

Original Article

Alpha basic crystallin expression in canine mammary tumors

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The aim of this study was to evaluate prognostic and/or diagnostic factors of canine mammary tumors by immunohistochemically analyzing the expression of alpha basic crystallin (α B-c). For this, formalin-fixed, paraffin-embedded blocks of 51 naturally-occurring canine mammary tumors (11 benign and 40 malignant) were used. Tissue from eight normal canine mammary glands were served as a control. Immunohistochemically, in the control mammary tissues, a few luminal epithelial cells were α B-c positive but myoepithelial cells were negative. In benign or simple type malignant tumors, α B-c expression was observed in luminal epithelial cells while the myoepithelial basal cells were negative. In benign or complex type malign tumors, positive staining was predominantly found in the cytoplasm of epithelial cells. Immunoreactivity of α B-c was also observed in neoplastic myoepithelial cells. Statistically, the number of cells immunolabeled with α B-c was found to be significantly different among tissues from normal canine mammary glands, benign lesions, and malignant tumors ($p < 0.05$). α B-c immunoreactivity was higher in malignant tumors than the control mammary tissues ($p < 0.001$). Data obtained in the current study revealed a strong association between high expression levels of α B-c and primary mammary gland tumors in canines.

Keywords: α B-crystallin, canine, mammary, neoplasms, tumors

Introduction

Heat shock proteins (HSPs) or stress proteins, which play an essential role in the maintenance of cellular homeostasis, are the products of several distinct gene families produced under both physiological and stress conditions [2]. Alpha basic-crystallin (α B-c) belongs to a family of small HSPs, and is expressed in response to different

stresses including heat shock, oxidative stress, metal ions, and cytokines [6,11]. α B-c is constitutively expressed in many humans and animals tissues such as the lens of the eyes, heart, and skeletal muscle [9,16]. α B-c has chaperone-like properties which prevent the aggregation of damaged or proteins misfolded due to cell stress, and has been shown to inhibit apoptosis [10].

Mammary tumors, the most common neoplasms in female dogs, may provide a useful model for human breast cancer research [15]. Both benign and malignant neoplasms account for approximately 50% of all tumors in dogs and show wide pathological and clinical heterogeneity [12,18]. After surgical intervention, approximately 48% of the affected dogs die or are euthanized within a 1-year period because of tumor recurrence or metastasis [7]. Therefore, additional and reliable prognostic tools are required for effective risk assessment. The aims of this study were to evaluate the prognostic and/or diagnostic factors associated with canine mammary tumors in dogs by analyzing the immunohistochemical expression of α B-c.

Materials and Methods

Formalin-fixed, paraffin-embedded blocks containing 51 naturally occurring canine mammary tumors (11 benign and 40 malignant) were selected retrospectively and acquired from the Veterinary Laboratory (Italy) and Department of Pathology, Faculty of Veterinary Medicine, Ondokuz Mayıs University (Turkey). Additionally, eight normal canine mammary glands obtained from the dogs that had died of causes unrelated to tumor development served as healthy control tissues. Three pathologists (TG, MYG and MS) independently diagnosed the tumors according to the World Health Organization classifications for canine mammary neoplasm [13].

Sections from all the tissue samples were cut (at a

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Table 1. Results of immunohistochemical staining for alpha basic crystallin (α B-c) expression and histopathologic diagnoses

Histopathologic diagnoses	Number of samples	Number and percent of samples for each α B-c immunoreactivity score			
		0*	1 [†]	2 [‡]	3 [§]
Malignant lesions	40	3 (7.5%)	10 (25%)	12 (30%)	15 (37.5%)
Complex carcinoma	14	–	3 (21.4%)	5 (35.7%)	6 (42.9%)
Carcinosarcoma	8	1 (12.5%)	1 (12.5%)	3 (37.5%)	3 (37.5%)
Tubulopapillary carcinoma	11	1 (9.1%)	3 (27.3%)	3 (27.3%)	4 (36.3%)
Spindle carcinoma	5	1 (20%)	2 (40%)	1 (20%)	1 (20%)
Simple carcinoma	2	–	1 (50%)	–	1 (50%)
Benign lesions	11	3 (27.3%)	5 (45.4%)	2 (18.2%)	1 (9.1%)
Complex adenoma	4	–	2 (50%)	1 (25%)	1 (25%)
Simple adenoma	3	1 (33.3%)	1 (33.3%)	1 (33.3%)	–
Benign mixed tumor	2	1 (50%)	1 (50%)	–	–
Hyperplasia	2	1 (50%)	1 (50%)	–	–

