

Surface expression of NMDA receptor changes during memory consolidation in the crab *Neohelice granulata*

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The aim of the present study was to analyze the surface expression of the NMDA-like receptors during the consolidation of contextual learning in the crab *Neohelice granulata*. Memory storage is based on alterations in the strength of synaptic connections between neurons. The glutamatergic synapses undergo various forms of *N*-methyl-D aspartate receptor (NMDAR)-dependent changes in strength, a process that affects the abundance of other receptors at the synapse and underlies some forms of learning and memory. Here we propose a direct regulation of the NMDAR. Changes in NMDAR's functionality might be induced by the modification of the subunit's expression or cellular trafficking. This trafficking does not only include NMDAR's movement between synaptic and extra-synaptic localizations but also the cycling between intracellular compartments and the plasma membrane, a process called surface expression. Consolidation of contextual learning affects the surface expression of the receptor without affecting its general expression. The surface expression of the GluN1 subunit of the NMDAR is down-regulated immediately after training, up-regulated 3 h after training and returns to naïve and control levels 24 h after training. The changes in NMDAR surface expression observed in the central brain are not seen in the thoracic ganglion. A similar increment in surface expression of GluN1 in the central brain is observed 3 h after administration of the competitive GABA_A receptor antagonist, bicuculline. These consolidation changes are part of a plasticity event that first, during the down-regulation, stabilizes the trace and later, at 3-h post-training, changes the threshold for synapse activation.

It is generally accepted that the storage of memory is based on alterations in the strength of synaptic connections between neurons. Synaptic plasticity is the ability to alter synaptic connections between neurons that underlies learning and memory processes. Glutamate receptors mediate the majority of excitatory neurotransmission in the vertebrate nervous system. *N*-Methyl-D aspartate receptors (NMDARs) are one of the three pharmacologically distinct subtypes of glutamatergic ionotropic receptors. They are formed by heteromeric complexes which are usually comprised of essential GluN1 subunits and distinct NR2 (A–D) or NR3 subunits (Dingledine et al. 1999; Villmann et al. 1999; Cull-Candy et al. 2001). The opening of NMDAR channels, which are permeable to potassium, sodium, and calcium, requires simultaneous glutamate binding and postsynaptic membrane depolarization (McBain and Mayer 1994; Dingledine et al. 1999). This voltage-dependent control of glutamate-induced calcium influx hints at the possibility that NMDARs serves as “Hebbian coincidence detectors” and as one of the molecular mechanisms eliciting synaptic plasticity (Tsien 2000).

Glutamatergic synapses undergo various forms of NMDAR-dependent changes in strength, a process thought to underlie some forms of learning and memory (Malenka and Nicoll 1999). One of the experimental models used to study the synaptic plasticity is based on long-term potentiation (LTP) of synaptic transmission. NMDA receptor antagonists block LTP induction and different forms of memory (Riedel et al. 2003). In addition, it

has also been demonstrated that there is a post-training increase in GluN1 subunit expression in hippocampal membranes of rats trained in an inhibitory avoidance task (Cammarota et al. 2000; Moyano et al. 2004).

LTP dependent on NMDARs has been reported in the invertebrate model *Aplysia californica* (Lin and Glanzman 1994; Roberts and Glanzman 2003). The role of NMDARs in learning and memory processes was also demonstrated in several other invertebrates (Müssig et al. 2010; Amano and Maruyama 2011; Miyashita et al. 2012; Hepp et al. 2013). In our laboratory, research performed in the crab *Neohelice granulata* (formerly *Chasmagnathus granulatus*) using context associative learning (context signal memory, CSM) has indicated that NMDA-like receptors are involved in consolidation and reconsolidation (Pedreira et al. 2002; Troncoso and Maldonado 2002). Pharmacological-behavioral experiments have shown that systemic administration of the vertebrate NMDAR antagonists MK-801 and AP5 disrupts both processes of the contextual associative memory paradigm developed in our laboratory (Pedreira et al. 2002; Troncoso and Maldonado 2002; Barreiro et al. 2013). The function of these receptors is also necessary for the acquisition of extinction memory but not for its consolidation (Pérez-Cuesta et al. 2007). In a recent study, we characterized the GluN1-like subunit, and describe the distribution of NMDA-like receptors in the crab central nervous system (CNS). We identified two immunoreactive bands for the

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GluN1-like subunit. The close proximity of the molecular weight of these bands suggests that they may represent post-translational modifications of one protein (Petralia et al. 1994; Rao and Craig 1997). These GluN1-like bands are present in all major ganglia of the CNS; in addition, the band intensities suggest anGluN1-like homogenous distribution among ganglia. At the subcellular level, the GluN1-like subunit is only detectable in synaptosomes that illustrate the synaptic membrane localization of this protein (Hepp et al. 2013).

