#### **Review Article**

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# Heterogeneity of Human $\gamma\delta$ T Cells and Their Role in Cancer Immunity

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#### **Conflict of Interest**

The authors declare no potential conflicts of interest.

#### Abbreviations

BTN3A, butyrophilin 3A; DETC, dendritic epidermal T cell; KIR, killer inhibitory receptor; NCR, natural cytotoxicity receptor; NKG2D, NK group 2 member D; Tfh, follicular Th; TME, tumor microenvironment; ULBP, UL16-binding protein

#### ABSTRACT

The  $\gamma\delta$  T cells are unconventional lymphocytes that function in both innate and adaptive immune responses against various intracellular and infectious stresses. The  $\gamma\delta$  T cells can be exploited as cancer-killing effector cells since  $\gamma\delta$  TCRs recognize MHC-like molecules and growth factor receptors that are upregulated in cancer cells, and  $\gamma\delta$  T cells can differentiate into cytotoxic effector cells. However,  $\gamma\delta$  T cells may also promote tumor progression by secreting IL-17 or other cytokines. Therefore, it is essential to understand how the differentiation and homeostasis of  $\gamma\delta$  T cells are regulated and whether distinct  $\gamma\delta$  T cell subsets have different functions. Human  $\gamma\delta$  T cells are classified into V $\delta$ 2 and non-V $\delta$ 2  $\gamma\delta$ T cells. The majority of V $\delta$ 2  $\gamma\delta$  T cells are V $\gamma$ 9 $\delta$ 2 T cells that recognize pyrophosphorylated isoprenoids generated by the dysregulated mevalonate pathway. In contrast, V $\delta$ 1 T cells expand from initially diverse TCR repertoire in patients with infectious diseases and cancers. The ligands of Vô1 T cells are diverse and include the growth factor receptors such as endothelial protein C receptor. Both V $\delta$ 1 and V $\delta$ 2  $\gamma\delta$  T cells are implicated to have immunotherapeutic potentials for cancers, but the detailed elucidation of the distinct characteristics of 2 populations will be required to enhance the immunotherapeutic potential of  $\gamma\delta$  T cells. Here, we summarize recent progress regarding cancer immunology of human  $\gamma\delta$ T cells, including their development, heterogeneity, and plasticity, the putative mechanisms underlying ligand recognition and activation, and their dual effects on tumor progression in the tumor microenvironment.

**Keywords:** T-lymphocyte subsets;  $\gamma \delta$  T cell; T Cell Receptors, gamma delta; Tumor microenvironment

#### INTRODUCTION

Among 3 main lineages of lymphocytes— $\alpha\beta$  T cells,  $\gamma\delta$  T cells, and B cells,  $\gamma\delta$  T cells are the most enigmatic lymphocytes that express TCRs rearranged from TCR  $\gamma$  and  $\delta$  genes (1-3). The  $\gamma\delta$  T cells are one of the innate immune cells that have a pivotal role in cancer immunosurveillance as the deficiency of  $\gamma\delta$  T cells increased the susceptibility to cancers (4-7). They can mediate potent direct cytotoxicity by recognizing transformed target cells via the  $\gamma\delta$  TCRs, but they may also detect cancer cells via activating NK cell receptors such as NK

#### **Author Contributions**

Conceptualization: Lee HW, Chung YS, Kim TJ; Funding acquisition: Kim TJ, Chung YS; Project administration: Kim TJ; Validation: Kim TJ; Writing - original draft: Lee HW, Kim TJ; Writing - review & editing: Lee HW, Chung YS, Kim TJ. group 2 member D (NKG2D) or natural cytotoxicity receptors (NCRs) (8,9). The application of  $\gamma\delta$  T cells, mostly V $\gamma$ 9V $\delta$ 2 T cells, for cancer immunotherapy has been explored against various tumors of hematological and epithelial origin (10-16). Many clinical trials have shown that those treatments are feasible and safe, but with some obvious limitations (4,12,17). Therefore, a better understanding of  $\gamma\delta$  T cell subset-specific responses during tumor immunity is vital to rationally develop optimal strategies for maximizing the anti-tumor activity of  $\gamma\delta$  T cells and inhibiting their pro-tumor activity. Here, we summarize the recent progress regarding the immunobiology of human  $\gamma\delta$  T cells, including the heterogeneity and plasticity, the putative mechanisms of ligand recognition and activation, their positive and negative effects on the cancer progression, and the future perspective of immunotherapy using  $\gamma\delta$  T cells.

#### ORIGIN AND DEVELOPMENT OF THE HUMAN $\gamma\delta$ T CELLS

The  $\gamma\delta$  T cells develop earlier than  $\alpha\beta$  T cells in the thymus and are exported to different peripheral tissues according to the chronological order of the thymic development (18,19). In the mouse, the first  $\gamma\delta$  T cells at the embryonic day of 14,  $V\gamma5^*V\delta1^*$  T cells are selected by selection and upkeep of intraepithelial T cells protein 1 presented on thymic epithelial cells and become dendritic epidermal T cells (DETCs) responsible for body-barrier surveillance (20,21). A later process of thymic T cell development generates  $V\gamma6^*V\delta1^*$  T cells that are destined for the female genital tract, peritoneal cavity and tongue, and other  $\gamma\delta$  T cells with diverse VDJ clonotypes containing  $V\gamma1$ , 2, 4, and 7 segments (18,22). Whereas the early developing  $\gamma\delta$  T cells have invariant TCRs, the  $\gamma\delta$  T cells appearing during the later period of the development are diverse in TCR repertoire (22-24).

Human  $\gamma\delta$  T cells are also present in the thymus as well as the periphery, suggesting the thymic development of human  $\gamma\delta$  T cells (25). Although adult blood  $\gamma\delta$  T cells are predominated by V $\gamma$ 9V $\delta$ 2 cells, neonatal cord blood  $\gamma\delta$  T cells express a diversity of V $\gamma$ and V $\delta$  chains paired in various combinations, and the majority of neonatal  $\gamma\delta$  T cells are V $\gamma$ 9·V $\delta$ 1<sup>+</sup> cells (26,27). Therefore, the adult blood V $\gamma$ 9V $\delta$ 2 cells appear to represent the post-natal expansion of V $\gamma$ 9V $\delta$ 2 cells expressing canonical CDR3s in response to microbial phosphoantigens that are described below (28-30). Human V $\gamma$ 9V $\delta$ 2 cells have been shown to expand rapidly after birth within 1 year of life (31). In the adult, V $\delta$ 1 and V $\delta$ 2  $\gamma\delta$  T cells are localized in the barrier tissues and the peripheral blood, respectively (32).

#### HETEROGENEITY OF THE HUMAN $\gamma\delta$ T CELLS

Although  $\gamma\delta$  T cells are cousins of  $\alpha\beta$  T cells,  $\gamma\delta$  T cells directly recognize Ags via their  $\gamma\delta$  TCRs without the need of MHC molecules similarly to B cells (1,33,34). The  $\gamma\delta$  T cells are sometimes referred to as innate lymphocytes since they can recognize microbial or stress-induced patterns and respond rapidly without previous exposure to the Ags (1,24). However, some  $\gamma\delta$  T cells exhibit highly adaptive features such as clonal expansion and differentiation from naïve cells to effector cells (35). The overall characteristics of  $\gamma\delta$  T cells may be positioned between NK cells and CD8<sup>+</sup> T cells (36). The  $\gamma\delta$  T cells are heterogeneous concerning functional features depending on the usage of TCR  $\gamma$  and  $\delta$  chains and the tissue localization. The  $\gamma\delta$  T cells account for 0.5%–5% of all peripheral blood T cells (2,7).

