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Susceptibility to Fosfomycin and Nitrofurantoin of ESBL-Positive Escherichia coli and Klebsiella pneumoniae Isolated From Urine of Pediatric Patients

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ABSTRACT

Background: Pediatric urinary tract infection (UTI) caused by extended-spectrum β -lactamase (ESBL)-positive gram-negative bacilli (GNB) has limited options for oral antibiotic treatment. The purpose of this study was to investigate the susceptibility of ESBL-positive *Escherichia coli* and *Klebsiella pneumoniae* isolates from pediatric urine samples to two oral antibiotics (fosfomycin and nitrofurantoin).

Methods: From November 2020 to April 2022, ESBL-positive *E. coli* and *K. pneumoniae* isolates from urine samples were collected at Samsung Medical Center, Seoul, Korea. Patients over 18 years of age or with malignancy were excluded. For repeated isolates from the same patient, only the first isolate was tested. Minimum inhibitory concentrations (MICs) were measured using agar (fosfomycin) or broth (nitrofurantoin) dilution methods. MIC₅₀ and MIC₉₀ were measured for fosfomycin and nitrofurantoin in both *E. coli* and *K. pneumoniae*.

Results: There were 117 isolates from 117 patients, with a median age of 7 months (range, 0.0–18.5 years). Among 117 isolates, 92.3% (108/117) were *E. coli* and 7.7% (9/117) were *K. pneumoniae*. Isolates from the pediatric intensive care unit (PICU) and general ward (GW) was 11.1% (13/117) and 88.9% (104/117), respectively. Among 108 *E. coli* isolates, MIC₅₀ and MIC₉₀ for fosfomycin were 0.5 µg/mL and 2 µg/mL, respectively. Fosfomycin susceptibility rate was 97.2% (105/108) with a breakpoint of 128 µg/mL. Fosfomycin susceptibility rate was significantly lower in PICU isolates than in GW isolates (81.8% vs. 99.0%, *P* = 0.027). For nitrofurantoin, both the MIC₅₀ and MIC₉₀ were 16 µg/mL. Nitrofurantoin susceptibility rate was 96.3% (104/108) with a breakpoint of 64 µg/mL based on Clinical and Laboratory Standards Institute guidelines. Among the nine *K. pneumoniae* isolates, the MIC₅₀ and MIC₉₀

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Disclosure

The authors have no potential conflicts of interest to disclose.

Author Contributions

Conceptualization: Park KS, Kim DR, Kim YJ. Data curation: Park KS, Kim DR, Shin A, Kim KR, Park H, Son S. Formal analysis: Kim DR, Park KS. Investigation: Kim YJ. Park KS, Kim DR. Methodology: Park KS, Baek JY, Kim YJ. Validation: Cho H. Writing - original draft: Park KS, Kim DR. Writing - review & editing: Park KS, Kim DR, Baek JY, Shin A, Kim KR, Park H, Son S, Cho H, Kim YJ. for fosfomycin was 2 μ g/mL and 32 μ g/mL, respectively. MIC₅₀ and MIC₉₀ for nitrofurantoin were 64 μ g/mL and 128 μ g/mL, respectively.

Conclusion: For uncomplicated UTI caused by ESBL-positive GNB in Korean children, treatment with fosfomycin and nitrofurantoin for *E. coli* infections can be considered as an effective oral therapy option.

Keywords: Pediatric; Urinary Tract Infection; Fosfomycin; Nitrofurantoin

INTRODUCTION

Urinary tract infection (UTI) is common in children and requires antibiotic treatment. Antibacterial resistance in UTIs is a growing problem in both children and adults.¹⁻³ It is of concern that UTIs caused by resistant pathogens such as extended-spectrum β -lactamase (ESBL) positive gram-negative bacilli (GNB) is rising in children without significant risk factors in the community.⁴⁻⁶

We previously reported the antibiotic resistance pattern in children less than 24 months of age with UTI and showed an increasing resistance rate of ESBL positivity among isolates of *Escherichia coli* and *Klebsiella pneumoniae* from 1.3% to 8.2% over two test periods, 2000–2004 and 2010–2014.⁷ In young febrile children with toxic appearance and significantly increased inflammatory markers, parenteral antibiotics are used as the empiric treatment. Oral antibiotic treatment is common practice in children with UTIs and there are several options for oral antibiotic treatment.⁸ However, pediatric UTI caused by ESBL-positive GNB has limited options for oral antibiotics.

