REVIEW ARTICLE



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Giardia duodenalis genetic assemblages and hosts

Martin F. Heyworth^{1,2,*}

¹ Research Service (151), Corporal Michael J. Crescenz Veterans Affairs (VA) Medical Center, University and Woodland Avenues, Philadelphia, PA 19104, USA

² Department of Medicine, Perelman School of Medicine, University of Pennsylvania, Philadelphia, PA 19104, USA

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Abstract – Techniques for sub-classifying morphologically identical *Giardia duodenalis* trophozoites have included comparisons of the electrophoretic mobility of enzymes and of chromosomes, and sequencing of genes encoding β -giardin, triose phosphate isomerase, the small subunit of ribosomal RNA and glutamate dehydrogenase. To date, *G. duodenalis* organisms have been sub-classified into eight genetic assemblages (designated A–H). Genotyping of *G. duodenalis* organisms isolated from various hosts has shown that assemblages A and B infect the largest range of host species, and appear to be the main (or possibly only) *G. duodenalis* assemblages that undeniably infect human subjects. In at least some cases of assemblage A or B infection in wild mammals, there is suggestive evidence that the infection had resulted from environmental contamination by *G. duodenalis* cysts of human origin.

Key words: Assemblage, Genotype, Giardia, Giardia infections, Giardiasis.

Résumé – Assemblages génétiques et hôtes de *Giardia duodenalis*. Les techniques pour sous-classer morphologiquement des trophozoïtes identiques de *Giardia duodenalis* ont inclus des comparaisons de la mobilité électrophorétique des enzymes et des chromosomes et le séquençage des gènes codant pour la β -giardine, la triose-phosphate isomérase, la petite sous-unité ribosomique de l'ARN et la glutamate déshydrogénase. À ce jour, *G. duodenalis* a été sous-classé en 8 assemblages génétiques (désignés par A-H). Le génotypage de *G. duodenalis* isolés à partir de divers hôtes a montré que les assemblages A et B infectent le grand plus grand nombre d'espèces d'hôtes, et semblent être les assemblages principaux (ou peut-être uniques) qui infectent les sujets humains de manière indéniable. Dans au moins certains cas d'infection chez les mammifères sauvages par les assemblages A ou B, des éléments indiquent que l'infection était due à la contamination de l'environnement par des kystes de *G. duodenalis* d'origine humaine.

Morphologically similar or identical *Giardia* organisms, designated *Giardia duodenalis* (synonyms *G. intestinalis* and *G. lamblia*) [54], can infect the intestine of numerous species of mammalian host. *G. duodenalis* is the only *Giardia* species that causes human infection; other currently recognised species in this genus include the following (hosts are mentioned in parentheses): *Giardia muris* (rodents) [15], *G. microti* (voles, muskrats) [57], *G. psittaci* (budgerigars) [11], *G. ardeae* (great blue herons) [12] and *G. agilis* (amphibians) [14].

From the 1980s onwards, increasingly precise methods have been developed to sub-classify morphologically identical *G. duodenalis* organisms. Early work of this type involved examination of the electrophoretic mobility of *G. duodenalis* enzymes [1, 23]. In the mid-1990s, such work delineated two

distinct sub-populations of *G. duodenalis*, designated assemblages A and B [32]. Additional evidence for heterogeneity of *G. duodenalis* emerged from study of the electrophoretic mobility of *Giardia* chromosomes [46]. Polymerase Chain Reaction (PCR) amplification of *G. duodenalis* DNA, and restriction fragment length polymorphism (RFLP) analysis and sequencing of the resulting PCR products, added further insight into the heterogeneity of the organism, confirming the existence of assemblages A and B, and – in conjunction with data from enzyme electrophoresis – delineating six additional assemblages (C–H) [5, 26, 27, 34–37].

Giardia duodenalis genes (genetic loci) used for genotyping the organisms include genes encoding β -giardin (bg), triose phosphate isomerase (tpi), the small subunit of ribosomal RNA (ssu) and glutamate dehydrogenase (gdh) [15]. Giardia duodenalis assemblages have been shown to be either relatively

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^{*}Corresponding author: martin.heyworth@va.gov

 Table 1. Giardia duodenalis assemblages and corresponding hosts.