In general, changes in the functionality of NMDARs might be induced by the modification of the subunit's expression that form the receptor, or by phosphorylation which in turn modulates its interactions with other proteins. In addition, cellular trafficking of NMDARs has also been extensively studied (Wenthold et al. 2003). This trafficking does not only include movement of NMDARs between synaptic and extra-synaptic localizations at the surface of neurons but also the cycle between intracellular compartments and the plasma membrane (Groc et al. 2009). Thus, it was demonstrated that the receptors might change their position by lateral diffusion in the plasma membrane (Tovar and Westbrook 2002) or by trafficking receptors from the intracellular vesicles to insertion into the plasma membrane, a process called surface expression (Bard and Groc 2011). The aim of the present study was to analyze the surface expression of NMDA-like receptors during consolidation of the contextual learning in the crab *N. granulata*. To reach such a goal we used differential cross-linking with bis(sulfosuccinimidyl)suberate (BS3) to establish a GluN1 membrane/intracellular ratio. The ratio was calculated at different time points after training to establish the time course of learning-induced changes in surface expression. The surface expression of the GluN1 subunit of the NMDA receptor is down-regulated immediately after training, up-regulated 3 h after training and returns to the naïve and control levels 24 h after training. This effect was independent of GluN1 synthesis and only occurred in the central brain and not in the thoracic ganglion. To further study the pathway that underlies the changes in surface expression of the NMDA receptor subunit GluN1 we used bicuculline (Tano et al. 2013) (antagonist of GABA_A receptors) to indirectly induce the activation of the glutamatergic pathway. There is evidence that bicuculline changes the GluN1 content at post-synaptic densities in cultured rat hippocampal neurons (Ehlers 2003). We observed a similar increment in surface expression of GluN1 in the central brain 3 h after the drug administration.

All in all these data suggest that in the crab *N. granulata*, NMDARs are involved in memory consolidation through fine regulation of their synaptic localization. These changes in surface expression of the NMDARs suggest a role in the stabilization of the learned task.

Results

Training with 30 trials generates a long-term context-signal memory trace in *N. granulata*

Presentation of a moving screen (visual danger stimulus, VDS) to each crab elicits an escape response that diminishes over 30 iterative presentations separated by

100 sec. During the pretraining trial, trained (Tr, $N = 17$) and control groups (Ctl, $N = 17$) show no significant differences in their escape response (Fig. 1A,B, PT). The response levels of the Tr group to the VDS reduces to levels below Ctl group basal activity (Fig. 1A, inset). This suggests a strategy change in the trained animals. Twenty-four hours after training, Tr and Ctl groups are evaluated with six VDS presentations in the same context where they were trained, and analyzed in two blocks: first testing trial and retraining from the second to the sixth trial. A significantly lower response is observed in the Tr animals for the first testing trial ($p < 0.05$) as for the remaining five trials, when compared with Ctl group animals ($p < 0.01$) (Fig. 1A,B). This response difference between groups in the testing session is defined as retention of the learned context–signal association. All the following experiments were performed with a replicate in which one was used for a biochemical experiment and the second was evaluated for memory retention. The corresponding biochemical data was used only if the replicate performed with animals from the same population showed significant memory retention at the first testing trial.

GluN1 subunit of the NMDA receptor in the crab *N. granulata* is detected by antibodies to the carboxyl-terminus of the protein

Previous physiological and immunohistochemical evidences indicate that the NMDAR is present in the crab nervous system (Hepp et al. 2013) and is necessary for the consolidation of the context signal memory (CSM) (Troncoso and Maldonado 2002; Pérez-Cuesta et al. 2007) In previous work from our laboratory, the NMDA receptor subunit GluN1 was characterized and its localization in the nervous system as well as its distribution at cellular level was described (Hepp et al. 2013). In that opportunity, a polyclonal antibody directed at a peptide corresponding to the amino acids 909–938 of the rat GluN1 subunit was used (Chemicon, AB1516). To further characterize the GluN1 receptor

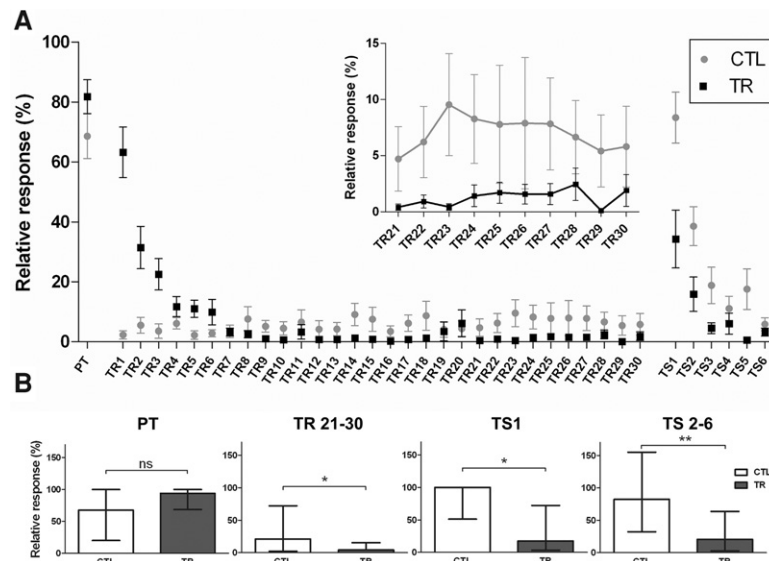


Figure 1. Training and retention testing of the context signal memory in the crab *N. granulata*. (A) Relative response (%) means and SEM for pretraining trial (one stimulus presentation, to equilibrate the response level of the groups), training (day 1) and testing (day 2), control group (Ctl, 30 recordings of basal activity with 100 sec interrecordings interval) and training groups (Tr, 30 recordings during stimulus presentations, with 100 sec intertrial interval). (B) Comparison of the response level for Ctl and Tr groups during pretraining (PT) (no significant differences), the block between training trial 21–30 (Tr 21–30) ($p < 0.05$), the first testing trial (TS1, $p < 0.05$) and the block between testing trial 2 and 6 (TS2–TS6, $p < 0.01$). Animals per group, $n = 17$.