*0: no positively stained tumor cells; [†]5 ~ 25%; [‡]26 ~ 50%; [§]greater than 51% positive tumor cells.

thickness of 4 μ m) and stained with haematoxylin and eosin. Additional sections were placed on slides coated with 3-aminopropyltriethoxysilane (Sigma, USA) and stained using a streptavidin-biotin-peroxidase complex technique (Histostain Plus kit; Zymed, USA) with a monoclonal anti- α B-c antibody (1/1,000 dilution, SPA-222; Stressgen, Canada). Incubation with amino ethyl carbazole (AEC substrate kit; Invitrogen, USA) or 3,3'-diaminobenzidine (DAB chromogen/substrate kit; Scytek, USA) as a chromogen in H₂O₂ was performed for 10 min. The sections were counterstained with Mayer's hematoxylin for 1 min, rinsed with tap water, and mounted with an aqueous mounting medium (Vision Mount; Lab Vision, USA).

The percentages of the total area of the immunohistochemically positive cells were assessed with a microscopy image analysis system (Bs200P; BAB Software, Turkey). A total of 10 high-power fields were randomly chosen and analyzed at \times 400. The findings were categorized as follows: (0) no positively stained tumor cells; (1) 5 ~ 25%; (2) 26 ~ 50%; and (3) greater than 51% positive tumor cells. Statistical analysis was carried out using a logistic regression analysis. The results were considered statistically significant if $p < 0.05$.

Results

The immunohistochemical results of α B-c expression and histopathologic diagnoses are summarized in Table 1. In the control mammary tissues, a few luminal epithelial cells were positive but the basal myoepithelial cells were negative (Fig. 1A). Muscles (*e.g.*, M. supramammarius) were always positive and were used as positive internal controls when present.

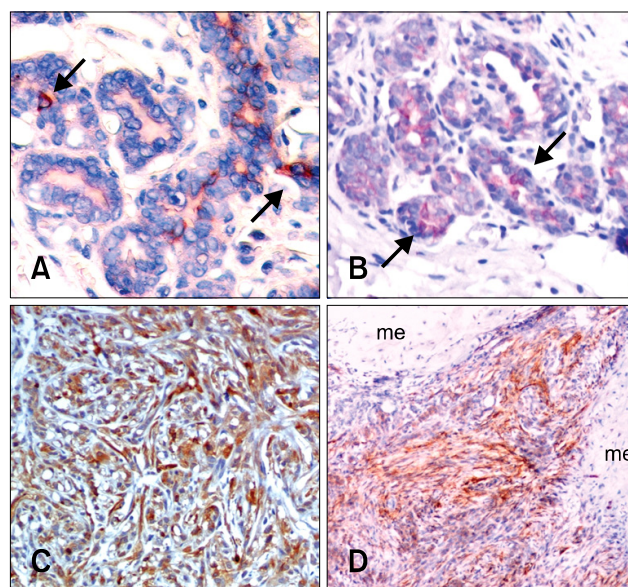


Fig. 1. (A) Some alpha basic crystallin (α B-c)-positive immunoreaction observed at the luminal epithelial cells in the control mammary tissues (arrows); myoepithelial cells were negative. (B) α B-c-immunopositive reaction of the luminal epithelial cells of the simple adenoma (arrows). (C) In the complex adenocarcinoma, positive staining was predominantly found in the cytoplasm of neoplastic epithelial cells and, to a lesser extent, in the neoplastic myoepithelial cells. (D) In carcinosarcoma, no immunopositive reaction was observed in the mesenchymal elements (me), but positive staining was seen in the neoplastic epithelial cells. Immunohistochemical staining was performed with streptavidin- biotin-peroxidase and amino ethyl carbazole chromogen. A: \times 220, B and C: \times 140, D: \times 90.