Assemblage	e Host(s)
A	Humans [15], dog [5, 8, 29, 58], cat [27, 51], cattle [15, 24], alpaca [19], deer [45, 48], ferret [39], pig [3], beaver [40], chinchilla [39], jaguar [47], horse [56], marsupials [38, 52], sheep [17, 27, 30, 62], goat [17], muskox [9], non-human primates [60, 61], cetacean(s) [41, 42], seals [26], Australian sea lion [10], moose [27, 43], reindeer [43], chicken [5], gull [26]
В	Humans [5, 15], cattle [15, 38], dog [8], gazelle [48], deer [38], horse [56], beaver [40, 50], muskrat [50], chinchilla [39, 47], ferret [39], rabbit [30, 39], Desmarest's hutia [39], marsupials [38, 52], guinea pig [27], rock hyrax [4], non-human primates [47, 60, 61], chicken [5], sheep [62], seals [26], pig [13], Australian sea lion [10], ostrich [47], dolphin [25, 41], porpoise [25], gull [26]
С	Dog [8, 29], kangaroo [38], cattle [33], pig [33] cetacean(s) [42]
D	Dog [5, 8], chinchilla [39], kangaroo [38], cattle [33], cetacean(s) [42], fox [38]
Е	Cattle [24, 30, 38], sheep [15, 17, 27, 30], pig [3, 13, 15], alpaca [19], goat [17, 62], horse [56], yak [27], fox [38], deer [27], cat [27]
F	Cat [27, 51], cetacean(s) [42], pig [3, 33]
G	Rat [27, 63], mouse [63]
Н	Grey seal [26], gull [26]

specific to certain hosts (assemblages C–H) or essentially unrestricted in terms of the species of host that they can infect (assemblages A and B; Table 1). Within a single "isolate" of *G. duodenalis*, different genetic loci may have DNA sequences typical of different assemblages (e.g., ssu typical of assemblage B, and tpi and bg typical of assemblage A) [39], a situation that may make it unrealistic to try to assign a given isolate of *G. duodenalis* exclusively to one or other assemblage. This point is pertinent to Table 1, which may present an oversimplified classification, in not discriminating between data obtained from a single genetic locus and from several loci [7]. A comprehensive review, published in 2011, includes detailed information about assemblages of *G. duodenalis*, and non-human hosts for the respective assemblages [15].

Unambiguous direct evidence that human giardiasis can be an example of a zoonosis, i.e. a human infection acquired from non-human hosts under "natural" conditions (via ingestion of G. duodenalis cysts excreted by animals), is limited. One study from the United Kingdom suggested that contact with farm animals (especially pigs) and with pets (especially dogs and cats) was a risk factor for giardiasis in human subjects [59]. Suggestive evidence that G. duodenalis can be transmitted between dogs and human subjects was obtained from a study in a tea-growing community in northeast India [55]. In this work, an association was found between the presence of G. duodenalis infection in human subjects and in dog(s) occupying the same household. For one such household, genetic identity between G. duodenalis in a dog and in human subject(s) was reported [55]. In this example, the direction of presumed inter-species transmission of G. duodenalis might have been either, or both, dog-to-human or human-to-dog. One caveat that applies to genetic studies of G. duodenalis that rely on faecal cysts as the starting material for molecular analysis is whether the presence of such cysts necessarily reflects infection, rather than resulting merely from coprophagy of faecal material containing cysts, and passage of these cysts through an animal's gastrointestinal tract without causing infection [22].

Dogs have been infected with *G. duodenalis* of human origin, by oral administration of trophozoites or cysts of this

organism [44]. There is an anecdotal report of an investigator developing giardiasis as a result of deliberately ingesting a gel capsule containing *Giardia* trophozoites that had originated from an animal host (a Gambian giant pouched rat) [31]. This work showed that animal-to-human transmission of *Giardia* infection can occur under experimental conditions. It is, however, unclear whether the result of the experiment just described constitutes evidence for the zoonotic transmissibility of *G. duodenalis*, under "normal field conditions".