Fosfomycin and nitrofurantoin are broad-spectrum cell wall inhibitors that can be useful oral treatment options in non-complicated UTIs of community origin due to their rapid oral absorption, high urine concentration, and bactericidal activity against a wide range of gram-negative and gram-positive bacteria.⁹⁻¹¹ The purpose of this study was to investigate the susceptibility of ESBL-positive *E. coli* and *K. pneumoniae* isolates cultured from pediatric urine samples to two oral antibiotics (fosfomycin and nitrofurantoin).

METHODS

This study included isolates from urine in pediatric patients at Samsung Medical Center from November 2020 to April 2022. Clinical information and microbial data were collected from electronic medical records. Patients over 18 years of age or with malignant diseases were excluded.

Bacterial isolates

ESBL-positive *E. coli* and *K. pneumoniae* isolated from urine samples were collected. For repeated isolates from the same patient, only the first isolate was tested. Bacterial species were identified using the VITEK 2 system (bioMérieux, Marcy-l'Etoile, France).

Determination of fosfomycin and nitrofurantoin minimal inhibitory concentration (MIC)

Antibiotic media were prepared with concentrations ranging from 0 mg/L to 256 mg/L through 0.5 serial dilution. Mueller Hinton Agar (BD, Franklin Lakes, NJ, USA) was used for fosfomycin

testing and Mueller Hinton Broth (BD) was used for nitrofurantoin.^{12,13} The final inoculation concentration was 10⁴ cfu/spot for fosfomycin testing and 10⁵ cfu/mL for nitrofurantoin. The cultures were incubated at 37°C for 16 hours. *E. coli* (ATCC 25922), *Enterococcus faecalis* (ATCC 29212), and *Pseudomonas aeruginosa* (ATCC 27853) were also tested as controls.

MIC analysis followed the Clinical & Laboratory Standards Institute (CLSI) guidelines,¹⁴ and the concentration at which growth was completely inhibited was measured as the MIC of the corresponding strain. According to CLSI guidelines, the *E. coli* and *K. pneumoniae* breakpoints for susceptibility was set to $\leq 64 \mu g/mL$ and $\leq 32 \mu g/mL$ for fosfomycin and nitrofurantoin, respectively.

Statistical analysis

Each *P* value was confirmed using the Fisher's exact test to compare the susceptibility rate between the general ward (GW) and pediatric intensive care unit (PICU).

Ethics statement

This study was approved by the Institutional Review Board of Samsung Medical Center, Seoul, Korea and the requirement for informed consent was waived (approval number: 2018-02-107 and 2023-02-091).

RESULTS

ESBL-positive isolates

A total of 117 ESBL-positive isolates were collected, of which 108 (92.3%) were *E. coli* and nine (7.7%) were *K. pneumoniae*. The median age of patients at the time of positive urine culture was seven months (range, 0.0–18.5 years) (**Supplementary Table 1**). Among these, 13 isolates (11.1%) were from patients in the PICU; *E. coli* (n = 11) and *K. pneumoniae* (n = 2). A total of 104 (88.9%) isolates were from patients in the GW; *E. coli* (n = 97) and *K. pneumoniae* (n = 7). Isolates showed high resistance to ampicillin, aztreonam, ciprofloxacin, gentamicin, and trimethoprim/sulfamethoxazole (TMP/SMX) (Table 1).