In humans, several functional V $\gamma$  gene segments (including V $\gamma$ 2, V $\gamma$ 3, V $\gamma$ 4, V $\gamma$ 5, V $\gamma$ 8, V $\gamma$ 9, and V $\gamma$ 11) rearrange into 5 J $\gamma$  segments and 2 C $\gamma$  segments on chromosome 7 to generate TCR $\gamma$  chains, whereas TCR $\delta$  chains are generated by the rearrangement of at least 7 V $\delta$ , 3 D $\delta$ , 3 J $\delta$ , and 1 C $\delta$  segments on chromosome 14 (2,3,7,37). Whereas V $\delta$ 1, V $\delta$ 2, and V $\delta$ 3 segments are used only in the rearrangement of the TCR  $\delta$  chains, V $\delta$ 4–V $\delta$ 7 segments are also used in the rearrangement of the TCR  $\alpha$  chains and have alternative gene names belonging to TCR V $\alpha$  gene segments (37). The functional features of  $\gamma\delta$  T cells are closely correlated with the usage of the TCR $\delta$  chains (24). Among 7 V $\delta$  segments, V $\delta$ 1 and V $\delta$ 2 segment-using  $\gamma\delta$  TCRs are the most common human  $\gamma\delta$  TCRs (2).

As the most abundant human  $\gamma\delta$  T cells are V $\gamma$ 9V $\delta$ 2 T cells that recognize unique phosphoantigens and V $\delta$ 1 T cells have adaptive features distinct from V $\gamma$ 9V $\delta$ 2 T cells,  $\gamma\delta$ T cells are commonly classified into V $\delta$ 2 and non-V $\delta$ 2  $\gamma\delta$  T cells (2,35,38). The V $\gamma$ 9V $\delta$ 2 T cells are the most well-known human  $\gamma\delta$  T cells and have been exploited for anti-cancer immunotherapy (10,11). The characteristics and the adaptive features of non-V $\delta$ 2  $\gamma\delta$  T cells, especially V $\delta$ 1  $\gamma\delta$  T cells, are recently recognized, and these V $\delta$ 1  $\gamma\delta$  T cells are also thought to be a candidate for anti-cancer immunotherapy (35).

# THE $\gamma\delta$ TCR STRUCTURE AND ACTIVATION OF THE HUMAN $\gamma\delta$ T CELLS

Although  $\gamma\delta$  T cells share TCR rearrangement mechanism and memory functions with  $\alpha\beta$  T cells, they differ in the immune response kinetics and mechanisms of target cell recognition (39). The  $\gamma\delta$  T cells do not recognize MHC molecules, but many  $\gamma\delta$  T cells respond to nonpeptide Ags or MHC-like molecules, such as MHC class I-related chain A (MICA), MICB, or UL16-binding protein (ULBP), that are upregulated in cells under stressed conditions such as infection or cancer transformation in MHC-unrestricted manner (2,3). Similarly to  $\alpha\beta$  TCRs,  $\gamma\delta$  TCRs are also associated with CD3 molecules, but differently from murine  $\alpha\beta$ TCRs, murine  $\gamma\delta$  TCRs contain only CD3 $\gamma\epsilon$  dimers, not CD3 $\delta\epsilon$  dimers (40). Notably, murine  $\gamma\delta$  TCR cells can develop in the absence of CD3 $\epsilon$  or CD3 $\delta$  (41,42), but the expression of CD3 $\gamma$ is indispensable for the murine  $\gamma\delta$  T cell development (43). Furthermore, CD3 $\zeta$  chain is not necessary for the  $\gamma\delta$  T cell development and FccRIy chain, a CD3 $\zeta$  chain family member that can dimerize with CD3 $\zeta$ , is expressed upon activation and then included in the  $\gamma\delta$  TCR complexes (44). On the other hand, human  $\gamma\delta$  TCR complex contains CD3 $\delta$  chain and shows a TCR $\gamma\delta$ CD3 $\epsilon_{2}\delta\gamma\zeta_{2}$  stoichiometry similarly to human  $\alpha\beta$  TCR complex, whereas mouse  $\gamma\delta$ TCR complex has a TCR $\gamma\delta$ CD3 $\epsilon_2\gamma_2\zeta_2$  stoichiometry (45). Human  $\gamma\delta$  TCR signaling is less dependent on CD3 $\gamma$  chain than CD3 $\delta$  chain as human patient lacking CD3 $\gamma$  have abundant peripheral blood  $\gamma\delta$  T cells expressing high levels of  $\gamma\delta$  TCR (46). Interestingly, forced expression of human, but not murine, CD3δ transgene rescue the γδ T cell development in mice deficient in both CD3 $\delta$  and CD3 $\gamma$  genes, suggesting the unique role of human CD3 $\delta$  in the TCR signaling (45).

The  $\gamma\delta$  TCR signaling is qualitatively different from the  $\alpha\beta$  TCR signaling (44). The  $\gamma\delta$  TCRs self-oligomerize and cause constitutive signaling in the absence of ligands (47). These  $\gamma\delta$  TCR signaling characteristics are similar to those of pre- $\alpha\beta$  TCR signaling responsible for the  $\beta$  selection during thymic T cell development (48). During the thymic  $\gamma\delta$  T cell development,  $\gamma\delta$  T cells that encounter strong agonistic ligands obtain the capability of secreting IFN- $\gamma$ . In contrast,  $\gamma\delta$  T cells that do not encounter strong agonists adopt IL-17-default position (1,47).

In the periphery, the stimulation of  $\gamma\delta$  T cells via  $\gamma\delta$  TCR and costimulatory receptors or NK cell receptors triggers  $\gamma\delta$  T cells to undergo clonal expansion and differentiation into effector cells and to produce large quantities of pro-inflammatory cytokines such as IFN- $\gamma$  or IL-17. Upon activation,  $\gamma\delta$  T cells can also exert a potent cytotoxic activity without the obligatory delay associated with clonal expansion and differentiation (49).

Although  $\gamma\delta$  TCR is regarded as an activating receptor,  $\gamma\delta$  TCR may act as an inhibitory receptor in certain contexts. The consequence of the constitutive  $\gamma\delta$  TCR signaling can be inhibition of  $\gamma\delta$  T cell activation when the ligands on target cells are constitutively presented (50). In NK cells, the constitutive inhibitory signaling through killer inhibitory receptor (KIR) sets up a threshold that NK cells are not easily activated, and a full activation of NK cell requires very high concentrations of activating ligands for NK cell-activating receptors and/ or downregulation of inhibitory ligand, MHC class I on the target cells (51). The V $\gamma5\delta1$  TCRs in mouse DETCs form constitutive immunological synapses with keratinocytes in the steady state and are argued to have a role similar to KIR on NK cells (20).

Since  $\gamma\delta$  T cells have a lot of NK cell-activating receptors such as NCRs and NKG2D (8,9), the functional roles of  $\gamma\delta$  TCR should be carefully investigated in heterogeneous subpopulations of  $\gamma\delta$  T cells since the NK receptors, not  $\gamma\delta$  TCR, could be main receptors for  $\gamma\delta$  T cell activation. The NK cell-activating receptors can be considered as costimulatory receptors if  $\gamma\delta$  TCR and NK cell receptors induce synergistic signaling for  $\gamma\delta$  T cell responses (52). The list of costimulatory receptors for  $\alpha\beta$  T cells has been expanded and includes a prototype costimulatory molecule CD28 (53). The relevance of costimulatory molecules for  $\alpha\beta$  T cells in  $\gamma\delta$  T cells remains debatable. About 40%–60% of  $\gamma\delta$  T cells express CD28, and the expression of CD28 is decreased upon the activation of  $\gamma\delta$  T cells (54,55). Since anti-CD28 agonistic Abs enhance human  $\gamma\delta$  T cell proliferation, the role of CD28 as a costimulatory molecule is valid in a subpopulation of human  $\gamma\delta$  T cells. Considering the phenotypes of memory and effector CD8<sup>+</sup>  $\alpha\beta$  T cells (56), it may be hypothesized that the expression of CD28 is lost upon the prolonged activation of a subpopulation of  $\gamma\delta$  T cells. It is noteworthy that a higher proportion of V $\delta$ 1  $\gamma\delta$  T cells do not express CD28 than that of V $\delta$ 2  $\gamma\delta$  T cells, but the most of V $\delta$ 2  $\gamma\delta$  T cells express CD28 similarly to naïve  $\alpha\beta$  T cells (35,57).

