

Salidroside suppresses the growth and invasion of human osteosarcoma cell lines MG63 and U2OS *in vitro* by inhibiting the JAK2/STAT3 signaling pathway

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Abstract. Previous research has reported that salidroside exerts antitumor properties on numerous types of tumor cells; however, its effect on osteosarcoma cells remains unknown. The present study aimed to investigate the effects of salidroside on the viability, apoptosis and invasion of osteosarcoma cells *in vitro*, and determine the underlying mechanism of action. The results of an MTT revealed that salidroside suppressed the viability of osteosarcoma cells (MG63 and U2OS cells) in a time- and concentration-dependent manner. The results of cell morphological analysis (profile observations and Hoechst 33258 staining) and the detection of apoptosis by flow cytometry further indicated that the decrease in osteosarcoma cell viability induced by salidroside was associated with cell apoptosis. Western blot analysis not only confirmed these results but also suggested that salidroside induced the apoptosis of osteosarcoma cells by activating the caspase-9-dependent apoptotic pathway. In addition, we reported that salidroside induced G₀/G₁ phase arrest and suppressed the invasion of osteosarcoma cells, as measured by flow cytometric cell cycle analysis and a Transwell invasion assay, respectively. Western blot analysis confirmed the aforementioned results. Furthermore, our findings demonstrated that salidroside induced the apoptosis, G₀/G₁ phase arrest and suppressed the invasion of osteosarcoma cells by inhibiting the janus kinase 2 (JAK2)/signal transducer and activator of transcription 3 (STAT3) signaling pathway, as determined by western blot analysis. In summary, the findings of the present study suggested that salidroside may inhibit the progression

of osteosarcoma by suppressing the growth and invasion of osteosarcoma cells. Furthermore, the investigations into the underlying mechanism demonstrated that salidroside exerted notable antitumor activity in osteosarcoma cells by inhibiting the JAK2/STAT3 signaling pathway.

Introduction

Osteosarcoma has a double-peak age distribution occurring in children and young adults <20 years of age (75%) and those >50-60 years of age (25%), known as secondary osteosarcoma (1). Osteosarcoma is the most common pediatric malignant bone tumor, accounting for ~5% of all pediatric tumors (2,3). The possible risk factors for secondary osteosarcoma are radiation, Paget's disease and Li-Fraumeni syndrome (4). Early pulmonary metastasis leads to the low survival rate of osteosarcoma (5). At present, surgery combined with chemotherapy is the most common treatment method for osteosarcoma. In 1979, Rosen *et al* proposed the concept of neoadjuvant chemotherapy, and after more than 40 years of development and improvement, this has become the preferred treatment plan for osteosarcoma (6). However, current chemotherapeutic drugs not only have strong cytotoxic effects, but also have toxic side effects and can induce therapeutic resistance (7). Therefore, novel treatment strategies are required.

Rhodiola rosea L., a perennial herbaceous plant, is the most type of common Chinese medicine and is widely used in the medical field (8). Among all of the effective components extracted from *Rhodiola rosea* L., salidroside exhibits powerful properties and has received notable attention. Recent studies have reported that salidroside has anti-fatigue, anti-aging, anti-oxidant, anti-inflammatory, neuroprotective and cardiovascular protective effects (9-12). A literature review revealed that salidroside exhibits antitumor effects in various tumors, including fibrosarcoma (13), bladder carcinoma (14), lung carcinoma (15), breast carcinoma (16) and renal cell carcinoma (17) *in vitro*; however, the association between salidroside and osteosarcoma requires further investigation. In the present study, we investigated the potential antitumor effects of salidroside *in vitro* and the underlying molecular mechanism.

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Materials and methods

Cell culture and treatment. Human osteosarcoma cell lines MG63 and U2OS (ZQXZBIO, Shanghai, China), were selected to assess the antitumor effects of salidroside. Cells were cultured in Dulbecco's modified Eagle's medium combined with high-glucose medium (DMEM-HG) containing 10% fetal bovine serum (FBS) and 1% penicillin/streptomycin (all Gibco; Thermo Fisher Scientific, Inc., Waltham, MA, USA), and were maintained in a 37°C humidified incubator with 5% CO₂. Cells were harvested with a 0.25% trypsin-0.02% EDTA solution and passaged when the cells attained ~80% confluence. Salidroside (Fig. 1A; purity >99%, MedChem Express, Monmouth Junction, NJ, USA) was dissolved in PBS at room temperature and filtered through a 0.22- μ m filter (Merck KGaA, Darmstadt, Germany) prior to use. To determine the potential role of salidroside in osteosarcoma cells, the cells were pretreated with different concentrations (0, 1, 2.5, 5, 7.5 and 10 mM) of salidroside at 37°C for 24, 48 and 72 h, respectively. Cells cultured without pretreatment were used as a control. Z-LEHD-FMK (50 μ M at 37°C for 2 h, Selleck Chemicals, Houston, TX, USA), a caspase-9 specific inhibitor, was used to explore whether salidroside induced osteosarcoma cell apoptosis via the caspase-9-dependent apoptotic pathway. Furthermore, FLLL32 (5 μ M at 37°C for 2 h), a specific inhibitor of JAK2/STAT3 phosphorylation, was used to further confirm whether salidroside induced osteosarcoma cell apoptosis via the JAK2/STAT3 signaling pathway.

Cell viability assay. Cell viability was assessed using an MTT assay (18). Briefly, the two cell lines were seeded in 96-well plates (6x10³ cells/well) for 24 h and when they reached ~80% confluence, they were treated with different concentrations of salidroside (0, 1, 2.5, 5, 7.5 and 10 mM) at 37°C for 24, 48, and 72 h, respectively. After treatment, cells were cultured in DMEM-HG medium containing MTT solution (10 μ l/well, 5 mg/ml in PBS) in a 37°C humidified incubator for 4 h. Then, dimethyl sulfoxide (150 μ l/well) was added to dissolve the formazan crystals. The absorbance value was measured at a wavelength of 490 nm using a SpectraMax Plus 384 microplate reader (Molecular Devices, LLC, Sunnyvale, CA, USA). Cell viability rate = (salidroside treatment group absorbance value - blank control group absorbance value) / (no salidroside treatment group absorbance value - blank control group absorbance value).

Morphology of apoptotic cells. An inverted phase contrast fluorescence microscope (Carl Zeiss, Heidenheimer, Germany) was used to directly observe morphological changes in the two cell lines treated with salidroside. Cells were separately seeded into 6-well (12x10⁴ cells/well) and 24-well (3x10⁴ cells/well) plates, cultured to confluence, and treated with (0, 1, 5 and 10 mM) salidroside at 37°C for 48 h. Firstly, we directly observed the morphology of apoptotic cells seeded in the 6-well plate (magnification, x100). Cells were counted from five random fields for each group, and the average was expressed as the number of apoptotic cells. Then, we observed the nuclear morphology of apoptotic cells seeded in the 24-well plate using Hoechst 33258 staining. In brief, cells were washed three times with PBS, fixed in 4% paraformaldehyde at 4°C for 20 min, stained with Hoechst 33258 staining solution

(125 μ l/well, Beyotime Institute of Biotechnology, Haimen, China) for 5 min, rewashed three times with PBS, covered with anti-fading solution, and then observed using the aforementioned fluorescence microscope (magnification, x400). Cells were counted from five random fields for each group and the number of apoptotic cells (Hoechst-positive cells) was expressed as a percentage (%) of the total number of counted cells.

