



The role of nitric oxide in pre-synaptic plasticity and homeostasis

Neil Hardingham[‡], James Dachtler^{†‡} and Kevin Fox^{*}

School of Biosciences, Cardiff University, Cardiff, UK

Edited by:

Jaichandar Subramanian,
Massachusetts Institute of
Technology, USA

Reviewed by:

Joern R. Steinert, University of
Leicester, UK
Bernardo Moreno-López, University
of Cadiz, Spain

*Correspondence:

Kevin Fox, School of Biosciences,
Cardiff University, Museum Avenue,
Cardiff CF10 3AX, UK
e-mail: foxkd@cardiff.ac.uk

† Present address:

James Dachtler, School of
Biomedical Sciences, University of
Leeds, West Yorkshire, UK

[‡] These authors are joint first
authors.

Since the observation that nitric oxide (NO) can act as an intercellular messenger in the brain, the past 25 years have witnessed the steady accumulation of evidence that it acts pre-synaptically at both glutamatergic and GABAergic synapses to alter release-probability in synaptic plasticity. NO does so by acting on the synaptic machinery involved in transmitter release and, in a coordinated fashion, on vesicular recycling mechanisms. In this review, we examine the body of evidence for NO acting as a retrograde factor at synapses, and the evidence from *in vivo* and *in vitro* studies that specifically establish NOS1 (neuronal nitric oxide synthase) as the important isoform of NO synthase in this process. The NOS1 isoform is found at two very different locations and at two different spatial scales both in the cortex and hippocampus. On the one hand it is located diffusely in the cytoplasm of a small population of GABAergic neurons and on the other hand the alpha isoform is located discretely at the post-synaptic density (PSD) in spines of pyramidal cells. The present evidence is that the number of NOS1 molecules that exist at the PSD are so low that a spine can only give rise to modest concentrations of NO and therefore only exert a very local action. The NO receptor guanylate cyclase is located both pre- and post-synaptically and this suggests a role for NO in the coordination of local pre- and post-synaptic function during plasticity at individual synapses. Recent evidence shows that NOS1 is also located post-synaptic to GABAergic synapses and plays a pre-synaptic role in GABAergic plasticity as well as glutamatergic plasticity. Studies on the function of NO in plasticity at the cellular level are corroborated by evidence that NO is also involved in experience-dependent plasticity in the cerebral cortex.

Keywords: LTP (Long Term Potentiation), synaptic plasticity, NOS1, experience-dependent plasticity, guanylate cyclase

INTRODUCTION

Nitric oxide is a ubiquitous signaling molecule in the brain and in other organs of the body. It is involved in an almost bewildering array of functions. Consequently, there have been many reviews over the years that have described its role in retrograde signaling (Brenman and Bredt, 1997), cellular function (Garthwaite, 2008), synaptic plasticity (Holscher, 1997), development (Contestabile, 2000), excitotoxicity (Calabrese et al., 2007), blood flow (Gordon et al., 2007) and mental health (Steinert et al., 2010). However, in this review we focus on the role of NO in synaptic plasticity and specifically its function as a retrograde messenger. It seems fitting to look at the evidence now as it is 25 years since the original discovery that NO (or endothelial derived relaxing factor) might act as an intercellular messenger in the brain (Garthwaite et al., 1988), during which time there has been a steady accumulation of evidence for the role of NO synthase in synaptic plasticity and homeostasis at both excitatory and inhibitory synapses. In the following sections we briefly review the main pathways by which NO acts and the distance over which it acts, before discussing the evidence for its role in synaptic signaling during plasticity and homeostasis.

MOLECULAR PATHWAYS FOR THE ACTION OF NO

Nitric oxide is generated by the enzyme NO synthase (NOS). NOS1 (nNOS or neuronal NOS) is one of three major isoforms of NO synthase, the others being NOS2 (iNOS or inducible NOS) and NOS3 (eNOS or endothelial NOS). Many cell types in the body can express NOS2, including immune response cells (Hickey, 2001), glial cells (Nomura and Kitamura, 1993) and neurons (Corsani et al., 2008). Unlike NOS1 and NOS3 that are expressed constitutively, NOS2 is induced by inflammatory cytokines (Saha and Pahan, 2006). Calcium/calmodulin has such a high affinity for NOS2 that its activity is not modulated by this route, which means that NOS2 activity is under the control of cytokines rather than calcium signaling. Antagonists of NOS2 have been reported to reduce synaptic plasticity and alter both spontaneous and evoked synaptic activity in the cortex (Buskila and Amitai, 2010), although NOS1 may also have been affected at the drug concentrations used in this study.

NOS3 was originally isolated from endothelial cells, and along with other NOS isoforms is present in the tissues of the cardiovascular system (Buchwalow et al., 2002). While early reports suggested NOS3 was located in neurons (Dinerman et al., 1994),

these findings were later rebutted by the same group (Blackshaw et al., 2003). NOS1 knockouts show that NOS1 is the source of 95% of the NO in the cortex (Huang et al., 1993) and plays a major role in synaptic plasticity (see Section NO Controls Pre-Synaptic Function and The Role of NO in Plasticity). However, tonic levels of NO produced by NOS3 may also play a role in the induction of plasticity (Hopper and Garthwaite, 2006).

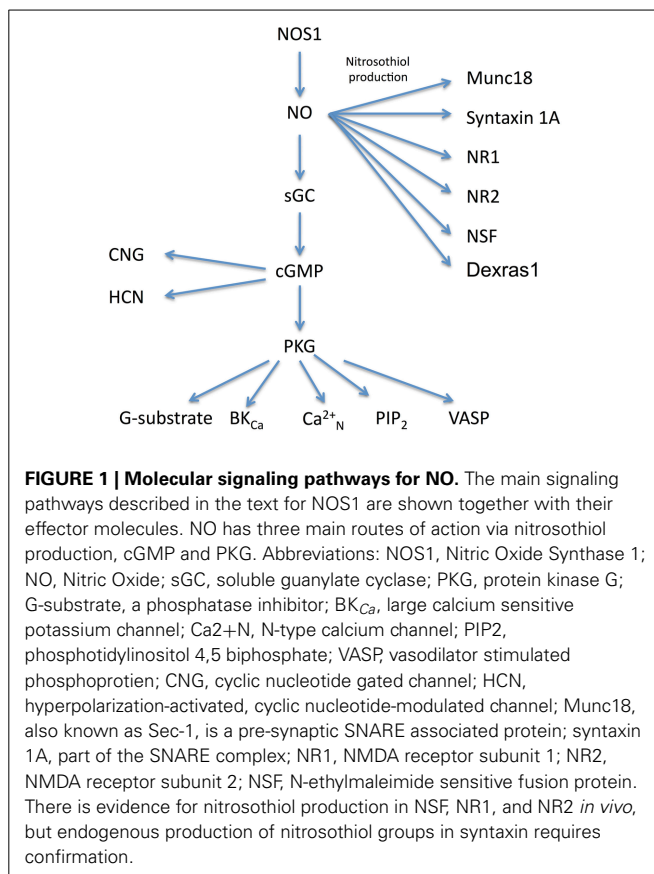
SOLUBLE GUANYLATE CYCLASE

Soluble guanylyl cyclase (sGC) is the most sensitive receptor for NO, with an EC_{50} in the low nanomolar (nM) range (Roy et al., 2008). A good deal of evidence has been gathered in recent years for its importance in mediating the actions of endogenous NO, predominantly at pre-synaptic locations (Garthwaite, 2010; Neitz et al., 2011; Eguchi et al., 2012; Bartus et al., 2013).

Soluble guanylyl cyclase mediates the production of cGMP from GTP. Three subunits of the protein have been identified, α_1 , α_2 , and β_1 . A functional receptor is a heterodimer consisting of one α and one β subunit. Two isoforms of the receptor exist ($\alpha_1\beta_1$ and $\alpha_2\beta_1$) with a complex regional expression. For example, the $\alpha_1\beta_1$ heteromer is dominant in the caudate-putamen and nucleus accumbens whilst $\alpha_2\beta_1$ is dominant in the hippocampus and olfactory bulb (Gibb and Garthwaite, 2001; Mergia et al., 2003). The $\alpha_2\beta_1$ receptor is present at the highest levels in the brain and the α_2 subunit has been shown to bind to the cell membrane through PSD95 (Russwurm et al., 2001; Mergia et al., 2003), which suggests a post-synaptic localization. The $\alpha_2\beta_1$ isoform can substitute for most functions of the more widely expressed $\alpha_1\beta_1$ isoform despite there being a 90% reduction in sGC in the α_1 KOs (Friebe and Koesling, 2009). However, deletion of the β_1 subunit eliminates expression of any sGC resulting in an 80% infant mortality within 2 days of birth (Friebe and Koesling, 2009). To date, the two α subunit isoforms have only been found to have distinct functions in the induction of LTP in the visual cortex where both isoforms are necessary (Haghikia et al., 2007).

The guanylyl cyclase receptor consists of a haem group of the type that binds O_2 in hemoglobin, but when associated with the receptor protein, it exhibits a substantial preference for NO, allowing detection of NO in the presence of at least 10,000 fold excess of O_2 , despite the molecular similarity of the two ligands (Martin et al., 2006).

The mechanism of activation of sGC by NO is complex and involves a conformational change via binding at the haem site, which enables increased conversion of GTP to cGMP (Roy et al., 2008). NO activates guanylyl cyclase within 20 ms and, following removal of NO, activity decays with a half life of 200 ms (Bellamy and Garthwaite, 2001). With formation of cGMP, a bifurcation occurs in the route of action (Figure 1); one route is for cGMP to affect cGMP-activated protein kinases (cGKs or PKGs). Multiple substrates for PKG have been identified including PKG activated phosphatases, leading indirectly to altered levels of phosphorylation of effector proteins (Schlossmann and Hofmann, 2005). The second major route of action for cGMP is to bind to agonist or regulatory sites on cyclic nucleotide-gated (CNG) ion channels or hyperpolarization-activated, cyclic nucleotide-modulated (HCN) channels.



PRODUCTION OF NITROSO THIOL GROUPS

There are a number of cases where NO signaling in the brain is transduced in a cGMP independent manner. The thiol side chains of cysteine residues in proteins can be modified by the addition of an NO group and this outcome could occur by two known routes: the thiol group can be oxidized to a thyl followed by addition of NO, which is known as oxidative nitrosylation, or NO can react with O_2 to produce N_2O_3 which then interacts with the thiol group to produce nitrosothiol, and this process is known as nitrosation (Heinrich et al., 2013). At present, the endogenous route for nitrosothiol production is not known.

A number of pre-synaptic proteins have been identified as potential targets for nitrosothiol production and therefore as a mechanism for mediating alterations in pre-synaptic strength (Figure 1). The t-snare protein synapsin has been identified as a target for nitrosothiol production in pancreatic cells (Wiseman et al., 2011) and syntaxin 1a and n-sec1 (also known as Munc18) have been shown to be a target for nitrosothiol production in neurons (Meffert et al., 1996; Prior and Clague, 2000; Palmer et al., 2008). A small GTPase known as Dexas1 (which can be induced by dexamethasone) is held in close proximity to NOS1 by CAPON (Jaffrey et al., 1998) and can be modified by production of nitrosothiol (Fang et al., 2000).

Nitrosothiol production requires much higher concentrations of NO than activation of sGC and proceeds with slower kinetics. For example, nitrosothiol production in syntaxin 1A occurs with an IC_{50} of 1.1 μ M NO (Palmer et al., 2008) compared with

the nM range of detection for sGC (Roy et al., 2008). It has been estimated that an NO concentration of 200 μ M would require 2 min to produce nitrosothiol groups in half the substrate (Ahern et al., 2002). The high concentrations and slow reaction kinetics of nitrosothiol production raise the question of whether it can occur naturally. Most of the experiments conducted on production of nitrosothiol groups in various proteins use NO donors at quite high levels [for example 100–1000 μ M for nitrosothiol production in SNAP25 (Di Stasi et al., 2002)]. However, a technique for detecting nitrosothiol groups in proteins known as the biotin switch method has been used to demonstrate the existence of endogenous nitrosothiol groups *in vivo* by comparing results in wild-type mice with NOS1 knockout mice (Jaffrey et al., 2001). The synaptic proteins that appear to have endogenous nitrosothiol groups using this method include NR1, NR2A (Jaffrey et al., 2001), and NSF (Huang et al., 2005).

It may not be coincidental that some of the molecules shown to have nitrosothiol groups *in vivo* are held in close proximity to NOS1 and thereby experience the higher source concentrations of NO. The NMDA receptor is local to NOS1 by virtue of them both binding to PSD95 and dexas1 is close to NOS1 because both bind to CAPON (Fang et al., 2000). It may also be relevant that nitrosothiol groups occur on molecules that tend to lie close to lipid membranes, in this case synaptic membranes. It has been suggested that the kinetics of the reaction between NO and O₂ to produce N₂O₃ could be increased by NO and O₂ becoming concentrated in lipid membranes (Heinrich et al., 2013). However, once again it should be emphasized that the endogenous routes for generating nitrosothiol groups on proteins are not known at present.

THE CELLULAR LOCATION OF NOS1

NOS1 is composed of several splice variants. The long form of NOS is α NOS1 which contains a PDZ binding domain that enables it to bind to the PDZ2 domain of PSD95 (Brenman et al., 1996; Eliasson et al., 1997) localizing NOS1 to the post-synaptic density (see Doucet et al., 2012). There are also shorter splice variants of NOS1 lacking the PDZ domain known as β NOS1 and γ NOS1. While the latter is not expressed very highly in the brain, β NOS1 is expressed quite highly in the ventral cochlear nuclei, the striatum and the lateral tegmental nuclei (Eliasson et al., 1997). In the cortex and hippocampus, the current evidence suggests that NOS1 is located in two very different neuronal compartments in two different cell types. On the one hand, NOS1 is located in the cytoplasm of a small subpopulation of GABAergic cells in the cortex and hippocampus and on the other, it is located in a far larger population of excitatory neurons, but highly restricted to the spine head. The ease with which NOS1 can be detected at the two locations depends on the techniques used as described below.

LIGHT MICROSCOPY

The light microscopy (LM) level is sufficient to demonstrate the presence of cytoplasmic NOS1 (Eliasson et al., 1997; Blackshaw et al., 2003; Kubota et al., 2011). LM antibody studies have shown that the strongest NOS1 staining in the neocortex and hippocampus occurs in a small subpopulation of GABAergic neurons

(Wendland et al., 1994; Aoki et al., 1997; Blackshaw et al., 2003) that co-express Somatostatin, Neuropeptide Y and the Substance P receptor (Kubota et al., 2011). The NOS1⁺ GABAergic neurons contain both α NOS1 and β NOS1. A significant component of the cytoplasmic staining is attributable to β NOS1 as it persists in α NOS1 knockouts (Eliasson et al., 1997). Weaker labeling of the cortical neuropil is also consistently reported in the same papers. Recent studies using targeted knockin of cre-recombinase into the NOS1 gene and subsequent crosses to GFP reporter lines clearly show two populations of NOS1⁺ GABAergic cells, one of neurogliaform morphology (type II) and the other characterized by long range axonal projections (type I) (Taniguchi et al., 2011). Again the neuropil can be seen throughout the cortical layers including clear axonal labeling (Figure 2). Pyramidal cell labeling is not seen in these cre lines, however, possibly due to the technique only showing high levels of NOS1 expression (Josh Huang personal communication). Weak labeling of CA1 pyramidal cells can be seen using NOS1 antibodies with the right fixative conditions (Burette et al., 2002; Blackshaw et al., 2003) and colocalization of NMDA, PSD95, and NOS1 shows that some of the punctate labeling seen with LM is due to NOS1 in spines (Burette et al., 2002).