Genotyping of *G. duodenalis* organisms obtained from human subjects with *Giardia* infection has shown that assemblages A and B appear to be the only ones that undeniably cause human infections [15], although there have been occasional reports of the isolation, from human subjects, of *G. duodenalis* organisms that have genetic markers characteristic of non-A, non-B, assemblages [6, 49]. Mixed infections of human and non-human hosts with more than one assemblage of *G. duodenalis* concurrently have been described [15, 16, 18, 28]. Table 1 of the present article does not identify which infections, among those documented in the references cited, were part of a mixed infection resulting from more than one assemblage of *G. duodenalis*.

Individual *G. duodenalis* organisms can show sequence differences between different copies of the same gene (allelic sequence heterozygosity) [2, 7].

Although much of the literature on inter-species transmission of *G. duodenalis* has focussed on actual or presumed animal-to-human transmission, there is increasing evidence that *Giardia* cysts of human origin can contaminate the environment and infect wild mammals (which, in turn, may act as a reservoir for future infection of human subjects) [53]. The ability to identify *G. duodenalis* genetic assemblages has provided a level of precision and specificity that was lacking when essentially the only tool was morphological examination of trophozoites. For example, excretion of assemblage B cysts by Australian sea lions, and relative proximity of colonies of these animals to human settlements at coastal sites, speaks to the probability of initial infection of the animals by cysts of human origin [10]. Similarly, presence of assemblage A and B *G. duodenalis* infection in freely ranging gorillas may reflect human-to-gorilla transmission of the parasite [20, 21].

Of two outbreaks of human G. duodenalis infection in Canadian communities during the 1990s, one ceased and the other diminished after, in each case, a beaver excreting G. duodenalis cysts was removed from the water source supplying the respective community [40]. These observations suggested that the human cases of giardiasis had resulted from ingesting G. duodenalis cysts excreted by the beavers. The respective studies predated current knowledge of G. duodenalis assemblages. Archival material (Giardia organisms) from these outbreaks was, however, available for study by modern molecular techniques some two decades later [40]. Using such techniques, it was found that, in one of the outbreaks, the beaver was infected with assemblage A G. duodenalis, whereas the water contained G. duodenalis of assemblage B, and assemblages A and B were isolated from the infected human subjects. In the other outbreak, the beaver was found to be infected with assemblage B, whereas the infected human subjects included one with assemblage A infection [40]. Consequently, a straightforward causal relationship between the beavers and all the human cases was not found. Anthropocentric historical assumptions, that a relationship between Giardia infection in beavers and in human subjects merely involves beaver-to-human transmission of the parasite, have yielded to a more nuanced appreciation of environmental contamination by human-derived G. duodenalis cysts that may infect beavers [53].

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References

- Andrews RH, Adams M, Boreham PFL, Mayrhofer G, Meloni BP. 1989. *Giardia intestinalis*: electrophoretic evidence for a species complex. International Journal for Parasitology, 19, 183–190.
- Ankarklev J, Svärd SG, Lebbad M. 2012. Allelic sequence heterozygosity in single *Giardia* parasites. BMC Microbiology, 12, 65.
- Armson A, Yang R, Thompson J, Johnson J, Reid S, Ryan UM. 2009. *Giardia* genotypes in pigs in Western Australia: prevalence and association with diarrhea. Experimental Parasitology, 121, 381–383.
- Beck R, Sprong H, Bata I, Lucinger S, Pozio E, Cacciò SM. 2011. Prevalence and molecular typing of *Giardia* spp. in captive mammals at the zoo of Zagreb, Croatia. Veterinary Parasitology, 175, 40–46.
- 5. Berrilli F, D'Alfonso R, Giangaspero A, Marangi M, Brandonisio O, Kaboré Y, Glé C, Cianfanelli C, Lauro R, Di Cave D. 2012. *Giardia duodenalis* genotypes and *Cryptosporidium* species in humans and domestic animals in Côte d'Ivoire: occurrence and evidence for environmental contamination. Transactions of the Royal Society of Tropical Medicine and Hygiene, 106, 191–195.
- Broglia A, Weitzel T, Harms G, Cacciò SM, Nöckler K. 2013. Molecular typing of *Giardia duodenalis* isolates from German travellers. Parasitology Research, 112, 3449–3456.