Flow cytometric analysis of cell apoptosis. To further verify the effect of salidroside on the apoptosis of osteosarcoma cells, an Annexin V-fluorescein isothiocyanate (FITC)/propidium iodide (PI) apoptosis detection kit (Nanjing KeyGen Biotech Co., Ltd., Nanjing, China) was used. Briefly, following treatment with (0, 1, 5 and 10 mM) salidroside for 48 h, MG63 cells (1x10⁵ cells/sample) were digested with trypsin (0.25%, room temperature, 1-3 min), centrifuged (300 x g, 4°C, 5 min), washed twice with PBS, and resuspended in binding buffer (500 μ l/sample, Nanjing KeyGen Biotech Co., Ltd.). Each sample was stained with Annexin V-FITC (5 μ l) and propidium iodide (PI; 5 μ l) in the dark at 4°C for 15 min and then immediately analyzed with a flow cytometer equipped with FACSComp software (BD Biosciences, Franklin Lakes, NJ, USA). Annexin V-FITC/PI⁻ cells were identified as viable cells, Annexin V-FITC⁺/PI⁻ cells were identified as early apoptotic cells, and Annexin V-FITC⁺/PI⁺ cells were identified as the sum of late apoptotic cells and necrotic cells.

Flow cytometric analysis of the cell cycle. A cell cycle detection kit (Nanjing KeyGen Biotech Co., Ltd.) was used to evaluate the cell cycle distribution of cells. In brief, after treatment with (0, 1, 5, 10 mM) salidroside for 24 h, MG63 cells (1x10⁵ cells/sample) were digested with trypsin (without EDTA, 0.25%, room temperature, 1-3 min), centrifuged (300 x g, 4°C, 5 min), washed three times with PBS, and fixed in 70% ethanol at 4°C overnight. Then, each sample was centrifuged (300 x g, 4°C, 5 min), washed three times with PBS, incubated with RNase A (100 μ l) at 37°C for 30 min, stained with PI (400 μ l) in the dark at 4°C for 30 min, and then immediately analyzed with a flow cytometer equipped with CellQuest software (BD Biosciences).

Cell invasion assay. Cell invasion was analyzed using a Transwell invasion assay. Matrigel (20 μ l/chamber, BD Biosciences) was evenly applied to the upper surface of the chamber (8.0 μ m, cat. no. 3422; Costar; Corning, Inc., Corning, NY, USA) and incubated in a 37°C humidified incubator for 30 min. The two cell lines (1x10⁵ cells/chamber) were suspended in preprocessed DMEM-HG medium (200 μ l/chamber, serum-free medium alone, or supplemented with 0, 1, 5 or 10 mM salidroside) and then seeded into the upper surface of the chamber. DMEM-HG medium (500 μ l/well, containing 10% FBS) was added to the 24-well plate. Following incubation in a 37°C humidified incubator for 24 h, the upper-chamber cells were wiped out with a cotton swab, and the lower-chamber cells were washed twice with PBS, fixed in 4% paraformaldehyde at 4°C for 20 min, stained with 0.1% crystal violet (Beyotime Institute of Biotechnology) at room temperature for 5 min, rewashed twice with PBS, then observed under an inverted phase contrast microscope (magnification, x400, Zeiss AG, Oberkochen, Germany). Cells were counted in five random fields for each

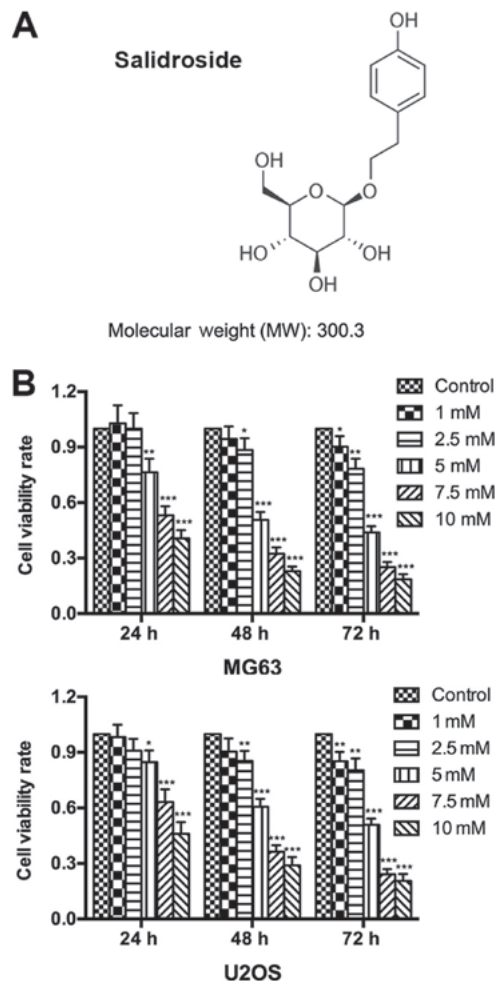


Figure 1. Salidroside inhibits osteosarcoma cell growth. (A) Molecular structure of salidroside. (B) The growth inhibitory effect of salidroside as determined by an MTT assay. * $P < 0.05$, ** $P < 0.01$, *** $P < 0.001$ vs. control group.

group, and the number of invasive cells (crystal violet-positive cells) was expressed as a percentage (%) of the total number of counted cells.

Western blot analysis. After treatment with (0, 1, 5 and 10 mM) salidroside for 48 h, MG63 cells were digested with trypsin and lysed by radioimmunoprecipitation assay lysis buffer (Beyotime Institute of Biotechnology). The lysates were centrifuged at $12,000 \times g$ at 4°C for 30 min, then the supernatants were collected and quantified by a bincinchoninic acid assay (CW Biotechnology, Beijing, China). Total protein ($30 \mu\text{g}$) was separated using 10 or 12% SDS-PAGE with a 5% stacking gel, and then transferred to polyvinylidene difluoride membranes ($0.22 \mu\text{m}$; Merck KGaA). The membranes were subsequently blocked in tris-buffered saline with 10% Tween-20 (TBS-T) containing 5% nonfat milk at room temperature for 2 h, incubated with primary antibodies: rabbit anti-B-cell lymphoma 2 (Bcl-2; cat. no. 4223; 1:1,000), Bcl-2-associated X protein (Bax; cat. no. 5023; 1:1,000), caspase-3 (cat. no. 9665; 1:1,000), caspase-7 (cat. no. 9491; 1:1,000), caspase-9 (cat. no. 9502; 1:1,000), cyclin D1 (cat. no. 2978; 1:1,000), p21 (cat. no. 2947; 1:1,000), matrix metalloproteinase 2 (MMP-2; cat. no. 40994; 1:1,000), MMP-9 (cat. no. 13667; 1:1,000), signal transducer and activator of transcription 3 (STAT3; cat. no. 12640;

1:1,000), phosphorylated (p)-STAT3 (cat. no. 9145; 1:2,000), JAK2 (cat. no. 3230; 1:1,000) and p-Janus kinase 2 (JAK2; cat. no. 3776; 1:1,000), all from Cell Signaling Technology, Inc., Danvers, MA, USA; mouse anti- β -actin (cat. no. CW0264; 1:5,000) and rabbit anti-GAPDH (CW0101, 1:5,000) were obtained from CWBIO (Beijing, China) in TBS-T containing 5% bovine serum albumin at 4°C overnight and washed three times with TBS-T. Then, the membranes were incubated with secondary antibodies (horseradish peroxidase-conjugated goat, anti-mouse (cat. no. CW0102; 1:10,000) or rabbit (cat. no. CW0156; 1:10,000; CWBIO) in TBS-T for 2 h. An enhanced chemiluminescence western blot detection kit (Thermo Fisher Scientific, Inc.), ChemiDoc™ XRS+ System and Image Lab™ 2.1 software (Bio-Rad Laboratories, Inc., Hercules, CA, USA) were used to observe the expression of all proteins.