ESBL-positive E. coli antibiotic susceptibility

The susceptibility of ESBL-positive *E. coli* to fosfomycin and nitrofurantoin is shown in Fig. 1. The MIC₅₀ and MIC₉₀ for fosfomycin against the 108 *E. coli* isolates were 0.5 μ g/mL and 2 μ g/mL,

Antibiotics	l	E. coli (n = 108))	K. pneumoniae (n = 9)			
	S	I	R	S	I	R	
Ampicillin	0.00%	0.00%	100.00%	0.00%	0.00%	100.00%	
Amox/clav	55.60%	25.00%	19.40%	33.30%	33.30%	33.30%	
Amikacin	100.00%	0.00%	0.00%	88.90%	0.00%	11.10%	
Aztreonam	0.00%	0.00%	100.00%	22.20%	0.00%	77.80%	
Ciprofloxacin	46.30%	0.90%	52.80%	44.40%	11.10%	44.40%	
Gentamicin	55.60%	1.90%	42.60%	66.70%	0.00%	33.30%	
Ertapenem	99.10%	0.00%	0.90%	100.00%	0.00%	0.00%	
Imipenem	100.00%	0.00%	0.00%	100.00%	0.00%	0.00%	
Pip/tazo	90.70%	4.60%	4.60%	66.70%	11.10%	22.20%	
Tigecycline	100.00%	0.00%	0.00%	88.90%	11.10%	0.00%	
TMP/SMX	39.80%	0.00%	60.20%	22.20%	0.00%	77.80%	

Table 1. Antibiotic susceptibility of isolates

Amox/clav = amoxicillin/clavulanate, Pip/tazo = piperacillin/tazobactam, TMP/SMX = trimethoprim/sulfamethoxazole, S = sensitive, I = intermediate, R = resistant.

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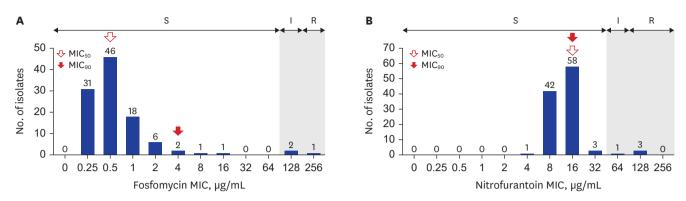


Fig. 1. Distribution of MICs and susceptibility of ESBL-positive *E. coli* isolates. (A) ESBL-positive *E. coli* fosfomycin MIC. (B) ESBL-positive *E. coli* nitrofurantoin MIC. MIC = minimum inhibitory concentration, ESBL = extended-spectrum β-lactamase, S = sensitive, I = intermediate, R = resistant.

respectively (**Fig. 1A**). The fosfomycin susceptibility rate was 97.2% (105/108). Fosfomycin susceptibility was significantly lower in isolates from the PICU than those from the GW (81.8% vs. 99.0%, P = 0.027; **Table 2**). The MIC₅₀ and MIC₉₀ of nitrofurantoin against the 108 *E. coli* isolates was 16 µg/mL for both MICs (**Fig. 1B**). The nitrofurantoin susceptibility rate was 96.3% (104/108). Although not significant, nitrofurantoin susceptibility in isolates from the PICU tended to be lower than those from the GW (81.8% vs. 97.9%, P = 0.051; **Table 2**).

ESBL-positive K. pneumoniae antibiotic susceptibility

The MIC₅₀ and MIC₉₀ of fosfomycin for the nine *K. pneumoniae* isolates were 2 μ g/mL and 32 μ g/mL, respectively (**Fig. 2A**). All *K. pneumoniae* isolates were susceptible to fosfomycin; however, only 33% of *K. pneumoniae* isolates were susceptible to nitrofurantoin. The MIC₅₀ and MIC₉₀ for nitrofurantoin in the nine *K. pneumoniae* isolates were 64 μ g/mL and 128 μ g/mL, respectively (**Fig. 2B**).

Table 2. Comparison	of antibiotic susceptibilit	y in E. coli isolated from G	Ws and PICU

Location		Fosfomycin			Nitrofurantoin		
		S	I	R	S	I	R
GW	97 (100)	96 (99.0)	1 (1.0)	0 (0.0)	95 (97.9)	0 (0.0)	2 (2.1)
PICU	11 (100)	9 (81.8)	1 (9.1)	1 (9.1)	9 (81.8)	1 (9.1)	1 (9.1)

Values are presented as number (%).

GW = general ward, PICU = pediatric intensive care unit, S = sensitive, I = intermediate, R = resistant.