- Cacciò SM, Ryan U. 2008. Molecular epidemiology of giardiasis. Molecular & Biochemical Parasitology, 160, 75–80.
- Covacin C, Aucoin DP, Elliot A, Thompson RCA. 2011. Genotypic characterisation of *Giardia* from domestic dogs in the USA. Veterinary Parasitology, 177, 28–32.
- Davidson RK, Amundsen H, Lie NO, Luyckx K, Robertson LJ, Verocai GG, Kutz SJ, Ytrehus B. 2014. Sentinels in a climatic outpost: endoparasites in the introduced muskox (*Ovibos moschatus wardi*) population of Dovrefjell, Norway. International Journal for Parasitology: Parasites and Wildlife, 3, 154–160.
- Delport TC, Asher AJ, Beaumont LJ, Webster KN, Harcourt RG, Power ML. 2014. *Giardia duodenalis* and *Cryptosporidium* occurrence in Australian sea lions (*Neophoca cinerea*) exposed to varied levels of human interaction. International Journal for Parasitology: Parasites and Wildlife, 3, 269–275.
- 11. Erlandsen SL, Bemrick WJ. 1987. SEM evidence for a new species, *Giardia psittaci*. Journal of Parasitology, 73, 623–629.
- Erlandsen SL, Bemrick WJ, Wells CL, Feely DE, Knudson L, Campbell SR, van Keulen H, Jarroll EL. 1990. Axenic culture and characterization of *Giardia ardeae* from the great blue heron (*Ardea herodias*). Journal of Parasitology, 76, 717–724.
- 13. Farzan A, Parrington L, Coklin T, Cook A, Pintar K, Pollari F, Friendship R, Farber J, Dixon B. 2011. Detection and characterization of *Giardia duodenalis* and *Cryptosporidium* spp. on swine farms in Ontario, Canada. Foodborne Pathogens and Disease, 8, 1207–1213.
- 14. Feely DE, Erlandsen SL. 1985. Morphology of *Giardia agilis*: observation by scanning electron microscopy and interference reflexion microscopy. Journal of Protozoology, 32, 691–693.
- Feng Y, Xiao L. 2011. Zoonotic potential and molecular epidemiology of *Giardia* species and giardiasis. Clinical Microbiology Reviews, 24, 110–140.
- Gelanew T, Lalle M, Hailu A, Pozio E, Cacciò SM. 2007. Molecular characterization of human isolates of *Giardia duodenalis* from Ethiopia. Acta Tropica, 102, 92–99.
- Geurden T, Thomas P, Casaert S, Vercruysse J, Claerebout E. 2008. Prevalence and molecular characterisation of *Cryptosporidium* and *Giardia* in lambs and goat kids in Belgium. Veterinary Parasitology, 155, 142–145.
- Geurden T, Levecke B, Cacciò SM, Visser A, de Groote G, Casaert S, Vercruysse J, Claerebout E. 2009. Multilocus genotyping of *Cryptosporidium* and *Giardia* in non-outbreak related cases of diarrhoea in human patients in Belgium. Parasitology, 136, 1161–1168.
- Gomez-Puerta LA, Lopez-Urbina MT, Alarcon V, Cama V, Gonzalez AE, Xiao L. 2014. Occurrence of *Giardia duodenalis* assemblages in alpacas in the Andean region. Parasitology International, 63, 31–34.
- Graczyk TK, Bosco-Nizeyi J, Ssebide B, Thompson RCA, Read C, Cranfield MR. 2002. Anthropozoonotic *Giardia duodenalis* genotype (assemblage) A infections in habitats of free-ranging human-habituated gorillas, Uganda. Journal of Parasitology, 88, 905–909.
- Hogan JN, Miller WA, Cranfield MR, Ramer J, Hassell J, Noheri JB, Conrad PA, Gilardi KVK. 2014. *Giardia* in mountain gorillas (*Gorilla beringei beringei*), forest buffalo (*Syncerus caffer*), and domestic cattle in Volcanoes National Park, Rwanda. Journal of Wildlife Diseases, 50, 21–30.
- 22. Inpankaew T, Schär F, Odermatt P, Dalsgaard A, Chimnoi W, Khieu V, Muth S, Traub RJ. 2014. Low risk for transmission of zoonotic *Giardia duodenalis* from dogs to humans in rural Cambodia. Parasites & Vectors, 7, 412.

