

Review

Tafenoquine for travelers' malaria: evidence, rationale and recommendations

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Abstract

Background: Endemic malaria occurring across much of the globe threatens millions of exposed travelers. While unknown numbers of them suffer acute attacks while traveling, each year thousands return from travel and become stricken in the weeks and months following exposure. This represents perhaps the most serious, prevalent and complex problem faced by providers of travel medicine services. Since before World War II, travel medicine practice has relied on synthetic suppressive blood schizontocidal drugs to prevent malaria during exposure, and has applied primaquine for presumptive anti-relapse therapy (post-travel or post-diagnosis of *Plasmodium vivax*) since 1952. In 2018, the US Food and Drug Administration approved the uses of a new hepatic schizontocidal and hypnozoitocidal 8-aminoquinoline called tafenoquine for the respective prevention of all malarias and for the treatment of those that relapse (*P. vivax* and *Plasmodium ovale*).

Methods: The evidence and rationale for tafenoquine for the prevention and treatment of malaria was gathered by means of a standard search of the medical literature along with the package inserts for the tafenoquine products Arakoda™ and Krintafel™ for the prevention of all malarias and the treatment of relapsing malarias, respectively.

Results: The development of tafenoquine—an endeavor of 40 years—at last brings two powerful advantages to travel medicine practice against the malaria threat: (i) a weekly regimen of causal prophylaxis; and (ii) a single-dose radical cure for patients infected by vivax or ovale malarias.

Conclusions: Although broad clinical experience remains to be gathered, tafenoquine appears to promise more practical and effective prevention and treatment of malaria. Tafenoquine thus applied includes important biological and clinical complexities explained in this review, with particular regard to the problem of hemolytic toxicity in G6PD-deficient patients.

Key words: Malaria, prevention, treatment, travelers, primaquine, tafenoquine, G6PD deficiency**Introduction**

Each of the five species of malaria-causing plasmodial parasites naturally infecting humans often progress to threatening clinical syndromes in malaria-naïve patients unless prompt diagnosis and appropriate therapy first occurs. Death as an outcome of infection is confirmed in all of these species: *Plasmodium falciparum*, *Plasmodium vivax*, *Plasmodium malariae*, *Plasmodium ovale* and *Plasmodium knowlesi*.^{1–5} Infections by some species may more rapidly and frequently progress to serious illness than others, but malaria in all its forms provokes a debilitating febrile

illness posing a potentially mortal threat in non-immune patients.⁶ The notion of intrinsically benign or malignant species of the plasmodia should be acknowledged as dangerous dogma and the diagnosis of any malaria managed as a clinical emergency.⁷ Successfully preventing such emergencies in travelers merits the relatively complex and difficult clinical task of doing so practically and effectively.

Naturally acquired immunity, community-based measures of prevention and control, along with local access to competent healthcare provided by malaria-aware governments, together

greatly mitigate the harm caused by these parasites in endemic areas.^{8–10} In contrast, protection of relatively vulnerable travelers almost wholly depends on the recommendations and practices of travel medicine providers—local protections for them effectively do not exist beyond the passive benefit of reduced transmission and risk. Among the agencies and experts offering the distinct advice to travelers and residents alike, strategic thinking has historically been focused on the species once known as ‘malignant tertian malaria’, *P. falciparum*. In contrast, ‘benign tertian malaria’, *P. vivax*, was deeply neglected, and the tools and advice for its prevention, treatment or control were inadequate.^{11–14} In 2015, the World Health Organization (WHO) acknowledged the mortal risk of vivax malaria and the neglect of it in public health and clinical medicine.¹⁵

A great deal of recent work and progress begins to correct the problem of neglect of vivax malaria in endemic communities,¹⁶ but travel medicine strategy and practices remain aimed principally at falciparum malaria.^{17–19} Up to the present day, suppressive chemoprophylaxis applying blood schizontocidal drugs dominates travel medicine practice.²⁰ A fundamental biological distinction between falciparum and vivax malarias—dormant liver stages called hypnozoites present in the latter and absent in the former—explains the inadequacy of suppressive chemoprophylaxis alone against the malarias.^{21–23} Latent malaria and the threat of relapse require additional (post-travel presumptive anti-relapse therapy (PART)) or alternative (causal prophylaxis) approaches to chemoprevention.

Two regulatory events in the USA in 2018 offer potentially transformative changes in how travel medicine deals with the malaria threat.²⁴ The Food and Drug Administration (FDA) approved a new 8-aminoquinoline drug called tafenoquine for uses in the treatment or prevention of malaria: Krintafel™ (GlaxoSmithKline®, USA) or Arakoda™ (60 Degrees Pharmaceuticals® LLC, USA), respectively (Figure 1). The US Army discovered tafenoquine in

1978 during an era of historic neglect of antimalarial drug development^{25,26} relative to the comparatively vigorous current efforts.²⁷ Tafenoquine thus lingered through fits and starts of clinical development in the three decades that followed.²⁸ Approximately 10 years ago, dawning realization of the clinical and public health importance of vivax malaria helped spur commitment to making tafenoquine available for use (Bill and Melinda Gates Foundation, Medicines for Malaria Venture and GSK).^{29,30}

Complex biology governs the rationale underpinning safe and appropriate use of tafenoquine in travel medicine. The class effect of hemolytic toxicity in patients having the X-linked trait of glucose-6-phosphate dehydrogenase (G6PD) deficiency substantially deepens the complexity of its use. This review aims to explain these complexities along with the evidence and rationale for potential roles of tafenoquine for the prevention or treatment of malaria.

Essential Biology

The life cycles of the plasmodia guide chemotherapeutic and chemopreventive strategies. The many stages of them are variably susceptible to antimalarial classes of drugs (Figure 2), most having class-specific therapeutic effects. Clinically applied blood schizontocidal drugs, for example, have no hypnozoitocidal activity. Nonetheless, cross-class effects among antimalarials occur, sometimes species-specific in manner; e.g. the blood schizonticide chloroquine also exerts gametocytocidal activity in *P. vivax* but not *P. falciparum*.³¹ Tafenoquine may be unique among registered antimalarial compounds in having demonstrable activity among all classes of antimalarials.^{32,33}

All malarias derive from the bite of infectious anopheline mosquitos (excepting congenital or transfusion/transplant malarias). Injected plasmodial tachysporozoites invade hepatic cells, multiply as hepatic schizonts and after a week or more

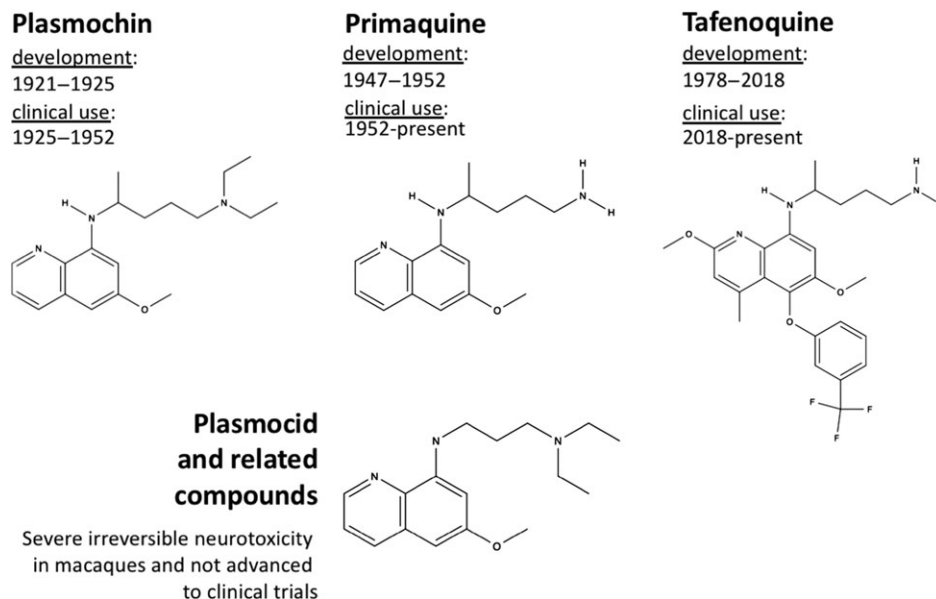


Figure 1. Evolution of the 8-aminoquinoline hypnozoitocides, including the winnowing out of irreversible severe neurotoxicity of plasmocid and related compounds distinguished by fewer than four methylene groups separating the amino groups of the alkyl chain at the defining 8-amino position. Plasmochin and others (including primaquine) having at least four methylene groups exhibited no such neurotoxicity but instead reversible toxicity at sub-lethal doses involving principally hepatic, hematological and gastrointestinal systems

