, there is no sufficient data of the association between rectal cancer risk and parameters of oxidative stress [17].

Our study highlighted that an oxidative stress was installed during radiochemotherapy. We also pointed the fact that at the beginning of radiochemotherapy all serum levels of the oxidative stress parameters (malondialdehyde, serum ceruloplasmin and albumin thiols) were higher than the normal ones, suggesting that their high pre-therapeutic serum level could be associated with an increased risk of rectal cancer.

Preoperative radiochemotherapy is established as a standard treatment for locally advanced rectal cancer based on the result of the CAO/ARO/AIO94 trial, which

demonstrated a better local control after neoadjuvant versus adjuvant radiochemotherapy [18]. In addition, in an editorial, Glimelius recalled that concomitant radiochemotherapy decreases the risk of local relapse with 60-65% in the neoadjuvant setting compared to 20-40% when administered postoperative [19]. This difference was probably related to a restocking with tumor cells in the period between surgery and the initiation of radiotherapy or to the increase in free radicals determined by the surgical intervention [19,20]. Other explanation can be the hypoxic status of the cancer cells localized in the surgical bed, because it is known that hypoxia is the most important cause of radioresistance and local treatment failure [21].

Conclusion

The cell is constantly exposed to the cytotoxic action of the reactive species. The cancer cell is the subject of a dual cause-effect relationship and that is the

reason why we cannot say with absolute certainty if oxidative stress is a cause or a result of a malignant transformation at this level.

Many in vivo and in vitro studies demonstrated that compared to normal ones, cancer cells are exposed to an intense, continuously, sublethal oxidative stress.

Source of founding

This work received financial support through the project entitled "CERO – Career profile: Romanian Researcher", grant number POSDRU/159/1.5/S/135760, co-financed by the European Social Fund for Sectoral Operational Programme Human Resources Development 2007-2013.

Disclosures

Authors declare that there is no conflict of interest regarding the publication of this paper.