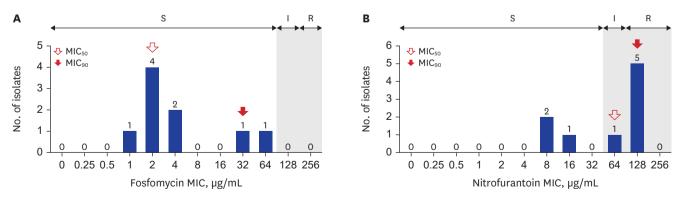


Fig. 2. Distribution of MICs and susceptibility of ESBL-positive K. pneumoniae isolates. (A) ESBL-positive K. pneumoniae fosfomycin MIC. (B) ESBL-positive K. pneumoniae nitrofurantoin MIC.
MIC = minimum inhibitory concentration, ESBL = extended-spectrum β-lactamase, S = sensitive, I = intermediate, R = resistant.

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DISCUSSION

In this study, we investigated fosfomycin and nitrofurantoin susceptibility in ESBL-positive *E. coli* and *K. pneumoniae* isolated from urine in pediatric patients (\leq 18 years). Susceptibility to both fosfomycin and nitrofurantoin in *E. coli* and susceptibility to fosfomycin in *K. pneumoniae* were high, above 95%, while only one-third of *K. pneumoniae* isolates were susceptible to nitrofurantoin. Infection caused by ESBL-positive GNB in children has increased with the increasing use of broad-spectrum antibiotics in recent years.¹⁵ In particular, in cases of pediatric UTIs caused by ESBL-positive *E. coli* and *K. pneumoniae*, broad-spectrum parenteral antibiotic administration is inevitable in many cases, which not only increases hospital length of stay and medical expenses but may result in the emergence of new resistant bacteria. Recently, it has been reported that fosfomycin and nitrofurantoin, which are old antimicrobial agents, are highly effective against GNB isolated from in urine, ^{10,11,16} and interest in these two drugs as a treatment option for uncomplicated UTI has increased.

Several previous studies have reported on ESBL-producing GNB susceptibility to fosfomycin and nitrofurantoin. In a Canadian study, ESBL-producing E. coli isolated from UTI patients (both children and adults included) reported 100% and 83.3% susceptibility to fosfomycin and nitrofurantoin, respectively, from 2010 to 2013.¹⁷ In the U.S., susceptibility of the E. coli ESBL-phenotype to fosfomycin and nitrofurantoin in 2012 was reported to be 98.7% and 82.9%, respectively; however, the susceptibility of 65 Klebsiella ssp. with the ESBL-phenotype (including K, pneumoniae 59 isolates) for nitrofurantoin was very low at 4.6%,¹⁸ The results from a 2015 to 2017 study by the East Sussex Healthcare NHS Trust in England showed that ESBLproducing *E. coli* showed high susceptibility to fosfomycin (98%) and nitrofurantoin (93%), whereas Klebsiella species showed rather low susceptibility at 62% and 58%, respectively.¹⁹ In a study conducted on *E. coli* isolated from adult UTI patients in China, the incidence of ESBLproducing strains ranged from 18% to 43%. The susceptibility to fosfomycin and nitrofurantoin in 332 ESBL-producing isolates (including ESBL-uncertain isolates) was 90.1% and 93.1%, respectively, in 2021.²⁰ In a study of pediatric UTI patients in India, among 271 isolated ESBLpositive strains, E. coli showed the highest proportion with 221 cases (81.6%). The susceptibility to fosfomycin and nitrofurantoin was 98.5% and 95.2% for E. coli and 96.8% and 75.1% for K. pneumoniae, respectively, in 2022 (including both ESBL-positive and ESBL-negative isolates).²¹ The susceptibility of 267 cases of ESBL-producing *E. coli* to fosfomycin isolated from urine in Korea was 99.6%, followed by ciprofloxacin (68.5%), trimethoprim-sulfamethoxazole (55.4%), and ampicillin (31.5%).²² However, it is not clear whether pediatric isolates were included in this study.