ELECTRON MICROSCOPY

Using electron microscopy (EM), much of the neuropil labeling present in LM studies can be seen to reside in the axons of NOS⁺ GABAergic neurons (Aoki et al., 1997). However, EM studies reveal a further component of the neuropil labeling to be due to the very precise and restricted localization of NOS1 in spines, spine heads, and occasionally the plasma membrane of dendrites

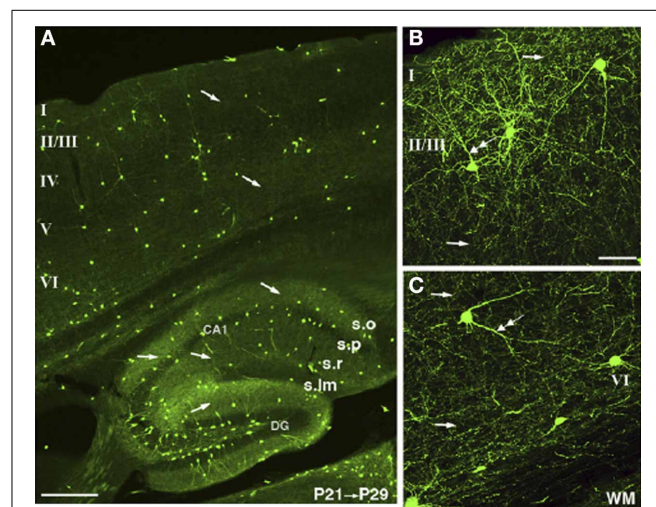


FIGURE 2 | NOS1 positive cells at the LM level in the Cortex and Hippocampus. Cells expressing TdTomato fluoresce in nNOS positive cells in an nNOS-CreER;Ai9 mouse. The TdTomato is rendered green in the images. **(A)** The nNOS positive cells make up a small population scattered in cortex and hippocampus. **(B)** A dense and diffuse plexus of neuropil can be seen throughout layer II/III and **(C)** throughout deeper layers of the cortex. Single arrows indicate axons and double arrows dendrites. Adapted from Taniguchi et al. (2011) with kind permission of the authors and Cell press.

(Aoki et al., 1997, 1998). The NOS1 visible in the heads of spines in the visual cortex and in some cases at the base of spines accounts for 30–75% of the punctate labeling in cortical electron micrographs (**Figure 3**). Although the NOS1⁺ GABAergic neurons are sparsely spiny and could theoretically account for some of the NOS1 spine labeling, the extent of the spine labeling seen in EM is too great to be due purely to GABAergic cells (Cheri Aoki personal communication); therefore a considerable amount of spine labeling must be attributable to excitatory pyramidal cells. Furthermore, the NOS1 labeling in spines is quite distinctive in that the labeled spines are joined to dendrites that do not contain NOS1 labeling (**Figure 3**); if these spines were located on GABAergic cells, the cytoplasm would be labeled as well. EM studies of cortical synapses also show that the gold particle distribution associated with NOS1 labeling is coextensive with that for PSD95 relative to the plasma membrane (Valtschanoff and Weinberg, 2001). Similarly, in the hippocampus, EM studies show that NOS1 is located in dendritic spines on pyramidal cells (Burette et al., 2002). The NO receptor sGC is found pre-synaptic and within 50–150 nm of the NOS (**Figure 4**). In conclusion, pyramidal cells in the neocortex and hippocampus contain NOS1 that is highly localized to the spine head, spine neck, or plasma membrane of the dendrites and is closely apposed to pre-synaptic sGC.

MOLECULAR AND FUNCTIONAL METHODS

The reason why NOS1 is localized to the spine head is due to the nature of the alpha sub-isoform of NOS1 which contains a PDZ binding domain that enables it to bind to the PDZ2 domain of PSD95 (Brenman et al., 1996; Eliasson et al., 1997). Using proteomic analysis of molecules associated with the NMDA receptor, it has been shown that NOS1 is part of the NMDA signaling complex (Husi et al., 2000). The authors used a combination of immunoaffinity chromatography, immunoprecipitation with an antibody directed against the NR1 subunit, and peptide affinity based on the structure of the NR2B subunit C terminus that binds to the NMDAR-binding protein PSD-95. The structure and binding partners of NOS1 and PSD95 are reviewed in (Zhou and Zhu, 2009) and (Doucet et al., 2012).

Functional assays also demonstrate the synaptic location of NOS1. The functional consequences of disrupting the interaction between NOS1 and PSD95 has been studied by expressing decoy proteins that code for amino acids constituting the PDZ binding domain of α NOS1. For example, glutamate induced activation of p38 normally leads to excitotoxic cell death, but this process can be prevented by expression of the first 300 amino acids of NOS1 (NOS1_{1–300}) (Cao et al., 2005). Similarly, cerebral ischemia induced by cerebral artery occlusion leads to cortical damage which can be reduced by

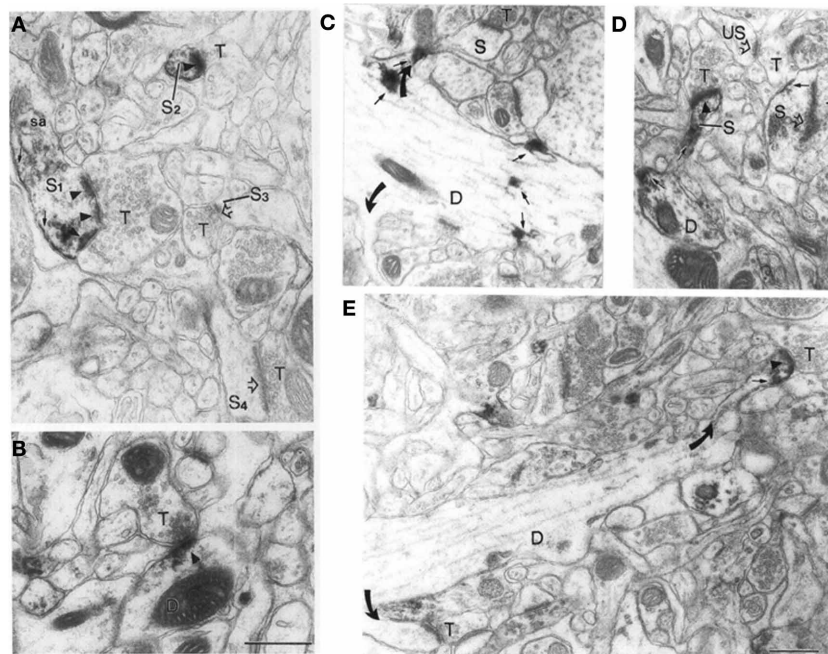
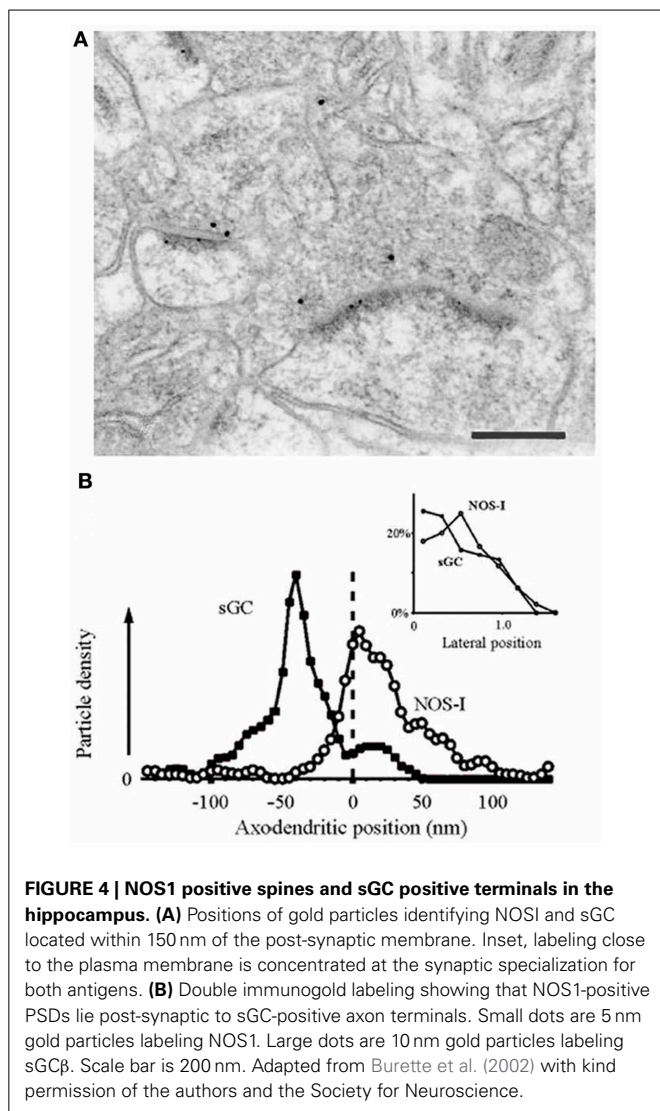


FIGURE 3 | NOS1 positive spines at the EM level in the Visual Cortex. (A) Large dendritic spine (S1) with a perforated PSD showing NOS1-immunoreactivity (arrowheads). NOS1 immunoreactivity is also present along the plasma membrane (small arrow) and near the spine apparatus (sa). A second small spine (S2) shows NOS1 immunoreactivity along the plasma membrane and over the PSD. Not all spines are labeled (S3 and S4). T represents unlabeled pre-synaptic terminals. Open arrows mark unlabeled PSDs. **(B)** Axodendritic synapse showing NOS1 labeling of a PSD (arrowhead). **(C)** NOS1 labeling occurs at the spine base (upper curved arrow) and dendritic

shaft (small arrows). Lower curved arrow points to an unlabeled spine. S is a spine head and D is a dendritic shaft where limited NOS1 labeling occurs along the plasma membrane. **(D)** NOS1 immunoreactivity over the spine neck (S), plasma membrane forming the spine head (small arrow) and the PSD (filled arrowhead). US marks an unlabeled spine and open arrowheads also mark unlabeled spines and T is the pre-synaptic terminal. **(E)** NOS1 immunoreactivity only in the spine head. Note that in all these cases there is no labeling of the dendritic cytoplasm. Calibration bar = 500 nm. Adapted from Aoki et al. (1998) with kind permission of the author and Elsevier press.



NOS1_{1–133} (Zhou et al., 2010) and pTAT-PDZ1-2 (Aarts et al., 2002). Thermal hyperalgesia and chronic mechanical allodynia can be inhibited by intrathecal application of IC8731 or tat-NOS1 (NOS1 1-299) (Florio et al., 2009). These molecules do not act to reduce the enzymatic activity of NOS1, but rather to decrease the coupling between NOS1 and NMDA receptors by disrupting the ability of NOS1 to bind to PSD95 (Florio et al., 2009).

Finally, studies on synaptic plasticity (as described in Section The Role of NO in Plasticity), show that the pre-synaptic NO-dependent component of LTP can be prevented by post-synaptic application of NOS antagonists to layer 2/3 pyramidal cells in the somatosensory cortex (Hardingham and Fox, 2006). Similar results have been demonstrated for layer 5 cortical cells (Sjostrom et al., 2007). This implies that the NO synthase exists in pyramidal cells in the cortex.

In conclusion, LM studies are able to demonstrate the presence of NOS1 in the NOS⁺ GABAergic cells of the neocortex and hippocampus but LM is at the limit for demonstrating its presence in

pyramidal cells, while EM, proteomic, and functional analysis are sensitive enough to demonstrate the presence of NOS1 at spines of pyramidal cells.

THE PHYSIOLOGICAL CONCENTRATION OF NO AND ITS DISTANCE OF ACTION

A theoretical consideration of the rate of production of NO at an individual synapse suggests that NO has a source concentration in the low nanomolar range. Working forward from a knowledge of the rate of NO production per NOS molecule *in vitro* of 20 per second (Santolini et al., 2001) and using an estimate of the number of NMDA receptors and therefore NOS molecules present at a single post-synaptic density, the concentration in the immediate vicinity of the NOS molecule can be estimated at 2.5 nM, falling 10 fold within approximately 700 nm (Hall and Garthwaite, 2009). Working backwards from a measure of NO concentration generated in a cerebellar slice stimulated with NMDA gives a similar rate of production of NO per NOS molecule (10 per second) and a source concentration at the synapse of approximately 0.01–0.1 nM (Wood et al., 2011). A number of studies have reported that NO is produced in the brain in the picomolar range (Wakatsuki et al., 1998; Sato et al., 2006; Wood et al., 2011) and several other labs in the low nM range (<10 nM) (Shibuki and Kimura, 1997; Kimura et al., 1998; Wu et al., 2001, 2002; Sammut et al., 2006, 2007a,b; Ondracek et al., 2008; Sammut and West, 2008).

If the concentration of NO produced at a synapse is in the pM to low nM range, then the rate of inactivation of NO with distance in the brain implies that it can only act over a relatively short range. The most sensitive target for NO is soluble guanylate cyclase (sGC), which can respond to as little as 1 pM NO (Batchelor et al., 2010). The EC₅₀ of sGC to NO is thought to be in the low nanomolar range at 1.7 nM (Griffiths et al., 2003). Physiological concentrations of ATP (1 mM) and GTP (0.1 mM), which antagonistically decrease and increase the sensitivity of sGC to NO, respectively, elevate the EC₅₀ to 3.4 nM (Roy et al., 2008). Taking into account both the likely concentration of NO at the synapses and the sensitivity of sGC suggests that NO is only likely to act over distances of less than 1 micron.

The lower estimate of NO evolution in the picomolar range would sit on the non-linear cusp of the NO/sGC binding curve (Roy et al., 2008). This raises the interesting possibility that a tonic level of NO production could interact with the NMDA receptor activated NO concentration to boost its effect on sGC. For example, a tonic level of 250 pM NO would move the operating point of the synapse onto the linear part of the NO/sGC curve [see Figure 7B of Roy et al. (2008)]. There is evidence for a tonic level of NO production in the brain originating from both NOS3 and NOS1 (Hopper and Garthwaite, 2006; Dachtler et al., 2011). Furthermore, tonic levels of NO have been found to influence the magnitude of LTP, giving further credence to this notion. NO donors can be shown to facilitate both post-synaptic potentials and LTP (Bohme et al., 1991; Malen and Chapman, 1997; Hardingham and Fox, 2006). The higher estimate of NO release in the nM range would not require background levels of NO to move sGC on to the linear part of its response curve. With either mode of action, NO would only be able to act over a distance of

less than about 1 micron, effectively making it a synapse specific signal.