# THE $\gamma\delta$ T Cells in the tumor microenvironment (TME)

#### Recruitment of human $\gamma\delta$ T cells into the TME

Cancer is characterized not only by transformed cancer cells but also by non-cancer cells, such as immune cells, fibroblasts, and endothelial cells, and the extracellular matrix that establishes the TME. Initially, the cellular stresses experienced by transformed cancer cells trigger the upregulation of ligands for NK cell receptors (58). Although initially recruited NK cells can kill cancer cells, the cytotoxic activity of NK cells is not sustained but exhausted when cancer cells outnumber NK cells in the advanced stage of cancer (59). The persistent chronic inflammation associated with cancer recruits many kinds of immune cells, including Treg cells and myeloid-derived suppressor cells into the TME. It is a common consensus that the TME inhibits the anti-tumor immune responses in most clinical situations (60-62).

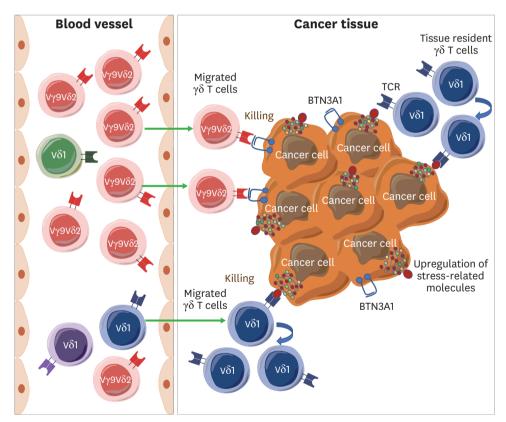
The  $\gamma\delta$  T cells also infiltrate into a variety of the tumors in the early and late stages of cancer development, where they are known to modulate the anti-tumor response through pro- or

anti-inflammatory cytokines and their interactions with different types of innate and adaptive immune cells in the TME (7,49,63,64). The  $\gamma\delta$  T cells migrate into the TME in response to CC chemokines such as MCP-1, regulated on activation normal T cell expressed and secreted, MIP-1 $\alpha$  and MIP-1 $\beta$  (11,35,65).

#### Major tumor-infiltrating $\gamma\delta$ T cell subsets: human V $\delta$ 1 and V $\gamma$ 9V $\delta$ 2 T cells

In humans, V $\delta$ 1 and V $\gamma$ 9V $\delta$ 2 T cells are 2 main populations of  $\gamma\delta$  T cells in the tissues and peripheral blood. In tumors, one subset can be predominant over the other depending on the types and origin of the tumors (7,11,49,64-67). Both V $\delta$ 1 and V $\gamma$ 9V $\delta$ 2 T cells have the cytotoxic capability and can have anti-cancer activity (11,36). The 2 subsets of  $\gamma\delta$  T cells express distinct chemokine receptors and cell adhesion molecules, suggesting different homing mechanisms that can be selectively utilized for cancer immunotherapy (35,68,69). A diagram is displayed in **Fig. 1**, which shows their differential involvement in the anti-cancer immunity.

The human V $\gamma$ 9V $\delta$ 2 T cells are the most predominant  $\gamma\delta$  T cells in the adult peripheral blood, but they are not a major  $\gamma\delta$  T cell population at the time of birth as the V $\delta$ 1  $\gamma\delta$  T cells are predominant during fetal and early life (24,31). The V $\gamma$ 9V $\delta$ 2 T cells expand postnatally in response to phoshoantigens by microbes. The canonical V $\gamma$ 9V $\delta$ 2 T cells with V $\gamma$ 9J $\gamma$ P sequences recognize phosphoantigens presented by butyrophilin 3A (BTN3A). Interestingly, prenyl pyrophosphates (phosphoantigens) bind to the intracellular B30.2 domain of BTN3A1,



**Figure 1.** Differential recruitment of V $\delta$ 1 and V $\gamma$ 9 $\delta$ 2  $\gamma\delta$  T cells into the tumor tissue. In blood, V $\gamma$ 9 $\delta$ 2  $\gamma\delta$  T cells are predominant over V $\delta$ 2  $\gamma\delta$  T cells in healthy individuals. Most of the V $\gamma$ 9 $\delta$ 2  $\gamma\delta$  T cells have canonical TCRs responding to prenyl pyrophosphates that are elevated in cancer cells and are recruited into the tumor via chemokine receptors. In contrast, some clonotypes of V $\delta$ 1  $\gamma\delta$ T cells are selected from a diverse V $\delta$ 1 TCR repertoire. Specific V $\delta$ 1  $\gamma\delta$ T cells migrate into the tumor tissues, expand, and kill cancer cells. The tissue-resident V $\delta$ 1  $\gamma\delta$ T cells may respond to the tissue stress and proliferate to kill cancer cells.

suggesting that the Vy9Vô2 TCR senses the internal changes of mevalonate or non-mevalonate metabolic pathways within cancer or infected cells, respectively (70,71). Elevated cytoplasmic prenyl pyrophosphate levels as a result of a dysregulation of the mevalonate pathway triggers an inside-out signaling leading to a structural change of the extracellular domain of BTN3A1, which enhances the binding force between BTN3A1 and  $V\gamma 9V\delta 2$  TCR. It is noteworthy that the RhoB activation in cancer cells is another determinant for the relocalization of BTN3A1 and the activation of V $\gamma$ 9V $\delta$ 2  $\gamma\delta$  T cells (72). Therefore, the canonical V $\gamma$ 9V $\delta$ 2  $\gamma\delta$  T cells can be cancerkilling lymphocytes through the recognition of the altered metabolism of cancer cells and the mutation of RhoB. However, the functional role and Ags of non-canonical Vy9V82 v8 T cells are not well understood. The non-canonical  $V\gamma 9V\delta 2 \gamma \delta T$  cells predominate over the canonical  $V\gamma 9V\delta 2 \gamma \delta T$  cells in the fetal tissues, but the canonical  $V\gamma 9V\delta 2 \gamma \delta T$  cells become predominant in the adult blood. Interestingly, glioblastoma-infiltrating  $v\delta$  T cells have non-canonical TCR repertoire using Cy2 segment (66). Regarding the ligands for Vy9V $\delta$ 2 y $\delta$  TCR, it is needed to address the role of a novel ligand for Vy9 TCR, BTN2A1, in the cancer immunity and whether BTN2A1 is upregulated in the cancer cells (73). The  $V\gamma 9V\delta 2$  T cells also recognize cancer cells through stress-related proteins that can be upregulated upon malignant transformation, including the F1-ATPase, MICA/B, heat shock protein 60, ULBP, human MutS homolog 2, and DNAX-associated molecule-1 through NK cell receptors (1,2,74,75).

Whereas most of the V $\gamma$ 9V $\delta$ 2 T cells have restricted canonical TCRs, the V $\delta$ 1 T cells have diverse repertoires that use various kinds of Vy chains (35). In most adults, a small number of specific clonotypes emerge from an initially unfocused neonatal Vô1 TCR repertoire, undergo pronounced clonal expansion, and ultimately dominate the V $\delta$ 1 T cell compartment (69). The TCR-diverse CD27<sup>high</sup> and highly TCR focused CD27<sup>low/-</sup> populations represent naïve and effector V81 T cell subsets, respectively. The transition from naïve to effector Vô1 T cells is accompanied by a switch from lymphoid to peripheral homing receptors. Although ligands detected by V $\delta$ 1 TCRs remain largely uncharacterized, some peripheral circulating and tissue-resident Vô1 T cells recognize CD1c, the lipid-presenting MHC-like molecule CD1d, or MHC-related protein 1 (2,76-78). In addition to the TCR, Vo1 T cells also respond to a distinct set of cellular stress signals expressed by cancerous cells, such as MICA/B, ULBPs, B7-H6, and BAT3 via NKG2D or NCRs (1,77,79). In many tumors, V $\delta 1 \gamma \delta$  T cells are predominant over the V $\gamma$ 9V $\delta 2 \gamma \delta$  T cells, which suggests that the V $\delta 1 \gamma \delta$ T cells expand responding to cancer Ags (80). The utilization of the V $\delta$ T cells is a promising option for cancer immunotherapy. Therefore, the relative importance of V $\delta$ 1 y $\delta$ T cells and Vy9V $\delta$ 2  $\gamma\delta$  T cells should be considered in the future immunotherapy using  $\gamma\delta$  T cells (4,10,12,14,16). Non-V $\delta$ 1 non-V $\gamma$ 9V $\delta$ 2  $\gamma\delta$  T cells should also be considered as the V $\gamma$ 4V $\delta$ 5  $\gamma\delta$  T cells are able to eliminate cancer cells by recognizing endothelial protein C receptor (81).

