Research Article

Schinus terebinthifolius Essential Oil Attenuates Scopolamine-Induced Memory Deficits via Cholinergic Modulation and Antioxidant Properties in a Zebrafish Model

Elena Todirascu-Ciornea,¹ Heba A.S. El-Nashar,² Nada M. Mostafa,² Omayma A. Eldahshan ⁽¹⁾,² Razvan Stefan Boiangiu,¹ Gabriela Dumitru,¹ Lucian Hritcu ⁽¹⁾,¹ and Abdel Nasser B. Singab ⁽¹⁾

¹Department of Biology, Alexandru Ioan Cuza University of Iasi, Iasi 700506, Romania ²Department of Pharmacognosy, Ain Shams University, Cairo 11566, Egypt

Correspondence should be addressed to Lucian Hritcu; hritcu@uaic.ro and Abdel Nasser B. Singab; dean@pharma.asu.edu.eg

Received 16 May 2019; Revised 23 September 2019; Accepted 31 October 2019; Published 2 December 2019

Academic Editor: Yoshiki Mukudai

Copyright © 2019 Elena Todirascu-Ciornea et al. This is an open access article distributed under the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original work is properly cited.

Schinus terebinthifolius is a plant well recognized for its therapeutic profile such as anti-inflammatory and antitumor activities, promoting antibacterial activity and antioxidant and antidiabetic properties. This study aimed at examining whether *Schinus terebinthifolius* memory-enhancing activities are mediated by cholinergic and brain antioxidant systems in a scopolamine zebrafish model. *Schinus terebinthifolius* essential oil (10, 25, and $50 \,\mu$ L/L) was delivered to zebrafish by immersion in water for 8 days. Memory deficits were induced by scopolamine ($100 \,\mu$ M) administration. Zebrafish were divided into seven groups (n = 15/ group): vehicle group, scopolamine ($100 \,\mu$ M) group, *Schinus terebinthifolius* essential oil groups (STF; 10, 25, and $50 \,\mu$ L/L), the imipramine group (IMP; 20 mg/L, as the positive control in the NTT test), and the donepezil group (DP; 10 mg/L, as the positive control in the Y-maze test). Memory status was estimated by the novel tank diving test (NTT) and the Y-maze test and finally was validated by comparison with imipramine (20 mg/L) and donepezil ($10 \,m$ g/L). Gas chromatography-mass spectrometry (GC-MS) was used to detect oil compounds. Brain levels of acetylcholinesterase (AChE) and antioxidant enzymes were measured. After being exposed to *Schinus terebinthifolius* essential oil, the scopolamine zebrafish exhibited an improvement of memory processes in the NTT and Y-maze tests. The essential oil attenuated the elevated level of AChE and brain oxidative stress. *Schinus terebinthifolius* essential oil was found to support memory formation through the inhibition of the AChE activity and decreasing oxidative stress in the scopolamine-treated zebrafish brains.

1. Introduction

Human and animal behavior is regulated by the central cholinergic system [1]. In humans, the memory impairment induced by scopolamine reproduced some aspects of cognitive symptomatology evidenced in Alzheimer's disease (AD) [2], closely related to the degeneration of the cholinergic neurons in the basal forebrain-induced reduction of neurotransmission. Supporting evidence suggested memory deficits induced by scopolamine in zebrafish [3–5]. AChE inhibitors such as donepezil and rivastigmine are involved in the treatment of cognitive symptoms seen in AD [6].

In recent years, the anti-Alzheimer's potential for natural products from medicinal plants has been recognized by the scientific community [7, 8]. Hence, a great deal of research studies focus recently on plants and other natural products used around the world for age-related central nervous system (CNS) diseases [9].

Schinus L. (Anacardiaceae) is a genus native to South America particularly to Brazil and comprises about 29 species [10]. Several medicinal properties have been assigned to these plants in addition to their use as ornamental plants [11]. Different parts of *Schinus* plants have been utilized

traditionally for the alleviation of several disorders [12]. The essential oils of different *Schinus* species have previously been taken from different locations, and some variations in their chemical components and biological activities have been observed for *Schinus terebinthifolius* [10, 13–15].

Several classes of natural constituents were obtained from this plant, mainly essential oils [16]. Many medicinal properties were reported to the essential oil of *Schinus terebinthifolius*, such as antioxidant, wound healing, antitumor, and antimicrobial activities [17–19]. Moreover, the ethanolic extract of Egyptian *Schinus terebinthifolius* aerial parts exhibited anticholinesterase activity [20]. The pink fruits of *Schinus terebinthifolius* are rich in essential oils (1.5–10%), which was reported to have monoterpenes with high abundance along with some sesquiterpene hydrocarbons [21]. The pharmaceutical industry utilizes the essential oil of *Schinus terebinthifolius* fruits in cosmetics due to its aroma [22], and the Food and Drug Administration (FDA) ranked this essential oil as generally safe (GRAS) [23].

Supporting evidence suggested various biological effects of *Schinus terebinthifolius* Raddi. dos Santos da Rocha et al. [24] reported that *Schinus terebinthifolius* extract exhibited antioxidant and antidiabetic profile in mice mediated by its compounds such as gallic acid, gallotannins, and glycosylated flavonols. Miller et al. [25] demonstrated the vasodilatory activity of *Schinus terebinthifolius* extract in rats and mice and also antioxidant activities, especially in the CNS. The authors attributed these effects mainly to phenolic compounds. Vigo-Pelfrey et al. [26] reported the hepatoprotective effects of *Schinus terebinthifolius* extract against carbon tetrachloride- (CCl₄-) induced acute hepatotoxicity in rats.

Zebrafish (Danio rerio) is a promising in vivo tool to model or combat AD [27]. Also, zebrafish exhibited high sensitivity to all major classes of clinically active neurotropic drugs in a similar fashion like humans [28]. Furthermore, complex brain disorders such as depression, autism, psychoses, and cognitive deficits may be successfully analyzed in zebrafish in a cheaper and more powerful manner than in rodents [28]. Supporting evidence suggested that, in zebrafish, scopolamine exhibited memory-reducing effects without causing locomotory deficits and is often used with nootropic and memory-enhancing drugs to study memory formation [29]. Also, recently, our group demonstrated that agathisflavone isolated from Schinus polygamus (Cav.) Cabrera leaves improved memory and decreased brain oxidative stress and AChE activity in the scopolamine zebrafish model [5].