The view of NO as a synapse specific signal does not fit with the notion of NO as a volume transmitter. Nevertheless, there is evidence for NO acting as a volume transmitter in the Calyx of Held (Steinert et al., 2008). Theoretically, all that would be required for higher concentrations of NO would be higher concentrations of the enzyme NOS. It is conceivable that the GABAergic inhibitory cells that express NOS1 at much higher levels than excitatory cells (**Figure 2**) throughout their cytoplasm could provide such a source. The NOS1⁺ GABAergic cells produce a plexus of fine NOS positive fibers that ramify throughout the cortex and hippocampus, which could aid spatial summation of NO levels. However, little is known of NO release from this small subpopulation of cells at present.

NO CONTROLS PRE-SYNAPTIC FUNCTION

The past two decades have seen a steady but decisive accumulation of evidence showing not only that NO acts pre-synaptically on neurotransmitter release, but how it does so (Feil and Kleppisch, 2008). **Table 1** is a compilation of papers showing some of the evidence for NO's pre-synaptic action, its retrograde route from post- to pre-synaptic site and its pre-synaptic action in plasticity.

Much of the detailed evidence for NO's role in transmitter release comes from studies on the glutamatergic system, but a body of work implicates NO in regulating transmitter release from GABAergic (Kawaguchi et al., 1997; Li et al., 2002; Moreno-Lopez et al., 2002; Wall, 2003; Szabadits et al., 2007; Yang et al., 2007; Bright and Brickley, 2008; Xue et al., 2011; Lange et al., 2012) dopaminergic (West et al., 2002) and noradrenergic synapses (Montague et al., 1994; Kodama and Koyama, 2006).

A number of the studies providing evidence for the retrograde action of NO have come from cell cultures. Cell culture preparations have a number of technical advantages that allow the retrograde action of NO to be demonstrated (**Table 1**). However, since cells in culture are immature, it raises the question of whether NO acts the same way in more mature cells. Nevertheless, a number of studies made on mature neurons in intact slices of hippocampus (O'Dell et al., 1991; Schuman and Madison, 1991), amygdala (Lange et al., 2012), neocortex (Hardingham and Fox, 2006; Sjostrom et al., 2007), the medial nucleus of the trapezoid body (Steinert et al., 2008; Eguchi et al., 2012), cerebellum (Qiu and Knopfel, 2007), and the ventral lateral medulla (Huang et al., 2003), lead to similar conclusions about the action of NO in mature cells, suggesting that NO retains its retrograde pre-synaptic action into adulthood.

In the following sections we briefly review the findings for NO's effects on four aspects of pre-synaptic function; actions at the active zone, on vesicle recycling, effects on the readily releasable pool and actions on pre-synaptic growth. When viewed in combination, these studies suggest that NO may regulate pre-synaptic release by acting in a coordinated and synergistic manner on several aspects of pre-synaptic release (**Figure 5**).

EFFECTS ON THE ACTIVE ZONE AND TRANSMITTER RELEASE

Nitric oxide can affect transmitter release by nitrosothiol generation in a number of constituents of the active zone (**Figure 5B**).

For example, nitrosothiol production in syntaxin at Cys(145) has a facilitatory effect on release because it prevents munc18 (also known as n-sec1) from binding to the closed conformation of syntaxin 1a. This allows syntaxin1a to unfold and bind to both VAMP on the vesicle and SNAP25 at the release site, which in turn enables the vesicle to dock to the membrane (Meffert et al., 1996; Palmer et al., 2008). SNAP25 can itself have nitrosothiol groups generated by NO, which may further enhance release (Di Stasi et al., 2002). However, it is not clear at present whether the concentrations of NO necessary for production of nitrosothiol groups are realized at the synapse (see Sections The Cellular Location of NOS1 and The Physiological Concentration of NO and Its Distance of Action).

EFFECTS ON ION CHANNELS

Voltage gated ion channels that reside in the pre-synaptic terminal and affect transmitter release have been shown to be NO sensitive. In the peptidergic synapse of the pituitary nerve, NO can increase pre-synaptic release by enhancing the activity of large conductance Ca²⁺ activated K⁺ channels (BK). PKG only activates BK at depolarized potentials, which means that the action potential after-hyperpolarization becomes larger without affecting the spike threshold. Consequently, during prolonged trains of action potentials, the enhanced hyperpolarization provided by BK channels accelerates Na⁺ channel recovery (Klyachko et al., 2001). It can be demonstrated that cytosolic calcium almost doubles in the presence of exogenous cGMP. A possible physiological role for this action is suggested by showing that the action potential success rate during a 25 Hz stimulus train is almost twice as great in the control condition when compared to that in the presence of the NO synthase inhibitor 7-NI or the sGC inhibitor ODQ (Klyachko et al., 2001). In the brainstem, synaptic potentials generated by glutamatergic synapses in the ventrolateral medulla can be enhanced by application of the NOS substrate L-arginine (200 μM) (Huang et al., 2003). This effect can be shown to be due to NO acting via a cGMP/protein kinase G-dependent pathway on N-type calcium channels (Huang et al., 2003). It is not known at present whether BK or N-type calcium channels are affected by NO in the cortex or hippocampus.

One other means by which NO may affect transmitter release in some types of neuron is by stimulating the production of cGMP, which directly gates cyclic nucleotide gated channels (Neitz et al., 2011). Cyclic nucleotide gated (CNG) channels are well known for their function in transmitter release in some classes of cell, for example photoreceptors (Rieke and Schwartz, 1994) and olfactory epithelial cells (Leinders-Zufall et al., 2007). However, the distribution of CNG channels is more widespread and roughly mirrors the distribution of the NO/cGMP system (Kingston et al., 1999). For example, CNG channels are present in the rat hippocampus (Kingston et al., 1999) and may be involved in the induction of theta burst LTP in mouse hippocampus (Parent et al., 1998). While native heteromeric CNG channels formed by alpha and beta subunits are gated by cGMP, homomeric channels comprising just the beta subunit are directly activated by NO (Broillet and Firestein, 1997), raising the possibility that NO might act on native CNG channels

Table 1 | Evidence that nitric oxide influences presynaptic function.

References	Title	Presynaptic action?	Retrograde messenger?	Effect on plasticity?	Transmitter	Structure (preparation)
Arancio et al., 1996a	Nitric oxide acts directly in the presynaptic neuron to produce long-term potentiation in cultured hippocampal neurons	✓	✓	✓	Glutamate	Hippocampus (cell culture)
Lange et al., 2012	Heterosynaptic long-term potentiation at interneuron-principal neuron synapses in the amygdala requires nitric oxide signaling	✓	✓	✓	GABA	Amygdala (slices)
O'Dell et al., 1991	Tests of the roles of two diffusible substances in long-term potentiation: evidence for nitric oxide as a possible early retrograde messenger	✓	✓	✓	Glutamate	Hippocampus (slices)
Sjostrom et al., 2007	Multiple forms of long-term plasticity at unitary neocortical layer 5 synapses	✓	✓	✓	Glutamate	Visual cortex (slices)
Hardingham and Fox, 2006	The role of nitric oxide and GluR1 in presynaptic and postsynaptic components of neocortical potentiation	✓	✓	✓	Glutamate	Barrel cortex (slices)
Schuman and Madison, 1991	A requirement for the intercellular messenger nitric oxide in long-term potentiation	✓	✓	✓	Glutamate	Hippocampus (slices)
Volgushev et al., 2000	Retrograde signaling with nitric oxide at neocortical synapses	✓	✓	✓	Glutamate	Visual cortex (slices)
Montague et al., 1994	Role of NO production in NMDA receptor-mediated neuro-transmitter release in cerebral cortex	✓	✓		Glutamate	Neocortex (synaptosomes)
Micheva et al., 2003	Retrograde regulation of synaptic vesicle endocytosis and recycling	✓	✓		Glutamate	Hippocampus (cell culture)
Eguchi et al., 2012	Maturation of a PKG-dependent retrograde mechanism for exocytotic coupling of synaptic vesicles	✓	✓		Glutamate	MNTB/Caylx of Held (slices)
Lindskog et al., 2010	Postsynaptic GluA1 enables acute retrograde enhancement of presynaptic function to coordinate adaptation to synaptic inactivity	✓		✓	Glutamate	Hippocampus (cell culture)

(Continued)

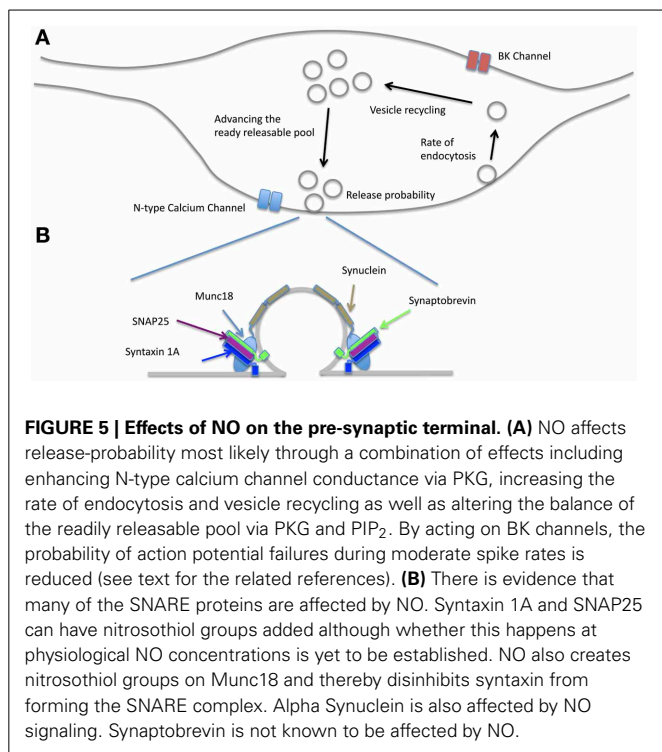
Table 1 | Continued

Qiu and Knopfel, 2007	An NMDA receptor/nitric oxide cascade in presynaptic parallel fiber-Purkinje neuron long-term potentiation	✓	✓	Glutamate	Cerebellum (slices)
Johnstone and Raymond, 2011	A protein synthesis and nitric oxide-dependent presynaptic enhancement in persistent forms of long-term potentiation	✓	✓	Glutamate	Hippocampus (slices)
Stanton et al., 2005	Imaging LTP of presynaptic release of FM1-43 from the rapidly recycling vesicle pool of Schaffer collateral-CA1 synapses in rat hippocampal slices	✓	✓	Glutamate	Hippocampus (slices)
Wang et al., 2005	Presynaptic and postsynaptic roles of NO, cGK, and RhoA in long-lasting potentiation and aggregation of synaptic proteins	✓	✓	Glutamate	Hippocampus (cell culture)
Arancio et al., 2001	Presynaptic role of cGMP-dependent protein kinase during long-lasting potentiation	✓	✓	Glutamate	Hippocampus (cell culture)
Huang et al., 2003	cGMP/protein kinase G-dependent potentiation of glutamatergic transmission induced by nitric oxide in immature rat rostral ventrolateral medulla neurons <i>in vitro</i>	✓		Glutamate	Ventrolateral medulla (slices)
Ratnayaka et al., 2012	Recruitment of resting vesicles into recycling pools supports NMDA receptor-dependent synaptic potentiation in cultured hippocampal neurons	✓		Glutamate	Hippocampus (cell culture)
Neitz et al., 2011	Presynaptic nitric oxide/cGMP facilitates glutamate release via hyperpolarization-activated cyclic nucleotide-gated channels in the hippocampus	✓		Glutamate	Hippocampus (slices)

by two routes. Finally, at the glutamatergic neuromuscular junction in *Drosophila*, calcium independent vesicular release can result from cGMP triggered by NO (Wildemann and Bicker, 1999), although the exact downstream processes by which this occurs are not known. Calcium independent vesicular release can also be observed in hippocampal synaptosomes (Meffert et al., 1994).

EFFECTS ON VESICLE RECYCLING

In order to sustain synaptic release over a period of time, the rate of vesicle recycling needs to at least equal the rate of vesicle exocytosis. This issue is particularly problematic for synapses that release transmitter at high rates, such as those located at the Calyx of Held that terminate on neurons of the medial nucleus of the trapezoid body (MNTB). Part of the solution to



this problem at the Calyx is provided by linking vesicle recycling to retrograde release of post-synaptic NO. Activation of the post synaptic MNTB neurons is related to the level of NO production (Steinert et al., 2008), which then drives the level of pre-synaptic cGMP production and hence the level of PKG activity (Eguchi et al., 2012). Finally, activation of PKG up-regulates PIP₂, which increases the rate of endocytosis (Eguchi et al., 2012). This homeostatic mechanism therefore links pre-synaptic rate of release (which is sensed by post-synaptic NOS1) to the rate of pre-synaptic vesicle recycling (Figure 5). Regulation of the recycling rate has also been demonstrated in the hippocampus, where a very similar retrograde NO—pre-synaptic cGMP/PIP₂ cascade regulates the rate of endocytosis and recycling (Micheva et al., 2003).

EFFECTS ON AVAILABILITY AND SIZE OF THE READILY RELEASABLE POOL (RRP)

Studies aimed at investigating the nature of synaptic plasticity have shown that LTP is accompanied by an increase (and LTD a decrease) in the rate of vesicular release from the readily releasable pool (RRP). The LTP process is NMDA receptor-, tyrosine kinase- and NO-dependent while the LTD process is NMDA-, NO- and PKG-dependent (Stanton et al., 2003, 2005). Studies have shown that the size of the RRP can be modulated by NO (Figure 5). For example, in the case of LTP, NMDA receptor activation leads to NO and calcineurin activation, which combine to increase the proportion of vesicles available for release (i.e., increase the RRP) (Ratnayaka et al., 2012). Once again this can be seen as a homeostatic response to an increase in release probability brought about by the process of LTP itself. The two

processes are coordinated because NO is involved both in increasing transmitter release and increasing the size of the readily releasable pool.

EFFECTS ON GROWTH OF PRE-SYNAPTIC TERMINALS

Nitric oxide also affects the growth and formation of new pre-synaptic terminals and can lead to the formation of multi-innervated spines. Long lasting potentiation leads to an increase in pre- and post-synaptic proteins in hippocampal cell cultures. GluA1 subunits of the AMPA receptor increase post-synaptically and synaptophysin increases pre-synaptically (Antonova et al., 2001). Furthermore, the two synaptic markers co-localize at higher frequency following long lasting potentiation, indicating that new synapses are formed. It has been shown that NMDA receptors, NO and actin are required for the pre-synaptic changes. NO acts via PKG to phosphorylate VASP (which acts on actin) and also via cGMP to act in parallel and downstream of RhoGTPase (Wang et al., 2005).