#### The $\gamma\delta$ T cells have both anti-tumor and pro-tumor activities

Upon migration to the TME,  $\gamma\delta$  T cells exert potent anti-tumor effects via multiple mechanisms (77,82). The  $\gamma\delta$  T cells can eliminate cancer cells via cytolytic receptor-ligand interactions including Fas ligand (83) and TNF-related apoptosis-inducing ligand (84) in addition to granzyme B and perforin and also have cytostatic anti-cancer activities by releasing IFN- $\gamma$  or TNF- $\alpha$  (82,85). The  $\gamma\delta$  T cells are also able to kill Ab-coated cancer cells by Ab-dependent cellular cytotoxicity using cell surface CD16 (Fc $\gamma$ RIII) similar to NK cells (86). Lastly,  $\gamma\delta$  T cells exhibit indirect anti-tumor responses by modulating different immune cell types including DCs, NK cells, neutrophils,  $\alpha\beta$  T cells and B cells in the TME (1,4,22,75,77,82,85).

In general, the extent of intratumoral  $\gamma\delta$  T cell infiltration is highly associated with the CD8<sup>+</sup> T cell signature and patients' prognosis, suggesting that  $\gamma\delta$  T cells largely perform anti-tumor

activity rather than pro-tumor activity (87,88). However, complex interactions between TME and intratumoral  $\gamma\delta$  T cells can result in the diversion of anti-tumor  $\gamma\delta$  T cells into protumor cells. Therefore, the precise role of  $\gamma\delta$  T cells in each individual patient may depend on the specific  $\gamma\delta$  T cell subsets and their functional polarization in the TME (63,77,82,85). Regarding T cell polarization, it is generally stated that Th1 and follicular Th (Tfh) cells have anti-tumor activity, whereas Th17 and Treg cells have pro-tumor activity (89). As the γδ Tfh cell-driven GC response tends to induce autoreactive B cells instead of pathogen-specific B cells, the anti-tumor activity of Tfh cells is not well established (90), but the involvement of Tfh cells in cancer tissues indicates an organized anti-tumor immunity with tertiary lymphoid tissue (91). Effector  $\gamma\delta$  T cells can also be classified as  $\gamma\delta$  Th1,  $\gamma\delta$  Th2,  $\gamma\delta$  Th17,  $\gamma\delta$  Tfh, and γδ Treg cells based on their functional polarization (1,7,22,35,49,75,77,85,92). Interestingly, in response to different cytokines,  $v\delta T$  cells can trans-differentiate from one phenotype to another (1,7,22,35,49,75,77,85,92). Both V $\delta$ 1 and V $\gamma$ 9V $\delta$ 2 T cells can be polarized into  $\gamma\delta$  Th1 cells, yo Tfh cells, yo T17 cells, yo Treg cells, and yo Th2 cells with distinct cytokines. It is important to investigate further whether  $\gamma\delta$  T cells are a primary driver of T cell polarization or whether the immunotherapy targeting  $\gamma\delta$  T cells can change the overall polarization of  $\alpha\beta$ T cells within the TME.

The  $\gamma\delta$  T cells are also subjected to immune exhaustion similarly to cancer-reactive cytotoxic T cells and NK cells. Although the nature of the  $\gamma\delta$  T cell immune exhaustion is not well reported, prolonged stimulation of  $\gamma\delta$  T cells appears to trigger their immune exhaustion. Since the immune exhaustion is reviewed extensively elsewhere (93-95), it will not be discussed here. It would be important and interesting to address how easily and deeply  $\gamma\delta$  T cells are exhausted and whether exhausted  $\gamma\delta$  T cells can be easily reawakened by strong stimuli, including cytokines or Ags such as phosphoantigens.

# FUTURE DIRECTIONS FOR OPTIMIZING ADOPTIVE $\gamma\delta$ T CELL TRANSFER AS AN ALTERNATIVE CANCER IMMUNOTHERAPY

The ability of  $\gamma\delta$  T cells to recognize the cellular stress via an MHC-independent mechanism and to potentiate other innate and adaptive immune cells makes them attractive mediators of cancer immunotherapy with potent and broad anti-tumor cytotoxicity (4,7,11,64,65,68,77,82,85,89). Especially, given their potent MHC-unrestricted anti-tumor activities,  $\gamma\delta$  T cells also can be considered as universal allogeneic adoptive T cell transfer for cancer patients. Accordingly, recent applications of  $\gamma\delta$  T cells to solid tumors have yielded promising results with associated clinical benefits, but issues of limited efficacy still remain with an average response ratio of only 21% and low proportion of complete remissions (7,14-16,85,96,97). Unfortunately, the tumor cells are effectively protected from tumor cell-killing immune activities in the immune-suppressive TME, which may also block the infiltration of infused  $\gamma\delta$  T cells. Furthermore, the anti-tumor function of  $\gamma\delta$  T cells can be limited by the pleiotropic effects of a mixture of heterogeneous populations of immune cells in the TME (4,7,14,16,63,64,85,97).

Therefore, current efforts in favor of a durable anti-tumor benefit from  $\gamma\delta$  T cell immunotherapy lie in the quest to minimize activation-induced  $\gamma\delta$  T cell death, anergy, and the polarization to specific  $\gamma\delta$  T cells with immunosuppressive function (4,7,14,15,92,98-100). Additionally, several cytokines such as IL-15, IL-18, and IL-21 have been found to have the ability to promote the

expansion of  $\gamma\delta$  T cells with a higher proliferative capacity, a more pronounced Th1 polarization, and an increased cytotoxic capacity and secretion of immune-stimulating paracrine factors such as GM-CSF, IFN- $\gamma$ , and TNF- $\alpha$  (101-103). In particular, disruption of the immunosuppressive TME could be a new strategy for improving the anti-tumor efficacy of  $\gamma\delta$  T cells. For example, IL-36 $\gamma$  acts synergistically with TCR signaling and is able to promote IFN- $\gamma$  production by CD8<sup>+</sup> T cells, NK cells, and  $\gamma\delta$  T cells by transforming the TME in favor of cancer eradication (104).

Up to date, clinical trials have been based on the adoptive transfer of peripheral circulating V $\gamma$ 9V $\delta$ 2 T cells after *ex vivo* expansion and activation (11,15,16,68,82,97,105). Given the accumulating pieces of evidence supporting the superior anti-tumor functionality of V $\delta$ 1 T cells compared with that of V $\gamma$ 9V $\delta$ 2 T cells, at least in the context of certain tumors (14,67,77,98,99,106-110), V $\delta$ 1 T cells may be a potent tool for clinical manipulation in cancer immunotherapy, and efforts have been put forth to explore strategies for clinical-grade expansion. An interesting property of V $\delta$ 1 T cells for the adoptive transfer approach is their CCL2-mediated chemotaxis toward tumors (67,111,112). V $\delta$ 1 T cells are also less susceptible to activation-induced cell death and could persist in the circulation for many years, which is in favor of a durable anti-tumor immunity (98,99,110). Intriguingly, IL-4 promotes the proliferation of V $\delta$ 1 T cells and simultaneously inhibits V $\delta$ 2 T-cell growth (77,80,113), thus providing a novel basis to develop the preferential expansion approaches for V $\delta$ 1 T cells.