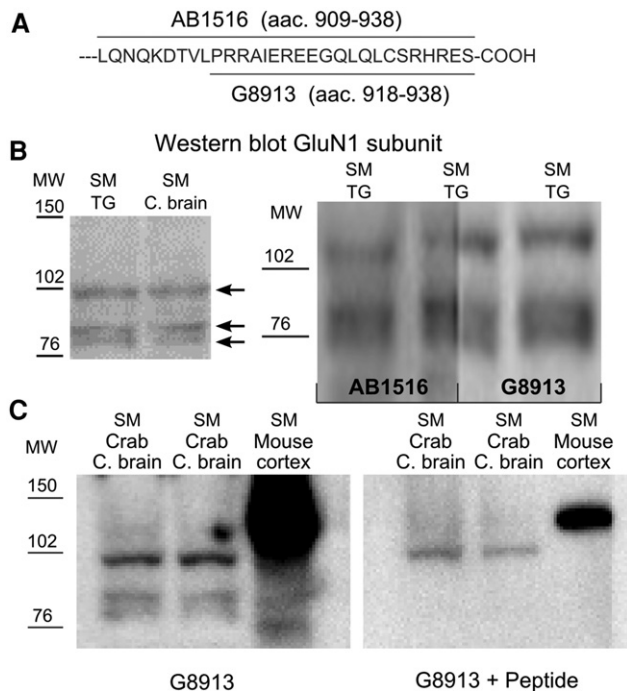


Figure 2. Antibodies against rat GluN1 cross-react with a GluN1 like protein in crab. (A) Comparison of the sequence corresponding to the carboxy-terminal GluN1 subunit of rat recognized by antibodies AB1516 and G8913. (B) Western blot assay using two anti-rat GluN1 antibodies. Synaptosomal membrane extracts from crab thoracic ganglion (SM TG) and from crab brain (SM Brain). The AB1516 antibody was used for the left panel and the first lane and half of the second lane of the right panel, while the G8913 antibody was used for the second half of the second lane and third lane of the right panel. Arrows indicate the bands for GluN1 membrane fraction at ~ 102 kDa and intracellular fraction at ~ 88 and ~ 84 kDa. (C) Western blots with the G8913 antibody left panel and with same antibody preincubated with the peptide that was used to make the antibody to discern the specific bands in the immunoblot, right panel, for both panels the samples were SM of Crab brain for the first two lanes and SM of mouse cortex for the third lane. Mw, molecular weight (kDa).

subunit in the crab and explore the receptor expression during long-term memory consolidation, we used an antibody directed to a sequence overlapping the one recognized by AB1516, G8913 Sigma-Aldrich (Fig. 2A). The signal for the AB1516 antibody is similar in both the central brain and thoracic ganglion (Fig. 2B, left panel). The G8913 antibody yields a similar signal when compared in the same lane of a Western blot, and two adjacent lanes, one for each antibody (Fig. 2B, right panel). The peptide described in Figure 2A was preincubated with the G8913 antibody and the specificity of the two lower bands evidenced by the antibody (at ~ 88 and ~ 84 kDa) was demonstrated (Fig. 2C). All of this evidence further supports that the evaluated protein is, in fact, a subunit of the crab homolog of the NMDA receptor.

Expression of the GluN1 subunit of the NMDA receptor in the central brain is not affected during consolidation

To evaluate the expression of the GluN1 subunit during consolidation of the CSM, we choose the central brain of *N. granulata*, a structure involved in this type of learning (Freudenthal et al. 1998). Total central brain homogenate from trained and control animals were obtained immediately after training or 3-h post-training, and were compared with that of Naïve animals (Nv) in a Western blot assay (Fig. 3). Actin was used as a loading control;

the signal obtained for the GluN1 of each animal was normalized to the actin content of the same line. The amount of the GluN1 subunit observed in the total central brain homogenates from Tr, Ctr, and Nv did not differ at the times evaluated (0 h: Tr $n = 14$, Ctr $n = 15$, Nv $n = 17$; 3 h: Tr $n = 5$, Ctr $n = 6$, Nv $n = 5$) (Fig. 3). This result suggests that the total amount of GluN1 does not change significantly during consolidation at the time points evaluated.