References

1. Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, Parkin DM, Forman D, Bray F. Cancer Incidence and Mortality Worldwide: Sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*. 2015 Mar 1; 136(5):E359-86. doi: 10.1002/ijc.29210.
2. Glimelius B, Pahlman L, Cervantes A. Cancerul rectal: Ghidurile de practica clinica ESMO pentru diagnostic, tratament si urmarire. *Journal of Radiotherapy & Medical Oncology*. 2014; 20 (Supplement 1):77.
3. Geisler DP. Local Treatment for Rectal Cancer. *Clin Colon Rectal Surg*. 2007 Aug; 20(3):182–189. doi: 10.1055/s-2007-984862.
4. Caravatta L, Picardi V, Tambaro R, Padula GD, Macchia G, Deodato F, Massaccesi M, Pacelli F, Berardi S, Ridolfini MP, Di Filippo L, Fabrizio G, Ingrosso M, Cellini N, Valentini V, Morganti AG. Neoadjuvant accelerated concomitant boost radiotherapy and multidrug chemotherapy in locally advanced rectal cancer: a dose-escalation study. *Am J Clin Oncol*. 2012 Oct; 35(5):424-31.
5. Schmiegel W, Reinacher-Schick A, Arnold D et al. Update S3-guideline "Colorectal cancer" 2008. *Zeitschrift für Gastroenterologie*. 2008; 46(8):799–840.
6. Glimelius B, Holm T, Blomqvist L. Chemotherapy in addition to preoperative radiotherapy in locally advanced rectal cancer - a systematic overview. *Rev Recent Clin Trials*. 2008 Sep; 3(3):204-11.
7. Funke S, Risch A, Nieters A, Hoffmeister M, Stegmaier C, Seiler C, Brenner H, Chang-Claude J. Genetic Polymorphisms in Genes Related to Oxidative Stress (*GSTP1*, *GSTM1*, *GSTT1*, *CAT*, *MnSOD*, *MPO*, *eNOS*) and Survival of Rectal Cancer Patients after Radiotherapy. *J Cancer Epidemiol*. 2009; 2009:302047. doi: 10.1155/2009/302047.
8. Ayala A, Muñoz MF, Argüelles S. Lipid Peroxidation: Production, Metabolism, and Signaling Mechanisms of Malondialdehyde and 4-Hydroxy-2-Nonenal. *Oxidative Medicine and Cellular Longevity*. 2014; 2014, Article ID 360438, 31 pages.
9. Long LH, Halliwell B. Oxidation and generation of hydrogen peroxide by thiols compounds in commonly used cell culture media. *Biochemical and Biophysical Research Communications*. 2001; 286:991–994.
10. Klaunig JE, Kamendulis LM. The role of oxidative stress in carcinogenesis. *Annu Rev Pharmacol Toxicol*. 2004; 44:239-67.
11. Valko M, Izakovic M. Role of oxygen radicals in DNA damage and cancer incidence. *Cancer Control*. 2006; 5:31–35.
12. Reuter S, Subasch C. Oxidative stress, inflammation and cancer. How are they link? *Free Radical Biology and Medicine*. 2010; 49:1603–1616.
13. Nitu R, Corol DI, Toma N. Free radicals in biological systems: their cytogenetics effects. *Progress in Biotechnology, University from Bucharest, Centrum of Researches in Enzymology, Biotechnology and Bioanalysis*, vol. 2, 2002, Ed. Ars Docendi, 17-24.
14. Halliwell B. Oxidative stress and cancer: have we moved forward?. *Biochem J*. 2007 Jan 1; 401(1):1-11.
15. Duracková Z. Some current insights into oxidative stress. *Physiol Res*. 2010; 59(4):459-69.
16. Valko M, Rhodes CJ. Free radicals, metals, and antioxidants in oxidative stress induced cancer. *Chemico-Biological Interactions*. 2006; 160:1–40.
17. Leufkens AM, van Duijnhoven FJ, Woudt SH, Siersema PD, Jenab M, Jansen EH, Pischon T, Tjønneland A, Olsen A, Overvad K, Boutron-Ruault MC, Clavel-Chapelon F, Morois S, Palli D, Pala V, Tumino R, Vineis P, Panico S, Kaaks R, Lukanova A, Boeing H, Aleksandrova K, Trichopoulou A, Trichopoulos D, Dilis V, Peeters PH, Skeie G, González CA, Argüelles M, Sánchez MJ, Dorronsoro M, Huerta JM, Ardanaz E, Hallmans G, Palmqvist R, Khaw KT, Wareham N, Allen NE, Crowe FL, Fedirko V, Norat T, Riboli E, Bueno-de-Mesquita HB. Biomarkers of oxidative stress and risk of developing colorectal cancer: a cohort-nested case-control study in the European Prospective Investigation Into Cancer and Nutrition. *Am J Epidemiol*. 2012 Apr 1; 175(7):653-63. doi: 10.1093/aje/kwr418.
18. Sauer R, Liersch T, Merkel S, Fietkau R, Hohenberger W, Hess C, Becker H, Raab HR, Villanueva MT, Witzigmann H, Wittekind C, Beissbarth T, Rödel C. Preoperative versus postoperative chemoradiotherapy for locally advanced rectal cancer: results of the German CAO/ARO/AIO-94 randomized phase III trial after a median follow-up of 11 years. *J*

- Clin Oncol. 2012 Jun 1; 30(16):1926-33. doi: 10.1200/JCO.2011.40.1836.
19. **Glimelius B.** Pre- or postoperative radiotherapy in rectal cancer – more to learn?. *Radiotherapy and Oncology*. 2001; 61:1-5.
20. **Potenza L, Calcabrini C, Bellis RD, Mancini U, Polidori E, Zeppa S, Alloni R, Cucchiari L, Dacha M.** Effect of surgical stress on nuclear and mitochondrial DNA from healthy sections of colon and rectum of patients with colorectal cancer. *J Biosci*. 2011 Jun; 36(2):243-51.
21. **Horsman MR, Overgaard J.** The impact of hypoxia and its modification of the outcome of radiotherapy. *J Radiat Res*. 2016 Aug; 57 Suppl 1:i90-i98. doi: 10.1093/jrr/rww007.