Further evidence for the role of NO in pre-synaptic growth comes from studies manipulating the PDZ2 domain of PSD95 (which is the PDZ domain that binds NOS1). Up-regulation of PSD95 in cultured hippocampal neurons or treatment with an NO donor leads to the formation of multi-innervated spines (MIS). However, if the PDZ2 domain on PSD95 is deleted, thereby dissociating NOS1 from PSD95, multi-innervated spines fail to form (Nikonenko et al., 2008). Similarly, down regulating NOS1 expression with iRNA also prevents MIS from forming (Nikonenko et al., 2008). Finally, increasing SAP97 expression leads to an increase in PSD95 and again an increase in MIS (Poglia et al., 2011). This effect is blocked by NOS antagonists (Poglia et al., 2011).

In conclusion, the studies cited above show that NO is not only involved in the relatively short term changes involved in transmitter release, such as recycling rates and availability of vesicles, but also, in the long-term, in increasing the availability of transmitter by formation of new pre-synaptic terminals, which results in dendritic spines receiving extra pre-synaptic terminals. Such processes could find application in synaptic plasticity. In the following section we review the function of NO in plasticity and examine to what extent the retrograde route of action is involved.

THE ROLE OF NO IN PLASTICITY

NO-DEPENDENT PRE-SYNAPTIC PLASTICITY

Some of the earliest studies on the role of NO in synaptic plasticity indicated that it might act at a pre-synaptic locus (O'Dell et al., 1991). Exogenous NO applied to neurons in a hippocampal slice increased spontaneous mini EPSCs and hemoglobin acting as an extracellular scavenger for NO was found to prevent LTP (O'Dell et al., 1991). Indeed, initial studies on the mechanisms of LTP itself provided evidence for a pre-synaptic locus of LTP expression (Malinow and Tsien, 1990). In a series of experiments on cultured hippocampal neurons, Arancio and colleagues showed that cGMP (the downstream effector of NO) needs to be pre-synaptic and NOS post-synaptic to produce plasticity. First, cGMP causes an increase in EPSC amplitude when injected into the pre-synaptic but not the post-synaptic cell (Arancio et al., 1995). Second, application of a PKG antagonist

peptide blocks tetanus induced LTP when injected into the pre-synaptic but not the post-synaptic neuron (Arancio et al., 2001). Third, application of a cGMP analogue increases miniature EPSC frequency and this effect is blocked by a post-synaptically but not pre-synaptically injected NOS inhibitor (Arancio et al., 1996a). Forth, a pre-synaptic injection of an NO scavenger also abolishes LTP (Arancio et al., 1996b). More recent work employing fluorescent markers of pre-synaptic function have visualized the pre-synaptic effect of NO in potentiation. Fluorescence imaging of FM-styryl dyes and synaptophysinI-pHluorin has shown that increases to the pre-synaptic recycling pool fraction following synaptic strengthening are dependent upon both NMDA receptor activation and NO release (Ratnayaka et al., 2012).

THE EFFECT OF INITIAL RELEASE-PROBABILITY ON THE LOCUS OF PLASTICITY

Early studies on hippocampal plasticity showed that the initial release-probability of the synapse influences whether a pre- or post-synaptic change occurs following LTP (Larkman et al., 1992). If the release-probability of the synapse is low initially then pre-synaptic plasticity occurs, whereas if the pre-synaptic release-probability is high, then a post-synaptic change occurs (Larkman et al., 1992). A similar principal operates at neocortical synapses. In visual cortex, the initial release-probability of the synapse, as judged by the paired pulse ratio (PPR), is predictive of whether NO-dependent potentiation occurs. Using a purely post-synaptic tetanus (without intentionally eliciting action potentials in the pre-synaptic terminals), potentiation occurs in synapses with a low initial PPR and depression or no change occurs in synapses with a high initial PPR (Volgushev et al., 2000). The same conclusion is arrived at if a paired pre- and post-spike conditioning protocol is used. Low release-probability synapses potentiate via changes in release-probability and high release-probability synapses depress (Hardingham et al., 2007). This normalization process causes the population of connections to adopt a more homogenous set of release probabilities after the protocol. These studies lead to two important conclusions; first, the direction of pre-synaptic plasticity acts in a homeostatic manner to move release-probability to an intermediate value and second, that pre-synaptic plasticity occurs provided that there is sufficient dynamic range for it to occur. There is less scope for increasing release-probability at a high release-probability synapse than at a low release-probability synapse. Potentially, a high release-probability synapse could show pre-synaptic potentiation by growth and/or production of MIS, which can occur and is NO-dependent (section Effects on Growth of Pre-Synaptic Terminals), but structural changes are unlikely within the timescale of an LTP experiment.

Since the initial release-probability of the synapse is an important determinant of the locus of plasticity and in which direction it operates, factors that control initial release-probability will determine the level and form of pre-synaptic plasticity. Adenosine is known to affect release-probability (Prince and Stevens, 1992) and a recent study in layer 5 of the somatosensory cortex has shown that adenosine levels are low early in development (P11-P22) and higher in older animals (P28-32) (Kerr et al., 2013).

This maturational change means that adenosine reduces release-probability in older animals, thereby increasing the dynamic range for pre-synaptic potentiation. Some mutant mice strains have unusually low initial release-probability synapses that can provide an increased dynamic range for LTP. For example, H-Ras^{G12V} mice have low release-probability synapses in the visual cortex, as judged by short-term dynamics and mini EPSP frequency, and consequently enhanced LTP with an increased pre-synaptic component (Kaneko et al., 2010).

PRE- AND POST-SYNAPTIC COMPONENTS OF PLASTICITY

Early studies on the role of NO in LTP using NO antagonists often found an absolute requirement for NO (Bohme et al., 1991; O'Dell et al., 1991; Schuman and Madison, 1991; Haley et al., 1993; Doyle et al., 1996; Malen and Chapman, 1997), whereas more recent studies have found LTP to be reduced rather than abolished in the absence of NO, both in the hippocampus (Phillips et al., 2008) and in the neocortex (Hardingham and Fox, 2006).

In the neocortex, LTP occurs as a mixture of pre- and post-synaptic changes, but the two components can be dissociated, either by blocking NOS post-synaptically or knocking out GluA1 (Hardingham and Fox, 2006). When a NOS inhibitor is introduced to the post-synaptic neuron via the electrode, plasticity proceeds by changes in quantal amplitude without changes in the variance of the response amplitude (Hardingham and Fox, 2006). Similarly, where single or double quantal release peaks are isolated in layer 2/3 neurons, LTP occurs by changing the quantal amplitude without changes in release-probability (Figure 6). Conversely, in GluA1 knockouts, LTP results in changes in release-probability (NP_r) without changes in quantal amplitude (Q) (Hardingham and Fox, 2006). Given that potentiation is NO-dependent in GluA1 knockouts this implies that NO acts via a pre-synaptic mechanism in neocortex (Hardingham and Fox, 2006).

The situation is similar in the mature hippocampus, in that the plasticity present in the GluA1 knockouts is largely NO-dependent (Phillips et al., 2008; Romberg et al., 2009), but it is not clear in this case whether the locus of NO-dependent plasticity is pre- or post-synaptic, or perhaps both. Phillips et al. (2008) suggested a pre-synaptic origin for NO-dependent LTP based on the decrease in PPR for 14/21 cases following potentiation, while Romberg et al. (2009) found no change in average PPR. As noted above, it may be that the initial release-probability present at a particular connection affects the likelihood of pre-synaptic plasticity at that synapse (see section The Effect of Initial Release-Probability on the Locus of Plasticity).

Nitric oxide is also known to affect post-synaptic AMPA receptor trafficking; NO increases GluA1 insertion acting via sGC and protein kinase G (PKG) (Serulle et al., 2008) and GluA2 heteromer insertion by production of nitrosothiol groups on NSF (N-ethylmaleimide-sensitive factor) (Huang et al., 2005). Furthermore, endogenous NSF does appear to contain nitrosothiol groups *in vivo*. (Huang et al., 2005). However, the GluA1 insertion mechanism cannot be the one operating in the GluA1 knockouts, leaving the GluA2 mechanism as the most likely to be operating in these studies. This view is given further support by

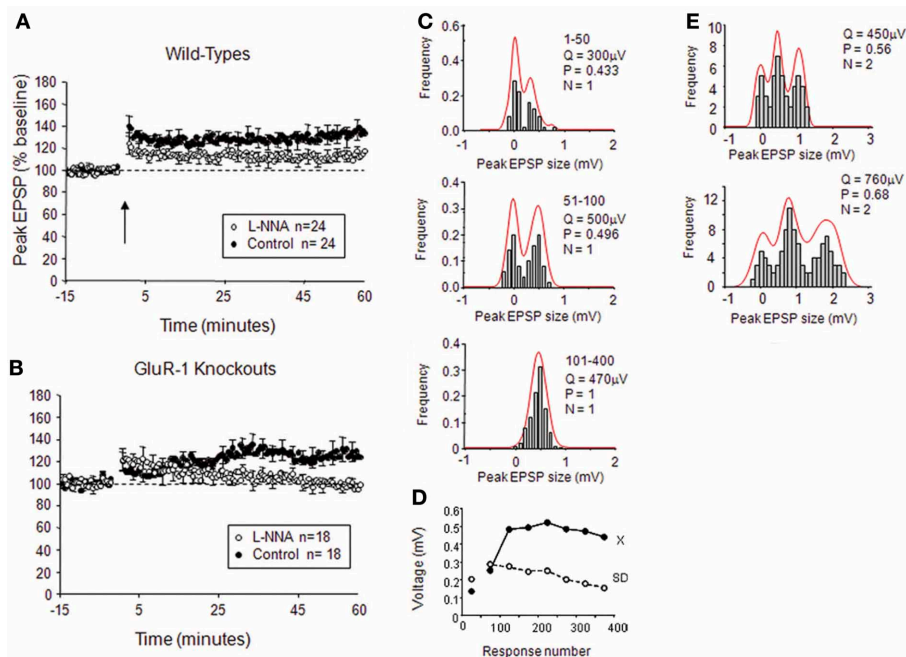


FIGURE 6 | Effects of NO on release-probability in cortical LTP. (A)

Intracellular application of the NOS antagonist L-NNA reduces but does not abolish spike pairing LTP in wild-type mice. **(B)** Intracellular application of L-NNA abolishes LTP in mice lacking the GluA1 subunit of the AMPA receptor. **(C)** Examples of quantal analysis from a single release site input onto a layer II/III neuron from a wild-type mouse; note that LTP occurs by an increase in release-probability and quantal amplitude. **(D)** The plot of EPSP amplitude and

standard deviation for the example in **(C)** during the course of LTP (x = mean, SD = standard deviation). **(E)** Example of quantal analysis from a double release site case in a wild-type treated with intracellular L-NNA; note that LTP occurs largely by an increase in quantal amplitude with a minor increase in release-probability. Q is quantal amplitude, P is release-probability, and N is the number of release sites. Adapted from Hardingham and Fox (2006) with permission of the Society for Neuroscience.

data showing the PKC dependence of LTP in the GluA1 knockout animals (Romberg et al., 2009). Nevertheless, in wild-types it is possible that both GluA1 and GluA2 are controlled by NO signaling. Accumulation of cGMP in hippocampal cells has recently been demonstrated using NO donors (Bartus et al., 2013) giving further credence to a post-synaptic role for NO. Furthermore, there is some evidence that dexas1 is activated by NO and is located post-synaptically due to CAPON binding dexas1 and NOS1 (Fang et al., 2000; Cheah et al., 2006). Together with the substantial evidence that NO acts pre-synaptically (Section NO Controls Pre-Synaptic Function), these findings raise the intriguing possibility that NO might play a role in coordinating pre- and post-synaptic changes at excitatory synapses during plasticity.

EVIDENCE FOR THE ROLE OF NO IN EXPERIENCE-DEPENDENT PLASTICITY

There is an extensive literature on the role of NO in learning and memory. Peripheral administration of NOS inhibitors have been shown to impair spatial memory acquisition or recall (Bohme et al., 1993; Chapman et al., 1992; Zou et al., 1998; Majlessi et al., 2008), social interactions (Bohme et al., 1993) and object recognition memory (Cobb et al., 1995). Central administration of NOS antagonists also alters behavior, including spatial learning in the Morris water maze and the passive avoidance test (Qiang et al., 1997; Majlessi et al., 2008; Li et al., 2012),

arguing against the peripheral effects of the drug. Inhibitors more specific to NOS1 have also shown sensitivity to behavioral performance in spatial reference and working memory (Holscher et al., 1996; Zou et al., 1998; Yildiz Akar et al., 2009). Furthermore, NOS1 knockout mice show impaired spatial memory, social interactions and contextual fear memory (Weitzdoerfer et al., 2004; Kelley et al., 2009; Tanda et al., 2009). In contrast, NOS3 knockout mice exhibit enhanced spatial learning, retention and reversal learning in the Morris water maze but increased anxiety-like behaviors in the plus maze and the open arena (Frisch et al., 2000). However, spatial learning is comparable to controls in the radial arm maze (Dere et al., 2001), suggesting that NOS3 knockout confers a specific deficit in spatial learning and may therefore play a particular role in hippocampal plasticity, where it has been shown to play a role in LTP in concert with NOS1 (Hopper and Garthwaite, 2006; Phillips et al., 2008).

A simpler form of experience-dependent plasticity that can be quantified by measuring neuronal responses rather than behavior is the plasticity that results from whisker deprivation in the barrel cortex. Depriving a single whisker for several days leads to expansion of the area of cortex dominated by that whisker (Fox, 1992; Wallace and Fox, 1999). NO is implicated in the potentiation component of this plasticity as α NOS1 knockouts exhibited reduced single whisker potentiation (Dachtler et al., 2011). In parallel with the LTP studies (Hardingham and Fox,

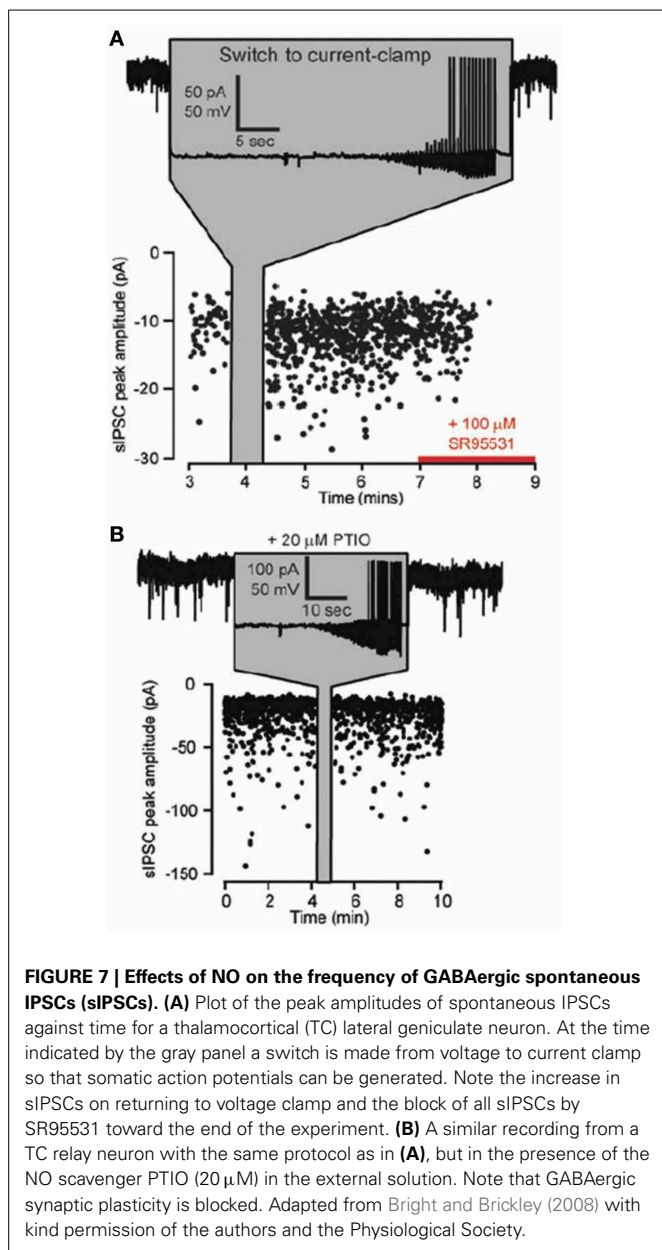


FIGURE 7 | Effects of NO on the frequency of GABAergic spontaneous IPSCs (sIPSCs). (A) Plot of the peak amplitudes of spontaneous IPSCs against time for a thalamocortical (TC) lateral geniculate neuron. At the time indicated by the gray panel a switch is made from voltage to current clamp so that somatic action potentials can be generated. Note the increase in sIPSCs on returning to voltage clamp and the block of all sIPSCs by SR95531 toward the end of the experiment. (B) A similar recording from a TC relay neuron with the same protocol as in (A), but in the presence of the NO scavenger PTIO (20 μ M) in the external solution. Note that GABAergic synaptic plasticity is blocked. Adapted from Bright and Brickley (2008) with kind permission of the authors and the Physiological Society.