Assessment of membrane NMDA receptor GluN1 subunit with surface expression assay

To identify the proportion of membrane GluN1 of the total GluN1 we use the differential cross-linking with bis(sulfosuccinimidyl)-suberate (BS3). Using BS3 on the tissue, on top of the original bands, a signal corresponding to a complex containing GluN1 appears approximately at the level of the 200 kDa marker (Fig. 4A,B). Similar bands appear when the BS3 cross-linking agent was applied to the central brain and the thoracic ganglion (Fig. 4A). Cross-linking for 1, 3, or 5 d, show similar intensities for the high molecular weight bands (Fig. 4A). Since BS3 does not cross the plasmatic membrane, cross-linking takes place only between proteins with an extracellular domain (Grosshans et al. 2002b). As shown in previous studies (Hall and Soderling 1997), when treating with BS3, the high molecular weight bands appear only when immunoblotting for a membrane protein (GluN1) but not for a protein that has no extracellular domains (actin) (Fig. 4B).

Training induces changes in surface expression of GluN1 in the central brain but not in the thoracic ganglion

We evaluated if learning affects the membrane/intracellular ratio of GluN1. We trained animals as described in Figure 1 and the localization of the GluN1 subunit was evaluated by Western blot analysis at different time points after training. The central brains were dissected and were treated with BS3 for the Nv, Tr, and Ctl animals at 0-, 3-, and 24-h post-training. The BS3 tissue treatment allowed us to identify the extracellular and intracellular GluN1 in total homogenates. Immediately after training there is a significant decrease of the GluN1 subunit expressed at the membrane surface for Tr animals compared with Ctl and Nv animals (ANOVA $F_{(2,15)} = 7.99$; $P = 0.004$; Fisher-LSD; Nv-Tr, $P < 0.01$; Ctl-Tr, $P < 0.05$; Ctl-Nv, NS.). At 3-h post-training there is an increase of the membrane/intracellular ratio for Tr animals compared with Nv animals (ANOVA, $F_{(2,14)} = 3.99$; $P = 0.042$; Fisher-LSD; Nv-Tr, $P < 0.05$) (Fig. 5A). At 24-h post-training there is no significant differences in the GluN1 surface expression between groups (Fig. 5A). The thoracic ganglia were extracted and treated with BS3 at 0- and 3-h post-training and the membrane/intracellular ratio was assessed. There was no significant difference for membrane surface expression of the GluN1 subunit in the thoracic ganglion during consolidation (Fig. 5B). These data suggest that the GluN1 subunit membrane exposure is affected during memory consolidation in the nervous system area described to be involved in learning and memory processes (Freudenthal and Romano 2000).

Surface expression of GluN1 is affected after systemic administration of bicuculline

To further study the pathway that underlies the membrane trafficking of the NMDA receptor subunit GluN1 we used bicuculline as an indirect way to induce glutamatergic activation and thus evaluate the effect on membrane surface exposure of the receptor. Three increasing doses of bicuculline were tested and the membrane/intracellular ratio was evaluated in the central brain 3 h

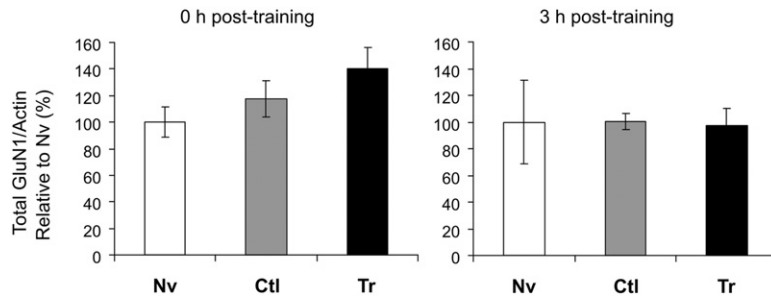


Figure 3. Total GluN1 subunit/actin in whole-central brain extracts relative to Naïve. The amount of total GluN1 was evaluated for the Naïve (Nv), Control (Ctl), and Trained (Tr) groups at different times post-training: 0 hours (0 h) and 3 hours (3 h). Actin was used as a loading control and values are shown relative to Nv (%).

after the systemic injection. As seen previously in Fig. 5, at 3-h post-training the membrane/intracellular NMDAR ratio increases, therefore this time was the one used to evaluate the effects of systemic bicuculline injection. A significant increase in the membrane/intracellular ratio was observed with a dose of 10 $\mu\text{g/g}$ 3-h post-injection (MEAN = 149.4, SEM = 14.5, $P < 0.05$) (Fig. 6). No significant increase was observed for the 3 $\mu\text{g/g}$ and the 15 $\mu\text{g/g}$ dose showed a tendency for increase that was not statistically confirmed. This suggests that increased membrane exposure of the NMDA receptor subunit GluN1 observed 3 h after neuronal activation may provide a mechanism through which the change in membrane exposure during consolidation is achieved.

Discussion

In this study, we show that memory consolidation affects surface expression of GluN1 subunit while not affecting the total expression of the protein.

CSM training in the crab *Neohelice* induces a memory with a strong retention at 24 h, when comparing Tr animals that were exposed to 30 iterative presentations of the VDS with Ctl animals that were exposed to the context with no VDS presentation (Fig. 1).