In our data, ESBL-positive *E. coli* showed good in vitro susceptibility for both oral fosfomycin and nitrofurantoin, and thus can be reserved for therapeutic candidates. On the other hand, we observed that ESBL-positive *K. pneumoniae* had a rather low susceptibility to nitrofurantoin, suggesting the importance of antimicrobial susceptibility testing before antibiotic choice. According to Infectious Disease Society of America guidance for the treatment of ESBL-positive *Enterobacterales*, single-dose oral fosfomycin is selectively recommended as an alternative option for uncomplicated ESBL-positive *E. coli* cystitis only, since other GNBs including *K. pneumoniae* retain an intrinsic *fosA* gene, the product of which can hydrolyze the drug.^{23,24} Nitrofurantoin on the other hand is suggested as the preferred choice for uncomplicated cystitis caused by general ESBL-positive *Enterobacterales* based on its safety and efficacy, even compared to oral fosfomycin.^{23,25} The guideline suggests that both drugs be avoided for pyelonephritis and complicated UTI because of their low penetration into the renal parenchyma. Intriguingly, recent adult studies have focused on extending multi-dose oral fosfomycin as the treatment option for pyelonephritis or complicated ESBL-positive *E. coli* UTI.^{26,27} Wald-Dickler et al.²⁶ conducted a multicenter retrospective cohort study that compared clinical efficacy of the outpatient regimen of oral fosfomycin with that of standard ertapenem. The authors found no statistical difference between the two groups regarding clinical success rate at 30-day and relapse rate, suggesting oral fosfomycin as a reasonable step-down option for complicated UTIs.

Unfortunately, only a few pediatric studies regarding oral fosfomycin or nitrofurantoin for UTI treatment have compared data to the adult population.^{28,29} Purcell et al.²⁸ reviewed the medical literature from 1946 to 2020 on oral fosfomycin treatment for pediatric UTI and only found eight related studies, most of which were published more than 20 years ago without reporting multidrug-resistant organisms. With recently increasing ESBL UTI cases among the pediatric population, further organized clinical studies for optimal dosing are in need, as supported by our in vitro clinical data.³⁰

According to the most recent Australian multicenter cohort study published in 2023, fosfomycin was effective for pediatric UTI (aged < 1 year: 1g/single dose or two doses, 1-12 years: 2g/single dose, > 12 years: 3 g/single dose).³¹ However, this study also mentions that further studies are needed to confirm the appropriate administration therapy and optimal duration of fosfomycin in pediatric population. There is not much literature on the side effects of fosfomycin. According to a clinical record analysis of the side effects of 35,481 fosfomycin conducted in Japan, the overall incidence of side effects was 3.5%. Side effects were higher when the daily dose exceeded 3 g, gastrointestinal disorders (2.7%) were the most common side effects, and there was one case of thrombocytopenia and one case of anemia.³² In a Turkish study of 50 pediatric patients (age 1-17), nitrofurantoin was found to be effective to treat the lower UTI due to ESBL-positive *E. coli* and no specific adverse effects were observed during the 10-day course of nitrofurantoin treatment (5 to 7 mg/kg/ day, 4 divided doses).²⁹ However, there is a report that pulmonary fibrosis was observed during long-term treatment (several years).³³ Therefore, additional studies are also needed to confirm the appropriate medication therapy and duration of nitrofurantoin.

In this study, the rate of fosfomycin resistance of *E. coli* in the PICU was significantly higher compared to the GW. Fosfomycin acts by inhibiting the initial enzymatic step of peptidoglycan biosynthesis in the cytoplasm. Antibiotic resistance mechanisms of fosfomycin are impaired drug uptake, enzymatic drug inactivation, and target modification. Worldwide spread of fosfomycin-modifying enzymes causing drug inactivation has drawn the attention as the emerging mechanism of resistance. This enzymatic drug inactivation is usually plasmid-mediated and fosA3 is most frequently found. There are several studies that reported fosA3 gene variants expressed with resistance to other antibiotic classes.34-36 In fact, a previous Korean study by Lee et al.³⁷ reported that among the 21 fosfomycin nonsusceptible isolates (E. coli and K. pneumoniae), 7 isolates harbored plasmid mediated fosA3 gene, which was connected to bla_{CTX-M} via insertion sequence, IS26-composite transposon. Therefore, we consider that the higher rate of fosfomycin resistance in PICU may have been influenced by greater exposure to other antibiotics in critically-ill patients in PICU and status of multidrug resistance in some isolates from PICU. However, only 11 E. coli isolates from PICU were included in this study. Therefore, we are cautious to mention any plausible conclusion at this point.