In benign or malignant simple type tumors, α B-c expression was observed in the luminal epithelial cells (Fig. 1B) while the myoepithelial cells were negative. Addi-

tionally, α B-c immunoreactivity of the epithelial cells in malignant tumors was more intense than that observed in the benign type. In both benign and malignant complex type tumors, positive staining was noted predominantly in the cytoplasm of the neoplastic epithelial cells (Fig. 1C). α B-c immunoreactivity was also found in neoplastic myoepithelial cells. However, α B-c-positive mesenchymal elements of the carcinosarcoma, such as cartilage or bone, were not observed (Fig. 1D).

The numbers of α B-c positive cells were statistically significant different between the control canine mammary tissues and benign tumors ($p < 0.05$). Statistically, significant differences of α B-c immunopositive cells were also found in between benign and malignant tumors ($p < 0.05$). Additionally, the most significant difference of immunopositive cell count was observed between normal mammary gland tissues and malignant tumors ($p < 0.001$).

Discussion

The results of the current study indicates that the number of α B-c-positive cells were increased in benign and malignant canine mammary tumors when compared to control mammary gland, and the number of α B-c-positive cells were higher in malignant tumors when compared with benign tumors.

α B-c is a member of the mammalian small HSP superfamily [8]. This factor is expressed in many tissues and organs, and acts as a molecular chaperon. It is upregulated by physiologic stress and neurodegenerative diseases [1,5,8,16]. Additionally, α B-c is expressed in breast and the other cancers, such as gliomas, and prostate and renal cell carcinomas, in humans [3,4,14,19]. However, the best of our knowledge, there is no such study performed in animals and the present report documents a connection between α B-c and mammary tumors in canines.

In the normal human breast tissue, α B-c is predominantly expressed in myoepithelial cell [14]. In contrast to this report, in the current study, very low level expression of α B-c was observed in the luminal epithelial cells of normal canine mammary tissues, but α B-c immunopositive reactivity in the myoepithelial cells was not observed. It is possible that this difference might be species-specific or related to the physiological stage of the control mammary gland tissues. Sitterding *et al.* [17] reported that α B-c may be used as a myoepithelial marker, but it does not have any special advantage over currently used markers such as smooth muscle myosine. In contrast, α B-c immune reactivity in the myoepithelial cells was not observed in our study. Therefore, α B-c immunostaining in the canine mammary gland tissues could not be suggested as a myoepithelial marker.

Chelouche-Lev *et al.* [3] have reported that out of 672 human breast tumors, 608 (90%) are positive for α B-c

immunostaining. In contrast, Moyano *et al.* [14] noted 39 positively immunostained samples out of 361 (11%) human breast cancer cases. Sitterding *et al.* [17] reported that α B-c expression is observed in human basal-like (81%) and metaplastic (86%) breast cancer. In the present study, the rate of α B-c expression in canine mammary gland tumors was 88.2% (45 out of 51). Although it is difficult to compare canines to humans, our data are similar to the findings of Chelouche-Lev *et al.* [3] and Sitterding *et al.* [17], but different from those of Moyano *et al.* [14].

Differences in the number of α B-c-immunolabelled cells were found to be statistically significant between the normal canine mammary tissues and the benign and malignant tumors. These differences were thought to be evidence that α B-c was involved in oncogenesis because of the anti-apoptotic effect of α B-c. This effect probably involves the suppression of caspase-3 activation and/or preventing the mitochondrial translocation of pro-apoptotic Bcl-2 family members such as Bax [10].

In conclusion, we believe that α B-c can play a role in the carcinogenesis of canine mammary glands because of the increased immunoreactivity of α B-c we observed in neoplastic tissues compared to the normal canine mammary gland tissues. α B-c has an anti-apoptotic effect and this anti-apoptotic effect can be important in chemotherapy resistance of the cancer. Additionally, targeted inhibition of α B-c may be a new strategy to the cancer therapy.