Here we find a specific signal for the NMDA receptor subunit GluN1. The specific bands are detected when using two different antibodies directed against the carboxy terminus of GluN1, as shown in our previous work (Hepp et al. 2013), and by the competition with the antigenic peptide shown in Figure 2C. This signal is detected both in the central brain and in the thoracic ganglion of the crab (Figs. 2B, left panel and 4A). The GluN1 comparison between these two areas is important because the central brain is relevant for memory consolidation as shown by Freudenthal and Romano (2000).

The total protein levels of the NMDA receptor subunit GluN1 in the central brain of Ctl and Tr animals do not differ from the levels in Nv animals, immediately after and 3-h post-training (Fig. 3). This led us to the idea that the regulation of the receptor was not at the protein expression level as seen in other memory models (Baez et al. 2013) but rather at the surface expression level. The regulation at the surface expression level has been repeatedly reported in memory and synaptic plasticity experiments (Camarota et al. 2000; Grosshans et al. 2002b; Moyano et al. 2004; Mukherjee et al. 2014).

To evaluate how much of the total protein was either exposed at the plasma membrane or was intracellular, we adapted the surface expression assay using BS3 cross-linking, previously used for AMPA and NMDA receptors (Hall and Soderling 1997; Grosshans et al. 2002b; Boudreau et al. 2012), to the crab nervous

system (Fig. 4). The increment in surface expression of the NMDA receptor measured by the BS3 technique positively correlates with changes in NMDA currents in rat CA1 region of the hippocampus (Grosshans et al. 2002a,b). Therefore, we consider that NMDA surface expression observed by BS3 technique corresponds to functional receptors. Furthermore, the BS3 technique is reliable to evaluate surface expression, yielding similar results as other techniques of surface modification of receptors such as chymotrypsin, biotinylation, and radioactive ligand binding (Hall and Soderling 1997; Grosshans et al. 2002b).

This protocol yielded separated signals for the membrane cross-linked GluN1 and the intracellular GluN1, and this cross-linking did not affect other intracellular proteins as shown with actin (Fig. 4B). Grosshans et al. (2002b) showed that the surface expression assay is able to detect increments in the NMDA receptor subunit GluN1 (indirectly through a decrease in intracellular GluN1) induced after a high-frequency stimulation that yields long-term potentiation. In our experiments, surface expression assays during consolidation indicate that the GluN1 subunit decreased in trained animals immediately after training when compared with Nv and Ctl animals in the central brain. This change was not general to the whole nervous system as shown by the unchanged levels shown among Nv, Ctl, and Tr animals in the thoracic ganglion (Fig. 5). Three hours post-training the surface expression of GluN1 in the central brain of Tr animals shows an increment when compared with Nv and Ctl animals, and again the change is seen only in the central brain and not in the thoracic ganglion (Fig. 5). The central brain levels of surface GluN1 are

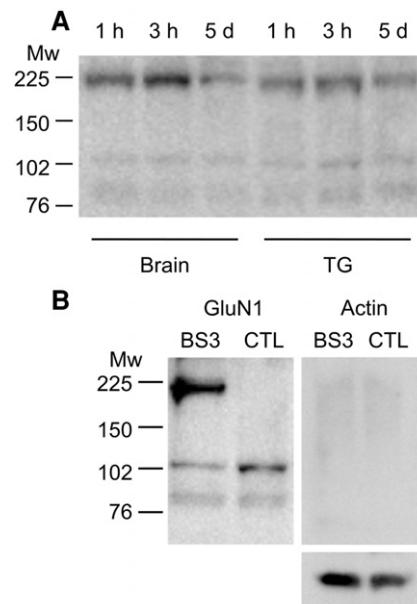


Figure 4. Membrane exposure assay with BS3. (A) Total extracts of central brain and thoracic ganglion treated with BS3 for different times: 1 hour (1 h), 3 hours (3 h), and 5 days (5 d). (B) Total extracts of thoracic ganglia treated with BS3 (BS3) or control (CTL). *Left panel*, AB1516 antibody for the GluN1 subunit; *right panel*, antibody for actin. Mw: molecular weight (kDa).