- Isaac-Renton JL, Cordeiro C, Sarafis K, Shahriari H. 1993. Characterization of *Giardia duodenalis* isolates from a waterborne outbreak. Journal of Infectious Diseases, 167, 431–440.
- 24. Khan SM, Debnath C, Pramanik AK, Xiao L, Nozaki T, Ganguly S. 2011. Molecular evidence for zoonotic transmission of *Giardia duodenalis* among dairy farm workers in West Bengal, India. Veterinary Parasitology, 178, 342–345.
- Lasek-Nesselquist E, Bogomolni AL, Gast RJ, Welch DM, Ellis JC, Sogin ML, Moore MJ. 2008. Molecular characterization of *Giardia intestinalis* haplotypes in marine animals: variation and zoonotic potential. Diseases of Aquatic Organisms, 81, 39–51.
- 26. Lasek-Nesselquist E, Welch DM, Sogin ML. 2010. The identification of a new *Giardia duodenalis* assemblage in marine vertebrates and a preliminary analysis of *G. duodenalis* population biology in marine systems. International Journal for Parasitology, 40, 1063–1074.
- Lebbad M, Mattsson JG, Christensson B, Ljungström B, Backhans A, Andersson JO, Svärd SG. 2010. From mouse to moose: multilocus genotyping of *Giardia* isolates from various animal species. Veterinary Parasitology, 168, 231–239.
- Levecke B, Geldhof P, Claerebout E, Dorny P, Vercammen F, Cacciò SM, Vercruysse J, Geurden T. 2009. Molecular characterisation of *Giardia duodenalis* in captive non-human primates reveals mixed assemblage A and B infections and novel polymorphisms. International Journal for Parasitology, 39, 1595–1601.
- Li W, Liu C, Yu Y, Li J, Gong P, Song M, Xiao L, Zhang X. 2013. Molecular characterization of *Giardia duodenalis* isolates from police and farm dogs in China. Experimental Parasitology, 135, 223–226.
- 30. Liu A, Yang F, Shen Y, Zhang W, Wang R, Zhao W, Zhang L, Ling H, Cao J. 2014. Genetic analysis of the *Gdh* and *Bg* genes of animal-derived *Giardia duodenalis* isolates in Northeastern China and evaluation of zoonotic transmission potential. PLoS One, 9(4), e95291.
- Majewska AC. 1994. Successful experimental infections of a human volunteer and Mongolian gerbils with *Giardia* of animal origin. Transactions of the Royal Society of Tropical Medicine and Hygiene, 88, 360–362.
- 32. Mayrhofer G, Andrews RH, Ey PL, Chilton NB. 1995. Division of *Giardia* isolates from humans into two genetically distinct assemblages by electrophoretic analysis of enzymes encoded at 27 loci and comparison with *Giardia muris*. Parasitology, 111, 11–17.
- 33. Minetti C, Taweenan W, Hogg R, Featherstone C, Randle N, Latham SM, Wastling JM. 2014. Occurrence and diversity of *Giardia duodenalis* assemblages in livestock in the UK. Transboundary and Emerging Diseases, 61, e60–e67.
- Monis PT, Mayrhofer G, Andrews RH, Homan WL, Limper L, Ey PL. 1996. Molecular genetic analysis of *Giardia intestinalis* isolates at the glutamate dehydrogenase locus. Parasitology, 112, 1–12.
- 35. Monis PT, Andrews RH, Mayrhofer G, Mackrill J, Kulda J, Isaac-Renton JL, Ey PL. 1998. Novel lineages of *Giardia intestinalis* identified by genetic analysis of organisms isolated from dogs in Australia. Parasitology, 116, 7–19.
- Monis PT, Andrews RH, Mayrhofer G, Ey PL. 1999. Molecular systematics of the parasitic protozoan *Giardia intestinalis*. Molecular Biology and Evolution, 16, 1135–1144.
- 37. Monis PT, Andrews RH, Mayrhofer G, Ey PL. 2003. Genetic diversity within the morphological species *Giardia intestinalis*