Statistical analysis. Significant differences between groups of data were analyzed by one-way analysis of variance and a Student-Newman-Keuls test using SPSS 18.0 software (SPSS, Inc., Chicago, IL, USA). Graphs were created using GraphPad Prism 6 software (GraphPad Software, Inc., La Jolla, CA, USA). All data were presented as the mean \pm standard error of the mean and three independent experiments were conducted. $P < 0.05$ was considered to indicate a statistically significant difference.

Results

Salidroside inhibits osteosarcoma cell growth. An MTT assay was performed to determine the growth inhibitory effects of salidroside on osteosarcoma cells. The results showed that salidroside significantly inhibited the viability of osteosarcoma cells in a time- and concentration-dependent manner. Furthermore, a significant inhibitory effect was detected with 5 mM salidroside at 24 h, which peaked at 10 mM salidroside at 72 h compared with the control (Fig. 1A). The half the maximal inhibitory concentration (IC_{50}) values were: MG63, 5.09 mM; U2OS, 9.79 mM; MG63, 4.72 mM; U2OS, 5.294 mM; MG63, 4.67 mM; and U2OS, 4.96 mM following treatment with salidroside for 24, 48, and 72 h, respectively. Based on the aforementioned results, we reported that salidroside inhibited osteosarcoma cell growth in a concentration-dependent manner. Thus, we selected 0, 1, 5 (the approximate IC_{50}) and 10 mM salidroside as effective drug concentrations for subsequent analysis.

Morphological changes in salidroside-treated osteosarcoma cells. In addition, we compared the growth inhibitory effect after treatment with 1, 5, and 10 mM salidroside for 48 h. Morphological changes in osteosarcoma cells were observed via microscopy. Cells in the control group presented a long spindle or polygon appearance and attached uniformly to the culture dish. Whereas, some cells became small and round, and were suspended in the medium in the salidroside-treated groups. The number of non-adherent cells significantly increased (MG63, 1 mM, 58.67 ± 3.51 ; 5 mM, 125.67 ± 6.03 and 10 mM, 171.00 ± 8.19 ; U2OS, 1 mM, 85.67 ± 4.04 ; 5 mM, 166.33 ± 12.22 and 10 mM, 257.00 ± 19.00) compared with the control group (MG63, 12.33 ± 2.52 ; U2OS, 12.67 ± 3.51) (Fig. 2A). Hoechst 33258

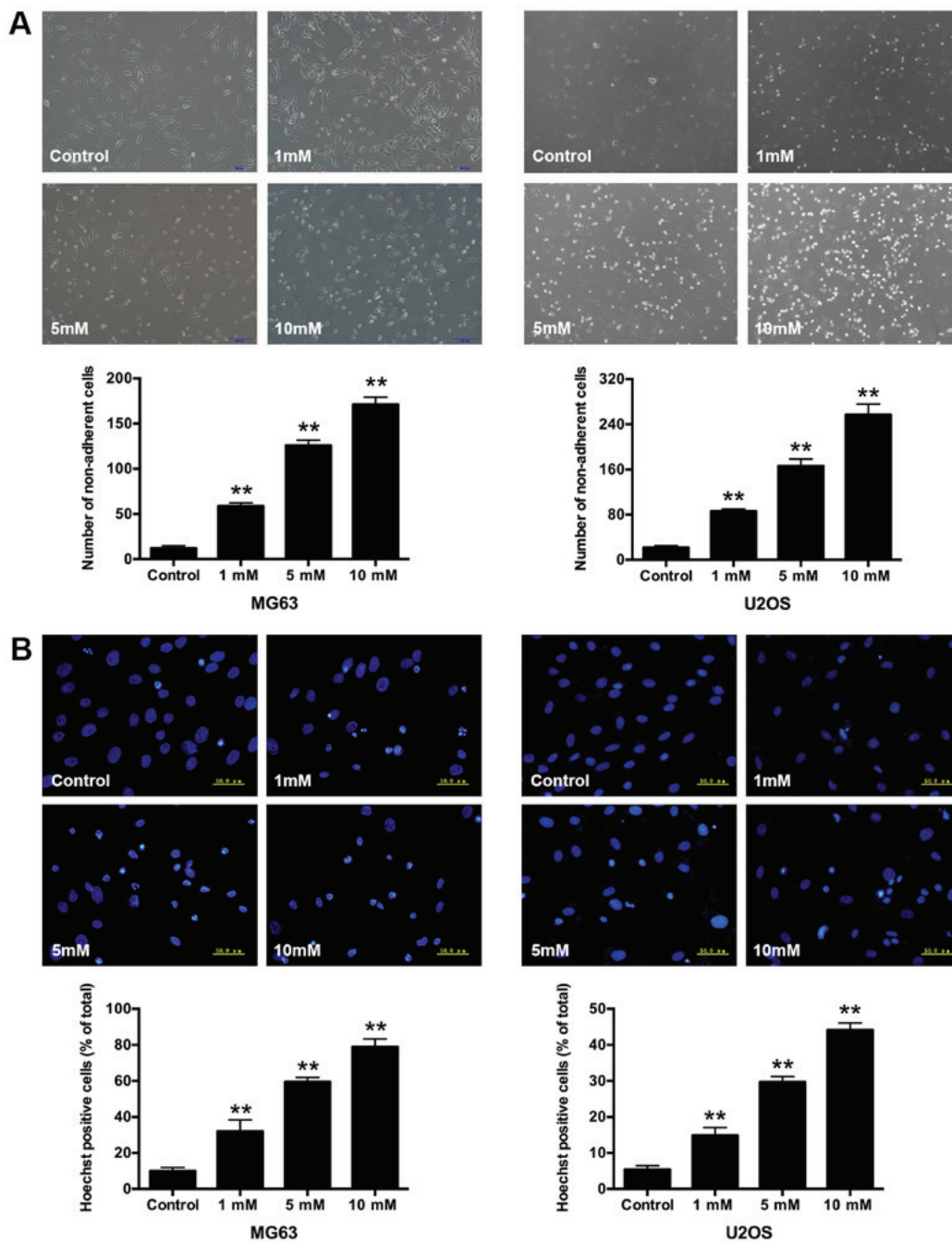


Figure 2. Morphological changes in osteosarcoma cells following treatment with salidroside. (A) Morphological appearance of MG63 and U2OS cells exposed to different concentrations of salidroside for 48 h was observed via inverted phase-contrast microscopy (magnification, x100). (B) Nuclear morphological appearance of MG63 and U2OS cells exposed to different concentrations of salidroside for 48 h was observed via Hoechst 33258 staining and inverted phase-contrast microscopy (magnification, x400). ** $P < 0.01$, vs. control group.

staining was performed to further observe the nuclear morphological changes in osteosarcoma cells. The non-apoptotic nuclei presented weaker blue fluorescence, while the apoptotic nuclei exhibited brighter fluorescence and the structure became condensed, fragmented and crescent-shaped. The percentage of apoptotic nuclei significantly increased (MG63, 1 mM, $32.03 \pm 6.38\%$; 5 mM, $59.53 \pm 2.47\%$ and 10 mM, $78.94 \pm 4.44\%$; U2OS, 1 mM, $14.91 \pm 2.16\%$; 5 mM, $29.72 \pm 1.47\%$ and 10 mM $44.13 \pm 1.91\%$); compared with the control groups (MG63, $10.00 \pm 1.90\%$; U2OS, $5.39 \pm 1.06\%$) (Fig. 2B). The results indicated that salidroside induced the apoptosis of osteosarcoma cells in a concentration-dependent manner.