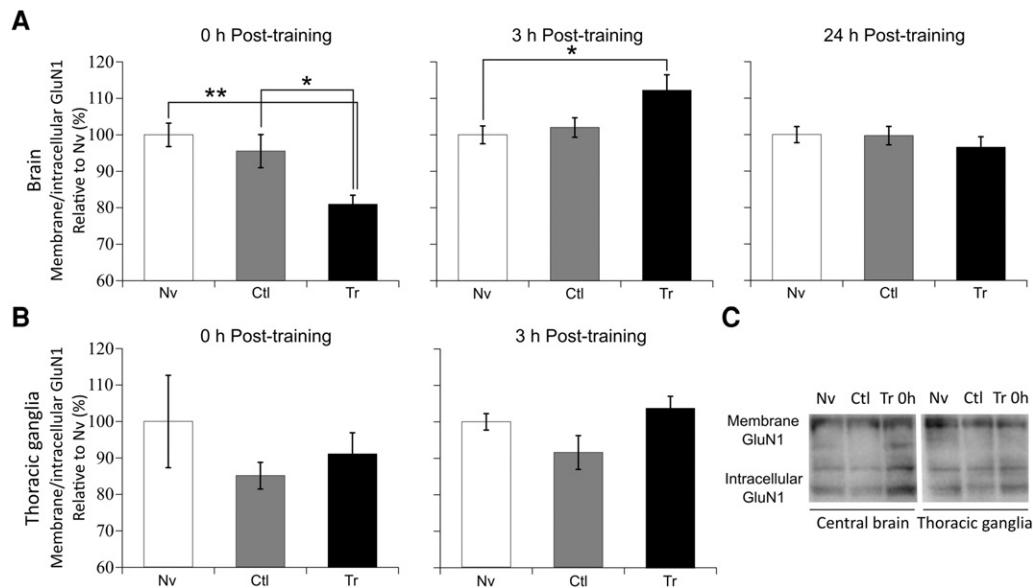


Figure 5. Membrane/intracellular GluN1 relation in the central brain and thoracic ganglion during consolidation. Bars represent mean and SEM of the relation between membrane GluN1 fraction and intracellular GluN1 fraction expressed as percentage of the average relation determined in simultaneously treated Naive (Nv) animals. Nv, control (Ctl) and trained (Tr) animals were treated with BS3 at different times post-training. (A) Central brain 0-h post-training (left panel, $n = 6$ for each group), 3-h post-training (center panel, $n = 6$ for each group), and 24-h post-training (right panel, $n = 10$ (Nv y Tr) and $n = 14$ (Ctl)). (*) $P < 0.05$ and (**) $P < 0.01$ for statistical differences between groups after ANOVA. (B) Thoracic ganglion 0-h post-training (left panel, $n = 6$ for each group) and 3-h post-training (right panel, $n = 6$ for each group). (C) Representative Western blot with the G8913 antibody of extracts obtained in Nv, Ctl, and Tr groups at 0-h post-training for central brain (left panel) and thoracic ganglion (right panel).

similar between the three animal groups when assessed 24-h post-training (Fig. 5).

During the initial part of LTP, the membrane ratio of splice variants of GluN1 dramatically changes (Grosshans et al. 2002b). One possibility is that the initial decrement in surface expression of GluN1 seen in the crab central brain immediately after training could be reflecting endocytosis of this subunit that is part of a splice variant rearrangement. Surface expression of GluN1 is augmented during LTP in rat CA1 area of the hippocampus from 30 min after high frequency stimulation until at least 3 h, in a time window that agrees with the plasticity seen in the crab during consolidation (Grosshans et al. 2002a). It has been proposed that NMDARs can significantly contribute to information transfer at synapses during periods of repetitive activity (Hunt and Castillo 2012), such as the CSM learning task, which relies on iterative presentations of the stimulus. These changes in the surface GluN1 of Tr animals during consolidation may be a form of metaplasticity, in which the changes in NMDA receptors suggest a shift in the threshold to undergo further plasticity.

In an attempt to address if the observed trafficking of GluN1 is elicited by neuronal activity, we used systemic bicuculline injections (antagonist of GABA_A receptors) in incremental doses and evaluated surface expression 3-h post-injection. The level of surface GluN1 in the central brain was incremented by injection of 10 $\mu\text{g/g}$ dose of bicuculline but not by a 3 $\mu\text{g/g}$ dose (Fig. 6A). The results from injected animals suggest that glutamatergic activity is a candidate for the changes seen in GluN1 surface expression during consolidation (Fig. 6). Mukherjee et al. (2015) describe an initial decrease of NMDAR surface expression after training followed by a later increase in rat anterior piriform cortex.

Neuronal activity in the central brain during consolidation could be changing the levels of functional NMDA receptor subunit GluN1. These changes could be reflecting time specific requirements at the synapses. A decrement in surface GluN1 immediately after training, would not allow further activation through

the receptor, calling for specificity on other synapses. On the contrary, the increment in surface GluN1 3-h post-training, once the consolidation processes is advanced, could reflect a higher probability for activation (Mukherjee et al. 2015).

This is the first time that a change in NMDA receptor surface expression during long-term memory consolidation is described in an invertebrate. Altogether, these results show a consolidation landscape where NMDA receptors, the coincidence detectors, have a more dynamic regulation than previously thought, thus contributing to mechanisms of memory consolidation. This work provides important groundwork and opens questions to investigate metaplasticity mechanisms in learning and memory.

Materials and Methods

Animals

We used adult male intertidal crabs (*N. granulata*, formerly *Chasmagnathus granulatus*), measuring 2.7–3.0 cm across the

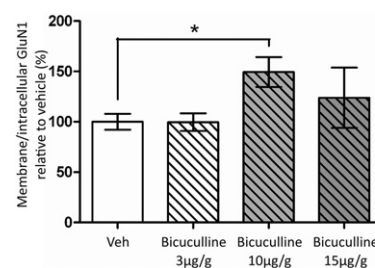


Figure 6. Membrane/intracellular relation of GluN1 after systemic injections bicuculline. Membrane/intracellular GluN1 relation in the central brain treated with BS3 at 3-h post-injection of different concentrations of bicuculline (3, 10, and 15 $\mu\text{g/g}$) or vehicle (Veh) ANOVA, (*) $P < 0.05$. $N = 14$.