and its relationship to host origin. Infection, Genetics and Evolution, 3, 29-38.

- Ng J, Yang R, Whiffin V, Cox P, Ryan U. 2011. Identification of zoonotic *Cryptosporidium* and *Giardia* genotypes infecting animals in Sydney's water catchments. Experimental Parasitology, 128, 138–144.
- 39. Pantchev N, Broglia A, Paoletti B, Globokar Vrhovec M, Bertram A, Nöckler K, Cacciò SM. 2014. Occurrence and molecular typing of *Giardia* isolates in pet rabbits, chinchillas, guinea pigs and ferrets collected in Europe during 2006–2012. Veterinary Record, 175, 18.
- 40. Prystajecky N, Tsui CK-M, Hsiao WWL, Uyaguari-Diaz MI, Ho J, Tang P, Isaac-Renton J. 2015. *Giardia* spp. are commonly found in mixed assemblages in surface water, as revealed by molecular and whole-genome characterization. Applied and Environmental Microbiology, 81, 4827–4834.
- Reboredo-Fernández A, Gómez-Couso H, Martínez-Cedeira JA, Cacciò SM, Ares-Mazás E. 2014. Detection and molecular characterization of *Giardia* and *Cryptosporidium* in common dolphins (*Delphinus delphis*) stranded along the Galician coast (Northwest Spain). Veterinary Parasitology, 202, 132–137.
- 42. Reboredo-Fernández A, Ares-Mazás E, Martínez-Cedeira JA, Romero-Suances R, Cacciò SM, Gómez-Couso H. 2015. *Giardia* and *Cryptosporidium* in cetaceans on the European Atlantic coast. Parasitology Research, 114, 693–698.
- 43. Robertson LJ, Forberg T, Hermansen L, Hamnes IS, Gjerde B. 2007. *Giardia duodenalis* cysts isolated from wild moose and reindeer in Norway: genetic characterization by PCR-RFLP and sequence analysis at two genes. Journal of Wildlife Diseases, 43, 576–585.
- 44. Rosa LAG, Gomes MA, Mundim AV, Mundim MJS, Pozzer EL, Faria ESM, Viana JC, Cury MC. 2007. Infection of dogs by experimental inoculation with human isolates of *Giardia duodenalis*: clinical and laboratory manifestations. Veterinary Parasitology, 145, 37–44.
- Santín M, Fayer R. 2015. Enterocytozoon bieneusi, Giardia, and Cryptosporidium infecting white-tailed deer. Journal of Eukaryotic Microbiology, 62, 34–43.
- Sarafis K, Isaac-Renton J. 1993. Pulsed-field gel electrophoresis as a method of biotyping of *Giardia duodenalis*. American Journal of Tropical Medicine and Hygiene, 48, 134–144.
- 47. Soares RM, de Souza SLP, Silveira LH, Funada MR, Richtzenhain LJ, Gennari SM. 2011. Genotyping of potentially zoonotic *Giardia duodenalis* from exotic and wild animals kept in captivity in Brazil. Veterinary Parasitology, 180, 344–348.
- Solarczyk P, Majewska AC, Słodkowicz-Kowalska A. 2014. Axenic *in vitro* culture and molecular characterization of *Giardia duodenalis* from red deer (*Cervus elaphus*) and Thomson's gazelle (*Gazella thomsonii*). Acta Parasitologica, 59, 763–766.
- 49. Soliman RH, Fuentes I, Rubio JM. 2011. Identification of a novel Assemblage B subgenotype and a zoonotic Assemblage C in human isolates of *Giardia intestinalis* in Egypt. Parasitology International, 60, 507–511.
- Sulaiman IM, Fayer R, Bern C, Gilman RH, Trout JM, Schantz PM, Das P, Lal AA, Xiao L. 2003. Triosephosphate isomerase gene characterization and potential zoonotic transmission of *Giardia duodenalis*. Emerging Infectious Diseases, 9, 1444–1452.
- 51. Suzuki J, Murata R, Kobayashi S, Sadamasu K, Kai A, Takeuchi T. 2011. Risk of human infection with *Giardia duodenalis* from cats in Japan and genotyping of the isolates to