Salidroside induces osteosarcoma cell apoptosis via the caspase-9-dependent apoptotic pathway. In addition, flow cytometry was performed to further verify the growth inhibitory effects of salidroside in osteosarcoma cells associated with apoptosis. The percentage of apoptotic cells was significantly increased in the presence of salidroside (MG63, 1 mM, $10.20 \pm 1.00\%$; 5 mM, $23.73 \pm 1.38\%$ and 10 mM, $45.77 \pm 3.07\%$; U2OS, 1 mM, $8.83 \pm 0.55\%$; 5 mM, $38.80 \pm 1.69\%$ and 10 mM, $49.10 \pm 0.78\%$), compared with the control groups (MG63, $6.90 \pm 0.40\%$; U2OS, $5.23 \pm 0.58\%$) (Fig. 3A). Then, the Bcl-2 and caspase family of proteins (Bax, Bcl-2, and caspase-3) were investigated to explore the potential

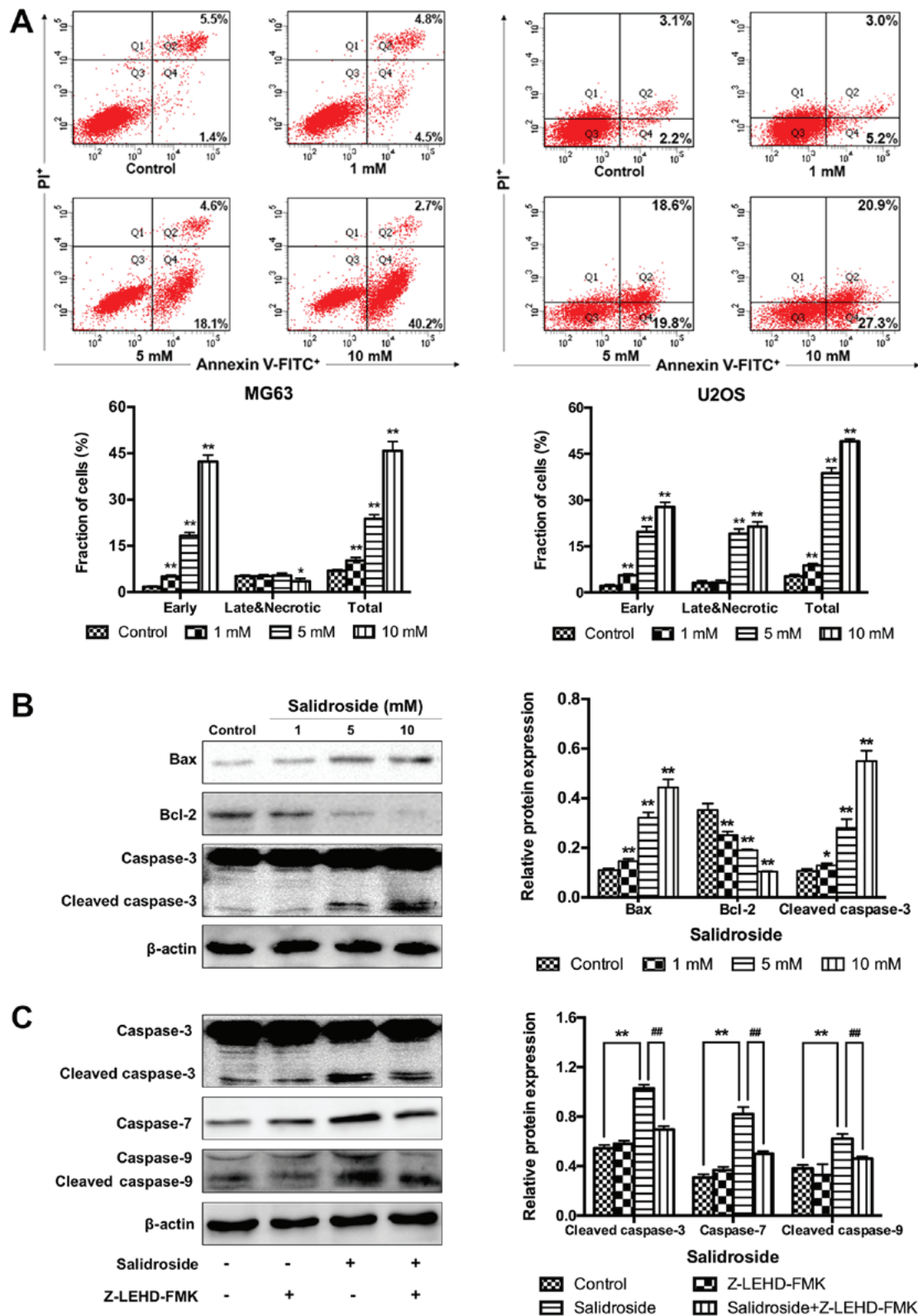


Figure 3. Salidoside induces osteosarcoma cell apoptosis via the caspase-9-dependent pathway. (A) Flow cytometry was used to assess the apoptosis of cells in response to salidoside apoptosis. (B) Apoptosis-related protein expression in MG63 cells in response to various concentrations of salidoside was detected via western blot analysis. (C) Whether the caspase-9-dependent pathway was involved in salidoside-induced apoptosis of osteosarcoma (MG63) cells. *P<0.05, **P<0.01, vs. control group. ##P<0.01, vs. salidoside group. Bcl-2, B-cell lymphoma 2; Bax, Bcl-2-associated X protein; FITC, fluorescein isothiocyanate; PI, propidium iodide.

molecular mechanism in osteosarcoma cells (MG63 cells were selected). Western blot analysis demonstrated that the expression of pro-apoptotic protein was significantly increased

and that of anti-apoptotic proteins was significantly decreased in the salidoside groups in a concentration-dependent manner, compared with the control group (Fig. 3B). Additionally, to