The present study is considering to characterize the chemical components of the essential oil isolated from *Schinus terebinthifolius* fruits grown in Egypt and to evaluate the antiamnesic and antioxidant effects of the essential oil against scopolamine using *in vivo* tests and to elucidate the main mechanism of action.

2. Materials and Methods

2.1. Plant Material. Schinus terebinthifolius Raddi fruits were collected during April 2017 from the Garden of

Mohamed Ali Palace, Giza, Egypt. Samples were identified by Mrs. Tereize Labib, the Taxonomy Specialist at El-Orman Botanical Garden, Giza, Egypt, and voucher specimen (no. PHG-P-ST-186) has been deposited at Pharmacognosy Department, Faculty of Pharmacy, Ain Shams University.

2.2. Extraction of the Essential Oil. The fruits of the plant were immediately hydrodistilled for 4 h using Clevenger-type apparatus. The resulting oil was dried over anhydrous sodium sulfate and stored at 4°C in the dark until use.

2.3. Gas Chromatography-Mass Spectrometry (GC-MS). The GC-MS analyses of the resulting oils were carried out at the Department of Medicinal and Aromatic Plants Research, National Research Center, using TRACE GC Ultra Gas Chromatographs (THERMO Scientific Corp., USA), coupled with a thermomass spectrometer detector (ISQ Single Quadrupole Mass Spectrometer). The GC-MS system was equipped with a TG-5MS column $(30 \text{ m} \times 0.25 \text{ mm i.d.},$ $0.25 \,\mu m$ film thickness). The analysis was carried out using helium as carrier gas at a flow rate of 1 mL/min and a split ratio of 1:10 using the following temperature program: 80°C for 2 min, rising at 5°C/min to 300°C, and is held for 5 min. The injector and detector were held at 280°C. 0.2 µL of diluted samples (1:10 hexane, v/v) were always injected. Mass spectra were obtained by electron ionization (EI) at 70 eV, using a spectral range of m/z 35–500. The components of essential oils were identified by comparison of their Kovats retention indices with those in the literature [30].

2.4. Experimental Animals and Maintenance. 105 adult male and female, wild-type short-thin zebrafish (Danio rerio), 3-4-month-old and 3-4 cm in length, were obtained from an authorized commercial supplier (Pet Product S.R.L., Bucharest, Romania). Animals were randomly divided into groups of 15 fish/10 L tank with a constant 14:10 h of the light/dark cycle at $26^{\circ}C \pm 2$ and fed twice daily with Norwin Norvitall flake. All fish used in this experiment were observed in guarantine at least one week before use in experimental studies. Acclimated zebrafish were randomly assigned into control, scopolamine (100 μ M), and three Schinus terebinthifolius essential oil treatment groups (STF; 10, 25, and 50 μ L/L). Also, the donepezil group (DP; 10 mg/ L) and imipramine (IMP; 20 mg/L) were used as positive controls. The research was accomplished by the Directive 2010/63/EU of the European Parliament and of the Council of 22 September 2010 on the protection of animals. All efforts were made to minimize animal suffering and to reduce the number of animals used.

The experimental procedure is described in Figure 1. Also, for the behavioral and biochemical parameters assays, we followed the methods of Dumitru et al. [31].

2.5. Acute Toxicity Study. An acute toxicity study was determined using 80 fish of both sexes divided into 4 groups as follows: a control group and STF (10, 25, and $50 \,\mu$ L/L)



FIGURE 1: Scheme of the experimental procedure.

groups. The fish were monitored for two weeks for any symptoms of mortality and toxicity [5].

2.6. Novel Tank Diving Test (NTT). The NTT was used to assess anxiety response in experimental fish according to a method described by Levin et al. [32]. ANY-maze® software (Stoelting Co, USA) and Logitech HD Webcam C922 Prostream camera were used for the automatic tracking of zebrafish behavior. Zebrafish were separately transferred into the tank, and for 6 min, the following variables were recorded: total distance (m), latency to the top (s), time spent in the top (s), time spent in the bottom (s), number of entries in the top, and number of entries in the bottom. In the NTT test, imipramine (IMP; 20 mg/L) was used as the reference drug.

2.7. Y-Maze Task. To explore the spatial memory, a Y-maze was used [5]. During 5 min, each fish was singly evaluated [4], and the behavioral endpoints analyzed were as follows: the total distance (m), the percent spontaneous alternation, and the locomotor activity (the number of arm entries). Donepezil (DP; 10 mg/L) was used as the reference drug in the Y-maze test.

2.8. Biochemical Parameter Assay. At the end of the experiment, all fish were euthanized by immersion for 10 min in ice water (2–4°C) until the disappearance of opercular movements [33]. The brain samples were dissected out in the ice. Samples were homogenized (1:10) in ice-cold 0.1 M potassium phosphate buffer (pH 7.4) and 1.15% KCl using a Potter Homogenizer coupled with Cole-Parmer Servodyne Mixer. The obtained supernatant was used for the biochemical assay, including acetylcholinesterase (AChE) [34], superoxide dismutase (SOD) [35], catalase (CAT) [36], glutathione peroxidase (GPX) [37] specific activities, and malondialdehyde (MDA) level [5]. Protein content was investigated using a bicinchoninic acid (BCA) protein assay kit (Sigma-Aldrich, Germany) using a method as described previously [38].

2.9. Statistical Analysis. The results are presented as the mean \pm standard error of the mean (SEM) and analyzed using a one-way analysis of variance (ANOVA) followed by

Tukey's *post hoc* multiple comparison test, using a p < 0.05 level of significance. Pearson correlation coefficient (r) was measured to assay the correlation between the behavioral scores, enzymatic activities, and lipid peroxidation. All values were calculated using GraphPad software (GraphPad Prism 7.0, La Jolla, CA, USA).