carapace and weighing ~17 g. The crabs were collected from water <1 m deep in narrow coastal inlets (rias) of San Clemente del Tuyú, Buenos Aires Province, Argentina. Once transported to the laboratory, they were housed in plastic tanks (35 × 48 × 27 cm) filled to a depth of 2 cm with diluted marine water (Red sea fish pharm) with a salinity of 1.0%–1.4% and a pH of 7.4–7.6. The water was changed every 2 d. The holding room was maintained on a 12-h light–dark cycle (lights on from 07:00–19:00 h) and the temperature was maintained between 22°C–24°C. Experiments are performed only in males to avoid disrupting the natural population or crabs. Females carry the fecundated eggs in the first stages of development and capturing them might affect the size of the population. Another factor considered is that a natural population has many sources of variability and selecting male animals restricts variables such as size range. The reported research was conducted in accordance with the local regulations for the care and use of laboratory animals. All experiments were done in accordance to local regulations to minimize animal suffering and the number of animals used.

Drugs and injection procedure

Dimethyl sulfoxide (DMSO) was used as the vehicle. Ten microliters of vehicle or drug solution was given through the right side of the dorsal cephalothoracic–abdominal membrane by means of a syringe fitted with a sleeve to control the depth of penetration to 4 mm, thus ensuring that the injected solution was released in the pericardial sac. Bicuculline, the competitive antagonist of GABA_A receptors, was purchased from Fluka Analytical and used in concentrations of 3, 10, and 15 µg/g.

Training and testing procedures

The apparatus used for training and measure of escape response is described in detail elsewhere (Romano et al. 1990). Briefly, the experimental unit was the actometer: a bowl shaped plastic container with a steep concave wall and a circular central flat floor 10 cm in diameter, covered to a depth of 0.5 cm with salt water. The crab was lodged in the container, which was suspended by three strings from an upper wooden framework 23 × 23 × 30 cm. and illuminated by a 5 W lamp placed 30 cm (Tano et al. 2013) above the animal. A motor-operated screen (US, an opaque rectangular strip of 25.0 × 7.5 cm) was moved horizontally over the animal from left to right, and vice versa. The screen's movements were cyclical. The screen displacements provoked the escape response of the crab and subsequent container vibrations. Four microphones were attached to the center of the outside base of the container. The microphones recorded the vibrations that were produced by the animal's response. These signals were amplified, integrated during the entire trial (9 sec) and translated into arbitrary numerical units ranging from 0 to 8000. Each trial lasted 5 sec and consisted of two cycles of stimulation (four screen presentations over the actometer), the recorded crab activity during the entire trial time is integrated. The experimental room has 40 actometers, isolated from each other by partitions. A computer is used to program trial sequences, trial duration (5 sec) and inter-trial interval (ITI, 100 sec), as well as to monitor experimental events. Trained animals received 30 trials of 5-sec duration, with 100-sec ITI and control animals were recorded during the same periods as the trained ones but without the screen presentation. All training sessions were preceded by 10 min of adaptation time in the actometer, after this adaptation period all animals (trained and control) receive one pretraining trial (PT) with screen presentation. This presentation response is used to distribute the animals in two groups (Tr and Ctl) with similar response levels and for the percent normalization of each crab. Training sessions lasted 50 min for both groups. The testing session is performed 24 h after training, with six trials. Crabs were individually housed during the entire inter-session interval in plastic containers, covered to a depth of 0.5 cm with seawater and kept inside dimly lighted drawers. The maximal response of an animal during a trial (PT, training or testing) is taken as a 100% response and all other responses are normalized to this value. This procedure minimizes differences

between animals due to factors other than the training (animal weight, motor vibration differences). The level of response at testing session was compared between trained and control groups. Retention of the learning acquired during training was considered when a statistically lower level of response was found in trained group in comparison to control group.

Protein extracts

The animals were anaesthetized by immersion in an ice/salt water mix for 5 min at 0-, 3-, or 24-h post-training or injection. The central brains and thoracic ganglia were dissected. For each sample, 4–6 central brains or thoracic ganglia were pooled in 1 mL of buffered crab saline solution (pH 7.4).

Total homogenates

Pooled central brains or thoracic ganglia were homogenized in 150 µL of hyposmotic solution containing 1% SDS and protease inhibitors (phenylmethylsulfonyl fluoride [PMSF] 0.5 mM, aprotinin 10 µg/mL, leupeptin 10 µg/mL, pepstatin A 1 µg/mL), using a glass–glass tight dounce homogenizer. Samples were centrifuged for 10 min at 10000g and the supernatant was separated and stored at –80°C until use.