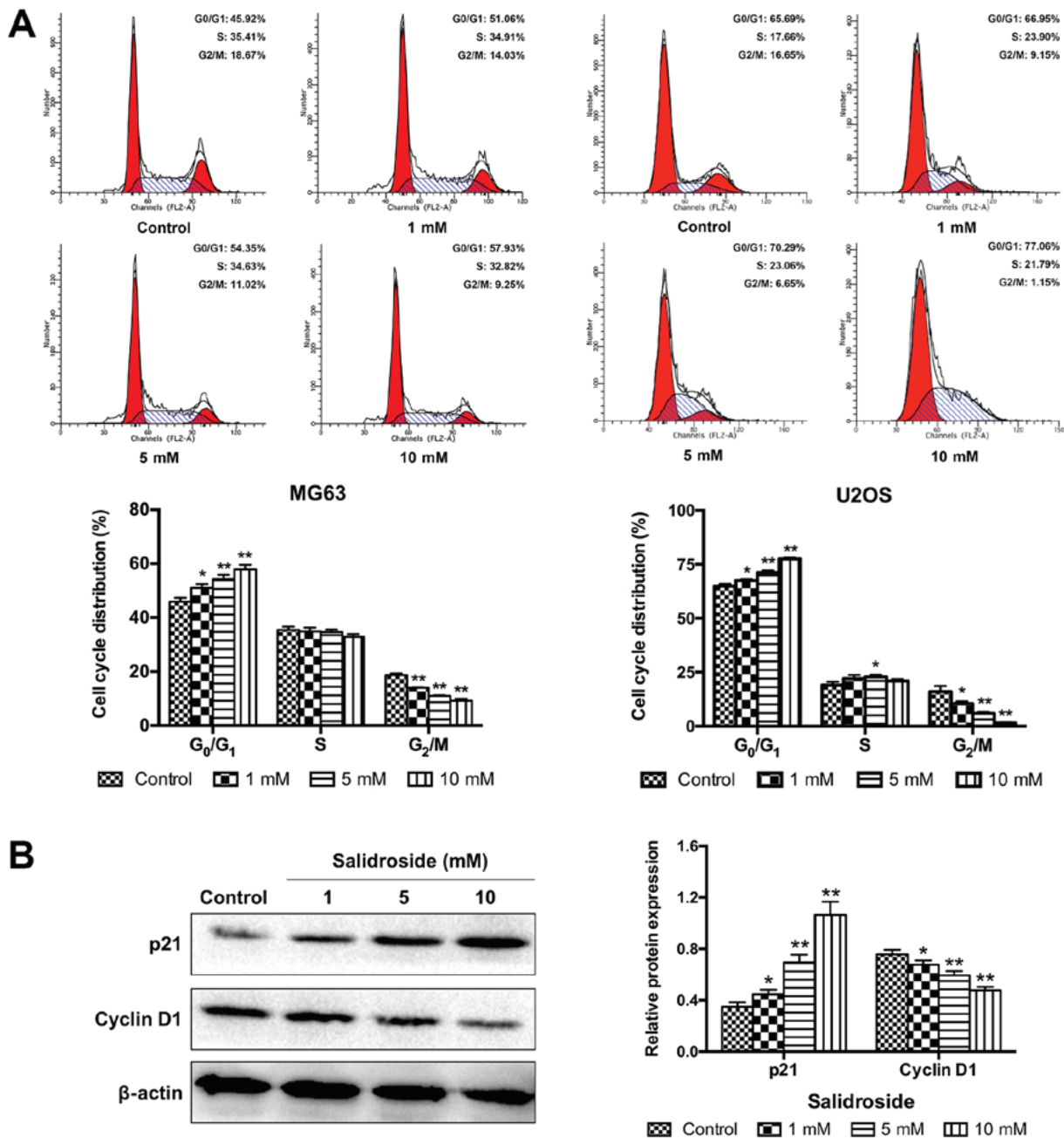


Figure 4. Salidroside induces G₀/G₁ phase cell cycle arrest in osteosarcoma cells. (A) Flow cytometry was used to assess the cell cycle distribution of MG63 and U2OS cells following treatment with different concentrations of salidroside for 24 h. (B) Cell cycle-related protein expression in MG63 cells was detected via western blot analysis. *P<0.05, **P<0.01, vs. control group.

evaluate whether the caspase-9-dependent apoptotic pathway was involved in salidroside-induced osteosarcoma cell apoptosis, Z-LEHD-FMK, a caspase-9 specific inhibitor, was used and the molecular mechanism was explored using western blot analysis. Cells were pretreated with or without Z-LEHD-FMK at 50 μM for 2 h, then 5 mM salidroside was added to the experimental group and cultured for another 48 h. The expression of cleaved caspase-3, caspase-7, and cleaved caspase-9 was significantly increased in the salidroside group, compared with the control group; however, the expression of these proteins was significantly reduced in the presence of Z-LEHD-FMK, compared with the salidroside group (Fig. 3C). These data not only confirmed that salidroside induced the apoptosis of osteosarcoma cells in a concentration-dependent

manner, but also indicated that salidroside induced MG63 cell apoptosis via the caspase-9-dependent apoptotic pathway.

Salidroside induces G₀/G₁ arrest in osteosarcoma cells.

To determine whether cell-cycle arrest was involved in the growth inhibitory effects of salidroside, we assessed the role of salidroside in the progression of the cell cycle in osteosarcoma cells using flow cytometry. As presented in Fig. 4A, salidroside significantly increased the percentage of cells in G₀/G₁ phase (MG63, 1 mM, 51.06±1.38%; 5 mM, 54.35±1.50% and 10 mM, 57.93±1.10%; U2OS, 1 mM 67.51±0.67%; 5 mM 71.22±0.88% and 10 mM 77.57±0.66%), but significantly decreased the percentage of cells in G₂/M phase (MG63, 1 mM, 14.02±0.24%; 5 mM, 11.01±0.21% and 10 mM, 9.25±0.56%;