3. Results and Discussion

3.1. Acute Toxicity Study. After the administration of STF (10, 25, and 50 μ L/L), the zebrafish did not show behavioral changes or other signs of toxicity and mortality, indicating the safety profile of the STF doses. Supporting data provide evidence that the administration of Schinus terebinthifolius extract does not induce any toxic effects, which could be an assurance for the medicinal use of this plant in traditional medicine. Lima et al. [39] demonstrated that acute and subacute administration (45 days) of the Schinus terebinthifolius bark extract does not produce toxic effects in Wistar rats of both sexes. Affonso et al. [40] reported no signs of toxicity of the essential oil from fruits of Schinus terebinthifolius on reproductive functions in male Wistar rats. Our results are following those described by other authors, demonstrating that STF was found to be nontoxic in the tested doses and experimental conditions.

3.2. Chemical Composition of the Essential Oil. A total of 27 chemicals were identified by GC-MS analysis (Figure 2), accounting for 97.22% of the total essential oil (Table 1). The most abundant chemical classes of the oil components were monoterpene hydrocarbons (78.17%), followed by oxygenated monoterpenes (17.54%), sesquiterpene hydrocarbons (0.35%), and oxygenated sesquiterpene (0.33%). The major components of the isolated essential oil were identified as β -phellandrene (32.40%), α -pinene (16.68%), terpinen-4-ol (11.01%), α -phellandrene (10.56%), and β -pinene (4.26%), followed by *dl*-limonene (3.74%), *y*-terpinene (3.03%), α -terpineol (2.59%), and α -terpinene (2.35%). The chemical composition of our essential oil showed a slight similarity to those reported by other authors. Cole et al. [41] studied the chemical composition of the essential oil from the ripe fruit of Schinus terebinthifolius collected on the campus of the Federal University of Espírito Santo (UFES), Goiabeiras, Vitória, Espírito Santo, Brazil. The authors identified 17



FIGURE 2: GC-MS chromatogram of essential oil isolated from *Schinus terebinthifolius* fruits grown in Egypt. The major components are highlighted in bold and marked by numbers in Table 1.

TABLE 1: The chemical composition of essential oil isolated from the fruits of Schinus terebinthifolius.

	Compound	Molecular formula	Retention index		7 1
No.			Calculated	Reported	Relative composition (%)
1.	Thujene	$C_{10}H_{16}$	923	930	1.08
2.	α-Pinene	$C_{10}H_{16}$	931	939	16.68
3.	Camphene	$C_{10}H_{16}$	950	954	0.17
4.	Sabinene	$C_{10}H_{16}$	972	975	0.68
5.	β -Pinene	$C_{10}H_{16}$	977	979	4.26
6.	α-Myrcene	$C_{10}H_{16}$	987	990	0.92
7.	Decane	$C_{10}H_{22}$	994	1000	0.56
8.	α-Phellandrene	$C_{10}H_{16}$	1005	1002	10.56
9.	α-Terpinene	$C_{10}H_{16}$	1017	1017	2.35
10.	o-Cymene	$C_{10}H_{14}$	1027	1026	0.74
11.	<i>dl</i> -Limonene	$C_{10}H_{16}$	1029	1029	3.74
12.	β -Phellandrene	$C_{10}H_{16}$	1030	1029	32.40
13.	α-Ocimene	$C_{10}H_{16}$	1045	1039	0.25
14.	γ-Terpinene	$C_{10}H_{16}$	1057	1059	3.03
15.	cis-Sabinene hydrate	$C_{10}H_{18}O$	1072	1070	0.09
16.	α-Terpinolene	$C_{10}H_{16}$	1083	1088	0.75
17.	Undecane	$C_{11}H_{24}$	1093	1100	0.85
18.	cis-p-Menth-2-en-1-ol	$C_{10}H_{18}O$	1122	1121	1.66
19.	Trans-1-Methyl-4-(1-methylethyl)-2-cyclohexen-1-ol	$C_{10}H_{18}O$	1142	1139	1.24
20.	Isoborneol	$C_{10}H_{18}O$	1156	1160	0.11
21.	Terpinen-4-ol	$C_{10}H_{18}O$	1178	1177	11.10
22.	α-Terpineol	$C_{10}H_{18}O$	1193	1188	2.59
23.	trans-Piperitol	$C_{10}H_{18}O$	1206	1208	0.48
24.	α -Terpinyl acetate	$C_{12}H_{20}O_2$	1345	1349	0.27
25.	Germacrene D	$C_{15}H_{24}$	1482	1485	0.11
26.	Germacrene B	$C_{15}H_{24}$	1553	1561	0.24
27.	10-Epi-y-eudesmol	$C_{15}H_{26}O$	1619	1623	0.31
Mond	oterpene hydrocarbons				78.17
Oxyg	Monoterpene hydrocarbons Oxygenated monoterpenes				17.54
Sesqu	Oxygenated monoterpenes Sesquiterpene hydrocarbons				0.35
Oxygenated sesquiterpenes					
Other	'S				0.85
Total	identified				97.22

compounds accounting for 91.15% of the total oil. Among them, the principal identified compounds were δ -3-carene (30.37%), limonene (17.44%), α -phellandrene (12.60%), α -pinene (12.59%), and *trans*-caryophyllene (1.77%). Maciel et al. [42] identified 37 compounds in the essential oil obtained from *Schinus terebinthifolia* leaves, germacrene B (14.9%), δ -cadinene (7.6%), β -elemene (7.2%), and α -murolol (5.1%) were the most abundant compounds. Ennigrou et al. [43] identified α -phellandrene (33.06–36.18%), α -pinene (14.85–15.18%), and limonene (6.62–8.79%), being the major components from the essential oil composition of *Schinus terebinthifolius* leaves and twigs collected in the region of Al

Ghazala, Ariana, Northern Tunisia. de Lima et al. [44] reported that *Schinus terebinthifolius* contains α - and β -pinene, Δ^3 -carene, limonene, α - and β -phellandrene, *p*-cymene, and terpinolene as the main components, together with low amounts of monoterpene alcohols and ketones, sesquiterpene hydrocarbons, alcohols and ketones, and triterpene alcohols and ketones [45]. Another study by Afifi et al. [46] described the α -pinene and the β -phellandrene as the major components in all the analyzed samples from the Schinus mole essential oil growing in Jordan. The observed variation in the chemical composition of the essential oil from Schinus terebinthifolius is related to the environmental conditions, the extraction protocols, and the part of the plant submitted to extraction. Based on these results, we found that our essential oil has a chemical composition comparable to those reported by other groups and could support its memory enhancer and antioxidant profile.