2006), experience-dependent potentiation was only abolished in double knockouts of α NOS1 and GluA1 (Dachtler et al., 2011). Plasticity was present in double knockouts of NOS3 and GluA1, suggesting that α NOS1 is the important isoform in the cortex, probably due to the close association between α NOS1 and the NMDA receptor (see section The Cellular Location of NOS1). In further support of this idea, NMDA-dependent release of NO is impaired in α NOS1 but not NOS3 knockout mice (Dachtler et al., 2011).

Further analysis of plasticity in α NOS1 knockout mice reveals both LTP and experience-dependent potentiation are abolished in male but not female mice (Dachtler et al., 2012). This could either mean that male mice rely solely on NO-dependent forms of potentiation, or that some form of compensation for the lack of NOS1 takes place in the female knockout mice that does not

occur in the males. The sex difference was not seen in wild-type animals suggesting that the latter is a possible explanation. The sex difference in the α NOS1 knockout mice may be of importance to interpreting stroke data because factors involved in LTP are often also involved in excitotoxicity. NOS1 has long been known to be a factor in ischemic damage in stroke (Huang et al., 1994), most likely through the association of α NOS1 and PSD-95 (Cao et al., 2005). However, the magnitude of ischemic damage differs depending upon sex. Male α NOS1 knockout mice have less ischemic damage than wild-types, while female α NOS1 knockout mice have more damage than their wild-type counterparts (McCullough et al., 2005).

NO AND PLASTICITY AT GABAERGIC SYNAPSES

Because NO can play a role in pre-synaptic plasticity, it also means that it is not restricted to act on a particular set of post-synaptic receptors or the protein trafficking machinery associated with them. Instead, in so far as the vesicular release machinery is common across transmitter systems, NO can potentially regulate release for several different neurotransmitters including GABA (Table 2).

Anatomical evidence implicates NO in regulation of pre-synaptic GABA release. In excitatory pyramidal cells in the hippocampus, NOS1 lies post synaptic to GABAergic synapses and the “NO receptor” (sGC) lies in the pre-synaptic terminals of those same GABAergic synapses, thereby providing both elements required for retrograde synaptic signaling in close assembly (Szabadits et al., 2007). In this case, rather than being associated with PSD95, which does not appear to localize at post-synaptic densities of symmetric synapses, GRIP1 may bind NOS1 at the post-synaptic site. The pre-synaptic terminals in question belong to parvalbumin- and CCK-containing cells that synapse onto somata and proximal dendrites of pyramidal cells. Consistent with this location, application of NO donors increases cGMP levels in GABAergic interneurons (Bartus et al., 2013). It is not clear how the endogenous signal arises to activate NOS1 at these inhibitory synapses, but one possibility is that action potentials could raise intracellular calcium via voltage gated calcium channels and the spatial localization of NOS1 immediately post-synaptic to the GABAergic terminals targets NO to the inhibitory terminals.

Physiological evidence further implicates NO in GABAergic synaptic plasticity. Evidence comes from observations on paraventricular neurons (Li et al., 2002), the prepositus hypoglossal nucleus (Moreno-Lopez et al., 2002), the amygdala (Lange et al., 2012) and thalamic projection neurons (Bright and Brickley, 2008). In the hippocampus, NMDA receptor activation in pyramidal cells causes an increase in spontaneous GABA_A receptor mediated IPSCs that are sensitive to an NO scavenger (Xue et al., 2011). In the ventral tegmental area, GABAergic synapses onto dopaminergic neurons express a pre-synaptic form of LTP that is dependent upon NMDA receptor activation, NO, GC, and PKG for its induction and maintenance and is selective to GABA_A synapses (Nugent et al., 2007, 2009). Furthermore, pre-synaptic GABAergic LTP from the lateral amygdala to the basolateral amygdala depends upon NO generated from glutamatergic neurons (Lange et al., 2012).

Table 2 | The role of Nitric oxide in GABAergic function.

References	Title	Presynaptic action?	Retrograde messenger?	Effect?	Structure (preparation)
Lange et al., 2012	Heterosynaptic long-term potentiation at interneuron-principal neuron synapses in the amygdala requires nitric oxide signaling	✓	✓	Effect on plasticity	Amygdala (slice)
Moreno-Lopez et al., 2002	Nitric oxide facilitates GABAergic neurotransmission in the cat oculomotor system: a physiological mechanism in eye movement control	✓	✓	Controls velocity responsiveness of PH neurons	Medial vestibular nucleus projection to prepositus hyperglossi (PH) neurons (<i>in vivo</i>)
Szabadits et al., 2007	Hippocampal GABAergic synapses possess the molecular machinery for retrograde nitric oxide signaling	✓	✓	Anatomical evidence: nNOS is post and sCG presynaptic	Hippocampus (<i>in vivo</i>)
Xue et al., 2011	NMDA receptor activation enhances inhibitory GABAergic transmission onto hippocampal pyramidal neurons via presynaptic and postsynaptic mechanisms	✓	✓	Increase in sIPSP frequency and amplitude	Hippocampal (slice)
Yang et al., 2007	Kv1.1/1.2 channels are downstream effectors of nitric oxide on synaptic GABA release to preautonomic neurons in the paraventricular nucleus	✓	✓	Nitric oxide acts on GABA via Kv1.1/1.2	Paraventricular nucleus of the hypothalamus (slices)
Yang et al., 2007	Kv1.1/1.2 channels are downstream effectors of nitric oxide on synaptic GABA release to preautonomic neurons in the paraventricular nucleus	✓	✓	Increases frequency but not amplitude of inhibitory minis	Paraventricular nucleus of the hypothalamus (slices)
Bright and Brickley, 2008	Acting locally but sensing globally: impact of GABAergic synaptic plasticity on phasic and tonic inhibition in the thalamus	✓	✓	Increases frequency of sIPSCs	Thalamus (slices)
Wall, 2003	Endogenous nitric oxide modulates GABAergic transmission to granule cells in adult rat cerebellum	✓	✓	NO modulates tonic GABA release	Cerebellum (slices)
Holmgren and Zilberter, 2001	Coincident spiking activity induces long-term changes in inhibition of neocortical pyramidal cells			Analogous to cases where nitric oxide is involved	Neocortical (slices)

The studies on GABAergic potentiation in the thalamus are particularly interesting because it only requires post-synaptic action potentials, which cause an increase in spontaneous GABAergic mIPSC frequency. This effect is blocked by the NO scavenger PTIO (Bright and Brickley, 2008) suggesting that the action potentials lead to release of NO that in turn produces changes in GABA release (Figure 7). The NO donor SNAP can also increase GABA mini frequency in these cells (Bright and Brickley, 2008). The sufficiency of post-synaptic action potentials in this study is reminiscent of the findings of Volgushev and colleagues in the visual cortex, who showed that post-synaptic action potentials produced NO-dependent potentiation in pyramidal cells (Volgushev et al., 2000) and Phillips et al. who showed that NO-dependent LTP in the hippocampus relies on

somatic post-synaptic action potentials (Phillips et al., 2008). These findings raise the possibility that post-synaptic action potentials may simultaneously produce NO-dependent plasticity at inhibitory synapses and spike timing-dependent plasticity at excitatory synapses on the same cell, a property that may be involved in maintenance of inhibitory-excitatory balance.

NO AND THE REGULATION OF EXCITATORY/INHIBITORY BALANCE

Cells in the cortex exhibit a balance between excitation and inhibition such the ratio between inhibitory and excitatory conductances is relatively constant for different inputs. For layer 5 pyramidal cells in the visual cortex, the ratio of excitatory to inhibitory conductance has been estimated at 20:80 (Le Roux et al., 2006) using the method of Monier et al. (2008). The

excitatory/inhibitory (E/I) balance would be expected to be a universal phenomenon as loss of E/I balance in favor of excitation leads to epilepsy. Consistent with this idea, inhibition has been shown to be matched to excitation in the visual (Anderson et al., 2000; Priebe and Ferster, 2005) auditory (Wehr and Zador, 2003, 2005) and somatosensory cortices (Wilent and Contreras, 2005). Studies have shown that NO may play a role in the maintenance of the E/I balance in the visual cortex. It can be demonstrated that the E/I balance is maintained in layer 5 pyramidal cells following potentiation by theta burst stimulation of cortical layers 2/3, 4 or 6 (Le Roux et al., 2006). Stimulating layer 4 and increasing endogenous levels of NO by dosing a cortical slice with L-arginine, or administration of an NO donor, also increases excitatory and inhibitory conductances in balance (Le Roux et al., 2009). These studies suggest that NO may play a homeostatic role in maintaining the balance between excitation and inhibition in the cortex.

CONCLUSIONS

Deciphering the role of NO in the brain has not been a simple matter and at times the results of different studies have been confusing. Nevertheless, a clearer picture is now emerging of how NO might act to regulate synaptic function in the brain. In excitatory cells, NOS1 is located discretely in spines and is tethered to the post-synaptic membrane by its interaction with PSD95 in complete contrast to its location in a subpopulation of NOS1⁺ NPY⁺ inhibitory cells, where NOS1 is located in the cytoplasm along axons and dendrites and appears to be expressed at higher levels. The low levels of NOS1 expression in excitatory cells of the cortex and hippocampus dictate that under normal physiological conditions low concentrations of NO are evolved during stimulation by calcium, which in turn means that it has a relatively small range and is therefore probably synapse-specific in its action. The only obvious receptor that is sensitive at the low nM to pM range is guanylate cyclase, although there is some evidence for endogenous levels of proteins with nitrosothiol groups that would require higher concentrations of NO. There is a substantial body of literature that suggests that NO acts in a retrograde manner on several aspects of vesicular release and recycling, so much so that it would seem perverse to argue that NO does not act pre-synaptically at this point. Present evidence suggests that NO acts

in a retrograde manner to affect not only glutamatergic synapses but also GABAergic synapses as well as other transmitter systems. Finally, there is substantial evidence in the literature that the retrograde route of action is important for plasticity in the cortex and hippocampus in both inhibitory and excitatory cells.

Nevertheless, a number of important questions remain about the action of NO at synapses. Two questions relate to the concentration of NO in the brain. First, are the levels of NO required for nitrosothiol production at the SNARE complex proteins actually achieved *in vivo*? Second, could higher levels of NO reported in some studies be generated by the higher NOS levels present in the NOS1⁺ GABAergic cells? A further set of questions relate to the action of NO at pre-synaptic GABAergic synapses. There is evidence that sGC is present in GABAergic terminals and that NOS1 lies post-synaptic to it (Szabadits et al., 2007). There is evidence that GABAergic mini EPSC frequency increases following somatic spiking in the LGn (Bright and Brickley, 2008). Therefore, what is the mechanism of post-synaptic spike-dependent potentiation of GABAergic transmission and is it indeed NO-dependent in the cortex and hippocampus? More generally, is this mechanism related to the post-synaptic spike potentiation present at excitatory synapses (Volgushev et al., 2000)? Unraveling this effect could help us understand whether the E/I balance is maintained by NO acting simultaneously on GABAergic and excitatory transmission (Le Roux et al., 2009). Finally, while we have concentrated on the pre-synaptic role of NO in this review, there is evidence that NO also has a post-synaptic action. In addition to activation of post-synaptic sGC, there is evidence that post-synaptic proteins have nitrosothiol groups, particularly those close to its PSD location (see Figure 1). If NO also has a post-synaptic role in plasticity it raises the additional question about whether it can play a homeostatic role in balancing or matching pre- and post-synaptic function. With a little good fortune, it will not take another 25 years of research to solve these and other related questions on the role NO plays in synaptic function in the brain.

ACKNOWLEDGMENTS

We should like to acknowledge the support of the MRC to Kevin Fox for the authors' own work. We should also like to thank John Garthwaite for critically reading an earlier draft of the manuscript.