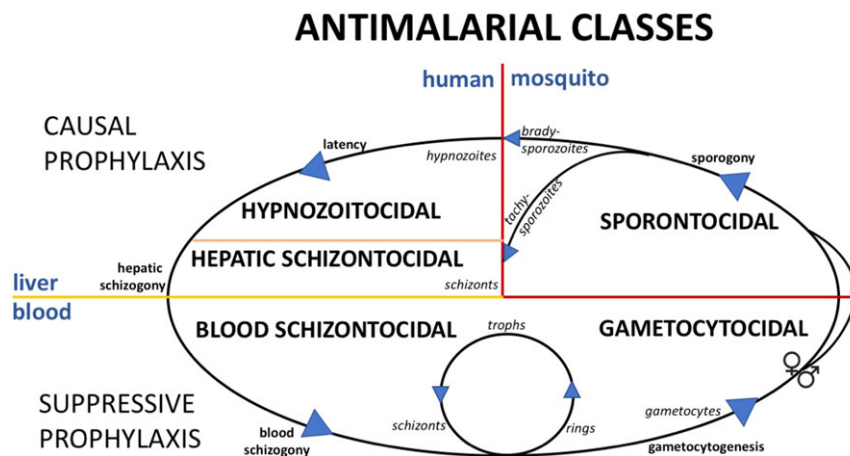


Figure 2. Antimalarial classes as guided by life cycle of the plasmodia

Table 1. Characteristics of relapsing and non-relapsing malarias

	Relapsing	Non-relapsing
Species	<i>P. vivax</i> , <i>P. ovale</i> , <i>P. cynomolgi</i> ^a	<i>P. falciparum</i> , <i>P. malariae</i> , <i>P. knowlesi</i> ^b
Hypnozoites	Present	Absent
Clinical attacks/infection	Variable, typically >3	1
Curative therapy	Blood schizontocidal, Hypnozoitocidal	Blood schizontocidal Gametocytocidal ^c
Suppressive chemoprophylaxis	Ineffective against relapses occurring post-chemoprophylaxis	Effective
Post-travel presumptive anti-relapse therapy	Not indicated after causal prophylaxis but necessary after suppressive prophylaxis	Not indicated
Causal chemoprophylaxis	Effective	Effective

^aA natural zoonosis of Southeast Asian macaques confirmed in only a single patient but perhaps more common than now appreciated.

^bA natural zoonosis of Southeast Asian macaques confirmed in thousands of patients.

^cA single dose of 0.25 mg/kg primaquine to prevent onward transmission. Not recommended in relapsing malarias because hypnozoitocidal therapy also gametocytocidal.

emerge as infectious merozoites into the bloodstream where they again multiply asexually (schizogony) in red blood cells. Repeated cycles of that reproduction provoke the non-specific cyclic symptoms of acute malaria; typically daily bouts of spiking fever and shaking chills, often accompanied by headache, nausea, vomiting and myalgia. Some of those parasites become circulating sexual forms called gametocytes that may infect feeding anophelines but provoke no illness.

The infective bite of the relapsing malarias, *P. vivax* and *P. ovale*, includes brady-sporozoites that become latent hepatic hypnozoites. The timing of their activation to hepatic schizogony and subsequent clinical attacks varies greatly, between a month and several years after infection. In general, attacks occurring less than a month after infection derive from tachysporozoite-borne active hepatic schizonts, whereas after 1 month attacks probably derive from the delayed hepatic schizogony of brady-sporozoite-borne activated hypnozoites. These clinical events are called primary attacks and relapses.

The malarias infecting humans may be divided into relapsing and non-relapsing species, i.e. *P. vivax* and *P. ovale*, and *P. falciparum*, *P. malariae*, and *P. knowlesi*, respectively (Table 1). This fundamental distinction defines essential features of the treatment of the malarias; therapy of non-relapsing acute malarials involves only blood schizontocidal drugs (and gametocytocidal single-dose

primaquine not considered here), whereas that of the relapsing malarials includes a hypnozoiticide. Strategy for the prevention of the malarials also invokes non-relapsing and relapsing biology and antimalarial drug classes; suppressive chemoprophylaxis employs blood schizontocides against asexual reproduction in blood, whereas causal chemoprophylaxis applies hepatic schizontocides or hypnozoitocides in killing parasites before they mature to either hepatic schizonts or hypnozoites (Figure 2). Widely used suppressive chemoprophylactic drugs do not interfere with hepatic development, with the exception of the causal activity of atovaquone against hepatic schizonts of *P. falciparum*^{34,35} but not against hypnozoites of *P. vivax*^{36,37} or those of *Plasmodium cynomolgi* in rhesus macaques.³⁸

This review specifically considers the role of the new 8-aminoquinoline called tafenoquine in travel medicine practice. In terms of chemotherapy, only the relapsing malarials and hypnozoitocidal activity are relevant here. On the other hand, chemoprevention engages all malarials and activity against the hepatic stages of any plasmodial species, be those active schizonts, latent hypnozoites, or, more probably, their respective earliest (<48 h) post-invasion forms.³⁹ The broad spectrum activity of tafenoquine includes relatively potent blood schizontocidal effects,⁴⁰ but its clinical use as such is not recommended.

Rationale for prioritized causal prophylaxis

Suppressive chemoprophylaxis of malaria with blood schizontocides like quinine, atabrine, chloroquine, doxycycline, mefloquine and atovaquone–proguanil has successively dominated practice in travel medicine for over a century.^{41,42} This strategy served the intended purpose of effectively preventing attacks by what had been considered the only intrinsically dangerous species, *P. falciparum*. The inadequacy of chemoprophylactic suppression alone against the delayed attacks of the relapsing malarias has long been understood and thoroughly demonstrated.^{43–46} Though not always prescribed or even recommended,²⁰ post-travel PART using hypnozoitocidal primaquine addressed that inherent inadequacy. However, that practice also imposed G6PD-deficiency risk management, along with the inconvenience and adherence issues of 14 daily doses. Some authorities and experts have recommended daily primaquine (0.5 mg/kg) during exposure under some circumstances as safe, well-tolerated and effective causal prophylaxis (in non-pregnant, G6PD-normal travelers),⁴⁷ but with the important drawback of off-label use. Further, primaquine having poor activity against the asexual blood stages of *P. falciparum*⁴⁸ raises the specter of unmitigated prophylaxis breakthroughs. Primaquine as primary causal prophylaxis has thus not been widely adopted in travel medicine.