The compounds are listed in order of their elution on the DB-5 column. RI, identification based on a comparison of published Kovats retention indices; MS, identification based on mass spectral data.

3.3. Schinus terebinthifolius Essential Oil Improves Memory Performance. One-way ANOVA demonstrated overall powerful changes of the total distance (F (5, 84) = 3.23, p < 0.01), on the latency to top (F(5, 84) = 4.56, p < 0.001), on the time spent in top (F(5, 84) = 12.11, p < 0.0001), on the time spent in bottom (F(5, 84) = 2.64, p < 0.01), on the number of entries in the top (F(5, 84) = 20.35, p < 0.0001), and on the number of entries to the bottom (F(5, 84) =44.98, p < 0.0001). In addition, zebrafish exposed to scopolamine $(100 \,\mu\text{M})$ displayed anxiogenic-like profile, as evidenced by a significant increase of the latency to top (p < 0.001) and of the number of entries to the bottom (p < 0.0001) and a significant decrease of the time spent in top (p < 0.001) and of the number of entries to the top (p < 0.001) as compared to the vehicle group (Figure 3). Furthermore, STF attenuates the effects induced by scopolamine in a dose-dependent manner as compared to imipramine effects, collectively suggesting anxiolytic effects. Accordingly, locomotion tracking plots in zebrafish exposed to scopolamine were altered, whereas STF restored locomotion patterns nearly like an imipramine-treated group.

Furthermore, to assess the effects of STF on memory, the Y-maze test was employed (Figure 4). One-way ANOVA revealed representative changes of the total distance (F (5, 84) = 2.66, p < 0.01), on the number of arm entries (F (5, 84) = 20.93, p < 0.001), and on the percentage of the spontaneous alternation (F (5, 84) = 13.45, p < 0.0001). Also, the scopolamine treatment-induced memory deficits as evidenced by a significant decrease in the spontaneous alternation percentage (p < 0.0001) as compared to the vehicle group. Also, the treatment with scopolamine promoted decreased locomotion tracking plots that were prevented by STF treatment nearly to the donepezil group. Our results suggested that *Schinus terebinthifolius* essential oil possesses anxiolytic and memory supporting profile, which is in

agreement with the literature where Schinus terebinthifolius administration exhibited the neuroprotective effects possibly mediated by its antioxidant activity in a rotenone rat model of Parkinson's disease [7]. In addition, the improvement of memory deficits as a result of STF administration could be attributed to not only the presence in high amounts of the active compounds such as β -phellandrene, α -pinene, terpinen-4-ol, and α -phellandrene but also to the presence of other components in small amounts but with high activity such as β -pinene, followed by dl-limonene, γ -terpinene, α -terpineol, and α -terpinene or synergy between them. Lee at al. [47] demonstrated that *a*-pinene exhibited neuroprotective effects against memory impairment induced by scopolamine in C57BL/6 mice by increasing the mRNA expression of choline acetyltransferase in the cortex and the activity of the antioxidant enzymes in the hippocampus via activation of NF-E2-related factor 2. α -Phellandrene, a cyclic monoterpene presented in our STF, was reported to attenuate the inflammatory response through neutrophil migration modulation and mast cell stabilization in male Wistar rats or Swiss mice [48]. Machado et al. [49] identified the presence of high content of triterpenes in the chemical composition of the Schinus molle leaves extract. Also, the authors evidenced the antidepressant-like effect of the Schinus molle leaves extract in mice being dependent by its

3.4. Schinus terebinthifolius Essential Oil Exhibited Antiacetylcholinesterase Activity. To further elucidate the potential mechanism of the STF against scopolamine-induced memory deficits, the brain AChE activity was firstly evaluated. One-way ANOVA demonstrated overall significant effects on AChE specific activity (F (5, 84) = 68.57), p < 0.0001) (Figure 5). Treatment with scopolamine generates a significant increase in AChE activity in the brain as compared to the vehicle group (p < 0.0001). In contrast, the administration of the STF significantly reversed the effects of scopolamine on AChE activity (p < 0.0001). The effects of STF were consistent with those of donepezil. Donepezil significantly improved memory in the Y-maze test and showed inhibition of AChE. These results suggest that STF could protect against scopolamine-induced dysfunction of the cholinergic system.

interaction with the serotonergic (5-HT $_{1\mathrm{A}}$, 5-HT $_{2\mathrm{A}}$, and 5-

HT₃ receptors), noradrenergic (α_1 and α_2 -receptors), and

dopaminergic (D_1 and D_2 receptors) systems. Piccinelli et al.

[50] demonstrated antidepressive actions of the essential oil

from Schinus terebinthifolius fruits in rats mediated by (R)-

(+)-limonene and α -phellandrene. However, recently, we

demonstrated that Schinus polygamus (Cav.) Cabrera leaves

extract reduced cognitive deficits in scopolamine-treated

zebrafish, mainly due the presence of the agathisflavone, a

natural biflavonoid [5]. Our research provides rational use of

the STF in the direction of memory deficits, anxiety, de-

pression, and connected dementia.

AChE overexpression was evidenced as an important indicator of the cholinergic system dysfunction that could play a pivotal role in AD pathogenesis [51]. However, AD therapy implies AChE inhibitors and thereby enhances



FIGURE 3: Effects of the *Schinus terebinthifolius* essential oil (STF; 10, 25, and $50 \,\mu$ L/L) and the locomotion tracking pattern and behavioral parameters in different groups in the NTT test. Values are means ± SEM (*n* = 15). For Tukey's *post hoc* analyses, ## Sco vs. STF ($50 \,\mu$ L/L): *p* < 0.01 for total distance in tank, ## Sco vs. STF ($10 \,\mu$ L/L): *p* < 0.001, ## Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.001, and ## Sco vs. STF ($50 \,\mu$ L/L): *p* < 0.001 for latency to top, ### Sco vs. STF ($10 \,\mu$ L/L): *p* < 0.001, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001, and ### Sco vs. STF ($50 \,\mu$ L/L): *p* < 0.0001 for time spent in top, #Co vs. STF ($25 \,\mu$ L/L): *p* < 0.001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 for time spent in bottom, ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 for number of entries in the top and ### Sco vs. STF ($25 \,\mu$ L/L): *p* < 0.0001 and ### Sco vs. STF ($50 \,\mu$ L/L): *p* < 0.0001 for number of entries to the bottom.

cholinergic neurotransmission [52]. Murray et al. [53] reported that the essential oils from *Schinus areira* L. and *Schinus longifolia* (Lindl.) Speg. showed AChE inhibition. Among them, *Schinus longifolia* essential oil exhibited high AChE inhibition with an IC₅₀ value of $20 \pm 1 \,\mu$ g/mL. The STF used in the present study contains high levels of monoterpenes, like α -pinene (16.68%), α -phellandrene (10.56%), and β -phellandrene (32.40%) that could explain the observed AChE inhibitory activity and antioxidant activity, respectively.