REFERENCES

- Aarts, M., Liu, Y., Liu, L., Besshoh, S., Arundine, M., Gurd, J. W., et al. (2002). Treatment of ischemic brain damage by perturbing NMDA receptor- PSD-95 protein interactions. *Science* 298, 846–850. doi: 10.1126/science.1072873
- Ahern, G. P., Klyachko, V. A., and Jackson, M. B. (2002). cGMP and S-nitrosylation: two routes for modulation of neuronal excitability by NO. *Trends Neurosci.* 25, 510–517. doi: 10.1016/S0166-2236(02)02254-3
- Anderson, J. S., Carandini, M., and Ferster, D. (2000). Orientation tuning of input conductance, excitation, and inhibition in cat primary visual cortex. *J. Neurophysiol.* 84, 909–926.
- Antonova, I., Arancio, O., Trillat, A. C., Wang, H. G., Zablow, L., Udo, H., et al. (2001). Rapid increase in clusters of presynaptic proteins at onset of long-lasting potentiation. *Science* 294, 1547–1550. doi: 10.1126/science.1066273
- Aoki, C., Bredt, D. S., Fenstemaker, S., and Lubin, M. (1998). The subcellular distribution of nitric oxide synthase relative to the NR1 subunit of NMDA receptors in the cerebral cortex. *Prog. Brain Res.* 118, 83–97. doi: 10.1016/S0079-6123(08)63202-1
- Aoki, C., Rhee, J., Lubin, M., and Dawson, T. M. (1997). NMDA-R1 subunit of the cerebral cortex co-localizes with neuronal nitric oxide synthase at pre- and post-synaptic sites and in spines. *Brain Res.* 750, 25–40. doi: 10.1016/S0006-8993(96)01147-X
- Arancio, O., Antonova, I., Gambaryan, S., Lohmann, S. M., Wood, J. S., Lawrence, D. S., et al. (2001). Presynaptic role of cGMP-dependent protein kinase during long-lasting potentiation. *J. Neurosci.* 21, 143–149.
- Arancio, O., Kandel, E. R., and Hawkins, R. D. (1995). Activity-dependent long-term enhancement of transmitter release by presynaptic 3',5'-cyclic GMP in cultured hippocampal neurons. *Nature* 376, 74–80. doi: 10.1038/376074a0
- Arancio, O., Kiebler, M., Lee, C. J., Lev-Ram, V., Tsien, R. Y., Kandel, E. R., et al. (1996a). Nitric oxide acts directly in the presynaptic neuron to produce long-term potentiation in cultured hippocampal neurons. *Cell* 87, 1025–1035. doi: 10.1016/S0092-8674(00)81797-3
- Arancio, O., Lev-Ram, V., Tsien, R. Y., Kandel, E. R., and Hawkins, R. D. (1996b). Nitric oxide acts as a retrograde messenger during long-term potentiation in cultured hippocampal neurons. *J. Physiol. Paris*

- 90, 321–322. doi: 10.1016/S0928-4257(97)87907-7
- Bartus, K., Pigott, B., and Garthwaite, J. (2013). Cellular targets of nitric oxide in the hippocampus. *PLoS ONE* 8:e57292. doi: 10.1371/journal.pone.0057292
- Batchelor, A. M., Bartus, K., Reynell, C., Constantinou, S., Halvey, E. J., Held, K. F., et al. (2010). Exquisite sensitivity to subsecond, picomolar nitric oxide transients conferred on cells by guanylyl cyclase-coupled receptors. *Proc. Natl. Acad. Sci. U.S.A.* 107, 22060–22065. doi: 10.1073/pnas.1013147107
- Bellamy, T. C., and Garthwaite, J. (2001). Sub-second kinetics of the nitric oxide receptor, soluble guanylyl cyclase, in intact cerebellar cells. *J. Biol. Chem.* 276, 4287–4292. doi: 10.1074/jbc.M006677200
- Blackshaw, S., Eliasson, M. J., Sawa, A., Watkins, C. C., Krug, D., Gupta, A., et al. (2003). Species, strain and developmental variations in hippocampal neuronal and endothelial nitric oxide synthase clarify discrepancies in nitric oxide-dependent synaptic plasticity. *Neuroscience* 119, 979–990. doi: 10.1016/S0306-4522(03)00217-3
- Bohme, G. A., Bon, C., Lemaire, M., Reibaud, M., Piot, O., Stutzmann, J. M., et al. (1993). Altered synaptic plasticity and memory formation in nitric oxide synthase inhibitor-treated rats. *Proc. Natl. Acad. Sci. U.S.A.* 90, 9191–9194. doi: 10.1073/pnas.90.19.9191
- Bohme, G. A., Bon, C., Stutzmann, J. M., Doble, A., and Blanchard, J. C. (1991). Possible involvement of nitric oxide in long-term potentiation. *Eur. J. Pharmacol.* 199, 379–381. doi: 10.1016/0014-2999(91)90505-K
- Brenman, J. E., and Bredt, D. S. (1997). Synaptic signaling by nitric oxide. *Curr. Opin. Neurobiol.* 7, 374–378. doi: 10.1016/S0959-4388(97)80065-7
- Brenman, J. E., Chao, D. S., Gee, S. H., McGeer, A. W., Craven, S. E., Santillano, D. R., et al. (1996). Interaction of nitric oxide synthase with the postsynaptic density protein PSD-95 and alpha1-syntrophin mediated by PDZ domains. *Cell* 84, 757–767. doi: 10.1016/S0092-8674(00)81053-3
- Bright, D. P., and Brickley, S. G. (2008). Acting locally but sensing globally: impact of GABAergic synaptic plasticity on phasic and tonic inhibition in the thalamus. *J. Physiol.* 586, 5091–5099. doi: 10.1113/jphysiol.2008.158576
- Broillet, M. C., and Firestein, S. (1997). Beta subunits of the olfactory cyclic nucleotide-gated channel form a nitric oxide activated Ca²⁺ channel. *Neuron* 18, 951–958. doi: 10.1016/S0896-6273(00)80334-7
- Buchwalow, I. B., Podzuweit, T., Bocker, W., Samoiloova, V. E., Thomas, S., Wellner, M., et al. (2002). Vascular smooth muscle and nitric oxide synthase. *FASEB J.* 16, 500–508. doi: 10.1096/fj.01-0842com
- Burette, A., Zabel, U., Weinberg, R. J., Schmidt, H. H., and Valtchanoff, J. G. (2002). Synaptic localization of nitric oxide synthase and soluble guanylyl cyclase in the hippocampus. *J. Neurosci.* 22, 8961–8970.
- Buskila, Y., and Amitai, Y. (2010). Astrocytic iNOS-dependent enhancement of synaptic release in mouse neocortex. *J. Neurophysiol.* 103, 1322–1328. doi: 10.1152/jn.00676.2009
- Calabrese, V., Mancuso, C., Calvani, M., Rizzarelli, E., Butterfield, D. A., and Stella, A. M. (2007). Nitric oxide in the central nervous system: neuroprotection versus neurotoxicity. *Nat. Rev. Neurosci.* 8, 766–775. doi: 10.1038/nrn2214
- Cao, J., Viholainen, J. I., Dart, C., Warwick, H. K., Leyland, M. L., and Courtney, M. J. (2005). The PSD95-nNOS interface: a target for inhibition of excitotoxic p38 stress-activated protein kinase activation and cell death. *J. Cell Biol.* 168, 117–126. doi: 10.1083/jcb.200407024
- Chapman, P. F., Atkins, C. M., Allen, M. T., Haley, J. E., and Steinmetz, J. E. (1992). Inhibition of nitric oxide synthesis impairs two different forms of learning. *Neuroreport* 3, 567–570. doi: 10.1097/00001756-199207000-00005
- Cheah, J. H., Kim, S. F., Hester, L. D., Clancy, K. W., Patterson, S. E. 3rd., Papadopoulos, V., et al. (2006). NMDA receptor-nitric oxide transmission mediates neuronal iron homeostasis via the GTPase Dexas1. *Neuron* 51, 431–440. doi: 10.1016/j.neuron.2006.07.011
- Cobb, B. L., Ryan, K. L., Frei, M. R., Guel-Gomez, V., and Mickle, G. A. (1995). Chronic administration of L-NAME in drinking water alters working memory in rats. *Brain Res. Bull.* 38, 203–207. doi: 10.1016/0361-9230(95)00095-V
- Contestabile, A. (2000). Roles of NMDA receptor activity and nitric oxide production in brain development. *Brain Res. Brain Res. Rev.* 32, 476–509. doi: 10.1016/S0165-0173(00)00018-7
- Corsani, L., Bizzoco, E., Pedata, F., Gianfriddo, M., Fausone-Pellegrini, M. S., and Vannucchi, M. G. (2008). Inducible nitric oxide synthase appears and is co-expressed with the neuronal isoform in interneurons of the rat hippocampus after transient ischemia induced by middle cerebral artery occlusion. *Exp. Neurol.* 211, 433–440. doi: 10.1016/j.expneurol.2008.02.008
- Dachtler, J., Hardingham, N. R., and Fox, K. (2012). The role of nitric oxide synthase in cortical plasticity is sex specific. *J. Neurosci.* 32, 14994–14999. doi: 10.1523/JNEUROSCI.3189-12.2012
- Dachtler, J., Hardingham, N. R., Glazewski, S., Wright, N. F., Blain, E. J., and Fox, K. (2011). Experience-dependent plasticity acts via GluR1 and a novel neuronal nitric oxide synthase-dependent synaptic mechanism in adult cortex. *J. Neurosci.* 31, 11220–11230. doi: 10.1523/JNEUROSCI.1590-11.2011
- Dere, E., Frisch, C., De Souza Silva, M. A., Godecke, A., Schrader, J., and Huston, J. P. (2001). Unaltered radial maze performance and brain acetylcholine of the endothelial nitric oxide synthase knockout mouse. *Neuroscience* 107, 561–570. doi: 10.1016/S0306-4522(01)00382-7
- Di Stasi, A. M., Mallozzi, C., Macchia, G., Maura, G., Petrucci, T. C., and Minetti, M. (2002). Peroxynitrite affects exocytosis and SNARE complex formation and induces tyrosine nitration of synaptic proteins. *J. Neurochem.* 82, 420–429. doi: 10.1046/j.1471-4159.2002.00980.x
- Dinerman, J. L., Dawson, T. M., Schell, M. J., Snowman, A., and Snyder, S. H. (1994). Endothelial nitric oxide synthase localized to hippocampal pyramidal cells: implications for synaptic plasticity. *Proc. Natl. Acad. Sci. U.S.A.* 91, 4214–4218. doi: 10.1073/pnas.91.10.4214
- Doucet, M. V., Harkin, A., and Dev, K. K. (2012). The PSD-95/nNOS complex: new drugs for depression? *Pharmacol. Ther.* 133, 218–229. doi: 10.1016/j.pharmthera.2011.11.005
- Doyle, C., Holscher, C., Rowan, M. J., and Anwyl, R. (1996). The selective neuronal NO synthase inhibitor 7-nitro-indazole blocks both long-term potentiation and depotentiation of field EPSPs in rat hippocampal CA1 *in vivo*. *J. Neurosci.* 16, 418–424.
- Eguchi, K., Nakanishi, S., Takagi, H., Taoufiq, Z., and Takahashi, T. (2012). Maturation of a PKG-dependent retrograde mechanism for exocytotic coupling of synaptic vesicles. *Neuron* 74, 517–529. doi: 10.1016/j.neuron.2012.03.028
- Eliasson, M. J., Blackshaw, S., Schell, M. J., and Snyder, S. H. (1997). Neuronal nitric oxide synthase alternatively spliced forms: prominent functional localizations in the brain. *Proc. Natl. Acad. Sci. U.S.A.* 94, 3396–3401. doi: 10.1073/pnas.94.7.3396
- Fang, M., Jaffrey, S. R., Sawa, A., Ye, K., Luo, X., and Snyder, S. H. (2000). Dexas1: a G protein specifically coupled to neuronal nitric oxide synthase via CAPON. *Neuron* 28, 183–193. doi: 10.1016/S0896-6273(00)00095-7
- Feil, R., and Kleppisch, T. (2008). NO/cGMP-dependent modulation of synaptic transmission. *Handb. Exp. Pharmacol.* 184, 529–560. doi: 10.1007/978-3-540-74805-2_16
- Florio, S. K., Loh, C., Huang, S. M., Iwamaye, A. E., Kitto, K. F., Fowler, K. W., et al. (2009). Disruption of nNOS-PSD95 protein-protein interaction inhibits acute thermal hyperalgesia and chronic mechanical allodynia in rodents. *Br. J. Pharmacol.* 158, 494–506. doi: 10.1111/j.1476-5381.2009.00300.x
- Fox, K. (1992). A critical period for experience-dependent synaptic plasticity in rat barrel cortex. *J. Neurosci.* 12, 1826–1838.
- Friebe, A., and Koesling, D. (2009). The function of NO-sensitive guanylyl cyclase: what we can learn from genetic mouse models. *Nitric Oxide* 21, 149–156. doi: 10.1016/j.niox.2009.07.004
- Frisch, C., Dere, E., Silva, M. A., Godecke, A., Schrader, J., and Huston, J. P. (2000). Superior water maze performance and increase in fear-related behavior in the endothelial nitric oxide synthase-deficient mouse together with monoamine changes in cerebellum and ventral striatum. *J. Neurosci.* 20, 6694–6700.
- Garthwaite, J. (2008). Concepts of neural nitric oxide-mediated transmission. *Eur. J. Neurosci.* 27, 2783–2802. doi: 10.1111/j.1460-9568.2008.06285.x
- Garthwaite, J. (2010). New insight into the functioning of nitric