While chemoprophylaxis of any sort against significant risk of malaria imposes some obstacles and pitfalls, it is certainly preferred over no protection and may be less problematic than standby emergency self-treatment practices.^{49–51} Figure 3 illustrates the practical protections and pitfalls of suppressive prophylaxis against non-relapsing (upper panel) and relapsing malarias (lower panel) relative to those of causal prophylaxis. The failure to properly load suppressive dosing before travel or to continue dosing sufficiently long after travel results in attacks during and after travel in both types of malarias. Fully compliant loading and post-exposure suppressive dosing successfully prevents non-relapsing but not relapsing malaria attacks delayed after travel. Causal prophylaxis during exposure (loading or post-exposure dosing is minimal), in contrast, effectively prevents both types of malarias. Causal prophylaxis exceeds suppressive approaches in terms of simplicity of use and thoroughness of protection, but the good efficacy of fully compliant suppressive prophylaxis against *P. falciparum* has been broadly accepted as the standard-of-care in travel medicine.

Acute falciparum malaria is unquestionably a dangerous infection that may rapidly progress to complicated and severe disease syndromes in malaria-naïve patients. It does so in travelers more often than the other plasmodia,⁵² with the possible exception of *P. knowlesi*.⁵³ However, the notion of *P. falciparum* as the only species capable of such harm has been discredited with evidence, much of it only recently gathered.^{2–5,54–59} When the malarias are allowed to progress to severe and complicated disease in travelers, the frequency of death among them appears essentially equal, ~5–10%.⁵² All of the plasmodia are intrinsically dangerous and potentially lethal. Chemoprophylaxis strategy aimed at some species but not others, unless absolutely necessary, fails reason and many patients. Broad spectrum chemoprophylaxis against attacks by any plasmodial species, be those primary or relapsing, would potentially offer a conspicuously superior option.

The fact that *P. falciparum* acquired in Africa indeed causes most (~70%) malaria in travelers^{19,60,61}—a problem solved by

appropriate suppressive chemoprophylaxis—tends to obscure the broader geographic dominance of *P. vivax*. Excepting relatively few and minor geographic areas (e.g. Haiti), endemic transmission of *P. vivax* occurs wherever *P. falciparum* occurs, including much of malarious Africa.^{62–64} Endemic transmission of *P. vivax* extends well beyond the tropical range of *P. falciparum* (e.g. to the Korean Peninsula).⁶⁵ Once travelers are deemed to be in need of chemoprevention against malaria by estimated weight of risk of exposure,^{66,67} most of them will be at risk of infection by the hypnozoites of *P. vivax*, *P. ovale* or both (Figure 4). There may thus be few travelers not benefiting from an approach to chemoprophylaxis that prevents the formation of latent hypnozoites and post-travel attacks.

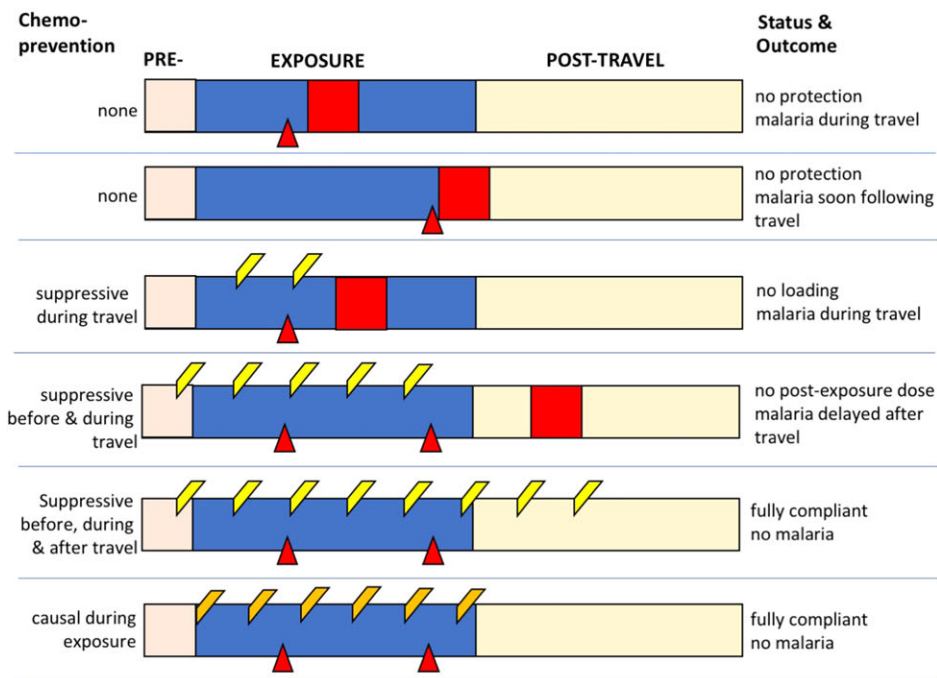
The availability of tafenoquine offers the critical strategic advantages of causal prophylaxis, along with practical advantages over primaquine for that indication. Tafenoquine overcomes three of the four key disadvantages of primaquine in comparison to most suppressive prophylaxis options: (i) chemoprophylaxis is an approved indication; (ii) dosing is weekly rather than daily; and (iii) blood schizontocidal activity may mitigate prophylaxis breakthroughs. The relatively very long plasma half-life of tafenoquine relative to primaquine (~15 days vs 6 h) confers many of its advantages. The key disadvantage is the 8-aminoquinoline liability of hemolytic toxicity in G6PD-deficient patients, and that problem is deepened by slow excretion. The safe use of tafenoquine or primaquine is nonetheless manageable by understanding G6PD deficiency and its diagnosis.

G6PD deficiency

The inherited X chromosome-linked G6PD deficiency trait is the most common human genetic abnormality and its genotypes and frequencies vary tremendously.⁶⁸ It tends to be absent in Native Americans, present at low frequencies (<1%) among most Caucasians and prevalent among people residing in malaria-endemic nations (averaging 8%).⁶⁹ The extent of harm caused by daily primaquine as hypnozoitocide depends on dose, the variant of G6PD deficiency involved, and whether hemi-, homo- or heterozygous.⁷⁰ Effects range from relatively mild and self-limiting to life-threatening. Caucasian, Middle Eastern and Asian peoples tend to have the most severely impaired G6PD deficiency variants.⁷¹ In moderately deficient (40–60% of normal activity) G6PD-deficient heterozygous females having the moderately impaired Asian Mahidol variant, a single 300-mg dose of tafenoquine proved slightly more hemolytic (nadir of ~23% Hb drop) than a 14-day daily regimen of 15-mg primaquine in that trial (~16% drop)⁷² or others (~13% Hb drop).⁷³ Prescribing tafenoquine for any indication requires ruling out any G6PD deficiency, excepting female heterozygotes having >70% of normal activity.

Conventional qualitative screening for G6PD deficiency prior to tafenoquine use may not suffice and quantitative testing is indicated by standard laboratory spectrophotometric assay. Patients having <70% of normal G6PD activity may not receive tafenoquine.⁷⁴ Qualitative screening, for example by the NADPH fluorescent spot test (FST) or newly available point-of-care rapid diagnostic tests for G6PD deficiency (RDT), lack sensitivity to deficiency above 30% of normal activity.^{75–77} Although qualitative screening offers nearly 100% sensitivity and specificity for male hemizygotes, female