This study has some limitations. First, the study was performed at a single institution, the number of isolates was relatively small at 117, and *K. pneumoniae* representation was very low at only nine isolates. Second, since this study focused on antimicrobial susceptibility tests of *E. coli* and *K. pneumoniae* only, we did not examine the treatment outcome or application of these two drugs in children with uncomplicated UTI. Third, other GNB were not tested.

In conclusion, we examined fosfomycin and nitrofurantoin susceptibility in *E. coli* and *K. pneumoniae* isolated from urine in Korean children. Our study implies that fosfomycin and nitrofurantoin are oral treatment options that can be used in pediatric patients with UTI caused by ESBL-positive *E. coli*.

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SUPPLEMENTARY MATERIAL

Supplementary Table 1

Baseline characteristics of pediatric patients with ESBL-positive GNB recovered from urine samples

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REFERENCES

- Fan NC, Chen HH, Chen CL, Ou LS, Lin TY, Tsai MH, et al. Rise of community-onset urinary tract infection caused by extended-spectrum β-lactamase-producing *Escherichia coli* in children. *J Microbiol Immunol Infect* 2014;47(5):399-405.
 PUBMED | CROSSREF
- Dayan N, Dabbah H, Weissman I, Aga I, Even L, Glikman D. Urinary tract infections caused by community-acquired extended-spectrum β-lactamase-producing and nonproducing bacteria: a comparative study. *J Pediatr* 2013;163(5):1417-21.
 PUBMED | CROSSREF
- Topaloglu R, Er I, Dogan BG, Bilginer Y, Ozaltin F, Besbas N, et al. Risk factors in community-acquired urinary tract infections caused by ESBL-producing bacteria in children. *Pediatr Nephrol* 2010;25(5):919-25.
 PUBMED | CROSSREF
- Saltoglu N, Karali R, Yemisen M, Ozaras R, Balkan II, Mete B, et al. Comparison of community-onset healthcare-associated and hospital-acquired urinary infections caused by extended-spectrum betalactamase-producing *Escherichia coli* and antimicrobial activities. *Int J Clin Pract* 2015;69(7):766-70.
 PUBMED | CROSSREF
- Ladhani S, Gransden W. Increasing antibiotic resistance among urinary tract isolates. Arch Dis Child 2003;88(5):444-5.
 PUBMED | CROSSREF
- Cheng MF, Chen WL, Huang IF, Chen JR, Chiou YH, Chen YS, et al. Urinary tract infection in infants caused by extended-spectrum beta-lactamase-producing *Escherichia coli*: comparison between urban and rural hospitals. *Pediatr Nephrol* 2016;31(8):1305-12.
 PUBMED | CROSSREF
- Lee YK, Lee H, Kim JM, Kang JM, Lee ST, Lee NY, et al. The antibiotic resistance pattern of gram-negative bacteria in children younger than 24 months with a urinary tract infection: a retrospective single-center study over 15 consecutive years. *Child Kidney Dis* 2015;19(2):148-53.
 CROSSREF