#### **CONCLUDING REMARKS**

Although  $\gamma\delta$  T cells are a small population of lymphocytes, they contribute significantly to rapid and sustained immune responses against cancer. In order to utilize the inherent activity of  $\gamma\delta$  T cells for cancer immunotherapy, it is critical to better characterize human  $\gamma\delta$  T cell subsets and the engaged mechanisms in various types of cancers. It is also necessary to understand the central paradigms that govern the tissue tropism, the stage of differentiation, the activation status, and the immune checkpoint receptor expression in  $\gamma\delta$  T cells so that  $\gamma\delta$  T cells can be durably activated with a potent anti-tumor phenotype. To maintain the anti-tumor activity of  $\gamma\delta$  T cells for a long period of time, the specific depletion of pro-tumor  $\gamma\delta$  T cells before the immunotherapy, the co-transfer of other immune cells that activate  $\gamma\delta$ T cells, and the modification of the cytokine balance in the TME should be considered in the immunotherapy using  $\gamma\delta$  T cells. In summary, as  $\gamma\delta$  T cells are heterogeneous, the pro-tumor or anti-tumor activities of different  $\gamma\delta$  T cell populations need to be thoroughly delineated and utilized to maximize the efficacy of the immunotherapy using  $\gamma\delta$  T cells.

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#### REFERENCES

 Vantourout P, Hayday A. Six-of-the-best: unique contributions of γδ T cells to immunology. Nat Rev Immunol 2013;13:88-100.
 PUBMED | CROSSREF

- Adams EJ, Gu S, Luoma AM. Human gamma delta T cells: evolution and ligand recognition. *Cell Immunol* 2015;296:31-40.
   PUBMED | CROSSREF
- 3. Chien YH, Meyer C, Bonneville M. γδ T cells: first line of defense and beyond. *Annu Rev Immunol* 2014;32:121-155.

- Stolk D, van der Vliet HJ, de Gruijl TD, van Kooyk Y, Exley MA. Positive & negative roles of innate effector cells in controlling cancer progression. *Front Immunol* 2018;9:1990.
- 5. Girardi M, Oppenheim DE, Steele CR, Lewis JM, Glusac E, Filler R, Hobby P, Sutton B, Tigelaar RE, Hayday AC. Regulation of cutaneous malignancy by gammadelta T cells. *Science* 2001;294:605-609. PUBMED | CROSSREF
- Mishra R, Chen AT, Welsh RM, Szomolanyi-Tsuda E. NK cells and gammadelta T cells mediate resistance to polyomavirus-induced tumors. *PLoS Pathog* 2010;6:e1000924.
   PUBMED | CROSSREF
- 7. Zhao Y, Niu C, Cui J. Gamma-delta (γδ) T cells: friend or foe in cancer development? *J Transl Med* 2018;16:3. PUBMED | CROSSREF
- Barrow AD, Martin CJ, Colonna M. The natural cytotoxicity receptors in health and disease. *Front Immunol* 2019;10:909.
  - PUBMED | CROSSREF
- Wrobel P, Shojaei H, Schittek B, Gieseler F, Wollenberg B, Kalthoff H, Kabelitz D, Wesch D. Lysis of a broad range of epithelial tumour cells by human gamma delta T cells: involvement of NKG2D ligands and T-cell receptor- versus NKG2D-dependent recognition. *Scand J Immunol* 2007;66:320-328.
   PUBMED | CROSSREF
- Braza MS, Klein B. Anti-tumour immunotherapy with Vγ9Vδ2 T lymphocytes: from the bench to the bedside. *Br J Haematol* 2013;160:123-132.
   PUBMED | CROSSREF
- Hannani D, Ma Y, Yamazaki T, Déchanet-Merville J, Kroemer G, Zitvogel L. Harnessing γδ T cells in anticancer immunotherapy. *Trends Immunol* 2012;33:199-206.
   PUBMED | CROSSREF
- Kunzmann V, Smetak M, Kimmel B, Weigang-Koehler K, Goebeler M, Birkmann J, Becker J, Schmidt-Wolf IG, Einsele H, Wilhelm M. Tumor-promoting versus tumor-antagonizing roles of γδ T cells in cancer immunotherapy: results from a prospective phase I/II trial. *J Immunother* 2012;35:205-213.
   PUBMED | CROSSREF
- Nicol AJ, Tokuyama H, Mattarollo SR, Hagi T, Suzuki K, Yokokawa K, Nieda M. Clinical evaluation of autologous gamma delta T cell-based immunotherapy for metastatic solid tumours. *Br J Cancer* 2011;105:778-786.
   PUBMED | CROSSREF
- Rei M, Pennington DJ, Silva-Santos B. The emerging protumor role of γδ T lymphocytes: implications for cancer immunotherapy. *Cancer Res* 2015;75:798-802.
   PUBMED | CROSSREF
- Fournié JJ, Sicard H, Poupot M, Bezombes C, Blanc A, Romagné F, Ysebaert L, Laurent G. What lessons can be learned from γδ T cell-based cancer immunotherapy trials? *Cell Mol Immunol* 2013;10:35-41.
   PUBMED | CROSSREF
- Lo Presti E, Pizzolato G, Gulotta E, Cocorullo G, Gulotta G, Dieli F, Meraviglia S. Current advances in γδ T cell-based tumor immunotherapy. *Front Immunol* 2017;8:1401.
   PUBMED | CROSSREF
- 17. Xiang Z, Tu W. Dual face of Vγ9Vδ2-T cells in tumor immunology: anti- versus pro-tumoral activities. *Front Immunol* 2017;8:1041.
   PUBMED | CROSSREF
- 18. Turchinovich G, Pennington DJ. T cell receptor signalling in γδ cell development: strength isn't everything. *Trends Immunol* 2011;32:567-573.
   PUBMED | CROSSREF
- Baker JE, Cado D, Raulet DH. Developmentally programmed rearrangement of T cell receptor Vgamma genes is controlled by sequences immediately upstream of the Vgamma genes. *Immunity* 1998;9:159-168.
   PUBMED | CROSSREF
- Chodaczek G, Papanna V, Zal MA, Zal T. Body-barrier surveillance by epidermal γδ TCRs. *Nat Immunol* 2012;13:272-282.
   PUBMED | CROSSREF



- Boyden LM, Lewis JM, Barbee SD, Bas A, Girardi M, Hayday AC, Tigelaar RE, Lifton RP. Skint1, the prototype of a newly identified immunoglobulin superfamily gene cluster, positively selects epidermal gammadelta T cells. *Nat Genet* 2008;40:656-662.
   PUBMED | CROSSREF
- Bonneville M, O'Brien RL, Born WK. Gammadelta T cell effector functions: a blend of innate programming and acquired plasticity. *Nat Rev Immunol* 2010;10:467-478.
- 23. O'Brien RL, Born WK. Gammadelta T cell subsets: a link between TCR and function? *Semin Immunol* 2010;22:193-198.

- Vermijlen D, Prinz I. Ontogeny of innate T lymphocytes some innate lymphocytes are more innate than others. *Front Immunol* 2014;5:486.
   PUBMED I CROSSREF
- Lanier LL, Weiss A. Presence of Ti (WT31) negative T lymphocytes in normal blood and thymus. *Nature* 1986;324:268-270.
   PUBMED | CROSSREF

#### Morita CT, Parker CM, Brenner MB, Band H. TCR usage and functional capabilities of human gamma delta T cells at birth. *J Immunol* 1994;153:3979-3988.