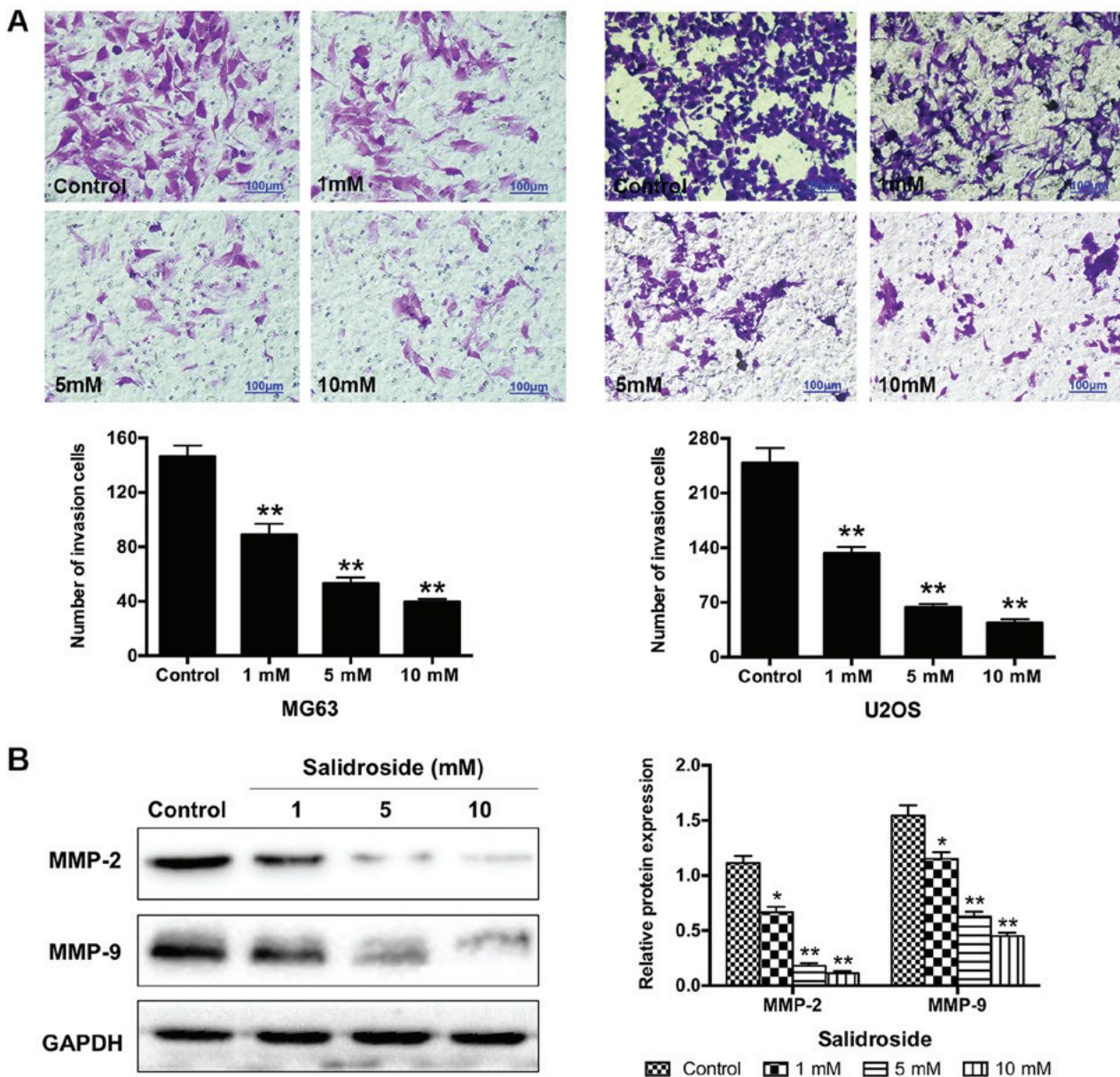


Figure 5. Salidroside suppresses osteosarcoma cell invasion. (A) Transwell assay was used to assess the invasion of MG63 and U2OS cells following treatment with different concentrations of salidroside for 48 h. Scale bar, 100 μ m. (B) Invasion-related protein expression in MG63 cells was detected via western blot analysis. * $P < 0.05$, ** $P < 0.01$, vs. control group. MMP, matrix metalloproteinase.

U2OS, 1 mM $10.42 \pm 1.11\%$; 5 mM, $5.93 \pm 0.63\%$ and 10 mM, $1.47 \pm 0.29\%$), compared with the control group (MG63, G_0/G_1 $45.92 \pm 1.36\%$ and G_2/M $18.68 \pm 0.65\%$; U2OS, G_0/G_1 $64.98 \pm 0.96\%$ and G_2/M $15.99 \pm 2.63\%$). Then, the expression of cell cycle-associated proteins were analyzed (cyclin D1 and p21) to elucidate the potential molecular mechanism. The expression of p21 was significantly increased and the expression of cyclin D1 was significantly decreased in the salidroside groups in a concentration-dependent manner, compared with the control group (Fig. 4B). These results demonstrated that salidroside induced G_0/G_1 phase arrest of osteosarcoma cells in a concentration-dependent manner.

Salidroside inhibits the invasion of osteosarcoma cells. We performed a Transwell assay to understand the effects of salidroside on the invasion of osteosarcoma cells. The

results showed that the number of invasive cells significantly decreased (MG63, 1 mM, 89.00 ± 7.94 ; 5 mM, 53.33 ± 4.16 and 10 mM, 39.67 ± 2.08 ; U2OS, 1 mM, 133.00 ± 8.00 ; 5 mM, 63.67 ± 4.16 and 10 mM, 44.00 ± 4.58) compared with the control groups (MG63, 146.33 ± 8.08 ; U2OS, 248.67 ± 19.04) (Fig. 5A). Additionally, we further investigated the potential molecular mechanism in MG63 cells using western blot analysis. Salidroside significantly decreased the expression of invasion-associated proteins (MMP-2 and MMP-9) in a concentration-dependent manner, compared with the control groups (Fig. 5B). In summary, these findings suggested that salidroside inhibited the invasion of osteosarcoma cells in a concentration-dependent manner.

Salidroside inhibits the JAK2/STAT3 signaling pathway in osteosarcoma cells. To further investigate the potential