Exposed to Non-Relapsing Malaria



Exposed to Relapsing Malaria

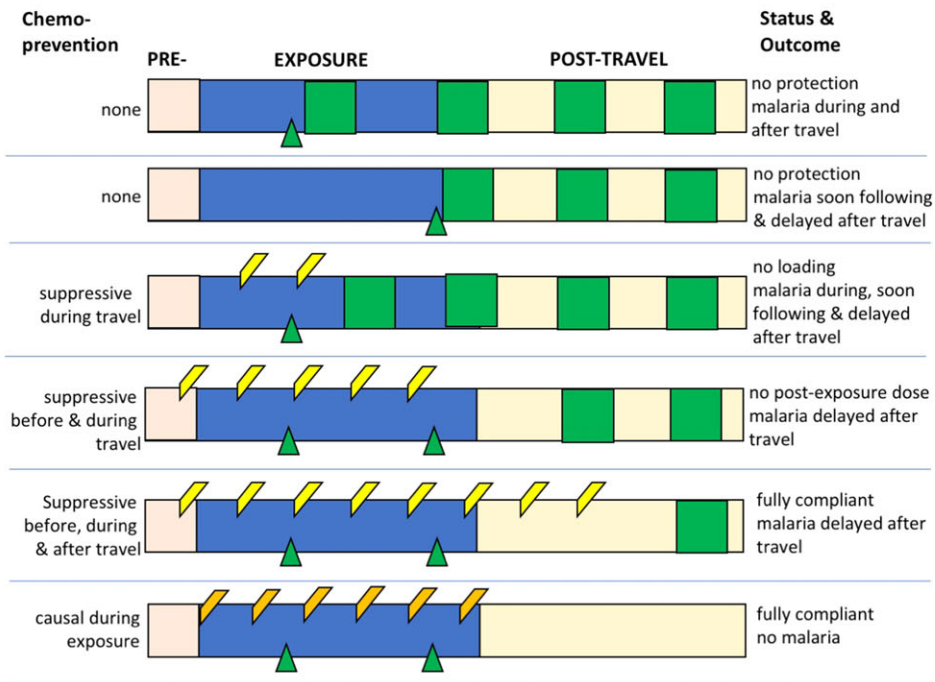


Figure 3. Schematic illustrating pitfalls and protections of suppressive (yellow dose indicators) or causal (orange dose indicators) chemoprevention of non-relapsing malaria like *P. falciparum* (top panel; red triangles and squares for inoculation and attack, respectively) or relapsing species like *P. vivax* (bottom panel; green triangles and squares)

homozygotes and female heterozygotes having <30% of normal activity,^{78,79} the latter having 30–70% of normal G6PD activity will often screen as normal.⁸⁰ The basis of this problem lies in the phenomenon of lyonization during embryonic development of female heterozygotes resulting in apparently random frequencies of

active/inactive normal vs abnormal X-chromosomes and red blood cell mosaicism for G6PD deficiency.⁷⁶ Recent efforts to develop simple and practical quantitative point-of-care test technologies may soon bear devices that greatly increase access to such testing and safe use of 8-aminoquinolines.⁸¹

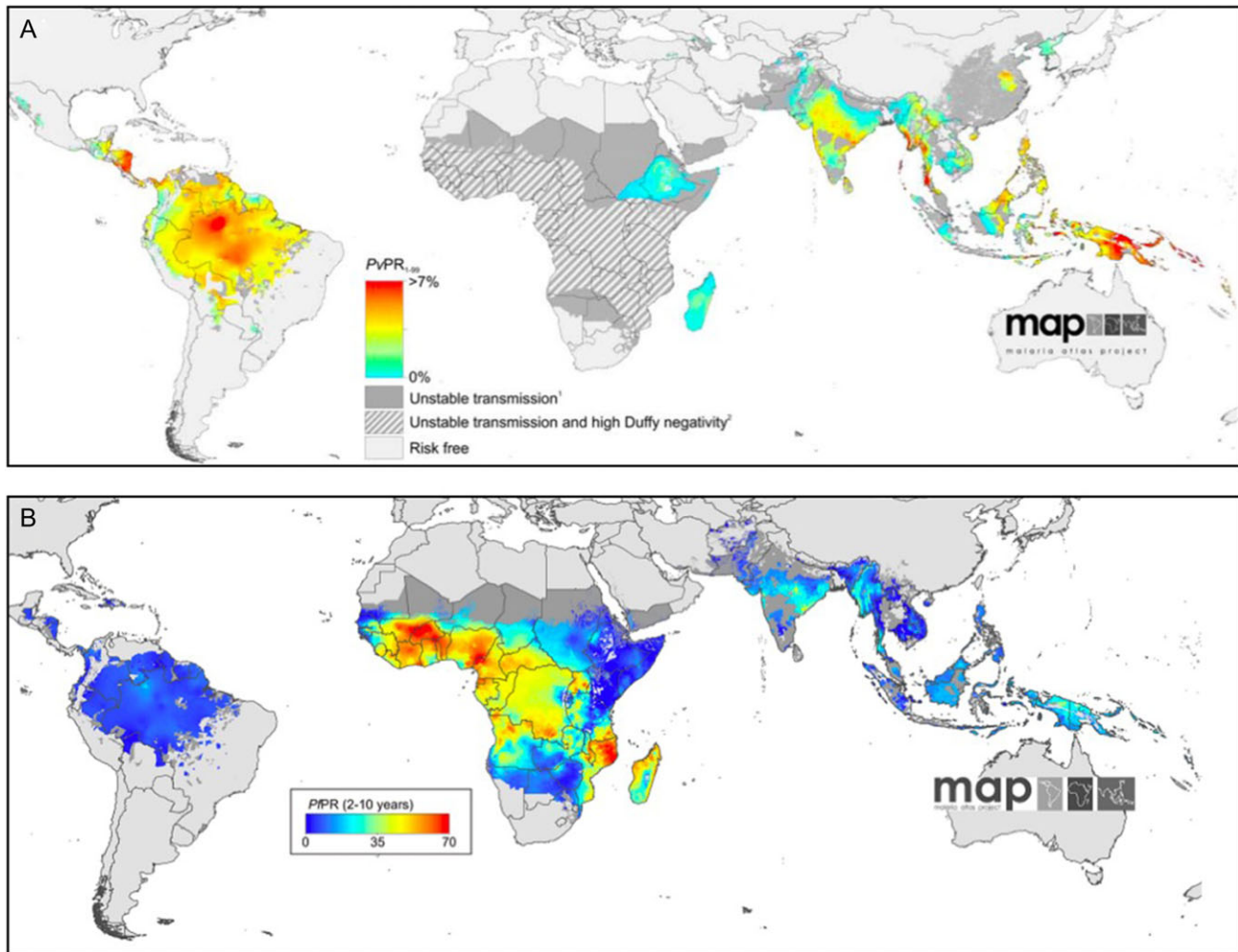


Figure 4. Geographic distribution and prevalence of *P. vivax* (A) and *P. falciparum* (B) in 2010^{65,120} reproduced here under Creative Commons license

Impaired CYP2D6 metabolism

Clinical and laboratory evidence suggested that the efficacy of primaquine may depend on natural variation in cytochrome P-450 2D6 (CYP2D6) isotype activity.^{82–84} In a trial of 177 Indonesian patients with vivax malaria given directly observed high-dose primaquine (0.5 mg/kg/day for 14 day) as PART in combination with artesunate, artesunate–pyronaridine or dihydroartemisinin–piperazine, 26 (15%) experienced relapses during 1 year of follow-up free of reinfection risk.⁸⁵ Among the 21 relapsing subjects evaluated for CYP2D6 genotype and dextromethorphan metabolism phenotype, 20 exhibited significantly impaired CYP2D6 activity.⁸⁶ Relatively common impaired CYP2D6 alleles like *10 (in Asian people) coupled with other less frequent impaired alleles (e.g. *4, *5 or *41) appeared to explain most therapeutic failures despite otherwise adequate dosing.

Although tafenoquine activity against rodent hepatic schizonts seems to also depend on CYP2D6 activity,⁸⁷ one randomized multi-center trial did not detect an association of CYP2D6 genotypes with tafenoquine efficacy (but did with the primaquine comparator arm).⁸⁸ The efficacy of tafenoquine in humans is not known to require metabolism by CYP2D6 or any other cytochrome P-450 isotype or monoamine oxidase, but this

body of evidence is as yet far from thorough or conclusive. Tafenoquine activity may or may not come with the liability of CYP2D6 dependency—decisive studies are needed to inform this important question.