- Vermijlen D, Brouwer M, Donner C, Liesnard C, Tackoen M, Van Rysselberge M, Twité N, Goldman M, Marchant A, Willems F. Human cytomegalovirus elicits fetal gammadelta T cell responses in utero. *J Exp Med* 2010;207:807-821.
   PUBMED | CROSSREF
- Parker CM, Groh V, Band H, Porcelli SA, Morita C, Fabbi M, Glass D, Strominger JL, Brenner MB. Evidence for extrathymic changes in the T cell receptor gamma/delta repertoire. *J Exp Med* 1990;171:1597-1612.
   PUBMED | CROSSREF
- Cairo C, Mancino G, Cappelli G, Pauza CD, Galli E, Brunetti E, Colizzi V. Vdelta2 T-lymphocyte responses in cord blood samples from Italy and Côte d'Ivoire. *Immunology* 2008;124:380-387.
- 30. Moens E, Brouwer M, Dimova T, Goldman M, Willems F, Vermijlen D. IL-23R and TCR signaling drives the generation of neonatal Vgamma9Vdelta2 T cells expressing high levels of cytotoxic mediators and producing IFN-gamma and IL-17. *J Leukoc Biol* 2011;89:743-752. PUBMED | CROSSREF
- De Rosa SC, Andrus JP, Perfetto SP, Mantovani JJ, Herzenberg LA, Herzenberg LA, Roederer M. Ontogeny of gamma delta T cells in humans. *J Immunol* 2004;172:1637-1645.
   PUBMED | CROSSREF
- S2. Cruz MS, Diamond A, Russell A, Jameson JM. Human αβ and γδ T cells in skin immunity and disease. *Front Immunol* 2018;9:1304.
   PUBMED | CROSSREF
- Allison TJ, Winter CC, Fournié JJ, Bonneville M, Garboczi DN. Structure of a human gammadelta T-cell antigen receptor. *Nature* 2001;411:820-824.
   PUBMED | CROSSREF
- 34. Legut M, Cole DK, Sewell AK. The promise of γδ T cells and the γδ T cell receptor for cancer immunotherapy. *Cell Mol Immunol* 2015;12:656-668.
  PUBMED | CROSSREF
- Davey MS, Willcox CR, Baker AT, Hunter S, Willcox BE. Recasting human Vô1 lymphocytes in an adaptive role. *Trends Immunol* 2018;39:446-459.
   PUBMED | CROSSREF
- 36. Pizzolato G, Kaminski H, Tosolini M, Franchini DM, Pont F, Martins F, Valle C, Labourdette D, Cadot S, Quillet-Mary A, et al. Single-cell RNA sequencing unveils the shared and the distinct cytotoxic hallmarks of human TCRVδ1 and TCRVδ2 γδ T lymphocytes. *Proc Natl Acad Sci U S A* 2019;116:11906-11915. PUBMED | CROSSREF
- 37. Kazen AR, Adams EJ. Evolution of the V, D, and J gene segments used in the primate gammadelta T-cell receptor reveals a dichotomy of conservation and diversity. *Proc Natl Acad Sci U S A* 2011;108:E332-E340.
  PUBMED | CROSSREF
- 38. Vavassori S, Kumar A, Wan GS, Ramanjaneyulu GS, Cavallari M, El Daker S, Beddoe T, Theodossis A, Williams NK, Gostick E, et al. Butyrophilin 3A1 binds phosphorylated antigens and stimulates human γδ T cells. *Nat Immunol* 2013;14:908-916.
   PUBMED | CROSSREF

- Simões AE, Di Lorenzo B, Silva-Santos B. Molecular determinants of target cell recognition by human γδ T cells. *Front Immunol* 2018;9:929.
   PUBMED I CROSSREF
- 40. Hayes SM, Love PE. Stoichiometry of the murine gammadelta T cell receptor. *J Exp Med* 2006;203:47-52. PUBMED | CROSSREF
- Malissen M, Gillet A, Ardouin L, Bouvier G, Trucy J, Ferrier P, Vivier E, Malissen B. Altered T cell development in mice with a targeted mutation of the CD3-epsilon gene. *EMBO J* 1995;14:4641-4653.
   PUBMED | CROSSREF
- 42. Dave VP, Cao Z, Browne C, Alarcon B, Fernandez-Miguel G, Lafaille J, de la Hera A, Tonegawa S, Kappes DJ. CD3 delta deficiency arrests development of the alpha beta but not the gamma delta T cell lineage. *EMBO J* 1997;16:1360-1370.
   PUBMED | CROSSREF
- 43. Haks MC, Krimpenfort P, Borst J, Kruisbeek AM. The CD3gamma chain is essential for development of both the TCRalphabeta and TCRgammadelta lineages. *EMBO J* 1998;17:1871-1882. PUBMED | CROSSREF
- 44. Hayes SM, Love PE. Distinct structure and signaling potential of the gamma delta TCR complex. *Immunity* 2002;16:827-838.

```
PUBMED | CROSSREF
```

- 45. Siegers GM, Swamy M, Fernández-Malavé E, Minguet S, Rathmann S, Guardo AC, Pérez-Flores V, Regueiro JR, Alarcón B, Fisch P, et al. Different composition of the human and the mouse gammadelta T cell receptor explains different phenotypes of CD3gamma and CD3delta immunodeficiencies. *J Exp Med* 2007;204:2537-2544.
  PUBMED | CROSSREF
- 46. Recio MJ, Moreno-Pelayo MA, Kiliç SS, Guardo AC, Sanal O, Allende LM, Pérez-Flores V, Mencía A, Modamio-Høybjør S, Seoane E, et al. Differential biological role of CD3 chains revealed by human immunodeficiencies. *J Immunol* 2007;178:2556-2564. PURMED | CROSSREE
- 47. Jensen KD, Su X, Shin S, Li L, Youssef S, Yamasaki S, Steinman L, Saito T, Locksley RM, Davis MM, et al. Thymic selection determines gammadelta T cell effector fate: antigen-naive cells make interleukin-17 and antigen-experienced cells make interferon gamma. *Immunity* 2008;29:90-100. PUBMED | CROSSREF
- Taghon T, Yui MA, Pant R, Diamond RA, Rothenberg EV. Developmental and molecular characterization of emerging beta- and gammadelta-selected pre-T cells in the adult mouse thymus. *Immunity* 2006;24:53-64.
   PUBMED | CROSSREF
- 49. Lafont V, Sanchez F, Laprevotte E, Michaud HA, Gros L, Eliaou JF, Bonnefoy N. Plasticity of γδ T cells: impact on the anti-tumor response. *Front Immunol* 2014;5:622. PUBMED | CROSSREF
- 50. Hayday A, Tigelaar R. Casting new light on the TCR. *Nat Immunol* 2012;13:209-211. PUBMED | CROSSREF
- Parham P, Moffett A. Variable NK cell receptors and their MHC class I ligands in immunity, reproduction and human evolution. *Nat Rev Immunol* 2013;13:133-144.
   PUBMED | CROSSREF
- 52. Wensveen FM, Jelenčić V, Polić B. NKG2D: a master regulator of immune cell responsiveness. *Front Immunol* 2018;9:441.

- 53. Ribot JC, debarros A, Silva-Santos B. Searching for "signal 2": costimulation requirements of γδ T cells. *Cell Mol Life Sci* 2011;68:2345-2355.
   PUBMED | CROSSREF
- 54. Testi R, Lanier LL. Functional expression of CD28 on T cell antigen receptor gamma/delta-bearing T lymphocytes. Eur J Immunol 1989;19:185-188.
  PUBMED | CROSSREF
- 55. Penninger JM, Timms E, Shahinian A, Jezo-Bremond A, Nishina H, Ionescu J, Hedrick SM, Mak TW. Alloreactive gamma delta thymocytes utilize distinct costimulatory signals from peripheral T cells. *J Immunol* 1995;155:3847-3855.
  PUBMED
- 56. Hamann D, Baars PA, Rep MH, Hooibrink B, Kerkhof-Garde SR, Klein MR, van Lier RA. Phenotypic and functional separation of memory and effector human CD8+ T cells. J Exp Med 1997;186:1407-1418. PUBMED | CROSSREF



- 57. De Rosa SC, Mitra DK, Watanabe N, Herzenberg LA, Herzenberg LA, Roederer M. Vdelta1 and Vdelta2 gammadelta T cells express distinct surface markers and might be developmentally distinct lineages. J Leukoc Biol 2001;70:518-526. PUBMED
- 58. Seelige R, Searles S, Bui JD. Mechanisms regulating immune surveillance of cellular stress in cancer. Cell Mol Life Sci 2018;75:225-240. PUBMED | CROSSREF

#### 59. Bassani B, Baci D, Gallazzi M, Poggi A, Bruno A, Mortara L. Natural killer cells as key players of tumor progression and angiogenesis: old and novel tools to divert their pro-tumor activities into potent antitumor effects. Cancers (Basel) 2019;11:461. PUBMED | CROSSREF

- 60. O'Donnell JS, Teng MW, Smyth MJ. Cancer immunoediting and resistance to T cell-based immunotherapy. Nat Rev Clin Oncol 2019;16:151-167. PUBMED | CROSSREF
- 61. Runa F, Hamalian S, Meade K, Shisgal P, Gray PC, Kelber JA. Tumor microenvironment heterogeneity: challenges and opportunities. Curr Mol Biol Rep 2017;3:218-229. PUBMED | CROSSREE
- 62. Nakamura K, Smyth MJ. Targeting cancer-related inflammation in the era of immunotherapy. Immunol Cell Biol 2017;95:325-332.