- oxide-receptive guanylyl cyclase: physiological and pharmacological implications. *Mol. Cell Biochem.* 334, 221–232. doi: 10.1007/s11010-009-0318-8
- Garthwaite, J., Charles, S. L., and Chess-Williams, R. (1988). Endothelium-derived relaxing factor release on activation of NMDA receptors suggests role as intercellular messenger in the brain. *Nature* 336, 385–388. doi: 10.1038/336385a0
- Gibb, B. J., and Garthwaite, J. (2001). Subunits of the nitric oxide receptor, soluble guanylyl cyclase, expressed in rat brain. *Eur. J. Neurosci.* 13, 539–544. doi: 10.1046/j.1460-9568.2001.01421.x
- Gordon, G. R., Mulligan, S. J., and Macvicar, B. A. (2007). Astrocyte control of the cerebrovasculature. *Glia* 55, 1214–1221. doi: 10.1002/glia.20543
- Griffiths, C., Wykes, V., Bellamy, T. C., and Garthwaite, J. (2003). A new and simple method for delivering clamped nitric oxide concentrations in the physiological range: application to activation of guanylyl cyclase-coupled nitric oxide receptors. *Mol. Pharmacol.* 64, 1349–1356. doi: 10.1124/mol.64.6.1349
- Haghikia, A., Mergia, E., Friebe, A., Eysel, U. T., Koesling, D., and Mittmann, T. (2007). Long-term potentiation in the visual cortex requires both nitric oxide receptor guanylyl cyclases. *J. Neurosci.* 27, 818–823. doi: 10.1523/JNEUROSCI.4706-06.2007
- Haley, J. E., Malen, P. L., and Chapman, P. F. (1993). Nitric oxide synthase inhibitors block long-term potentiation induced by weak but not strong tetanic stimulation at physiological brain temperatures in rat hippocampal slices. *Neurosci. Lett.* 160, 85–88. doi: 10.1016/0304-3940(93)90919-C
- Hall, C. N., and Garthwaite, J. (2009). What is the real physiological NO concentration in vivo? *Nitric Oxide* 21, 92–103. doi: 10.1016/j.niox.2009.07.002
- Hardingham, N., and Fox, K. (2006). The role of nitric oxide and GluR1 in presynaptic and postsynaptic components of neocortical potentiation. *J. Neurosci.* 26, 7395–7404. doi: 10.1523/JNEUROSCI.0652-06.2006
- Hardingham, N. R., Hardingham, G. E., Fox, K. D., and Jack, J. J. (2007). Presynaptic efficacy directs normalization of synaptic strength in layer 2/3 rat neocortex after paired activity. *J. Neurophysiol.* 97, 2965–2975. doi: 10.1152/jn.01352.2006
- Heinrich, T. A., Da Silva, R. S., Miranda, K. M., Switzer, C. H., Wink, D. A., and Fukuto, J. M. (2013). Biological nitric oxide signalling: chemistry and terminology. *Br. J. Pharmacol.* 169, 1417–1429. doi: 10.1111/bph.12217
- Hickey, M. J. (2001). Role of inducible nitric oxide synthase in the regulation of leucocyte recruitment. *Clin. Sci. (Lond.)* 100, 1–12. doi: 10.1042/CS20000135
- Holmgren, C. D., and Zilberter, Y. (2001). Coincident spiking activity induces long-term changes in inhibition of neocortical pyramidal cells. *J. Neurosci.* 21, 8270–8277.
- Holscher, C. (1997). Nitric oxide, the enigmatic neuronal messenger: its role in synaptic plasticity. *Trends Neurosci.* 20, 298–303. doi: 10.1016/S0166-2236(97)01065-5
- Holscher, C., Mcglinchey, L., Anwyl, R., and Rowan, M. J. (1996). 7-Nitro indazole, a selective neuronal nitric oxide synthase inhibitor *in vivo*, impairs spatial learning in the rat. *Learn. Mem.* 2, 267–278. doi: 10.1101/lm.2.6.267
- Hopper, R. A., and Garthwaite, J. (2006). Tonic and phasic nitric oxide signals in hippocampal long-term potentiation. *J. Neurosci.* 26, 11513–11521. doi: 10.1523/JNEUROSCI.2259-06.2006
- Huang, C. C., Chan, S. H., and Hsu, K. S. (2003). cGMP/protein kinase G-dependent potentiation of glutamatergic transmission induced by nitric oxide in immature rat rostral ventrolateral medulla neurons *in vitro*. *Mol. Pharmacol.* 64, 521–532. doi: 10.1124/mol.64.2.521
- Huang, P. L., Dawson, T. M., Bredt, D. S., Snyder, S. H., and Fishman, M. C. (1993). Targeted disruption of the neuronal nitric oxide synthase gene. *Cell* 75, 1273–1286. doi: 10.1016/0092-8674(93)90615-W
- Huang, Y., Man, H. Y., Sekine-Aizawa, Y., Han, Y., Juluri, K., Luo, H., et al. (2005). S-nitrosylation of N-ethylmaleimide sensitive factor mediates surface expression of AMPA receptors. *Neuron* 46, 533–540. doi: 10.1016/j.neuron.2005.03.028
- Huang, Z., Huang, P. L., Panahian, N., Dalkara, T., Fishman, M. C., and Moskowitz, M. A. (1994). Effects of cerebral ischemia in mice deficient in neuronal nitric oxide synthase. *Science* 265, 1883–1885. doi: 10.1126/science.7522345
- Husi, H., Ward, M. A., Choudhary, J. S., Blackstock, W. P., and Grant, S. G. (2000). Proteomic analysis of NMDA receptor-adhesion protein signaling complexes. *Nat. Neurosci.* 3, 661–669. doi: 10.1038/76615
- Jaffrey, S. R., Erdjument-Bromage, H., Ferris, C. D., Tempst, P., and Snyder, S. H. (2001). Protein S-nitrosylation: a physiological signal for neuronal nitric oxide. *Nat. Cell Biol.* 3, 193–197. doi: 10.1038/35055104
- Jaffrey, S. R., Snowman, A. M., Eliasson, M. J., Cohen, N. A., and Snyder, S. H. (1998). CAPON: a protein associated with neuronal nitric oxide synthase that regulates its interactions with PSD95. *Neuron* 20, 115–124. doi: 10.1016/S0896-6273(00)80439-0
- Johnstone, V. P., and Raymond, C. R. (2011). A protein synthesis and nitric oxide-dependent presynaptic enhancement in persistent forms of long-term potentiation. *Learn. Mem.* 18, 625–633. doi: 10.1101/lm.2245911
- Kaneko, M., Cheetham, C. E., Lee, Y. S., Silva, A. J., Stryker, M. P., and Fox, K. (2010). Constitutively active H-ras accelerates multiple forms of plasticity in developing visual cortex. *Proc. Natl. Acad. Sci. U.S.A.* 107, 19026–19031. doi: 10.1073/pnas.1013866107
- Kawaguchi, Y., Aosaki, T., and Kubota, Y. (1997). Cholinergic and GABAergic interneurons in the striatum. *Nihon Shinkei Seishin Yakurigaku Zasshi* 17, 87–90.
- Kelley, J. B., Balda, M. A., Anderson, K. L., and Itzhak, Y. (2009). Impairments in fear conditioning in mice lacking the nNOS gene. *Learn. Mem.* 16, 371–378. doi: 10.1101/lm.1329209
- Kerr, M. I., Wall, M. J., and Richardson, M. J. (2013). Adenosine A1-receptor activation mediates the developmental shift at layer-5 pyramidal-cell synapses and is a determinant of mature synaptic strength. *J. Physiol.* 591, 3371–3380. doi: 10.1113/jphysiol.2012.244392
- Kimura, S., Uchiyama, S., Takahashi, H. E., and Shibuki, K. (1998). cAMP-dependent long-term potentiation of nitric oxide release from cerebellar parallel fibers in rats. *J. Neurosci.* 18, 8551–8558.
- Kingston, P. A., Zufall, F., and Barnstable, C. J. (1999). Widespread expression of olfactory cyclic nucleotide-gated channel genes in rat brain: implications for neuronal signalling. *Synapse* 32, 1–12. doi: 10.1002/(SICI)1098-2396(199904)32:1<1::AID-SYNI>3.0.CO;2-M
- Klyachko, V. A., Ahern, G. P., and Jackson, M. B. (2001). cGMP-mediated facilitation in nerve terminals by enhancement of the spike afterhyperpolarization. *Neuron* 31, 1015–1025. doi: 10.1016/S0896-627300449-4
- Kodama, T., and Koyama, Y. (2006). Nitric oxide from the laterodorsal tegmental neurons: its possible retrograde modulation on norepinephrine release from the axon terminal of the locus coeruleus neurons. *Neuroscience* 138, 245–256. doi: 10.1016/j.neuroscience.2005.10.063
- Kubota, Y., Shigematsu, N., Karube, F., Sekigawa, A., Kato, S., Yamaguchi, N., et al. (2011). Selective coexpression of multiple chemical markers defines discrete populations of neocortical GABAergic neurons. *Cereb. Cortex* 21, 1803–1817. doi: 10.1093/cercor/bhq252
- Lange, M. D., Doengi, M., Lesting, J., Pape, H. C., and Jungling, K. (2012). Heterosynaptic long-term potentiation at interneuron-principal neuron synapses in the amygdala requires nitric oxide signalling. *J. Physiol.* 590, 131–143. doi: 10.1113/jphysiol.2011.221317
- Larkman, A., Hannay, T., Stratford, K., and Jack, J. (1992). Presynaptic release probability influences the locus of long-term potentiation. *Nature* 360, 70–73. doi: 10.1038/360070a0
- Le Roux, N., Amar, M., Baux, G., and Fossier, P. (2006). Homeostatic control of the excitation-inhibition balance in cortical layer 5 pyramidal neurons. *Eur. J. Neurosci.* 24, 3507–3518. doi: 10.1111/j.1460-9568.2006.05203.x
- Le Roux, N., Amar, M., Moreau, A. W., and Fossier, P. (2009). Roles of nitric oxide in the homeostatic control of the excitation-inhibition balance in rat visual cortical networks. *Neuroscience* 163, 942–951. doi: 10.1016/j.neuroscience.2009.07.010
- Leinders-Zufall, T., Cockerham, R. E., Michalakis, S., Biel, M., Garbers, D. L., Reed, R. R., et al. (2007). Contribution of the receptor guanylyl cyclase GC-D to chemosensory function in the olfactory epithelium. *Proc. Natl. Acad. Sci. U.S.A.* 104, 14507–14512. doi: 10.1073/pnas.0704965104
- Li, C. H., Wang, S. Z., Cai, Z. L., Liu, W. X., Xu, S. T., and Xiao, P.

- (2012). Effects of intrahippocampal L-NAME treatment on the behavioral long-term potentiation in dentate gyrus. *Neurosci. Lett.* 528, 201–204. doi: 10.1016/j.neulet.2012.08.056
- Li, D. P., Chen, S. R., and Pan, H. L. (2002). Nitric oxide inhibits spinally projecting paraventricular neurons through potentiation of presynaptic GABA release. *J. Neurophysiol.* 88, 2664–2674. doi: 10.1152/jn.00540.2002
- Lindskog, M., Li, L., Groth, R. D., Poburko, D., Thiagarajan, T. C., Han, X., et al. (2010). Postsynaptic GluA1 enables acute retrograde enhancement of presynaptic function to coordinate adaptation to synaptic inactivity. *Proc. Natl. Acad. Sci. U.S.A.* 107, 21806–21811. doi: 10.1073/pnas.1106399107
- Majlessi, N., Choopani, S., Bozorgmehr, T., and Azizi, Z. (2008). Involvement of hippocampal nitric oxide in spatial learning in the rat. *Neurobiol. Learn. Mem.* 90, 413–419. doi: 10.1016/j.nlm.2008.04.010
- Malen, P. L., and Chapman, P. F. (1997). Nitric oxide facilitates long-term potentiation, but not long-term depression. *J. Neurosci.* 17, 2645–2651.
- Malinow, R., and Tsien, R. W. (1990). Presynaptic enhancement shown by whole-cell recordings of long-term potentiation in hippocampal slices. *Nature* 346, 177–180. doi: 10.1038/346177a0
- Martin, E., Berka, V., Bogatenkova, E., Murad, F., and Tsai, A. L. (2006). Ligand selectivity of soluble guanylyl cyclase: effect of the hydrogen-bonding tyrosine in the distal heme pocket on binding of oxygen, nitric oxide, and carbon monoxide. *J. Biol. Chem.* 281, 27836–27845. doi: 10.1074/jbc.M601078200
- McCullough, L. D., Zeng, Z., Blizzard, K. K., Debchoudhury, I., and Hurn, P. D. (2005). Ischemic nitric oxide and poly (ADP-ribose) polymerase-1 in cerebral ischemia: male toxicity, female protection. *J. Cereb. Blood Flow Metab.* 25, 502–512. doi: 10.1038/sj.jcbfm.9600059
- Meffert, M. K., Calakos, N. C., Scheller, R. H., and Schulman, H. (1996). Nitric oxide modulates synaptic vesicle docking fusion reactions. *Neuron* 16, 1229–1236. doi: 10.1016/S0896-6273(00)80149-X
- Meffert, M. K., Premack, B. A., and Schulman, H. (1994). Nitric oxide stimulates Ca(2+)-independent synaptic vesicle release. *Neuron* 12, 1235–1244. doi: 10.1016/0896-6273(94)90440-5
- Mergia, E., Russwurm, M., Zoidl, G., and Koesling, D. (2003). Major occurrence of the new alpha2beta1 isoform of NO-sensitive guanylyl cyclase in brain. *Cell Signal.* 15, 189–195. doi: 10.1016/S0898-6568(02)00078-5
- Micheva, K. D., Buchanan, J., Holz, R. W., and Smith, S. J. (2003). Retrograde regulation of synaptic vesicle endocytosis and recycling. *Nat. Neurosci.* 6, 925–932. doi: 10.1038/nn1114
- Monier, C., Fournier, J., and Fregnac, Y. (2008). *In vitro* and *in vivo* measures of evoked excitatory and inhibitory conductance dynamics in sensory cortices. *J. Neurosci. Methods* 169, 323–365. doi: 10.1016/j.jneumeth.2007.11.008
- Montague, P. R., Gancayco, C. D., Winn, M. J., Marchase, R. B., and Friedlander, M. J. (1994). Role of NO production in NMDA receptor-mediated neurotransmitter release in cerebral cortex. *Science* 263, 973–977. doi: 10.1126/science.7508638
- Moreno-Lopez, B., Escudero, M., and Estrada, C. (2002). Nitric oxide facilitates GABAergic neurotransmission in the cat oculomotor system: a physiological mechanism in eye movement control. *J. Physiol.* 540, 295–306. doi: 10.1113/jphysiol.2001.013308
- Neitz, A., Mergia, E., Eysel, U. T., Koesling, D., and Mittmann, T. (2011). Presynaptic nitric oxide/cGMP facilitates glutamate release via hyperpolarization-activated cyclic nucleotide-gated channels in the hippocampus. *Eur. J. Neurosci.* 33, 1611–1621. doi: 10.1111/j.1460-9568.2011.07654.x
- Nikonenko, I., Boda, B., Steen, S., Knott, G., Welker, E., and Muller, D. (2008). PSD-95 promotes synaptogenesis and multiinnervated spine formation through nitric oxide signaling. *J. Cell Biol.* 183, 1115–1127. doi: 10.1083/jcb.2008.05132
- Nomura, Y., and Kitamura, Y. (1993). Inducible nitric oxide synthase in glial cells. *Neurosci Res* 18, 103–107. doi: 10.1016/0168-0102(93)90013-G
- Nugent, F. S., Niehaus, J. L., and Kauer, J. A. (2009). PKG and PKA signaling in LTP at GABAergic synapses. *Neuropsychopharmacology* 34, 1829–1842. doi: 10.1038/npp.2009.5
- Nugent, F. S., Penick, E. C., and Kauer, J. A. (2007). Opioids block long-term potentiation of inhibitory synapses. *Nature* 446, 1086–1090. doi: 10.1038/nature05726
- O'Dell, T. J., Hawkins, R. D., Kandel, E. R., and Arancio, O. (1991). Tests of the roles of two diffusible substances in long-term potentiation: evidence for nitric oxide as a possible early retrograde messenger. *Proc. Natl. Acad. Sci. U.S.A.* 88, 11285–11289. doi: 10.1073/pnas.88.24.11285
- Ondracek, J. M., Dec, A., Hoque, K. E., Lim, S. A., Rasouli, G., Indorkar, R. P., et al. (2008). Feed-forward excitation of striatal neuron activity by frontal cortical activation of nitric oxide signaling *in vivo*. *Eur. J. Neurosci.* 27, 1739–1754. doi: 10.1111/j.1460-9568.2008.06157.x
- Palmer, Z. J., Duncan, R. R., Johnson, J. R., Lian, L. Y., Mello, L. V., Booth, D., et al. (2008). S-nitrosylation of syntaxin 1 at Cys(145) is a regulatory switch controlling Munc18-1 binding. *Biochem. J.* 413, 479–491. doi: 10.1042/BJ20080069
- Parent, A., Schrader, K., Munger, S. D., Reed, R. R., Linden, D. J., and Ronnett, G. V. (1998). Synaptic transmission and hippocampal long-term potentiation in olfactory cyclic nucleotide-gated channel type 1 null mouse. *J. Neurophysiol.* 79, 3295–3301.
- Phillips, K. G., Hardingham, N. R., and Fox, K. (2008). Postsynaptic action potentials are required for nitric oxide-dependent long-term potentiation in CA1 neurons of adult GluR1 knock-out and wild-type mice. *J. Neurosci.* 28, 14031–14041. doi: 10.1523/JNEUROSCI.3984-08.2008
- Poglia, L., Muller, D., and Nikonenko, I. (2011). Ultrastructural modifications of spine and synapse morphology by SAP97. *Hippocampus* 21, 990–998. doi: 10.1002/hipo.20811
- Priebe, N. J., and Ferster, D. (2005). Direction selectivity of excitation and inhibition in simple cells of the cat primary visual cortex. *Neuron* 45, 133–145. doi: 10.1016/j.neuron.2004.12.024
- Prince, D. A., and Stevens, C. F. (1992). Adenosine decreases neurotransmitter release at central synapses. *Proc. Natl. Acad. Sci. U.S.A.* 89, 8586–8590. doi: 10.1073/pnas.89.18.8586
- Prior, I. A., and Clague, M. J. (2000). Detection of thiol modification following generation of reactive nitrogen species: analysis of synaptic vesicle proteins. *Biochim. Biophys. Acta* 1475, 281–286. doi: 10.1016/S0304-4165(00)00078-7
- Qiang, M., Chen, Y. C., Wang, R., Wu, F. M., and Qiao, J. T. (1997). Nitric oxide is involved in the formation of learning and memory in rats: studies using passive avoidance response and Morris water maze task. *Behav. Pharmacol.* 8, 183–187.
- Qiu, D. L., and Knopfel, T. (2007). An NMDA receptor/nitric oxide cascade in presynaptic parallel fiber-Purkinje neuron long-term potentiation. *J. Neurosci.* 27, 3408–3415. doi: 10.1523/JNEUROSCI.4831-06.2007
- Ratnayaka, A., Marra, V., Bush, D., Burden, J. J., Branco, T., and Staras, K. (2012). Recruitment of resting vesicles into recycling pools supports NMDA receptor-dependent synaptic potentiation in cultured hippocampal neurons. *J. Physiol.* 590, 1585–1597. doi: 10.1113/jphysiol.2011.226688
- Rieke, F., and Schwartz, E. A. (1994). A cGMP-gated current can control exocytosis at cone synapses. *Neuron* 13, 863–873. doi: 10.1016/0896-6273(94)90252-6
- Romberg, C., Raffel, J., Martin, L., Sprengel, R., Seeburg, P. H., Rawlins, J. N., et al. (2009). Induction and expression of GluA1 (GluR-A)-independent LTP in the hippocampus. *Eur. J. Neurosci.* 29, 1141–1152. doi: 10.1111/j.1460-9568.2009.06677.x
- Roy, B., Halvey, E. J., and Garthwaite, J. (2008). An enzyme-linked receptor mechanism for nitric oxide-activated guanylyl cyclase. *J. Biol. Chem.* 283, 18841–18851. doi: 10.1074/jbc.M801712200
- Russwurm, M., Wittau, N., and Koesling, D. (2001). Guanylyl cyclase/PSD-95 interaction: targeting of the nitric oxide-sensitive alpha2beta1 guanylyl cyclase to synaptic membranes. *J. Biol. Chem.* 276, 44647–44652. doi: 10.1074/jbc.M105587200
- Saha, R. N., and Pahan, K. (2006). Signals for the induction of nitric oxide synthase in astrocytes. *Neurochem. Int.* 49, 154–163. doi: 10.1016/j.neuint.2006.04.007
- Sammur, S., Bray, K. E., and West, A. R. (2007a). Dopamine D2 receptor-dependent modulation of striatal NO synthase activity. *Psychopharmacology (Berl.)* 191,