assess the route of infection in cats. Parasitology, 138, 493-500.

- Thompson J, Yang R, Power M, Hufschmid J, Beveridge I, Reid S, Ng J, Armson A, Ryan U. 2008. Identification of zoonotic *Giardia* genotypes in marsupials in Australia. Experimental Parasitology, 120, 88–93.
- Thompson RCA, Monis P. 2012. *Giardia* from genome to proteome. Advances in Parasitology, 78, 57–95.
- 54. Thompson RCA, Monis PT. 2004. Variation in *Giardia*: implications for taxonomy and epidemiology. Advances in Parasitology, 58, 69–137.
- 55. Traub RJ, Monis PT, Robertson I, Irwin P, Mencke N, Thompson RCA. 2004. Epidemiological and molecular evidence supports the zoonotic transmission of *Giardia* among humans and dogs living in the same community. Parasitology, 128, 253–262.
- 56. Traversa D, Otranto D, Milillo P, Latrofa MS, Giangaspero A, Di Cesare A, Paoletti B. 2012. *Giardia duodenalis* subassemblage of animal and human origin in horses. Infection, Genetics and Evolution, 12, 1642–1646.
- 57. van Keulen H, Feely DE, Macechko PT, Jarroll EL, Erlandsen SL. 1998. The sequence of *Giardia* small subunit rRNA shows that voles and muskrats are parasitized by a unique species *Giardia microti*. Journal of Parasitology, 84, 294–300.
- 58. Volotão ACC, Ramos NMD, Fantinatti M, de Moraes MVP, Netto HA, Storti-Melo LM, de Godoy EAM, Rossit ARB,

Fernandes O, Machado RLD. 2011. Giardiasis as zoonosis: between proof of principle and paradigm in the Northwestern region of São Paulo State, Brazil. Brazilian Journal of Infectious Diseases, 15, 382–383.

- Warburton ARE, Jones PH, Bruce J. 1994. Zoonotic transmission of giardiasis: a case control study. Communicable Disease Report. CDR Review, 4, R32–R36.
- Ye J, Xiao L, Ma J, Guo M, Liu L, Feng Y. 2012. Anthroponotic enteric parasites in monkeys in public park, China. Emerging Infectious Diseases, 18, 1640–1643.
- Ye J, Xiao L, Li J, Huang W, Amer SE, Guo Y, Roellig D, Feng Y. 2014. Occurrence of human-pathogenic *Enterocytozoon bieneusi*, *Giardia duodenalis* and *Cryptosporidium* genotypes in laboratory macaques in Guangxi, China. Parasitology International, 63, 132–137.
- 62. Zhang W, Zhang X, Wang R, Liu A, Shen Y, Ling H, Cao J, Yang F, Zhang X, Zhang L. 2012. Genetic characterizations of *Giardia duodenalis* in sheep and goats in Heilongjiang Province, China and possibility of zoonotic transmission. PLoS Neglected Tropical Diseases, 6(9), e1826.
- Zhao Z, Wang R, Zhao W, Qi M, Zhao J, Zhang L, Li J, Liu A. 2015. Genotyping and subtyping of *Giardia* and *Cryptosporidium* isolates from commensal rodents in China. Parasitology, 142, 800–806.

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