Synaptosomal extracts

The extracts enriched in synaptosomal membranes were obtained as follows: The dissected tissues were homogenized in 750 µL of cold homogenization buffer (20 mM Tris–HCl, 900 mM sucrose, 1 mM EDTA, pH 7.8), with protease inhibitors (as in total homogenates) with a glass–glass homogenizer. The tissue homogenates were centrifuged (4°C, 1000g, 10 min) to produce a pellet (P1) and a supernatant (S1). S1 was collected and the P1 was resuspended in 500 µL of the homogenization buffer and centrifuged (4°C, 1000g, 10 min) to obtain a pellet (P10) and supernatant (S10). P10 was resuspended in 30 µL of cold hyperosmotic buffer (20 mM HEPES, pH 7.9; 1.2 M KCl; 1.5 mM MgCl₂; 0.4 mM EDTA; 0.5 mM DTT; 50% glycerol, pepstatin A, 1 µg/mL; leupeptin, 10 µg/mL; 0.5 mM PMSF; and aprotinin, 10 µg/mL) and centrifuged (4°C, 10,000g, 15 min). The supernatant corresponds to the nuclear extract and the pellet was discarded. S1 and S10 were combined and centrifuged at 13,000g for 30 min to obtain a pellet (P2, enriched in synaptosomes) and a supernatant (S2, cytoplasmatic extract). P2 was resuspended in 200 µL of cold hyposmotic buffer (5 mM Tris–HCl, 50 mM CaCl₂, pH 8.1), sonicated at 49% for 3 sec and centrifuged (15,800g, 50 min) to produce a pellet (P3, enriched in synaptosomal membranes) and a supernatant (S3, enriched in synaptosomal content). P3 was resuspended in 30 mL of hyposmotic buffer-1% sodium dodecyl sulfate (SDS).

BS3-treated extracts

Pooled central brains or thoracic ganglia were washed twice by placing them in HSE buffer (2 mM) with protease inhibitors on a shaker and centrifuging for 2.5 min at 1000g. Central brains or thoracic ganglia were transferred to a new tube and incubated in 150 µL (for thoracic ganglia) or 100 µL (for central brains) in HSE buffer with 3 mM of BS3 for 1, 3 h or 5 d on a refrigerated (4°C) shaker. These fixation times were used for experiment in Figure 4, all other experiments involving BS3 were incubated for 1 h with the fixating agent. After incubation with BS3 samples were washed twice with HSE buffer as described in the first step of this protocol. Samples were transferred to a new tube and homogenized in 200 µL (for thoracic ganglia) or 75 µL (for central brains) of HSE buffer with 1% SDS and protease inhibitors. Samples were stored at –80°C until use.

Western blots

Loading buffer (4×) was added to each sample (35 µg of protein), which was then denatured (boiled for 5 min at 100°C). Then the samples were electrophoresed using 10% SDS-polyacrylamide gel electrophoresis (PAGE) at 100 V for 1 h and then electroblotted

to PVDF membrane (Bio-Rad) at 100 V for 1 h. The blots were blocked in blocking buffer (10% low-fat dry milk in Tris-buffered saline–0.1% Tween 20 [TBS-T], pH 7.6) for 1 h at room temperature. After two 15-min washes with TBS-T, the blots were incubated overnight (4°C) with a primary antibody, for either NMDA GluN1 or actin. For the AB1516 antibody (Chemicon), a 1:100 dilution in PBS-T was used. For the G8913 antibody (Sigma-Aldrich) a 1:400 dilution with T-TBS was used. For the actin antibody (Santa Cruz Biotechnology) a 1:1000 dilution in T-TBS was used. After six 10-min washes with TBS-T, the blots were incubated with horseradish peroxidase (HRP)-conjugated goat anti rabbit IgG (1:5000 dilution in TBS-T, 1 h, 25°C; Santa Cruz Biotechnology; sc-2030), followed by four washes of 10 min each with TBS-T. Detection was made with the Advanced ECL detection kit (GE Amersham) as described by the manufacturer, and the signals were digitized by a FUJI FILM-Intelligent Dark Box II apparatus with image reader LAS-1000 software. The protein size was estimated by its relative mobility using ImageJ 1.293 software (NIH) and correlating these measurements with the molecular weights of prestained standard proteins (Full-Range Rainbow Molecular Weight Markers: 12–225 kDa, Amersham).

Data analysis

Mann-Whitney test was used to analyze the Pretraining trial, the sum of the training trials from 21 to 30 (TR 21–30), the first testing trial (TS1) and the sum of testing trials 2–6 (TS 2–6).

For Western blots, optic density (OD) was measured using ImageJ software and statistical analysis was performed using one-way ANOVA, with Fisher-LSD post hoc multiple comparisons.

The data submitted complies with ANOVA requirements such as normality and homogeneity of variances. When normality and homogeneity was not met, nonparametric Kruskal–Wallis analysis was used.

Competing interest statement

The authors have no conflict of interest.

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Authors' contributions: All authors had full access to all the data in the study and take responsibility for the integrity of the data and the accuracy of the data analysis. Study concept and design: M.E.P. and R.F. Acquisition of data: Y.H., A.S., M.C.T., and R.F. Analysis and interpretation of data: Y.H., A.S., M.E.P., and R.F. Drafting of the article: Y.H., A.S., M.E.P., and R.F. Critical revision of the article for important intellectual content: Y.H., A.S., M.C.T., M.E.P., and R.F. Statistical analysis: Y.H., A.S. and R.F. Funding: M.E.P. and R.F. Administrative, technical, and material support: M.E.P. and R.F. Study supervision: M.E.P. and R.F.

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