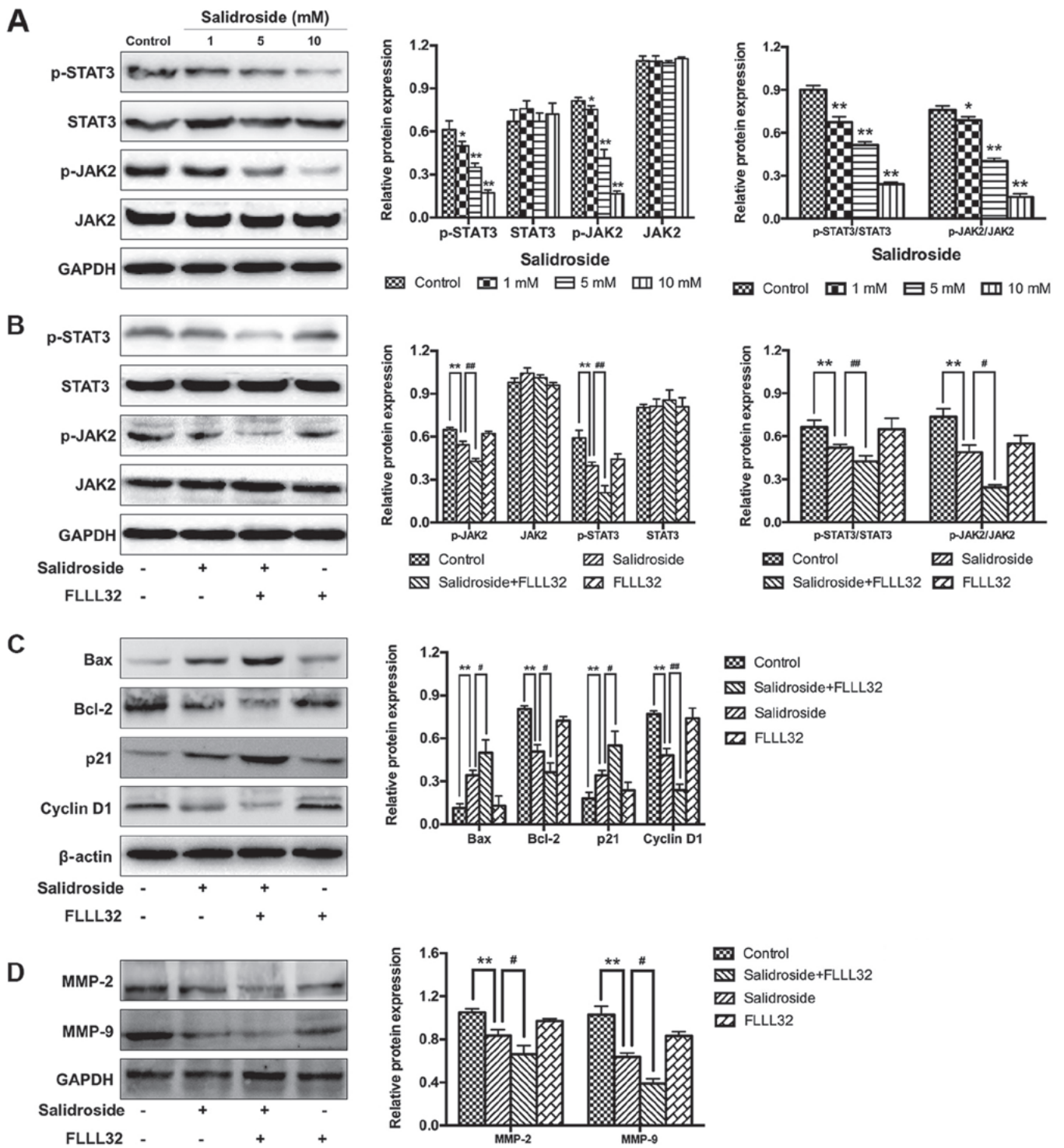


Figure 6. Salidroside inhibits the JAK2/STAT3 signaling pathway. (A) Western blot analysis was used to investigate whether the JAK2/STAT3 signaling pathway was associated with the growth inhibitory effect induced by salidroside. (B) FLLL32 (a specific JAK2/STAT3 inhibitor) was used to confirm the involvement of the JAK2/STAT3 signaling pathway. (C) Western blot analysis was used to verify the role of the JAK2/STAT3 pathway in the regulation of apoptosis- and cell cycle-associated proteins in salidroside-treated osteosarcoma cells. (D) Western blot analysis was used to verify the role of the JAK2/STAT3 pathway in the regulation of invasion-associated proteins in salidroside-treated osteosarcoma cells. *P<0.05, **P<0.01, vs. control group. #P<0.05, ##P<0.01, vs. salidroside group. Bcl-2, B-cell lymphoma 2; Bax, Bcl-2-associated X protein; JAK2, Janus kinase 2; MMP, matrix metalloproteinase; p, phosphorylated; STAT3, signal transducer and activator of transcription 3.

mechanism by which salidroside suppressed the growth of osteosarcoma (MG63) cells, the JAK2/STAT3 signaling pathway was analyzed. The results of western blot analysis demonstrated that salidroside significantly decreased the expression of p-JAK2 and p-STAT3 in a concentration-dependent manner, compared

with the control groups (Fig. 6A). Furthermore, FLLL32 (5 μ M), a specific inhibitor of JAK2/STAT3 phosphorylation, was used to further confirm these findings. As presented in Fig. 6B, FLLL32 significantly decreased the expression of p-JAK2 and p-STAT3, compared with the salidroside group. Similarly, apoptosis-, cell

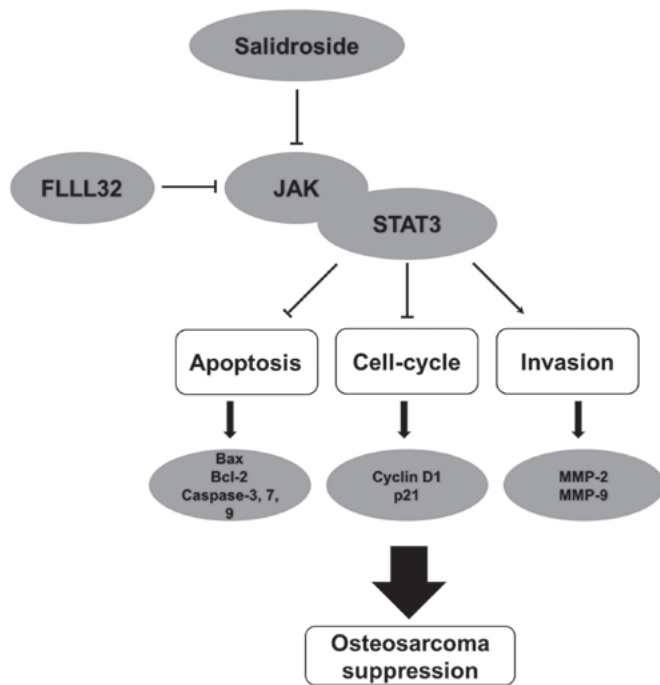


Figure 7. Diagram of the mechanism of salidroside-induced apoptosis, cell cycle arrest and suppressed invasion of osteosarcoma cells via inhibition of the JAK2/STAT3 signaling pathway. Bcl-2, B-cell lymphoma 2; Bax, Bcl-2-associated X protein; JAK, Janus kinase; MMP, matrix metalloproteinase; STAT3, signal transducer and activator of transcription 3.

cycle- and invasion-related proteins were accordingly altered in the salidroside + FLLL32 group, compared with the salidroside group (Fig. 6C and D). Collectively, the aforementioned results indicated that the JAK2/STAT3 signaling pathway was involved in salidroside-induced apoptosis and cell cycle arrest in MG63 cells. Collectively, the aforementioned results indicated that salidroside induced apoptosis and cell cycle arrest, and suppressed the invasion of osteosarcoma cells by inhibiting the JAK2/STAT3 signaling pathway.

Discussion

Osteosarcoma is a common aggressive malignant bone tumor. Despite considerable developments in the treatment of osteosarcoma, current treatments for osteosarcoma still have major limitations; the long-term survival (5-year survival rate is ~60%) and mortality rates remained unchanged (19). Therefore, novel therapeutic strategies are urgently required that can effectively act via various anticancer mechanisms. The positive effects of phytochemical drugs on antitumor activity have been demonstrated, as well as their protective effects against the side effects of traditional chemotherapeutic drugs, indicating that they are safer compounds for use in normal cells (20,21). However, the complex mechanism of action of phytochemical drugs complicates their application in treating human malignancies (22). Salidroside, a glucoside of tyrosol, was considered an effective ingredient of *Rhodiola*, which has been demonstrated to have anti-proliferative and pro-apoptotic effects in previous reports (8-12). Interestingly, research has reported that salidroside also has antitumor effects in several human tumor cells, such as neuroblastoma cells, bladder

cancer cells, glioma cells and lung cancer cells (14,15,20,23). Furthermore, Li *et al* (24) reported that salidroside combined with antitumor agents exerted excellent antitumor effects in colorectal cancer. Qi *et al* (25) revealed that salidroside had a direct inhibitory effect on the proliferation, migration and invasion of gastric cancer cells. In the present study, we first assessed the antitumor effects of salidroside in the treatment of osteosarcoma. We demonstrated that salidroside induced the growth and invasion of osteosarcoma cells, which indicated its therapeutic potential. The pharmacological mechanism of salidroside may be related to the JAK2/STAT3 signaling pathway (Fig. 7).

