



Draft Genome Sequence of *Paucibacter aquatile* CR182^T, a Strain with Antimicrobial Activity Isolated from Freshwater of Nakdong River in South Korea

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ABSTRACT This report details a draft genome sequence of *Paucibacter aquatile* CR182^T, isolated from river water, which contains 5,523,543 bp, has a G+C content of 66.3%, and harbors 4,544 protein-coding genes in 4 contigs. These genome data provide insights into the genetic basis of this strain's antibacterial activity and adaptive mechanisms.

The genus *Paucibacter* was first proposed in 2005 (1) with the type species *Paucibacter toxinivorans*, which belongs to the family *Comamonadaceae* in the phylum *Betaproteobacteria*. Members of the genus *Paucibacter* characteristically are Gram stain negative, rod shaped, and often resistant to antibiotics or toxic compounds (1–3). Here, we report the draft genome sequence of *Paucibacter aquatile* CR182^T, isolated from freshwater of the Nakdong River, South Korea, during a screen for bacteria with antimicrobial activity (3). Strain CR182^T has been deposited in the Korean Culture Center of Microorganisms (KCCM) and the Nite Biological Resource Center (NBRC) under the accession numbers KCCM 9028 and NBRC 113032, respectively.

Genomic DNA was extracted using a genomic DNA purification kit (Promega). A 20-kb SMRTbell genome library was sequenced using the PacBio RS II single-molecule real-time (SMRT) sequencing platform at ChunLab (Seoul, South Korea). The bacterial genome was assembled *de novo* into four contigs, with an average genome coverage of 230.9×, using the PacBio SMRT Portal (2.3.0) and the Hierarchical Genome Assembly Process (HGAP) (4). The Prokaryotic Genome Annotation System (Prokka) pipeline was used for genome annotation (5). The data were submitted to the Rapid Annotations using Subsystem Technology (RAST) server (6) and the National Center for Biotechnology Information (NCBI) genome sequence database. We searched for potential coding sequences using the Basic Local Alignment Search Tool (BLAST) against the UniProt (7), Pfam (8), and Clusters of Orthologous Groups of proteins (COG) (9) databases. Signal peptides and transmembrane helices were predicted using SignalP 4.1 (10) and TMHMM v2.0 (11). rRNA, tRNA, and other miscellaneous features were predicted using RNAmmer v1.2 (12), tRNAscan-SE v1.21 (13), and Rfam v12.0 (14).

The draft genome sequence of *P. aquatile* CR182^T is 5,523,543 bp, with a G+C content of 66.3%. The genome is predicted to have 4,544 coding sequences (CDSs), 72 tRNA genes, and 28 rRNA genes. We identified various antibacterial genes in the genome of *P. aquatile* CR182^T; antiSMASH shell analysis (15) revealed that the genome harbors 9 gene clusters involved in the biosynthesis of lantipeptide, hserlactone, bacteriocin, terpene, and a nonribosomal peptide synthetase (NRPS), a kind of amyelin. Analysis using the Antibiotic Resistance Genes Database (ARDB) (16) led to the identification of several genes (*marB*, *mecR*, *pbp2b*, *catB2*, *bacA*, *smeD*, and *oprM*) putatively involved in antibiotic resistance, some conferring specific resistance to

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macrolides, methicillin, penicillin, chloramphenicol, bacitracin, fluoroquinolone, and/or aminoglycosides.

The genome sequence and analysis of *P. aquatile* CR182^T in this study provide information to further explore and reveal the mechanisms of antimicrobial action in aquatic environments.

Accession number(s). This whole-genome shotgun project has been deposited at DDBJ/ENA/GenBank under the accession number [POSP00000000](https://www.ncbi.nlm.nih.gov/BioProject/PRJNA429314) (BioProject number PRJNA429314). The version described in this paper is the first version.