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References

1. **Bajramović JJ, Lassmann H, van Noort JM.** Expression of α B-crystallin in glia cells during lesional development in multiple sclerosis. *J Neuroimmunol* 1997, **78**, 143-151.
2. **Calderwood SK, Khaleque MA, Sawyer DB, Ciocca DR.** Heat shock proteins in cancer: chaperones of tumorigenesis. *Trends Biochem Sci* 2006, **31**, 164-172.
3. **Chelouche-Lev D, Kluger HM, Berger AJ, Rimm DL, Price JE.** α B-crystallin as a marker of lymph node involvement in breast carcinoma. *Cancer* 2004, **100**, 2543-2548.
4. **Chin D, Boyle GM, Williams RM, Ferguson K, Pandeya N, Pedley J, Campbell CM, Theile DR, Parsons PG, Coman WB.** Alpha B-crystallin, a new independent marker for poor prognosis in head and neck cancer. *Laryngoscope* 2005, **115**, 1239-1242.
5. **Dabir DV, Trojanowski JQ, Richter-Landsberg C, Lee VM, Forman MS.** Expression of the small heat-shock protein α B-crystallin in tauopathies with glial pathology.

- Am J Pathol 2004, **164**, 155-166.
6. **Goldbaum O, Richter-Landsberg C.** Stress proteins in oligodendrocytes: differential effects of heat shock and oxidative stress. *J Neurochem* 2001, **78**, 1233-1242.
 7. **Graham JC, Myers RK.** The prognostic significance of angiogenesis in canine mammary tumors. *J Vet Intern Med* 1999, **13**, 416-418.
 8. **Head MW, Goldman JE.** Small heat shock proteins, the cytoskeleton, and inclusion body formation. *Neuropathol Appl Neurobiol* 2000, **26**, 304-312.
 9. **Iwaki T, Wisniewski T, Iwaki A, Corbin E, Tomokane N, Tateishi J, Goldman JE.** Accumulation of alpha B-crystallin in central nervous system glia and neurons in pathologic conditions. *Am J Pathol* 1992, **140**, 345-356.
 10. **Kamradt MC, Lu M, Werner ME, Kwan T, Chen F, Strohecker A, Oshita S, Wilkinson JC, Yu C, Oliver PG, Duckett CS, Buchsbaum DJ, LoBuglio AF, Jordan VC, Cryns VL.** The small heat shock protein α B-crystallin is a novel inhibitor of TRAIL-induced apoptosis that suppresses the activation of caspase-3. *J Biol Chem* 2005, **280**, 11059-11066.
 11. **Konishi H, Matsuzaki H, Tanaka M, Takemura Y, Kuroda S, Ono Y, Kikkawa U.** Activation of protein kinase B (Akt/RAC-protein kinase) by cellular stress and its association with heat shock protein Hsp27. *FEBS Lett* 1997, **410**, 493-498.
 12. **Kumaraguruparan R, Karunagaran D, Balachandran C, Manohar BM, Nagini S.** Of humans and canines: a comparative evaluation of heat shock and apoptosis-associated proteins in mammary tumors. *Clin Chim Acta* 2006, **365**, 168-176.
 13. **Misdorp W, Else RW, Hellmén E, Lipscomb TP.** *Histological Classification of Mammary Tumors of the Dog and Cat.* 2nd ed. pp.1-59, Armed Forces Institute of Pathology, Washington, 1999.
 14. **Moyano JV, Evans JR, Chen F, Lu M, Werner ME, Yehiely F, Diaz LK, Turbin D, Karaca G, Wiley E, Nielsen TO, Perou CM, Cryns VL.** α B-crystallin is a novel oncoprotein that predicts poor clinical outcome in breast cancer. *J Clin Invest* 2006, **116**, 261-270.
 15. **Munson L, Moresco A.** Comparative pathology of mammary gland cancers in domestic and wild animals. *Breast Dis* 2007, **28**, 7-21.
 16. **Richter-Landsberg C, Goldbaum O.** Stress proteins in neural cells: functional roles in health and disease. *Cell Mol Life Sci* 2003, **60**, 337-349.
 17. **Sitterding SM, Wiseman WR, Schiller CL, Luan C, Chen F, Moyano JV, Watkin WG, Wiley EL, Cryns VL, Diaz LK.** α B-crystallin: A novel marker of invasive basal-like and metaplastic breast carcinomas. *Ann Diagn Pathol* 2008, **12**, 33-40.
 18. **Sorenmo K.** Canine mammary gland tumors. *Vet Clin North Am Small Anim Pract* 2003, **33**, 573-596.
 19. **Takashi M, Katsuno S, Sakata T, Ohshima S, Kato K.** Different concentrations of two small stress proteins, α B crystallin and HSP27 in human urological tumor tissues. *Urol Res* 1998, **26**, 395-399.