- Subcommittee on Urinary Tract Infection, Steering Committee on Quality Improvement and Management, Roberts KB. Urinary tract infection: clinical practice guideline for the diagnosis and management of the initial UTI in febrile infants and children 2 to 24 months. *Pediatrics* 2011;128(3):595-610.
 PUBMED | CROSSREF
- Doi A, Shimada T, Harada S, Iwata K, Kamiya T. The efficacy of cefinetazole against pyelonephritis caused by extended-spectrum beta-lactamase-producing Enterobacteriaceae. *Int J Infect Dis* 2013;17(3):e159-63.
 PUBMED | CROSSREF
- Tasbakan MI, Pullukcu H, Sipahi OR, Yamazhan T, Ulusoy S. Nitrofurantoin in the treatment of extendedspectrum β-lactamase-producing *Escherichia coli*-related lower urinary tract infection. *Int J Antimicrob Agents* 2012;40(6):554-6.
 PUBMED | CROSSREF
- 11. Neuner EA, Sekeres J, Hall GS, van Duin D. Experience with fosfomycin for treatment of urinary tract infections due to multidrug-resistant organisms. *Antimicrob Agents Chemother* 2012;56(11):5744-8.
- European Committee for Antimicrobial Susceptibility Testing (EUCAST) of the European Society of Clinical Microbiology and Infectious Diseases (ESCMID). EUCAST Definitive Document E.DEF 3.1, June 2000: Determination of minimum inhibitory concentrations (MICs) of antibacterial agents by agar dilution. *Clin Microbiol Infect* 2000;6(9):509-15.
 PUBMED | CROSSREF
- Vachvanichsanong P, McNeil EB, Dissaneewate P. Extended-spectrum beta-lactamase *Escherichia coli* and *Klebsiella pneumoniae* urinary tract infections. *Epidemiol Infect* 2020;149:e12.
 PUBMED | CROSSREF
- 14. CLSI. Performance Standards for Antimicrobial Susceptibility Testing, 32nd Edition. CLSI Guideline M100. Wayne, PA, USA: Clinical and Laboratory Standards Institute; 2022.
- Raphael E, Glymour MM, Chambers HF. Trends in prevalence of extended-spectrum beta-lactamaseproducing *Escherichia coli* isolated from patients with community- and healthcare-associated bacteriuria: results from 2014 to 2020 in an urban safety-net healthcare system. *Antimicrob Resist Infect Control* 2021;10(1):118.
 PUBMED | CROSSREF
- Seo MR, Kim SJ, Kim Y, Kim J, Choi TY, Kang JO, et al. Susceptibility of *Escherichia coli* from communityacquired urinary tract infection to fosfomycin, nitrofurantoin, and temocillin in Korea. *J Korean Med Sci* 2014;29(8):1178-81.
 PUBMED | CROSSREF
- Karlowsky JA, Denisuik AJ, Lagacé-Wiens PR, Adam HJ, Baxter MR, Hoban DJ, et al. In vitro activity of fosfomycin against *Escherichia coli* isolated from patients with urinary tract infections in Canada as part of the CANWARD surveillance study. *Antimicrob Agents Chemother* 2014;58(2):1252-6.
 PUBMED | CROSSREF
- Keepers TR, Gomez M, Celeri C, Krause KM, Biek D, Critchley I. Fosfomycin and comparator activity against select Enterobacteriaceae, Pseudomonas, and Enterococcus urinary tract infection isolates from the United States in 2012. *Infect Dis Ther* 2017;6(2):233-43.
 PUBMED | CROSSREF
- Raja NS. Oral treatment options for patients with urinary tract infections caused by extended spectrum βeta-lactamase (ESBL) producing Enterobacteriaceae. J Infect Public Health 2019;12(6):843-6.
 PUBMED | CROSSREF
- Jia P, Zhu Y, Li X, Kudinha T, Yang Y, Zhang G, et al. High prevalence of extended-spectrum betalactamases in *Escherichia coli* strains collected from strictly defined community-acquired urinary tract infections in adults in China: a multicenter prospective clinical microbiological and molecular study. *Front Microbiol* 2021;12:663033.
 PUBMED | CROSSREF
- Perween N, Rai S, Nandwani S, Kumar SK 2nd. Retrospective analysis of urinary tract infection in the pediatric population at a tertiary care centre. *Cureus* 2022;14(5):e24796.
 PUBMED | CROSSREF
- Ko KS, Suh JY, Peck KR, Lee MY, Oh WS, Kwon KT, et al. In vitro activity of fosfomycin against ciprofloxacin-resistant or extended-spectrum beta-lactamase-producing *Escherichia coli* isolated from urine and blood. *Diagn Microbiol Infect Dis* 2007;58(1):111-5.
 PUBMED | CROSSREF
- 23. Tamma PD, Aitken SL, Bonomo RA, Mathers AJ, van Duin D, Clancy CJ. Infectious Diseases Society of America 2022 guidance on the treatment of extended-spectrum β-lactamase producing Enterobacterales (ESBL-E), carbapenem-resistant Enterobacterales (CRE), and *Pseudomonas aeruginosa* with difficult-to-treat resistance (DTR-*P. aeruginosa*). *Clin Infect Dis* 2022;75(2):187-212.
 PUBMED | CROSSREF