Weekly tafenoquine for causal prophylaxis

Tafenoquine was registered with the US FDA under the trade-name Arakoda™ by 60 Degrees Pharmaceuticals® (USA) in 2018 with a labeled indication for chemoprevention of malaria in adult patients (≥18 year) confirmed to be G6PD-normal (>70% of normal activity) and not pregnant, lactating or having a history of psychoses.⁷⁴ The drug is available as tablets containing 100 mg base. A loading dose of 200 mg tafenoquine daily for 3 days during the week before travel is recommended, followed by weekly maintenance doses of 200 mg commencing 7 days after the last loading dose. Upon return from travel, the final dose should occur 7 days after the last maintenance dose taken in the malarious area.⁷⁴

The label for Arakoda™ includes an indication for ‘terminal prophylaxis’, an antiquated term for post-travel PART in connection with suppressive prophylaxis during travel.⁸⁹ The term is not particularly apt for tafenoquine as Arakoda™ because it

is no more than a final weekly dose after travel rather than the distinct dosing for PART with tafenoquine (i.e. 300 mg rather than 200 mg). Post-travel PART, i.e. terminal prophylaxis, is not necessary with tafenoquine (or primaquine) causal prophylaxis. On the other hand, when suppressive chemoprophylaxis is used and post-travel PART is indicated, tafenoquine as a single 300-mg dose may suffice in lieu of 14 days of primaquine (Table 2).

The clinical experience with 200-mg weekly tafenoquine prophylaxis is now limited to trials conducted in 462 non-immune subjects naturally exposed to falciparum and vivax malaria in Southeast Asia⁹⁰; 152 semi-immune subjects exposed to falciparum malaria in holoendemic sub-Saharan Africa^{91,92} and 12 non-immune, malaria-naïve volunteers experimentally challenged with blood stages of *P. falciparum*.⁷⁴ Comparators in these trials included mefloquine (with or without post-travel PART with primaquine) or placebo (Table 3). There was no placebo control in Trial 1 (Australian soldiers in Timor Leste), but a comparator of weekly mefloquine followed by post-travel PART with primaquine; four post-exposure attacks occurred among subjects taking tafenoquine, and one also occurred in that period among mefloquine-treated subjects. Another analysis of this trial mathematically derived a hypothetical malaria attack rate (8%) and estimated 100% protective efficacies of tafenoquine or mefloquine against primary attacks.⁹³ The placebo-controlled trial of tafenoquine prophylaxis in Kenyan adults⁹¹ showed 86% protective efficacy during 15 weeks of heavy exposure to risk of *P. falciparum* (Trial 2, Table 3). Another trial in Ghana also included a placebo control but with a mefloquine comparator (Trial 3, Table 3):⁹² after 12 weeks the protective efficacy of tafenoquine or mefloquine was 87% for each for *P. falciparum*. A separate analysis of these African trials estimated 94% and 95% protective efficacies for tafenoquine and mefloquine, respectively.⁹⁴ The African studies did not assess efficacy against late attacks by relapsing malarial. Shanks⁹⁵ explained the limitations and obstacles to conducting chemoprophylaxis trials. While head-to-head trials of the chemoprophylactic options against primary and delayed attacks would be ideal, they are also unlikely to be possible.

No clinical trial of tafenoquine has definitively demonstrated a causal vs suppressive prophylaxis mechanism. An early human challenge trial demonstrated a single 600 mg dose of tafenoquine successfully prevented *P. falciparum* in three of four subjects challenged.⁹⁶ At such a dose, slowly eliminated tafenoquine would have exerted blood schizontocidal activity over the normal

incubation period of *P. falciparum* (i.e. less than several weeks) if hepatic schizontocidal activity (causal) had been inadequate. Nonetheless, given the proven causal activity of primaquine against acute *P. falciparum* and acute or latent *P. vivax* malaria,³⁹ the structural relatedness of primaquine to tafenoquine (Figure 1), and evidence from an experiment in rhesus macaques challenged with *P. cynomolgi* sporozoites,³⁸ a causal mechanism of prophylaxis very likely pre-empts the suppressive activity of tafenoquine. Nonetheless, some workers argue that tafenoquine prophylaxis may include a significant suppressive activity component.⁹⁷ A randomized, placebo-controlled trial at Gabon measured the durability of post-treatment prophylaxis of tafenoquine at variable daily doses administered for only 3 days: after 77 days, 14 of 82 placebos experienced *P. falciparum*, whereas 16/79, 3/86, 1/79 and 0/84 subjects did with daily doses of 31.25, 62.5, 125 and 250 mg tafenoquine, respectively.⁹⁸ Such protection very long after dosing logically hints at suppressive prophylaxis, but this is not relevant with weekly tafenoquine dosing. Efficacious monthly dosing of tafenoquine during long-term travel, perhaps exploiting both causal and suppressive activities, may yet be demonstrated.

The label for Arakoda™ warns that adverse reactions may be delayed in onset or prolonged in duration due to the relatively very long plasma half-life of tafenoquine.⁹⁹ The listed warnings and precautions include hemolytic anemia, G6PD deficiency in pregnancy and lactation, methemoglobinemia, psychiatric effects and hypersensitivity reactions. An integrated safety analysis by the developers of Arakoda™ reported that diarrhea, nausea, vomiting, sinusitis, gastroenteritis and back/neck pain occurred at higher frequencies ($\geq 1\%$) relative to placebo; only the latter two occurring at $>5\%$.¹⁰⁰ Two trials followed up on the observed high rate (93%) of mild reversible vortex keratopathy and retinal abnormalities (39%) in the subjects of the trial in Southeast Asia and Australia⁹¹ and reported no concerns with regard to functional visual impairment.^{101,102} The 6-month limitation on tafenoquine prophylaxis in the Arakoda™ label stems from a lack of data rather than any indication of harm beyond that period. Necessity in practice with tafenoquine will likely extend that exposure period, and the reporting of adverse events in practice will later inform evidence-based limitations of use (<https://www.fda.gov/safety/MedWatch/default.htm>).

For most G6PD-normal, non-pregnant adult travelers at substantial risk of any malaria, weekly tafenoquine as causal prophylaxis provisionally (pending greater clinical experience with it) offers a superior option to either causal daily primaquine or any

Table 2. Chemoprophylactic strategies and agents

	Chemoprophylaxis strategy				
	Suppressive			Causal	
Agent	Mefloquine	Doxycycline	Atovaquone-proguanil	Primaquine	Tafenoquine
Dosing	Weekly	Daily	Daily	Daily	Weekly
Post-exposure PART required	Yes	Yes	Yes	No	No
Pregnancy	Yes	No	No	No	No
G6PD-deficient safety	Yes	Yes	Yes	No	No
Children	Yes	No	Yes	Yes	Insufficient evidence
Parasite resistance	Yes	Yes	Yes	No	Improbable
CYP-dependent	No evidence	No evidence	No evidence	Yes	Insufficient evidence

Table 3. Human trials of 200 mg weekly tafenoquine for prophylaxis against malaria

	Trial 1	Trial 2	Trial 3	Trial 4
Location	Timor Leste/Australia	Kenya	Ghana	Australia
Exposure	6mo meso-endemic <i>P. falciparum</i> and <i>P. vivax</i> ; 6mo post-exposure	15 weeks exposure to holoendemic <i>P. falciparum</i>	12 weeks exposure to holoendemic <i>P. falciparum</i>	Experimental <i>P. falciparum</i> blood stages
Subjects	Australian soldiers	Resident adults	Resident adults (excluding reproductive age females)	Malaria-naïve adults
Number of subjects and arms ^a	TQ = 462 MQ + PQ = 153	TQ = 61 Placebo = 62	TQ = 91 MQ = 46 Placebo = 94	TQ = 12 Placebo = 4
Protective Efficacy	Not estimable without placebo; 5 attacks occurred, all post-exposure; 4 in TQ group	86%	TQ = 87% MQ = 87%	100%
Reference	87	88	89	71

^aTQ, tafenoquine administered weekly 200 mg; MQ, mefloquine administered weekly 250 mg; PQ, primaquine administered daily 30 mg for 14 days immediately following travel.