PUBMED | CROSSREF

- 63. Lo Presti E, Di Mitri R, Pizzolato G, Mocciaro F, Dieli F, Meraviglia S. γδ cells and tumor microenvironment: a helpful or a dangerous liason? J Leukoc Biol 2018;103:485-492. PUBMED | CROSSREF
- 64. Paul S, Lal G. Regulatory and effector functions of gamma-delta ( $\gamma\delta$ ) T cells and their therapeutic potential in adoptive cellular therapy for cancer. Int J Cancer 2016;139:976-985. PUBMED | CROSSREF
- 65. Silva-Santos B, Serre K, Norell H. γδ T cells in cancer. Nat Rev Immunol 2015;15:683-691. PUBMED | CROSSREF
- 66. Lee M, Park C, Woo J, Kim J, Kho I, Nam DH, Park WY, Kim YS, Kong DS, Lee HW, et al. Preferential infiltration of unique Vy9Jy2-V82 T cells into glioblastoma multiforme. Front Immunol 2019;10:555. PUBMED | CROSSREF
- 67. Cordova A, Toia F, La Mendola C, Orlando V, Meraviglia S, Rinaldi G, Todaro M, Cicero G, Zichichi L, Donni PL, et al. Characterization of human  $\gamma\delta$  T lymphocytes infiltrating primary malignant melanomas. PLoS One 2012;7:e49878. PUBMED | CROSSREF
- 68. Deniger DC, Moyes JS, Cooper LJ. Clinical applications of gamma delta T cells with multivalent immunity. Front Immunol 2014;5:636. PUBMED | CROSSREF

#### 69. Davey MS, Willcox CR, Joyce SP, Ladell K, Kasatskaya SA, McLaren JE, Hunter S, Salim M, Mohammed F, Price DA, et al. Clonal selection in the human Vδ1 T cell repertoire indicates γδ TCR-dependent adaptive immune surveillance. Nat Commun 2017;8:14760. PUBMED | CROSSREF

- 70. Sandstrom A, Peigné CM, Léger A, Crooks JE, Konczak F, Gesnel MC, Breathnach R, Bonneville M, Scotet E, Adams EJ. The intracellular B30.2 domain of butyrophilin 3A1 binds phosphoantigens to mediate activation of human Vy9V82 T cells. Immunity 2014;40:490-500. PUBMED | CROSSREF
- 71. Yang Y, Li L, Yuan L, Zhou X, Duan J, Xiao H, Cai N, Han S, Ma X, Liu W, et al. A structural change in butyrophilin upon phosphoantigen binding underlies phosphoantigen-mediated  $V\gamma 9V\delta 2$  T cell activation. Immunity 2019;50:1043-1053.e5. PUBMED | CROSSREF
- 72. Sebestyen Z, Scheper W, Vyborova A, Gu S, Rychnavska Z, Schiffler M, Cleven A, Chéneau C, van Noorden M, Peigné CM, et al. RhoB mediates phosphoantigen recognition by Vγ9Vδ2 T cell receptor. Cell Reports 2016;15:1973-1985. PUBMED | CROSSREF
- 73. Rigau M, Ostrouska S, Fulford TS, Johnson DN, Woods K, Ruan Z, McWilliam HE, Hudson C, Tutuka C, Wheatley AK, et al. Butyrophilin 2A1 is essential for phosphoantigen reactivity by γδ T cells. Science 2020:367:eaav5516.

PUBMED | CROSSREF



- 74. Dai Y, Chen H, Mo C, Cui L, He W. Ectopically expressed human tumor biomarker MutS homologue 2 is a novel endogenous ligand that is recognized by human γδ T cells to induce innate anti-tumor/virus immunity. *J Biol Chem* 2012;287:16812-16819. PUBMED | CROSSREF
- 75. Tyler CJ, Doherty DG, Moser B, Eberl M. Human Vγ9/Vδ2 T cells: innate adaptors of the immune system. *Cell Immunol* 2015;296:10-21.
   PUBMED I CROSSREF
- 76. Uldrich AP, Le Nours J, Pellicci DG, Gherardin NA, McPherson KG, Lim RT, Patel O, Beddoe T, Gras S, Rossjohn J, et al. CD1d-lipid antigen recognition by the γδ TCR. *Nat Immunol* 2013;14:1137-1145.
  PUBMED | CROSSREF
- 77. Wu D, Wu P, Qiu F, Wei Q, Huang J. Human γδT-cell subsets and their involvement in tumor immunity. *Cell Mol Immunol* 2017;14:245-253.
  PUBMED | CROSSREF
- 78. Le Nours J, Gherardin NA, Ramarathinam SH, Awad W, Wiede F, Gully BS, Khandokar Y, Praveena T, Wubben JM, Sandow JJ, et al. A class of γδ T cell receptors recognize the underside of the antigenpresenting molecule MR1. *Science* 2019;366:1522-1527.
  PURMED L CROSSREE
- 79. Hudspeth K, Silva-Santos B, Mavilio D. Natural cytotoxicity receptors: broader expression patterns and functions in innate and adaptive immune cells. *Front Immunol* 2013;4:69.
  PUBMED | CROSSREF
- Almeida AR, Correia DV, Fernandes-Platzgummer A, da Silva CL, da Silva MG, Anjos DR, Silva-Santos B. Delta one t cells for immunotherapy of chronic lymphocytic leukemia: clinical-grade expansion/ differentiation and preclinical proof of concept. *Clin Cancer Res* 2016;22:5795-5804.
   PUBMED | CROSSREF
- Willcox CR, Pitard V, Netzer S, Couzi L, Salim M, Silberzahn T, Moreau JF, Hayday AC, Willcox BE, Déchanet-Merville J. Cytomegalovirus and tumor stress surveillance by binding of a human γδ T cell antigen receptor to endothelial protein C receptor. *Nat Immunol* 2012;13:872-879.
   PUBMED | CROSSREF
- Deniger DC, Maiti SN, Mi T, Switzer KC, Ramachandran V, Hurton LV, Ang S, Olivares S, Rabinovich BA, Huls MH, et al. Activating and propagating polyclonal gamma delta T cells with broad specificity for malignancies. *Clin Cancer Res* 2014;20:5708-5719.
   PUBMED | CROSSREF
- 83. Li Z, Xu Q, Peng H, Cheng R, Sun Z, Ye Z. IFN-γ enhances HOS and U2OS cell lines susceptibility to γδ T cell-mediated killing through the Fas/Fas ligand pathway. *Int Immunopharmacol* 2011;11:496-503.
   PUBMED | CROSSREF
- 84. Tawfik D, Groth C, Gundlach JP, Peipp M, Kabelitz D, Becker T, Oberg HH, Trauzold A, Wesch D. TRAILreceptor 4 modulates γδ T cell-cytotoxicity toward cancer cells. *Front Immunol* 2019;10:2044. PUBMED | CROSSREF
- Pauza CD, Liou ML, Lahusen T, Xiao L, Lapidus RG, Cairo C, Li H. Gamma delta T cell therapy for cancer: it is good to be local. *Front Immunol* 2018;9:1305.
   PUBMED I CROSSREF
- 86. Angelini DF, Borsellino G, Poupot M, Diamantini A, Poupot R, Bernardi G, Poccia F, Fournié JJ, Battistini L. FcgammaRIII discriminates between 2 subsets of Vgamma9Vdelta2 effector cells with different responses and activation pathways. *Blood* 2004;104:1801-1807.
  PUBMED I CROSSREF
- Gentles AJ, Newman AM, Liu CL, Bratman SV, Feng W, Kim D, Nair VS, Xu Y, Khuong A, Hoang CD, et al. The prognostic landscape of genes and infiltrating immune cells across human cancers. *Nat Med* 2015;21:938-945.
  - PUBMED | CROSSREF
- Meraviglia S, Lo Presti E, Tosolini M, La Mendola C, Orlando V, Todaro M, Catalano V, Stassi G, Cicero G, Vieni S, et al. Distinctive features of tumor-infiltrating γδ T lymphocytes in human colorectal cancer. Oncolmmunology 2017;6:e1347742.
   PUBMED | CROSSREF
- Kim JS, Kim YG, Park EJ, Kim B, Lee HK, Hong JT, Kim Y, Han SB. Cell-based immunotherapy for colorectal cancer with cytokine-induced killer cells. *Immune Netw* 2016;16:99-108.
   PUBMED I CROSSREF
- 90. Park C, Kim TJ. Expansion and sub-classification of T cell-dependent antibody responses to encompass the role of innate-like T cells in antibody responses. *Immune Netw* 2018;18:e34. PUBMED | CROSSREF