Cell proliferation is an important marker for tumor development. Therefore, inhibiting tumor growth (by promoting tumor cell apoptosis) is the most important objective in preventing tumor progression (26). The MTT assay is widely used in bioactive factor activity assays, large-scale antitumor drug screening and cytotoxicity assays (27). In the present study, the results of the MTT assay revealed that salidroside significantly inhibited the viability of osteosarcoma cells in a time- and concentration-dependent manner. The results of cell morphological observations and flow cytometric apoptosis detection further indicated that the decrease in cell viability induced by salidroside was associated with cell apoptosis. We investigated whether the expression of apoptotic-related proteins via western blot analysis, and the expression of the Bcl-2 and caspase families, critical apoptosis-related proteins, were regulated by salidroside. The Bcl-2 and caspase families are specific regulatory proteins of the mitochondrial apoptosis pathway, which is one of the main pathways of apoptosis (28). Our results indicated that the mitochondrial apoptosis pathway is involved in salidroside-mediated apoptosis of osteosarcoma cells. In addition, dysregulated cell cycle distribution is another feature of tumor development, and the induction of cell apoptosis is accompanied with cell cycle arrest (29). Flow cytometric cell cycle analysis is widely used for evaluating changes in cell cycle distribution (30). We reported that salidroside triggered G₀/G₁ phase arrest in osteosarcoma cells, which was consistent with previous reports (16,31). Then, the present study investigated the expression of cell cycle-related proteins using western blot analysis; the expression of cyclin D1 and p21 were revealed to be regulated by salidroside. Therefore, we concluded that salidroside induced the apoptosis of osteosarcoma cells by inducing G₀/G₁ phase arrest. We suggested that salidroside may function as an agonist to induce the apoptosis and G₀/G₁ phase arrest of osteosarcoma cells, and may represent as an alternative therapeutic strategy for the treatment of osteosarcoma.

Invasion, which generally leads to metastasis, can be used to predict tumor malignancy (32). Previous research reported that early metastasis (particularly pulmonary metastasis) remains as the main cause of mortality in ~40% of patients with osteosarcoma (33). The results of Transwell assays demonstrated that salidroside significantly inhibited the invasive ability of osteosarcoma cells in a concentration-dependent manner. We further investigated the molecular mechanism using western blot analyses and reported that salidroside significantly decreased the expression of MMP-2 and MMP-9 in a concentration-dependent manner. MMP-2 and MMP-9, the two most important proteins in the

MMP family, are involved in extracellular matrix degradation by effectively decomposing collagen IV and laminin (34). The MMP family of proteins facilitates tumor metastasis by degrading the basement membrane of the extracellular matrix, and is an effective marker for predicting tumor malignancy (35). Thus, our findings indicated that salidroside reduced the metastatic capabilities of osteosarcoma cells by suppressing the expression of MMPs.

Increasing evidence has suggested that the inhibitory effect of salidroside on different tumor cells is associated with different signal pathways. Sun *et al* (13) showed that salidroside inhibited the metastasis of human fibrosarcoma cells by downregulating the reactive oxygen species (ROS)/protein kinase C/extracellular signal-regulated kinase 1/2 pathway. Zhao *et al* (16) revealed that salidroside reduced oxidative stress and suppressed breast cancer growth by inhibiting the formation of ROS and activating the mitogen-activated protein kinase pathway. Fan *et al* (36) found that salidroside induced the apoptosis and autophagy of human colorectal cancer cells via inhibiting the PI3K/Akt/mammalian target of rapamycin pathway. Lv *et al* (17) reported that salidroside suppressed proliferation in renal cell carcinoma by modulating the JAK2/STAT3 pathway. Kang *et al* (37) demonstrated that salidroside inhibited the mobility and angiogenesis of breast cancer cells by regulating epidermal growth factor receptor/Jak2/STAT3 signaling via MMP2. To investigate the potential signaling pathway involved in salidroside-mediated cell apoptosis of osteosarcoma cells, western blot analysis was performed. We showed that the JAK2/STAT3 signaling pathway participated in salidroside-mediated cell apoptosis of osteosarcoma cells, which was consistent with previous reports of signaling in other tumors, including human renal carcinoma cells, human melanoma cells and colon cancer cells (38,39). JAK family proteins have been implicated in different cellular processes, such as cell proliferation, differentiation, apoptosis, invasion and angiogenesis (40). JAKs are activated by cytokines to phosphorylate Y residues and subsequently phosphorylate the downstream molecule STAT3 (40). STAT3 translocates to the nucleus and binds to DNA, regulating the expression of apoptosis-related genes, such as Bcl-2 and other members of this family (41). STAT3 is therefore considered an essential anti-apoptotic factor. The JAK2/STAT3 pathway is an evolutionarily conserved pathway that induces tissue homeostasis modulation and decreases the extent of damage during cellular stress; it is therefore considered a critical signaling pathway in cancer formation and progression (27,42). Our results revealed that salidroside reduced the phosphorylation of JAK2 and STAT3. In addition, we reported that FLLL32 (a specific JAK2/STAT3 inhibitor) significantly enhanced the inhibitory effects of salidroside on JAK2 and STAT3 phosphorylation in osteosarcoma cells. Taken together, these results demonstrated that salidroside induces cell apoptosis, cell cycle arrest and suppresses invasion of osteosarcoma cells via inhibiting the JAK2/STAT3 signaling pathway.

ROS can activate multiple pathways involved in metastasis, invasion, and apoptosis of tumor cells. It has been reported that ROS and several oxidative substances produced by tumor cells can promote tumor growth (43). Salidroside, an antioxidant, had been reported to exert anti-cancer effects in breast cancer treatment (16); however, whether autophagy and ROS are

involved in salidroside-induced apoptosis and cell cycle arrest in osteosarcoma cells is yet to be determined and further studies are required to investigate the mechanism of action of salidroside. In addition, a limitation of our study is that only an MTT assay was used to determine the growth inhibition by salidroside; further methods are required to verify our findings in the future. By consulting the literature, we found that low concentrations of salidroside could suppress the growth and invasion of several human tumor cells (15,17,20,31). Our preliminary research was based on the concentration at the micromolar level; however, the results showed that the activity of osteosarcoma cells was not affected. This could be due to the relatively mild efficacy of salidroside; thus, investigations were conducted with salidroside at the millimolar level in the present study. Additionally, several studies also used high concentrations (mM) of salidroside (36,44); we used similar concentrations of salidroside for analysis. Furthermore, the normal osteoblast cell line hFOB1.19 was employed to explore whether salidroside induced osteoblast apoptosis with the same concentrations. We reported that the concentration of salidroside significantly induced the apoptosis of normal osteoblasts (hFOB1.19) from ≥ 8.2 mM at 24 h (data not shown). Therefore, we considered that the sensitivity of cells to salidroside may differ between cell lines. Our future study aims to further narrow the concentration gradient of salidroside.

In the present study, it was demonstrated that salidroside was a critical inhibitor of osteosarcoma growth and invasion, and was associated with the induction of cell apoptosis, cell cycle arrest and suppressed invasion. Investigations into the molecular mechanism of action of salidroside suggested that apoptosis may be induced via the mitochondrial apoptosis pathway, and the JAK2/STAT3 pathway was reported to be involved in salidroside-mediated inhibition of osteosarcoma growth and invasion. In conclusion, the findings of the present study may provide insight into the molecular mechanism underlying the effects of salidroside and may be considered as a potential therapeutic agent for the treatment of osteosarcoma.

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Availability of data and materials

All data generated or analyzed during this study are included in this published article.

Authors' contributions

CL made substantial contributions to the design of the study for the analysis of the role of salidroside in modulating osteosarcoma cell growth and invasion. LH, ZH and LW performed experiments including the MTT assay, and cell apoptosis and cell cycle analysis. WL, XC and XZ performed the experiments to analyze molecular mechanisms, such as western blot analysis. SY conducted statistical analysis of the experimental data. CL

and LH were major contributors in writing the manuscript. All authors read and approved the final manuscript.

Ethics approval and consent to participate

Not applicable.

Patient consent for publication

Not applicable.

Competing interests

The authors declare that they have no competing interests.

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