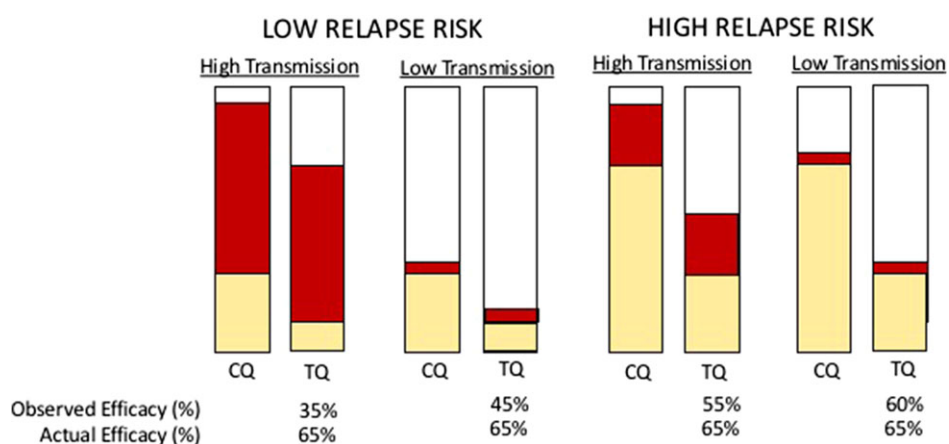


Figure 5. Hypothesized relative attack rates in the months following radical cure illustrate possible impacts of variable risks of relapse or reinfection on the estimation hypnozoitocidal efficacy of tafenoquine (TQ) fixed at a presumed 'actual' 95% rate compared to a chloroquine (CQ) arm without hypnozoitocidal therapy (relapse and reinfection attacks)

suppressive malaria prophylactic regimen (weekly or daily with or without post-travel PART). It is compatible with both short-notice or short-duration travel and particularly favored where endemic vivax or ovale malaria transmission occurs. Mainstream use of tafenoquine for the prevention of malaria in travelers offers a potential solution to the problem of delayed attacks by the relapsing malaras.

Single-dose tafenoquine for radical cure of relapsing malaria

The introduction of tafenoquine into practice as a hypnozoitocidal 8-aminoquinoline requires examination of the therapeutic principles at work. The primaquine standard-of-care, problematic as it may be, defines those with decades of experience and many millions of patients.^{103–106} Primaquine nonetheless imposes the

SUMMARY BOX 1. KEY POINTS ON TAFENOQUINE PROPHYLAXIS IN TRAVEL MEDICINE

- Suppressive malaria prophylaxis standard-of-care is not adequate to the threat of delayed attacks after travel by the relapsing malaras.
- Relapsing malaras occur wherever there is falciparum malaria, with few and minor exceptions.
- Causal prophylaxis is effective against all malaras and prevents delayed attacks after travel.
- Causal prophylaxis is suitable for both short-notice and short-duration travel.
- Tafenoquine is a new drug that offers the advantages of causal prophylaxis with a weekly dosing regimen.
- Tafenoquine is hemolytically toxic to patients having inherited G6PD deficiency, so is prohibited in those patients along with pregnant and lactating women. Safety in children is not yet established.

Table 4. Randomized clinical trials of tafenoquine for PART against vivax malaria

	Trial 1	Trial 2	Trial 3
Location	Multi-centers in Asia, Africa, and Americas	Multi-centers in Asia, Africa and Americas	Multi-centers in Asia, Africa and Americas
Subjects	Adult non-pregnant G6PD-normal residents with acute vivax malaria	Adult non-pregnant G6PD-normal residents with acute vivax malaria	Adult non-pregnant G6PD-normal residents with acute vivax malaria
Treatment arms, and numbers of subjects ^a	TQ + CQ = 57 PQ + CQ = 50 Placebo + CQ = 54	TQ + CQ = 260 PQ + CQ = 129 Placebo + CQ = 133	TQ + CQ = 166 PQ + CQ = 85
% Recurrence-free after 6 months	TQ + CQ = 89 PQ + CQ = 77 Placebo + CQ = 38	TQ + CQ = 62 PQ + CQ = 70 Placebo + CQ = 28	TQ + CQ = 73 PQ + CQ = 75
Reference	105	106	106

^aTQ, 300 mg single dose tafenoquine; CQ, 1500 mg chloroquine in three daily doses; PQ, 15 mg primaquine daily for 14 days.

difficulties of unknown mechanism of therapeutic activity against a cryptic and highly nuanced stage of some plasmodia—the hypnozoite—coupled with a vitally important hemolytic toxicity problem also of unknown mechanism in patients having a highly prevalent and diverse genetic abnormality, G6PD deficiency. Estimates of primaquine efficacy as impacted by parasite biology, epidemiology and partner blood schizontocides imposes great complexity of interpretation.¹⁰⁷ These issues all also bear on tafenoquine and its use in radical cure of the relapsing malaria.

Estimates of the efficacy of hypnozoitocides like tafenoquine are subject to important confounding factors. The natural activation of hypnozoites typically occurs over months following infection.¹⁰⁸ When relapse occurs in the presence of risk of reinfection, these two sources of acute malaria temporally mingle and no molecular laboratory technology differentiates them. Post-hypnozoitocidal recurrences in endemic areas may thus be represented by both therapeutic failures (relapse) and the primary attacks of mosquito-borne reinfection—recrudescence with blood schizontocidal failure may also occur but is not considered here. The rates of both relapse and reinfection naturally vary widely across endemic zones and each may impact inherently variable estimates of hypnozoitocidal efficacy. Figure 5 presents hypothetical rates of each in high and low transmission settings in order to illustrate these potential impacts. High transmission with low relapse risk (e.g. <30%) may greatly underestimate efficacy (left panels), an effect mitigated by high relapse risk (e.g. >70%), especially where there is low risk of reinfection (right panels). Reported estimates of efficacy from endemic areas are thus not absolute but reported as the fraction of patients not experiencing a recurrent parasitemia during months of follow-up, often relative to a hypnozoitocidal comparator or placebo control group (also called a relapse control). Conducting treatment and follow-up where reinfection does not occur and with a relapse control arm largely resolves these ambiguities.^{85,109} Such a trial for tafenoquine has yet to be completed, though one is in progress in Indonesia in 2018.

The two multi-center, double-blind and placebo-controlled randomized clinical trials estimating efficacy of tafenoquine at a single dose of 300 mg combined with standard chloroquine therapy (1500 mg base over 3 days) included 317 subjects thus dosed against naturally acquired *P. vivax* infections in Brazil, Peru,

Ethiopia, Thailand, Cambodia and the Philippines (Trials 1 and 2, Table 4).^{110,111} A total of 187 subjects in those trials received chloroquine and a placebo of tafenoquine. Subjects were followed for recurrent infections for six months. A total of 226 of 317 (71%) subjects did not experience recurrence within 6 months of tafenoquine and chloroquine therapy, whereas 79 of 187 (42%) subjects treated with chloroquine and placebo did so. In a third trial lacking a placebo control, tafenoquine ($n = 166$) or primaquine ($n = 85$) combined with chloroquine resulted in 73% and 75% remaining free of recurrence for 6 months (Trial 3, Table 4), consistent with non-inferiority of single-dose tafenoquine relative to daily 15 mg primaquine for 14 days.¹¹⁰

An important factor regarding hypnozoitocidal therapy bearing upon both efficacy and safety is co-administration with varied blood schizontocidal therapies. Indeed, the discovery effort leading to primaquine stemmed from an unexpected drug–drug interaction (DDI) between atabrine (mepacrine) and plasmochin (pamaquine) disqualifying co-administration for radical cure.¹¹² The developers of plasmochin and primaquine each reported DDI phenomena with varied partner blood schizontocides impacting efficacy, safety or both. Tafenoquine has thus far been examined only in combination with chloroquine in vivax malaria patients. However, it was evaluated with several distinct partner blood schizontocides against *P. cynomolgi* relapses in rhesus macaques.¹¹³ Those investigators reported a 10-fold increase in tafenoquine efficacy when administered with chloroquine, mefloquine or artemether–lumefantrine compared to tafenoquine alone. Over 60 years ago, Alving *et al.* reported essentially similar findings with primaquine given concurrent vs consecutive quinine or chloroquine.¹¹⁴ How these purely blood schizontocidal drugs so dramatically impact the hypnozoitocidal efficacy of 8-aminoquinolines remains unknown.