- 793–803. doi: 10.1007/s00213-006-0681-z
- Sammur, S., Park, D. J., and West, A. R. (2007b). Frontal cortical afferents facilitate striatal nitric oxide transmission *in vivo* via a NMDA receptor and neuronal NOS-dependent mechanism. *J. Neurochem.* 103, 1145–1156. doi: 10.1111/j.1471-4159.2007.04811.x
- Sammur, S., Dec, A., Mitchell, D., Linardakis, J., Ortigueira, M., and West, A. R. (2006). Phasic dopaminergic transmission increases NO efflux in the rat dorsal striatum via a neuronal NOS and a dopamine D(1/5) receptor-dependent mechanism. *Neuropsychopharmacology* 31, 493–505. doi: 10.1038/sj.npp.1300826
- Sammur, S., and West, A. R. (2008). Acute cocaine administration increases NO efflux in the rat prefrontal cortex via a neuronal NOS-dependent mechanism. *Synapse* 62, 710–713. doi: 10.1002/syn.20537
- Santolini, J., Adak, S., Curran, C. M., and Stuehr, D. J. (2001). A kinetic simulation model that describes catalysis and regulation in nitric-oxide synthase. *J. Biol. Chem.* 276, 1233–1243. doi: 10.1074/jbc.M006858200
- Sato, M., Nakajima, T., Goto, M., and Umezawa, Y. (2006). Cell-based indicator to visualize picomolar dynamics of nitric oxide release from living cells. *Anal. Chem.* 78, 8175–8182. doi: 10.1021/ac061791b
- Schlossmann, J., and Hofmann, F. (2005). cGMP-dependent protein kinases in drug discovery. *Drug Discov. Today* 10, 627–634. doi: 10.1016/S1359-6446(05)03406-9
- Schuman, E. M., and Madison, D. V. (1991). A requirement for the intercellular messenger nitric oxide in long-term potentiation. *Science* 254, 1503–1506. doi: 10.1126/science.1720572
- Serulle, Y., Arancio, O., and Ziff, E. B. (2008). A role for cGMP-dependent protein kinase II in AMPA receptor trafficking and synaptic plasticity. *Channels (Austin)* 2, 230–232. doi: 10.4161/chan.2.4.6391
- Shibuki, K., and Kimura, S. (1997). Dynamic properties of nitric oxide release from parallel fibres in rat cerebellar slices. *J. Physiol.* 498 (Pt 2), 443–452.
- Sjostrom, P. J., Turrigiano, G. G., and Nelson, S. B. (2007). Multiple forms of long-term plasticity at unitary neocortical layer 5 synapses. *Neuropharmacology* 52, 176–184. doi: 10.1016/j.neuropharm.2006.07.021
- Stanton, P. K., Winterer, J., Bailey, C. P., Kyrozi, A., Raginov, I., Laube, G., et al. (2003). Long-term depression of presynaptic release from the readily releasable vesicle pool induced by NMDA receptor-dependent retrograde nitric oxide. *J. Neurosci.* 23, 5936–5944.
- Stanton, P. K., Winterer, J., Zhang, X. L., and Muller, W. (2005). Imaging LTP of presynaptic release of FM1-43 from the rapidly recycling vesicle pool of Schaffer collateral-CA1 synapses in rat hippocampal slices. *Eur. J. Neurosci.* 22, 2451–2461. doi: 10.1111/j.1460-9568.2005.04437.x
- Steinert, J. R., Chernova, T., and Forsythe, I. D. (2010). Nitric oxide signaling in brain function, dysfunction, and dementia. *Neuroscientist* 16, 435–452. doi: 10.1177/1073858410366481
- Steinert, J. R., Kopp-Scheinflug, C., Baker, C., Challiss, R. A., Mistry, R., Hausteiner, M. D., et al. (2008). Nitric oxide is a volume transmitter regulating postsynaptic excitability at a glutamatergic synapse. *Neuron* 60, 642–656. doi: 10.1016/j.neuron.2008.08.025
- Szabadits, E., Cserep, C., Ludanyi, A., Katona, I., Gracia-Llanes, J., Freund, T. F., et al. (2007). Hippocampal GABAergic synapses possess the molecular machinery for retrograde nitric oxide signaling. *J. Neurosci.* 27, 8101–8111. doi: 10.1523/JNEUROSCI.1912-07.2007
- Tanda, K., Nishi, A., Matsuo, N., Nakanishi, K., Yamasaki, N., Sugimoto, T., et al. (2009). Abnormal social behavior, hyperactivity, impaired remote spatial memory, and increased D1-mediated dopaminergic signaling in neuronal nitric oxide synthase knockout mice. *Mol. Brain* 2, 19. doi: 10.1186/1756-6606-2-19
- Taniguchi, H., He, M., Wu, P., Kim, S., Paik, R., Sugino, K., et al. (2011). A resource of Cre driver lines for genetic targeting of GABAergic neurons in cerebral cortex. *Neuron* 71, 995–1013. doi: 10.1016/j.neuron.2011.07.026
- Valtschanoff, J. G., and Weinberg, R. J. (2001). Laminar organization of the NMDA receptor complex within the postsynaptic density. *J. Neurosci.* 21, 1211–1217.
- Volgushev, M., Balaban, P., Chistiakova, M., and Eysel, U. T. (2000). Retrograde signalling with nitric oxide at neocortical synapses. *Eur. J. Neurosci.* 12, 4255–4267. doi: 10.1046/j.0953-816X.2000.01322.x
- Wakatsuki, H., Gomi, H., Kudoh, M., Kimura, S., Takahashi, K., Takeda, M., et al. (1998). Layer-specific NO dependence of long-term potentiation and biased NO release in layer V in the rat auditory cortex. *J. Physiol.* 513(Pt 1), 71–81. doi: 10.1111/j.1469-7793.1998.071by.x
- Wall, M. J. (2003). Endogenous nitric oxide modulates GABAergic transmission to granule cells in adult rat cerebellum. *Eur. J. Neurosci.* 18, 869–878. doi: 10.1046/j.1460-9568.2003.02822.x
- Wallace, H., and Fox, K. (1999). The effect of vibrissa deprivation pattern on the form of plasticity induced in rat barrel cortex. *Somatosens. Mot. Res.* 16, 122–138. doi: 10.1080/08990229970564
- Wang, H. G., Lu, F. M., Jin, I., Udo, H., Kandel, E. R., De Vente, J., et al. (2005). Presynaptic and postsynaptic roles of NO, cGK, and RhoA in long-lasting potentiation and aggregation of synaptic proteins. *Neuron* 45, 389–403. doi: 10.1016/j.neuron.2005.01.011
- Wehr, M., and Zador, A. M. (2003). Balanced inhibition underlies tuning and sharpens spike timing in auditory cortex. *Nature* 426, 442–446. doi: 10.1038/nature02116
- Wehr, M., and Zador, A. M. (2005). Synaptic mechanisms of forward suppression in rat auditory cortex. *Neuron* 47, 437–445. doi: 10.1016/j.neuron.2005.06.009
- Weitzdoerfer, R., Hoeger, H., Engidawork, E., Engelmann, M., Singewald, N., Lubec, G., et al. (2004). Neuronal nitric oxide synthase knock-out mice show impaired cognitive performance. *Nitric Oxide* 10, 130–140. doi: 10.1016/j.niox.2004.03.007
- Wendland, B., Schweizer, F. E., Ryan, T. A., Nakane, M., Murad, F., Scheller, R. H., et al. (1994). Existence of nitric oxide synthase in rat hippocampal pyramidal cells. *Proc. Natl. Acad. Sci. U.S.A.* 91, 2151–2155. doi: 10.1073/pnas.91.6.2151
- West, A. R., Galloway, M. P., and Grace, A. A. (2002). Regulation of striatal dopamine neurotransmission by nitric oxide: effector pathways and signaling mechanisms. *Synapse* 44, 227–245. doi: 10.1002/syn.10076
- Wildemann, B., and Bicker, G. (1999). Nitric oxide and cyclic GMP induce vesicle release at *Drosophila* neuromuscular junction. *J. Neurobiol.* 39, 337–346. doi: 10.1002/(SICI)1097-4695(19990605)39:3<337::AID-NEU1>3.0.CO;2-9
- Wilent, W. B., and Contreras, D. (2005). Dynamics of excitation and inhibition underlying stimulus selectivity in rat somatosensory cortex. *Nat. Neurosci.* 8, 1364–1370. doi: 10.1038/nn1545
- Wiseman, D. A., Kalwat, M. A., and Thurmond, D. C. (2011). Stimulus-induced S-nitrosylation of Syntaxin 4 impacts insulin granule exocytosis. *J. Biol. Chem.* 286, 16344–16354. doi: 10.1074/jbc.M110.214031
- Wood, K. C., Batchelor, A. M., Bartus, K., Harris, K. L., Garthwaite, G., Vernon, J., et al. (2011). Picomolar nitric oxide signals from central neurons recorded using ultrasensitive detector cells. *J. Biol. Chem.* 286, 43172–43181. doi: 10.1074/jbc.M111.289777
- Wu, W. C., Wang, Y., Kao, L. S., Tang, F. I., and Chai, C. Y. (2002). Nitric oxide reduces blood pressure in the nucleus tractus solitarius: a real time electrochemical study. *Brain. Res. Bull.* 57, 171–177. doi: 10.1016/S0361-9230(01)00737-7
- Wu, W. C., Wang, Y., Su, C. K., and Chai, C. Y. (2001). The nNOS/cGMP signal transducing system is involved in the cardiovascular responses induced by activation of NMDA receptors in the rostral ventrolateral medulla of cats. *Neurosci. Lett.* 310, 121–124. doi: 10.1016/S0304-3940(01)02100-0
- Xue, J. G., Masuoka, T., Gong, X. D., Chen, K. S., Yanagawa, Y., Law, S. K., et al. (2011). NMDA receptor activation enhances inhibitory GABAergic transmission onto hippocampal pyramidal neurons via presynaptic and postsynaptic mechanisms. *J. Neurophysiol.* 105, 2897–2906. doi: 10.1152/jn.00287.2010
- Yang, Q., Chen, S. R., Li, D. P., and Pan, H. L. (2007). Kv1.1/1.2 channels are downstream effectors of nitric oxide on synaptic GABA release to preautonomic neurons in the paraventricular nucleus. *Neuroscience* 149, 315–327. doi: 10.1016/j.neuroscience.2007.08.007
- Yildiz Akar, F., Celikyurt, I. K., Ulak, G., and Mutlu, O. (2009). Effects of L-arginine on 7-nitroindazole-induced reference and working memory performance of rats. *Pharmacology* 84, 211–218. doi: 10.1159/000235997

- Zhou, L., Li, F., Xu, H. B., Luo, C. X., Wu, H. Y., Zhu, M. M., et al. (2010). Treatment of cerebral ischemia by disrupting ischemia-induced interaction of nNOS with PSD-95. *Nat. Med.* 16, 1439–1443. doi: 10.1038/nm.2245
- Zhou, L., and Zhu, D. Y. (2009). Neuronal nitric oxide synthase: structure, subcellular localization, regulation, and clinical implications. *Nitric Oxide* 20, 223–230. doi: 10.1016/j.niox.2009.03.001
- Zou, L. B., Yamada, K., Tanaka, T., Kameyama, T., and Nabeshima, T. (1998). Nitric oxide synthase inhibitors impair reference memory formation in a radial arm maze task in rats. *Neuropharmacology* 37, 323–330. doi: 10.1016/S0028-3908(98)00042-2
- Conflict of Interest Statement:** The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.
- Received: 13 August 2013; paper pending published: 11 September 2013; accepted: 05 October 2013; published online: 31 October 2013.*
- Citation: Hardingham N, Dachtler J and Fox K (2013) The role of nitric oxide in pre-synaptic plasticity and homeostasis. Front. Cell. Neurosci. 7:190. doi: 10.3389/fncel.2013.00190*
- This article was submitted to the journal Frontiers in Cellular Neuroscience.*
- Copyright © 2013 Hardingham, Dachtler and Fox. This is an open-access article distributed under the terms of the Creative Commons Attribution License (CC BY). The use, distribution or reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.*