3.5. Schinus terebinthifolius Essential Oil Restored the Brain Antioxidant Status. To support the memory enhancement and anti-AChE activities of STF in the scopolamine-treated rats, the brain antioxidant status was assessed secondly. Antioxidant enzymes including SOD, CAT, and GPX play a decisive role in the reduction of oxidative stress [54].

One-way ANOVA demonstrated overall significant effects on SOD (F(5, 84) = 50.33, p < 0.0001), CAT (F(5, 84) = 60.20, p < 0.0001), and GPX (F(5, 84) = 59.50, p < 0.0001) specific activities (Figure 5).

SOD, CAT, and GPX enzyme activity levels dramatically decreased as compared to the vehicle group after exposure to scopolamine (p < 0.0001), whereas the administration of the STF could restore the antioxidant status in the scopolamine-treated groups in a dose-dependent manner (p < 0.0001). Also, one-way ANOVA revealed significant effects on the MDA level (F (5, 84) = 24.28, p < 0.0001). At the same time, the level of MDA increased in the scopolamine-treated



FIGURE 4: Effects of the *Schinus terebinthifolius* essential oil (STF; 10, 25, and 50 μ L/L) on the locomotion tracking pattern and behavioral parameters in different groups in the Y-maze test. Values are means ± SEM. (n = 15). For Tukey's post hoc analyses, ## Sco vs. STF (10 μ L/L): p < 0.001, ## Sco vs. STF (25 μ L/L): p < 0.001, and ### Sco vs. STF (50 μ L/L): p < 0.001 for number of arm entries, ## Sco vs. STF (10 μ L/L): p < 0.001, ## Sco vs. STF (25 μ L/L): p < 0.001, and ## Sco vs. STF (50 μ L/L): p < 0.001 for spontaneous alternation %.

groups (p < 0.0001) as compared to the vehicle group, while exposure to STF dramatically decreased the level of MDA in a dose-dependent manner in the scopolamine-treated groups (p < 0.0001).

It has been reported that the brain of the patients suffering from AD presents a significant extent of oxidative damage [30]. Moreover, oxidative stress can induce cellular apoptosis in brain and cognitive impairment [55]. The activation of the antioxidant enzymes such as SOD and GPX could scavenge reactive oxygen species and protect cells against oxidative damage [56]. In our study, scopolamine decreased the antioxidant enzyme activity levels such as SOD, CAT, and GPX and increased the MDA level in the zebrafish brain. Thus, scopolamine created an imbalance between antioxidant and oxidant defense systems which may be responsible for observed impairment of memory in zebrafish. However, the essential oil restored the antioxidant status as evidenced by increased SOD, CAT, and GPX activity in scopolamine-treated zebrafish that parallels with a decrease of MDA level in the zebrafish brain. The results indicated that STF exhibits neuroprotective effects against oxidative stress, which is correlated with the previous studies. Silva et al. [57] demonstrated in vitro antioxidant, antiproliferative, and in vivo anti-inflammatory activities of Schinus terebinthifolius due to the presence of phenolic constituents. However, Dumitru et al. [31] already reported that Schinus polygamus (Cav.) Cabrera leaves extract is capable to control brain oxidative status in a scopolamine zebrafish model through agathisflavone, a natural biflavonoid that could restore the antioxidant enzyme activities. Our results agree with the behavioral results. Thus, we presumed that the modification of the cholinergic neuronal

system and the reduction of oxidative stress is the underlying mechanism to the cognitive enhancement effect of STF in scopolamine-treated rats.

Using the Pearson correlation coefficient, which quantifies the linear association between two quantitative variables, it was possible to estimate the relationships between different behavioral scores and biochemical parameters [58]. Our results revealed strong correlations between behavioral scores, enzymatic activities, and lipid peroxidation. Moreover, when the Pearson correlation coefficient (r) was evaluated, significant correlations between the spontaneous alternation percentage vs. MDA, the time spent in top vs. MDA, time spent in bottom vs. MDA, and the number of entries to the top vs. MDA, AChE vs. MDA, SOD vs. MDA, CAT vs. MDA, and GPX vs. MDA were reported (Table 2).

The negative correlation coefficient indicates that the lower is the value of MDA and the higher are the values of behavioral scores and antioxidant enzymes.

A significant correlation between potential antioxidant and anxiolytic effects of (+)-limonene epoxide has been evidenced in mice [59]. Alzate et al. [60] evidenced a correlation between the free radical-scavenging activity exhibited by *Bomarea* species and the phenolic content in the extracts.

The correlation coefficients that were assessed in the current study indicate that the antioxidant profile possessed by STF as a result of high amounts of monoterpenes is well correlated with the improvement of memory in behavioral tests (NTT and Y-maze) along with the decrease of AChE activity. Thus, STF exhibits neuroprotection against scopolamine-induced oxidative stress generation in the zebrafish brain.



TABLE 2: Correlation data between the different parameters measured in the study (behavioral and biochemical parameters).