- Elliott ZS, Barry KE, Cox HL, Stoesser N, Carroll J, Vegesana K, et al. The role of *fosA* in challenges with fosfomycin susceptibility testing of multispecies *Klebsiella pneumoniae* carbapenemase-producing clinical isolates. *J Clin Microbiol* 2019;57(10):e00634-19.
 PUBMED | CROSSREF
- Huttner A, Kowalczyk A, Turjeman A, Babich T, Brossier C, Eliakim-Raz N, et al. Effect of 5-day nitrofurantoin vs single-dose fosfomycin on clinical resolution of uncomplicated lower urinary tract infection in women: a randomized clinical trial. *JAMA* 2018;319(17):1781-9.
 PUBMED | CROSSREF
- 26. Wald-Dickler N, Lee TC, Tangpraphaphorn S, Butler-Wu SM, Wang N, Degener T, et al. Fosfomycin vs ertapenem for outpatient treatment of complicated urinary tract infections: a multicenter, retrospective cohort study. *Open Forum Infect Dis* 2021;9(1):ofab620.
 PUBMED | CROSSREF
- Hatlen TJ, Flor R, Nguyen MH, Lee GH, Miller LG. Oral fosfomycin use for pyelonephritis and complicated urinary tract infections: a 1 year review of outcomes and prescribing habits in a large municipal healthcare system. *J Antimicrob Chemother* 2020;75(7):1993-7.
- Purcell R, Wang N, Gwee A. Can fosfomycin be used for the treatment of Gram-negative urinary tract infections in children? *Arch Dis Child* 2021;106(9):925-8.
- Kara A, Gurgoze MK. The use of nitrofurantoin for children with acute cystitis caused by extendedspectrum B-lactamase-producing *Escherichia coli*. J Pediatr Urol 2019;15(4):378.e1-378.e5.
 PUBMED | CROSSREF
- Murray TS, Peaper DR. The contribution of extended-spectrum β-lactamases to multidrug-resistant infections in children. *Curr Opin Pediatr* 2015;27(1):124-31.
 PUBMED | CROSSREF
- Purcell R, Yeoh D, Bowen A, Britton PN, Carr JP, Chen M, et al. A multicentre, retrospective audit of fosfomycin use for urinary tract infections in Australian children and adolescents. *J Antimicrob Chemother* 2023;78(7):1616-21.
 PUBMED | CROSSREF
- 32. Mayama T, Yokota M, Shimatani I, Ohyagi H. Analysis of oral fosfomycin calcium (Fosmicin) side-effects after marketing. *Int J Clin Pharmacol Ther Toxicol* 1993;31(2):77-82.
- Mikkelsen LF, Rubak S. Reversible lung fibrosis in a 6-year-old girl after long term nitrofurantoin treatment. *BMC Pulm Med* 2020;20(1):313.
 PUBMED | CROSSREF
- 34. Falagas ME, Vouloumanou EK, Samonis G, Vardakas KZ. Fosfomycin. *Clin Microbiol Rev* 2016;29(2):321-47. PUBMED | CROSSREF
- Yang TY, Lu PL, Tseng SP. Update on fosfomycin-modified genes in Enterobacteriaceae. J Microbiol Immunol Infect 2019;52(1):9-21.
 PUBMED | CROSSREF
- 36. Cattoir V, Guérin F. How is fosfomycin resistance developed in *Escherichia coli? Future Microbiol* 2018;13:1693-6. PUBMED | CROSSREF
- 37. Lee SY, Park YJ, Yu JK, Jung S, Kim Y, Jeong SH, et al. Prevalence of acquired fosfomycin resistance among extended-spectrum β-lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* clinical isolates in Korea and IS26-composite transposon surrounding fosA3. *J Antimicrob Chemother* 2012;67(12):2843-7.
 PUBMED | CROSSREF