- Gu-Trantien C, Loi S, Garaud S, Equeter C, Libin M, de Wind A, Ravoet M, Le Buanec H, Sibille C, Manfouo-Foutsop G, et al. CD4<sup>+</sup> follicular helper T cell infiltration predicts breast cancer survival. *J Clin Invest* 2013;123:2873-2892.
   PUBMED | CROSSREF
- 92. Caccamo N, La Mendola C, Orlando V, Meraviglia S, Todaro M, Stassi G, Sireci G, Fournié JJ, Dieli F. Differentiation, phenotype, and function of interleukin-17-producing human Vγ9Vδ2 T cells. *Blood* 2011;118:129-138. PUBMED | CROSSREF
- 93. Thommen DS, Schumacher TN. T cell dysfunction in cancer. *Cancer Cell* 2018;33:547-562. PUBMED | CROSSREF
- 94. Xia A, Zhang Y, Xu J, Yin T, Lu XJ. T cell dysfunction in cancer immunity and immunotherapy. *Front Immunol* 2019;10:1719.
   PUBMED | CROSSREF
- 95. Kim PS, Ahmed R. Features of responding T cells in cancer and chronic infection. *Curr Opin Immunol* 2010;22:223-230.

- 96. Meraviglia S, Caccamo N, Guggino G, Tolomeo M, Siragusa S, Stassi G, Dieli F. Optimizing tumorreactive γδ T cells for antibody-based cancer immunotherapy. *Curr Mol Med* 2010;10:719-726. PUBMED | CROSSREF
- 97. Zou C, Zhao P, Xiao Z, Han X, Fu F, Fu L. γδ T cells in cancer immunotherapy. *Oncotarget* 2017;8:8900-8909.
- 98. Wu D, Wu P, Wu X, Ye J, Wang Z, Zhao S, Ni C, Hu G, Xu J, Han Y, et al. *Ex vivo* expanded human circulating Vδ1 γδT cells exhibit favorable therapeutic potential for colon cancer. *Oncolmmunology* 2015;4:e992749.

PUBMED | CROSSREF

- 99. Siegers GM, Dhamko H, Wang XH, Mathieson AM, Kosaka Y, Felizardo TC, Medin JA, Tohda S, Schueler J, Fisch P, et al. Human Vδ1 γδ T cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemia-derived cells. *Cytotherapy* 2011;13:753-764.
  PUBMED | CROSSREF
- 100. Gonçalves-Sousa N, Ribot JC, deBarros A, Correia DV, Caramalho I, Silva-Santos B. Inhibition of murine gammadelta lymphocyte expansion and effector function by regulatory alphabeta T cells is cell-contact-dependent and sensitive to GITR modulation. *Eur J Immunol* 2010;40:61-70.
  PUBMED | CROSSREF
- 101. Li W, Kubo S, Okuda A, Yamamoto H, Ueda H, Tanaka T, Nakamura H, Yamanishi H, Terada N, Okamura H. Effect of IL-18 on expansion of gammadelta T cells stimulated by zoledronate and IL-2. *J Immunother* 2010;33:287-296.
  PUBMED | CROSSREF
- 102. Li W, Yamamoto H, Kubo S, Okamura H. Modulation of innate immunity by IL-18. *J Reprod Immunol* 2009;83:101-105.
   PUBMED | CROSSREF
- 103. Van Acker HH, Anguille S, Willemen Y, Van den Bergh JM, Berneman ZN, Lion E, Smits EL, Van Tendeloo VF. Interleukin-15 enhances the proliferation, stimulatory phenotype, and antitumor effector functions of human gamma delta T cells. *J Hematol Oncol* 2016;9:101. PUBMED I CROSSREF
- 104. Wang X, Zhao X, Feng C, Weinstein A, Xia R, Wen W, Lv Q, Zuo S, Tang P, Yang X, et al. IL-36γ transforms the tumor microenvironment and promotes type 1 lymphocyte-mediated antitumor immune responses. *Cancer Cell* 2015;28:296-306. PUBMED | CROSSREF
- 105. Thedrez A, Harly C, Morice A, Salot S, Bonneville M, Scotet E. IL-21-mediated potentiation of antitumor cytolytic and proinflammatory responses of human V gamma 9V delta 2 T cells for adoptive immunotherapy. *J Immunol* 2009;182:3423-3431.
  PUBMED | CROSSREF
- 106. Chen J, Niu H, He W, Ba D. Antitumor activity of expanded human tumor-infiltrating gammadelta T lymphocytes. Int Arch Allergy Immunol 2001;125:256-263.
  PUBMED | CROSSREF
- 107. Choudhary A, Davodeau F, Moreau A, Peyrat MA, Bonneville M, Jotereau F. Selective lysis of autologous tumor cells by recurrent gamma delta tumor-infiltrating lymphocytes from renal carcinoma. *J Immunol* 1995;154:3932-3940.
  PUBMED

https://immunenetwork.org



- 108. Couzi L, Levaillant Y, Jamai A, Pitard V, Lassalle R, Martin K, Garrigue I, Hawchar O, Siberchicot F, Moore N, et al. Cytomegalovirus-induced gammadelta T cells associate with reduced cancer risk after kidney transplantation. J Am Soc Nephrol 2010;21:181-188.
  PUBMED | CROSSREF
- 109. Knight A, Arnouk H, Britt W, Gillespie GY, Cloud GA, Harkins L, Su Y, Lowdell MW, Lamb LS. CMVindependent lysis of glioblastoma by *ex vivo* expanded/activated Vδ1+ γδ T cells. *PLoS One* 2013;8:e68729. PUBMED | CROSSREF
- 110. Knight A, Mackinnon S, Lowdell MW. Human Vdelta1 gamma-delta T cells exert potent specific cytotoxicity against primary multiple myeloma cells. *Cytotherapy* 2012;14:1110-1118.
  PUBMED | CROSSREF
- 111. Lança T, Costa MF, Gonçalves-Sousa N, Rei M, Grosso AR, Penido C, Silva-Santos B. Protective role of the inflammatory CCR2/CCL2 chemokine pathway through recruitment of type 1 cytotoxic γδ T lymphocytes to tumor beds. *J Immunol* 2013;190:6673-6680. PUBMED | CROSSREF
- 112. Peng G, Wang HY, Peng W, Kiniwa Y, Seo KH, Wang RF. Tumor-infiltrating gammadelta T cells suppress T and dendritic cell function via mechanisms controlled by a unique toll-like receptor signaling pathway. *Immunity* 2007;27:334-348.
  PUBMED | CROSSREF
- 113. Mao Y, Yin S, Zhang J, Hu Y, Huang B, Cui L, Kang N, He W. A new effect of IL-4 on human γδ T cells: promoting regulatory Vδ1 T cells via IL-10 production and inhibiting function of Vδ2 T cells. *Cell Mol Immunol* 2016;13:217-228.
  PUBMED | CROSSREF