	Biochemical parameters	Pearson correlation coefficient (r)	Significance level (<i>p</i>)
Behavioral scores	I I I I I I I I I I I I I I I I I I I		8
Spontaneous alternation (%)	MDA	-0.563	p < 0.01
Time spent in top (s)	MDA	-0.653	p < 0.001
Time spent in bottom (s)	MDA	0.541	p < 0.01
Number of entries to the top	MDA	-0.512	p < 0.01
Biochemical parameters			1
AChE	MDA	0.724	<i>p</i> < 0.001
SOD	MDA	-0.586	p < 0.01
CAT	MDA	-0.713	p < 0.001
GPX	MDA	-0.630	<i>p</i> < 0.001

4. Conclusions

In conclusion, our results contribute to clarify the pharmacology of *Schinus terebinthifolius* essential oil in the zebrafish model of scopolamine, by demonstrating its memory-enhancing effects via cholinergic modulation and antioxidant properties. Further studies are required for a

possible application of *Schinus terebinthifolius* essential oil to improve cognition.

Data Availability

The data used to support the findings of this study are available from the corresponding author upon request.

Conflicts of Interest

The authors declare that there are no conflicts of interest regarding the publication of this paper.

Acknowledgments

This project was funded by the Ministry of Research and Innovation within Program 1—Development of the National RD System, Subprogram 1.2—Institutional Performance—RDI excellence funding projects (Contract no. 34PFE/19.10.2018).

References

- A. V. Terry and J. J. Buccafusco, "The cholinergic hypothesis of age and Alzheimer's disease-related cognitive deficits: recent challenges and their implications for novel drug development," *Journal of Pharmacology and Experimental Therapeutics*, vol. 306, no. 3, pp. 821–827, 2003.
- [2] I. Misane and S. O. Ögren, "Selective 5-HT1A antagonists WAY 100635 and NAD-299 attenuate the impairment of passive avoidance caused by scopolamine in the rat," *Neuropsychopharmacology*, vol. 28, no. 2, pp. 253–264, 2003.
- [3] J. M. Bailey, A. N. Oliveri, and E. D. Levin, "Pharmacological analyses of learning and memory in zebrafish (*Danio rerio*)," *Pharmacology Biochemistry and Behavior*, vol. 139, pp. 103– 111, 2015.
- [4] G. d. P. Cognato, J. W. Bortolotto, A. R. Blazina et al., "Y-Maze memory task in zebrafish (*Danio rerio*): the role of glutamatergic and cholinergic systems on the acquisition and consolidation periods," *Neurobiology of Learning and Memory*, vol. 98, no. 4, pp. 321–328, 2012.
- [5] K. Kim, Y. Bu, S. Jeong et al., "Memory-enhancing effect of a supercritical carbon dioxide fluid extract of the needles of *Abies koreana* on scopolamine-induced amnesia in mice," *Bioscience, Biotechnology, and Biochemistry*, vol. 70, no. 8, pp. 1821–1826, 2006.
- [6] P. T. Francis, A. M. Palmer, M. Snape, and G. K. Wilcock, "The cholinergic hypothesis of Alzheimer's disease: a review of progress," *Journal of Neurology, Neurosurgery, and Psychiatry*, vol. 66, no. 2, pp. 137–147, 1999.
- [7] A. Sereniki, C. F. B. Linard-Medeiros, S. N. Silva et al., "Schinus terebinthifolius administration prevented behavioral and biochemical alterations in a rotenone model of Parkinson's disease," *Revista Brasileira de Farmacognosia*, vol. 26, no. 2, pp. 240–245, 2016.
- [8] G. Karthivashan, S.-Y. Park, M.-H. Kweon et al., "Ameliorative potential of desalted Salicornia europaea L. extract in multifaceted Alzheimer's-like scopolamine-induced amnesic mice model," *Scientific Reports*, vol. 8, no. 1, p. 7174, 2018.
- [9] A. V. Terry, P. M. Callahan, B. Hall, and S. J. Webster, "Alzheimer's disease and age-related memory decline (preclinical)," *Pharmacology Biochemistry and Behavior*, vol. 99, no. 2, pp. 190–210, 2011.
- [10] R. Anand, K. D. Gill, and A. A. Mahdi, "Therapeutics of Alzheimer's disease: past, present and future," *Neuropharmacology*, vol. 76, pp. 27–50, 2014.
- [11] J. B. Martin, "Molecular basis of the neurodegenerative disorders," *New England Journal of Medicine*, vol. 340, no. 25, pp. 1970–1980, 1999.
- [12] E. Bruel-Jungerman, P. J. Lucassen, and F. Francis, "Cholinergic influences on cortical development and adult

neurogenesis," Behavioural Brain Research, vol. 221, no. 2, pp. 379-388, 2011.