While chloroquine or artemether–lumefantrine did not significantly impact tafenoquine pharmacokinetics in healthy subjects, dihydroartemisinin–piperaquine increased the C_{max} of tafenoquine by 38%, the area under the concentration (AUC) curve by 12%, and the plasma half-life by 29%.^{115,116} Tafenoquine did not appear to impact the pharmacokinetics or dynamics of chloroquine, artemether–lumefantrine, or dihydroartemisinin–piperaquine. The FDA label for Krintafel™ cites chloroquine as an example of appropriate companion therapy,

implicitly allowing for other partner blood schizontocides for radical cure.¹¹⁰ The data from *P. cynomolgi* in macaques seem to affirm that view so far as mefloquine and artemether–lume-fantrine are concerned.¹¹²

The package insert for Krintafel™ expresses an indicated use in radical cure of *P. vivax* malaria in patients at least 16 years of age who are also receiving companion blood schizontocidal therapy.¹¹¹ The warnings and precautions expressed therein are essentially similar to those for Arakoda™ (see above). Both labels warn of serious psychotic adverse reactions having occurred at the indicated dose (for Krintafel™) or higher dosing (for Arakoda™) in patients with a history of psychoses, along with serious hypersensitivity events (e.g. angioedema).^{74,111} Tafenoquine (as Arakoda™ or Krintafel™) may or may not be suited to patients with psychiatric histories; the evidence needed to definitively inform that question is lacking. In the instance of primaquine, there have been no significant clinical neurotoxicity signals after decades of use.^{117,118} Indeed, in the defining neurotoxicological studies of 8-aminoquinolines in rhesus macaques, severe irreversible brainstem neuronal injury occurred only among compounds of the plasmocid (or Rhodoquine) subclass (Figure 1).¹¹⁹ Among the plasmochin (or pamaquine) subclass of 8-aminoquinolines (all 8-aminoquinolines that advanced to human clinical trials, including primaquine and, later, tafenoquine), no such neurotoxicity occurred.

In summary, adult G6PD-normal non-pregnant or lactating patients diagnosed with acute *P. vivax* malaria, or those returning from travel of risk without causal prophylaxis, a single 300 mg dose of tafenoquine provides safe, well-tolerated, and efficacious PART. Post-diagnosis PART may be confidently combined with chloroquine, mefloquine, or artemether–lume-fantrine. Post-travel PART should consider the apparently conspicuous dependency of tafenoquine efficacy on the presence of select blood schizontocides as convincingly demonstrated in the *P. cynomolgi* animal model. Tafenoquine without a companion blood schizontocide possibly not killing hypnozoites at prescribed dose merits clinical caution and scientific attention. More details are available in the FDA Advisory Committee Briefing Document for Krintafel™: <https://www.fda.gov/downloads/advisorycommittees/committeesmeetingmaterials/drugs/anti-infectivedrugsadvisorycommittee/ucm612875.pdf>

Conclusions

The availability of tafenoquine for the prevention and treatment of malaria appears to offer potentially transformative new options in the practice of travel medicine. These applications strictly require reliable screening for G6PD deficiency, like the current standard of responsible care involving primaquine for causal prophylaxis or for post-travel or post-diagnosis PART in travelers. Excepting travel to the very few malarious areas where infection by hypnozoites is highly improbable, G6PD screening should be acknowledged as indicated in any traveler taking any chemoprophylactic option. Avoidance of G6PD screening with non-hemolytic suppressive chemoprophylactics (without post-travel PART) invites risk of post-travel attacks. No species of plasmodia is intrinsically benign. They all merit the diligence and relative difficulty of preventing them. Tafenoquine offers G6PD-normal and non-pregnant adults a convenient, safe, well-tolerated and efficacious means of preventing all malarias during travel or treating those that relapse after travel. The important work needed to assure the safety of tafenoquine in children is in progress, along with appropriate formulation for them. Far broader clinical experience with tafenoquine will have to accrue before fully understanding both its advantages and limitations, but its promise certainly merits such accrual.

Conflict of interest: None declared.

Disclaimer

The author served as a paid consultant to GlaxoSmithKline® (GSK, UK) in support of the application of Krintafel™ for registration with the US FDA. His laboratory in Jakarta, Indonesia, today conducts a pivotal clinical trial of Krintafel™ for radical cure of vivax malaria for which GSK is the sponsor. He receives no personal financial incentive or award for that work from GSK or any other organization. The author holds no financial interest in GSK or any of its products. At the invitation of 60 Degrees Pharmaceuticals® (USA), the author provided testimony to the US FDA favorable to the registration of Arakoda™ for chemoprevention of malaria, for which he received no financial

SUMMARY BOX 2. KEY POINTS ON TAFENOQUINE RADICAL CURE IN TRAVEL MEDICINE

- Suppressive malaria prophylaxis standard-of-care requires post-travel presumptive anti-relapse therapy (PART) to destroy latent hypnozoites and prevent delayed attacks in the months following travel.
- A diagnosis of acute relapsing malaria (*P. vivax* or *P. ovale*) in any patient requires PART to destroy latent hypnozoites and prevent subsequent attacks by them.
- Primaquine has been the standard-of-care for PART as 14 daily doses of 0.5 mg/kg for the past 66 years.
- Tafenoquine is a new drug with an indication for post-travel or post-diagnosis PART against relapsing malarias as a single adult dose of 300 mg.
- Tafenoquine is, like primaquine, hemolytically toxic to patients having inherited G6PD deficiency, so is prohibited in those patients along with pregnant and lactating women. Safety in children is not yet established.

compensation or award. He holds no financial interest in that drug, its manufacturer or any of its products.