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REFERENCES

- Rapala J, Berg KA, Lyra C, Niemi M, Manz W, Suomalainen S, Paulin L, Jahti K. 2005. *Paucibacter toxinivorans* gen. nov., sp. nov., a bacterium that degrades cyclic cyanobacterial hepatotoxins microcystins and nodularin. *Int J Syst Evol Microbiol* 55:1563–1568. <https://doi.org/10.1099/ijs.0.63599-0>.
- Pheng S, Lee JJ, Eom MK, Lee KH, Kim S. 2017. *Paucibacter oligotrophus* sp. nov., isolated from fresh water, and emended description of the genus *Paucibacter*. *Int J Syst Evol Microbiol* 67:2231–2235. <https://doi.org/10.1099/ijs.0.001931>.
- Nam YH, Choi A, Hwang JM, Yim KJ, Kim J-H, Choi G-G, Chung EJ. 2018. *Paucibacter aquatile* sp. nov., isolated from freshwater of the Nakdong River, Republic of Korea. *Arch Microbiol in press*. <https://doi.org/10.1007/s00203-018-1494-2>.
- Chin C-S, Alexander DH, Marks P, Klammer AA, Drake J, Heiner C, Clum A, Copeland A, Huddleston J, Eichler EE, Turner SW, Korlach J. 2013. Nonhybrid, finished microbial genome assemblies from long-read SMRT sequencing data. *Nat Methods* 10:563–569. <https://doi.org/10.1038/nmeth.2474>.
- Seemann T. 2014. Prokka: rapid prokaryotic genome annotation. *Bioinformatics* 30:2068–2069. <https://doi.org/10.1093/bioinformatics/btu153>.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paccian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: Rapid Annotations using Subsystems Technology. *BMC Genomics* 9:75. <https://doi.org/10.1186/1471-2164-9-75>.
- Wu CH, Apweiler R, Bairoch A, Natale DA, Barker WC, Boeckmann B, Ferro S, Gasteiger E, Huang H, Lopez R, Magrane M, Martin MJ, Mazumder R, O'Donovan C, Redaschi N, Suzek B. 2006. The Universal Protein Resource (UniProt): an expanding universe of protein information. *Nucleic Acids Res* 34:D187–D191. <https://doi.org/10.1093/nar/gkj161>.
- Punta M, Coghill PC, Eberhardt RY, Mistry J, Tate J, Boursnell C, Pang N, Forslund K, Ceric G, Clements J, Heger A, Holm L, Sonnhammer EL, Eddy SR, Bateman A, Finn RD. 2012. The Pfam protein families database. *Nucleic Acids Res* 40:D290–D301. <https://doi.org/10.1093/nar/gkr1065>.
- Tatusov RL, Fedorova ND, Jackson JD, Jacobs AR, Kiryutin B, Koonin EV, Krylov DM, Mazumdr R, Mekhedov SL, Nikolskaya AN, Rao BS, Smirnov S, Sverdlov AV, Vasudevan S, Wolf YI, Yin JJ, Natale DA. 2003. The COG database: an updated version includes eukaryotes. *BMC Bioinformatics* 4:41. <https://doi.org/10.1186/1471-2105-4-41>.
- Petersen TN, Brunak S, von Heijne G, Nielsen H. 2011. SignalP 4.0: discriminating signal peptides from transmembrane regions. *Nat Methods* 8:785–786. <https://doi.org/10.1038/nmeth.1701>.
- Krogh A, Larsson B, Von Heijne G, Sonnhammer EL. 2001. Predicting transmembrane protein topology with a hidden Markov model: application to complete genomes. *J Mol Biol* 305:567–580. <https://doi.org/10.1006/jmbi.2000.4315>.
- Lagesen K, Hallin P, Rødland EA, Stærfeldt HH, Rognes T, Ussery DW. 2007. RNAmmer: consistent and rapid annotation of ribosomal RNA genes. *Nucleic Acids Res* 35:3100–3108. <https://doi.org/10.1093/nar/gkm160>.
- Lowe TM, Eddy SR. 1997. tRNAscan-SE: a program for improved detection of transfer RNA genes in genomic sequence. *Nucleic Acids Res* 25:955–964.
- Griffiths-Jones S, Moxon S, Marshall M, Khanna A, Eddy SR, Bateman A. 2005. Rfam: annotating non-coding RNAs in complete genomes. *Nucleic Acids Res* 33:D121–D124. <https://doi.org/10.1093/nar/gki081>.
- Medema MH, Blin K, Cimermancic P, de Jager V, Zakrzewski P, Fischbach MA, Weber T, Takano E, Breitling R. 2011. antiSMASH: rapid identification, annotation and analysis of secondary metabolite biosynthesis gene clusters in bacterial and fungal genome sequences. *Nucleic Acids Res* 39:W339–W346. <https://doi.org/10.1093/nar/gkr466>.
- Liu B, Pop M. 2009. ARDB—antibiotic resistance genes database. *Nucleic Acids Res* 37:D443–D447. <https://doi.org/10.1093/nar/gkn656>.