- [13] S. Kotani, T. Yamauchi, T. Teramoto, and H. Ogura, "Pharmacological evidence of cholinergic involvement in adult hippocampal neurogenesis in rats," *Neuroscience*, vol. 142, no. 2, pp. 505–514, 2006.
- [14] N. Solanki, A. Salvi, G. Patki, and S. Salim, "Modulating oxidative stress relieves stress-induced behavioral and cognitive impairments in rats," *The International Journal of Neuropsychopharmacology*, vol. 20, no. 7, pp. 550–561, 2017.
- [15] M. Valko, D. Leibfritz, J. Moncol, M. T. D. Cronin, M. Mazur, and J. Telser, "Free radicals and antioxidants in normal physiological functions and human disease," *The International Journal of Biochemistry & Cell Biology*, vol. 39, no. 1, pp. 44–84, 2007.
- [16] V. López, S. Martín, M. P. Gómez-Serranillos, M. E. Carretero, A. K. Jäger, and M. I. Calvo, "Neuroprotective and neurochemical properties of mint extracts," *Phytotherapy Research*, vol. 24, no. 6, pp. 869–874, 2010.
- [17] M. Božović, A. Pirolli, and R. Ragno, "Mentha suaveolens ehrh. (Lamiaceae) essential oil and its main constituent piperitenone oxide: biological activities and chemistry," *Molecules (Basel, Switzerland)*, vol. 20, no. 5, pp. 8605–8633, 2015.
- [18] T. Kihara and S. Shimohama, "Alzheimer's disease and acetylcholine receptors," *Acta Neurobiol Exp (Wars)*, vol. 64, no. 1, pp. 99–105, 2004.
- [19] J. Shen, "Impaired neurotransmitter release in Alzheimer's and Parkinson's diseases," *Neuro-degenerative Diseases*, vol. 7, no. 1–3, pp. 80–83, 2010.
- [20] J. M. Ellis, "Cholinesterase inhibitors in the treatment of dementia," *The Journal of the American Osteopathic Association*, vol. 105, no. 3, pp. 145–158, 2005.
- [21] D. A. El-Sherbiny, A. E. Khalifa, A. S. Attia, and E. E.-D. S. Eldenshary, "Hypericum perforatum extract demonstrates antioxidant properties against elevated rat brain oxidative status induced by amnestic dose of scopolamine," *Pharmacology Biochemistry and Behavior*, vol. 76, no. 3, pp. 525–533, 2003.
- [22] V. V. Giridharan, R. A. Thandavarayan, S. Sato, K. M. Ko, and T. Konishi, "Prevention of scopolamine-induced memory deficits by schisandrin B, an antioxidant lignan from Schisandra chinensis in mice," *Free Radical Research*, vol. 45, no. 8, pp. 950–958, 2011.
- [23] A. Nunomura, R. J. Castellani, X. Zhu, P. I. Moreira, G. Perry, and M. A. Smith, "Involvement of oxidative stress in Alzheimer disease," *Journal of Neuropathology & Experimental Neurology*, vol. 65, no. 7, pp. 631–641, 2006.
- [24] P. dos Santos da Rocha, A. P. de Araújo Boleti, M. do Carmo Vieira et al., "Microbiological quality, chemical profile as well as antioxidant and antidiabetic activities of Schinus terebinthifolius Raddi," Comparative Biochemistry and Physiology Part C: Toxicology and Pharmacology, vol. 220, pp. 36–46, 2018.
- [25] D. L. Miller, I. A. Papayannopoulos, J. Styles et al., "Peptide compositions of the cerebrovascular and senile plaque core amyloid deposits of Alzheimer's disease," *Archives of Biochemistry and Biophysics*, vol. 301, no. 1, pp. 41–52, 1993.
- [26] C. Vigo-Pelfrey, D. Lee, P. Keim, I. Lieberburg, and D. B. Schenk, "Rapid communication: characterization of β -amyloid peptide from human cerebrospinal fluid," *Journal of Neurochemistry*, vol. 61, no. 5, pp. 1965–1968, 1993.

- [27] K. H. C. Baser, "Biological and pharmacological activities of carvacrol and carvacrol bearing essential oils," *Current Pharmaceutical Design*, vol. 14, no. 29, pp. 3106–3119, 2008.
- [28] M. Jukic, O. Politeo, M. Maksimovic, M. Milos, and M. Milos, "In vitro acetylcholinesterase inhibitory properties of thymol, carvacrol and their derivatives thymoquinone and thymohydroquinone," *Phytotherapy Research*, vol. 21, no. 3, pp. 259–261, 2007.
- [29] T. J. Hamilton, A. Morrill, K. Lucas et al., "Establishing zebrafish as a model to study the anxiolytic effects of scopolamine," *Scientific Reports*, vol. 715081 pages, 2017.
- [30] W.-J. Huang, X. Zhang, and W.-W. Chen, "Role of oxidative stress in Alzheimer's disease," *Biomedical Reports*, vol. 4, no. 5, pp. 519–522, 2016.
- [31] G. Dumitru, H. A. S. El-Nashar, N. M. Mostafa et al., "Agathisflavone isolated from Schinus polygamus (Cav.) Cabrera leaves prevents scopolamine-induced memory impairment and brain oxidative stress in zebrafish (Danio rerio)," *Phytomedicine*, vol. 58, Article ID 152889, 2019.
- [32] E. D. Levin, Z. Bencan, and D. T. Cerutti, "Anxiolytic effects of nicotine in Zebrafish," *Physiology & Behavior*, vol. 90, no. 1, pp. 54–58, 2007.
- [33] F. L. A. Batista, L. M. G. Lima, I. A. Abrante et al., "Antinociceptive activity of ethanolic extract of *Azadirachta indica* A. Juss (Neem, Meliaceae) fruit through opioid, glutamatergic and acid-sensitive ion pathways in adult zebrafish (*Danio rerio*)," *Biomedicine & Pharmacotherapy*, vol. 108, pp. 408–416, 2018.
- [34] B. Srikumar, K. Ramkumar, T. Raju, and B. Shankaranarayana Rao, "Assay of acetylcholinesterase activity in the brain," in *Brain and Behavior*, T. Raju, B. Kutty, T. Sathyaprabha, and B. Shanakranarayana Rao, Eds., pp. 142–144, National Institute of Mental Health and Neurosciences, Bangalore, India, 2004.
- [35] C. Winterbourn, R. Hawkins, M. Brian, and R. Carrell, "The estimation of red cell superoxide dismutase activity," *Journal* of Laboratory and Clinical Medicine, vol. 85, no. 2, p. 337, 1975.
- [36] A. K. Sinha, "Colorimetric assay of catalase," Analytical Biochemistry, vol. 47, pp. 389–394, 1972.
- [37] M. Sharma and Y. K. Gupta, "Chronic treatment with trans resveratrol prevents intracerebroventricular streptozotocin induced cognitive impairment and oxidative stress in rats," *Life Sciences*, vol. 71, no. 21, pp. 2489–2498, 2002.
- [38] P. K. Smith, R. I. Krohn, G. T. Hermanson et al., "Measurement of protein using bicinchoninic acid," *Analytical Biochemistry*, vol. 150, no. 1, pp. 76–85, 1985.
- [39] L. B. Lima, C. F. B. Vasconcelos, H. M. L. Maranhão et al., "Acute and subacute toxicity of *Schinus terebinthifolius* bark extract," *Journal of Ethnopharmacology*, vol. 126, no. 3, pp. 468–473, 2009.
- [40] C. R. G. Affonso, R. M. Fernandes, J. M. G. de Oliveira et al., "Effects of the essential oil from fruits of *Schinus ter-ebinthifolius* Raddi (Anacardiaceae) on reproductive functions in male rats," *Journal of the Brazilian Chemical Society*, vol. 23, pp. 180–185, 2012.
- [41] E. R. Cole, R. B. dos Santos, V. Lacerda Júnior, J. D. L. Martins, S. J. Greco, and A. Cunha Neto, "Chemical composition of essential oil from ripe fruit of *Schinus terebinthifolius* Raddi and evaluation of its activity against wild strains of hospital origin," *Brazilian journal of microbiology*, vol. 45, no. 3, pp. 821–828, 2014.
- [42] A. J. Maciel, C. P. Lacerda, L. J. Danielli, S. A. L. Bordignon, A. M. Fuentefria, and M. A. Apel, "Antichemotactic and

antifungal action of the essential oils from *Cryptocarya* aschersoniana, Schinus terebinthifolia, and Cinnamomum amoenum," Chemistry and Biodiversity, vol. 16, no. 8, pp. 597–604, Article ID e1900204, 2019.

- [43] A. Ennigrou, H. Casabianca, E. Vulliet, B. Hanchi, and K. Hosni, "Assessing the fatty acid, essential oil composition, their radical scavenging and antibacterial activities of *Schinus terebinthifolius* Raddi leaves and twigs," *Journal of Food Science and Technology*, vol. 55, no. 4, pp. 1582–1590, 2018.
- [44] M. R. F. de Lima, J. de Souza Luna, A. F. dos Santos et al., "Anti-bacterial activity of some Brazilian medicinal plants," *Journal of Ethnopharmacology*, vol. 105, no. 1, pp. 137–147, 2006.
- [45] Q. Xu, Y. Park, X. Huang et al., "Physical activities and future risk of Parkinson disease," *Neurology*, vol. 75, no. 4, pp. 341–348, 2010.
- [46] F. Afifi, N. H. Aboalhaija, O. Awwad, E. Khalil, R. Abbassi, and I. F. Abaza, "Chemodiversity and antiproliferative activity of the essential oil of *Schinus molle L.* growing in Jordan," *Chemistry and Biodiversity*, vol. 16, Article ID e1900388, 2019.
- [47] G.-Y. Lee, C. Lee, G. H. Park, J.-H. Jang, Amelioration of scopolamine-induced learning and memory impairment by α-pinene in C57BL/6 mice," *Evidence-Based Complementary* and Alternative Medicine, vol. 2017, Article ID 4926815, 9 pages, 2017.
- [48] H. D. A. S. Siqueira, B. S. Neto, D. P. Sousa et al., " α -Phellandrene, a cyclic monoterpene, attenuates inflammatory response through neutrophil migration inhibition and mast cell degranulation," *Life Sciences*, vol. 160, pp. 27–33, 2016.
- [49] D.G. Machado, M. P. Kaster, R. W. Binfaré et al., "Antidepressant-like effect of the extract from leaves of *Schinus molle* L. in mice: evidence for the involvement of the monoaminergic system," *Progress in Neuro-Psychopharmacology* and Biological Psychiatry, vol. 31, no. 2, pp. 421–428, 2007.
- [50] A. C. Piccinelli, J. A. Santos, E. C. Konkiewitz et al., "Antihyperalgesic and antidepressive actions of (R)-(+)-limonene, α-phellandrene, and essential oil from Schinus terebinthifolius fruits in a neuropathic pain model," *Nutritional Neuroscience*, vol. 18, no. 5, pp. 217–224, 2015.
- [51] X.-W. Jiang, J.-P. Bai, Q. Zhang et al., "Caffeoylquinic acid derivatives protect SH-SY5Y neuroblastoma cells from hydrogen peroxide-induced injury through modulating oxidative status," *Cellular and Molecular Neurobiology*, vol. 37, no. 3, pp. 499–509, 2017.
- [52] P. . d.S. . d. Rocha, J. F. Campos, V. Nunes-Souza et al., "Antioxidant and protective effects of Schinus terebinthifolius Raddi against doxorubicin-induced toxicity," *Applied Biochemistry and Biotechnology*, vol. 184, no. 3, pp. 869–884, 2018.
- [53] A. Murray, M. Gurovic, S. Rodriguez, M. Murray, and A. Ferrero, "Acetylcholinesterase inhibition and antioxidant activity of essential oils from *Schinus areira* L. and *Schinus longifolia* (Lindl.) Speg.," *Natural Product Communications*, vol. 4, no. 6, pp. 873–876, 2009.
- [54] Z. Merali, C. Cayer, P. Kent et al., "Sacred Maya incense, copal (Protium copal - burseraceae), has antianxiety effects in animal models," *Journal of Ethnopharmacology*, vol. 216, pp. 63–70, 2018.
- [55] H. Javed, M. M. Khan, A. Khan et al., "S-allyl cysteine attenuates oxidative stress associated cognitive impairment and neurodegeneration in mouse model of streptozotocin-induced experimental dementia of Alzheimer's type," *Brain Research*, vol. 1389, pp. 133–142, 2011.

- [56] X. Zhang, A. Zhang, B. Jiang, Y. Bao, J. Wang, and L. An, "Further pharmacological evidence of the neuroprotective effect of catalpol from Rehmannia glutinosa," *Phytomedicine*, vol. 15, no. 6, pp. 484–490, 2008.
- [57] M. M. d. Silva, E. K. K. Iriguchi, C. A. L. Kassuya et al., "Schinus terebinthifolius: phenolic constituents and in vitro antioxidant, antiproliferative and in vivo anti-inflammatory activities," *Revista Brasileira de Farmacognosia*, vol. 27, pp. 445–452, 2017.
- [58] H. Haddadi, Z. Rajaei, H. Alaei, and S. Shahidani, "Chronic treatment with carvacrol improves passive avoidance memory in a rat model of Parkinson's disease," *Arquivos de Neuro-Psiquiatria*, vol. 76, no. 2, pp. 71–77, 2018.
- [59] A. A. C. de Almeida, R. B. F. de Carvalho, O. A. Silva, D. P. de Sousa, and R. M. de Freitas, "Potential antioxidant and anxiolytic effects of (+)-limonene epoxide in mice after marble-burying test," *Pharmacology Biochemistry and Behavior*, vol. 118, pp. 69–78, 2014.
- [60] F. A. Alzate G, J. A. Gil Q, N. d. S. Jiménez U, G. J. Arango A, and B. Weniger, "Antioxidant potential of some species of the genus *Bomarea* (Alstroemeriaceae)," *Vitae*, vol. 18, pp. 201– 207, 2011.