References

- Newton CR, Taylor TE, Whitten RO. Pathophysiology of fatal falciparum malaria in African children. *Am J Trop Med Hyg* 1998; 58:673–83.
- Park SW, Kim DW, Park JW *et al.* A case of fatal *Plasmodium vivax* malaria with multiple-organ failure. *Infect Chemother* 2005; 37:111–5.
- Groger M, Fischer HS, Veletzky L, Lalremruata A, Ramharter M. A systematic review of the clinical presentation, treatment, and relapse characteristics of human *Plasmodium ovale* malaria. *Malar J* 2017; 16:112.
- Collins WE, Jeffery GM. *Plasmodium malariae*: parasite and disease. *Clin Microbiol Rev* 2007; 20:579–92.
- Rajahram G, Barber BE, William T, Menon J, Anstey NM, Yeo TW. Deaths due to *Plasmodium knowlesi* malaria in Sabah, Malaysia: association with reporting as *P. malariae* and delayed parenteral artesunate. *Malar J* 2012; 11:284.
- Kain KC, Harrington MA, Tennyson S, Keystone JS. Imported malaria: prospective analysis of problems in diagnosis and management. *Clin Infect Dis* 1998; 27:142–9.
- Baird JK, Nelwan J, Taylor WR. Approach to the patient with malaria. In: Keystone JS, Kozarsky PE, Connor BA, Nothdurft HD, Leder K, Mendelson M (eds). *Travel Medicine*, 4th edn. Elsevier, 2019 Chapter 17.
- Tediosi F, Lengeler C, Castro M *et al.* Chapter 13: Malaria control. In: Holmes KK, Bertozzi S, Bloom BR, and Jha P (eds). *Major Infectious Diseases. Disease Control Priorities*, 3rd edn, Vol. 6. Washington: DC: World Bank, 2017, pp. 347–364.
- Bhatt S, Weiss DJ, Cameron E *et al.* The effect of malaria control on *Plasmodium falciparum* in Africa between 2000 and 2015. *Nature* 2015; 526:207–11.
- Cibulskis RE, Alonso PE, Aponte J *et al.* Malaria: global progress 2000–2015 and future challenges. *Infect Dis Poverty* 2016; 9:61.
- Mendis K, Sina BJ, Marchesini P, Carter R. The neglected burden of *Plasmodium vivax* malaria. *Am J Trop Med Hyg* 2001; 64:97–106.
- Baird JK. Neglect of *Plasmodium vivax* malaria. *Trends Parasitol* 2007; 23(11). PMID: 17933585.
- Price RN, Tjitra E, Guerra CA, Yeung S, White NJ, Anstey NM. Vivax malaria: neglected and not benign. *Am J Trop Med Hyg* 2007; 77:79–87.
- Bassat Q, Velarde M, Mueller I *et al.* Key knowledge gaps for *Plasmodium vivax* control and elimination. *Am J Trop Med Hyg* 2016; 95:62–71.
- Carlton JM, Sina BJ, Adams JH. Why is *Plasmodium vivax* a neglected tropical disease? *PLoS Negl Trop Dis* 2011; 5:e1160.
- World Health Organization. *World Malaria Report 2017*. Geneva; 2018. Geneva: World Health Organization. ISBN: 978 92 4 156552 3.
- Freedman DO, Chen LH, Kozarsky PE. Medical considerations before international travel. *N Engl J Med* 2016; 375:247–60.
- Boggild A, Brophy J, Charlebois P *et al.* Summary of recommendations for the prevention of malaria by the Committee to Advise on Tropical Medicine and Travel (CATMAT). *Can Commun Dis Rep* 2014; 40:118–32.
- PHE Advisory Committee for Malaria Prevention for UK Travelers. *Guidelines for malaria prevention in travelers from the UK: 2017; 2017*. London: Public Health England; 146pp.
- Baird JK. Management of *Plasmodium vivax* risk and illness in travelers. *Trop Dis Travel Med Vaccines* 2017; 3:7.
- Mühlberger N, Jelinek T, Gascon J *et al.* Epidemiology and clinical features of vivax malaria imported to Europe: sentinel surveillance data from TropNetEurop. *Malar J* 2004; 3:5.
- Steinhardt LC, Magill AJ, Arguin PM. Review: malaria chemoprophylaxis for travellers to Latin America. *Am J Trop Med Hyg* 2011; 85:1015–24.
- Schwartz E, Parise M, Kozarsky P, Cetron M. Delayed onset of malaria – implications for chemoprophylaxis in travellers. *N Engl J Med* 2003; 349:1510–16.
- Tan KR, Hwang J. Tafenoquine receives regulatory approval in U. S. for prophylaxis of malaria and radical cure of *Plasmodium vivax*. *J Travel Med* 2018. doi:10.1093/jtm/tay071.
- Gutteridge WE. Antimalarial drugs currently in development. *J R Soc Med* 1989; 17:63–6.
- Davidson DE Jr, Ager AL, Brown JL, Chapple FE, Whitmire RE, Rossan RN. New tissue schizontocidal antimalarial drugs. *Bull World Health Organ* 1981; 59:463–79.
- Ashley EA, Phyto AP. Drugs in development for malaria. *Drugs* 2018; 78:861–79.
- Peters W. The evolution of tafenoquine – antimalarial for a new millennium? *J R Soc Med* 1999; 92:345–52.
- Kitchener S, Nasveld P, Edstein MD. Tafenoquine for the treatment of current *Plasmodium vivax* malaria. *Am J Trop Med Hyg* 2007; 76:494–6.
- Crockett M, Kain KC. Tafenoquine: a promising new antimalarial agent. *Expert Opin Investig Drugs* 2007; 16:705–15.
- Jeffery GM. Infectivity of mosquitoes of *Plasmodium vivax* following treatment with chloroquine and other antimalarials. *Am J Trop Med Hyg* 1958; 7:207–11.
- Peters W, Robinson BL, Milhous WK. The chemotherapy of rodent malaria. II. Studies on a new 8-aminoquinoline, WR 238,605. *Ann Trop Med Parasitol* 1993; 87:547–52.
- Coleman RE. Sporontocidal activity of the antimalarial WR-238605 against *Plasmodium berghei* ANKA in *Anopheles stephensi*. *Am J Trop Med Hyg* 1990; 42:196–2015.
- Shapiro TA, Ranasinha CD, Kumar N, Barditch-Crovo P. Prophylactic activity of atovaquone against *Plasmodium falciparum* in humans. *Am J Trop Med Hyg* 1999; 60:831–6.
- Berman JD, Nielsen R, Chulay JD *et al.* Causal prophylactic activity of atovaquone-proguanil (Malarone) in a human challenge model. *Trans R Soc Trop Med Hyg* 2001; 95:429–32.
- Maguire JD, Llewellyn DM. Relapsing malaria after 6 months of daily atovaquone-proguanil in Afghanistan: the case for expanded use of primaquine as a causal prophylactic. *J Travel Med* 2007; 14:411–14.
- Metzler E, Rahav G, Schwartz E. Vivax malaria chemoprophylaxis: the role of atovaquone-proguanil compared to other options. *Clin Infect Dis* 2018; 66:1751–5.
- DiTusa C, Kozar MP, Pybus B *et al.* Causal prophylactic efficacy of primaquine, tafenoquine, and atovaquone-proguanil against *Plasmodium cynomolgi* in a rhesus monkey model. *J Parasitol* 2014; 100:671–3.
- Baird JK, Fryauff DJ, Hoffman SL. Primaquine for the prevention of malaria in travelers. *Clin Infect Dis* 2003; 37:1659–67.
- Nasveld P, Kitchener S. Treatment of acute vivax malaria with tafenoquine. *Trans R Soc Trop Med Hyg* 2005; 99:2–5.
- Shanks GD. Historical review: problematic malaria prophylaxis with quinine. *Am J Trop Med Hyg* 2016; 95:269–72.
- Arguin PM, Magill AJ. For the record: a history of malaria chemoprophylaxis. <https://wwwnc.cdc.gov/travel/yellowbook/2018/infectious-diseases-related-to-travel/emfor-the-record-a-history-of-malaria-chemoprophylaxisem>. Visited 13 September 2018.

43. Sinton JA, Smith S, Pottinger D. Studies on malaria with special reference to treatment. XII. Further researches into the treatment of chronic benign tertian malaria with plasmoquine and quinine. *Indian J Med Res* 1929; 20:793–814.
44. Shannon JA. Chemotherapy in malaria. *Bull N Y Acad Med* 1946; 22:345–57.
45. Centers for Disease Control. Malaria among US military personnel returning from Somalia, 1993. *MMWR Morb Mortal Wkly Rep* 1993; 42:524–6.
46. Krafts K, Hempelmann E, Skorska-Stania A. From methylene blue to chloroquine: a brief review of the development of antimalarial therapy. *Parasitol Res* 2012; 111:1–6.
47. Kolifarhood G, Raeisi A, Ranjbar M *et al.* Prophylactic efficacy of primaquine for preventing *Plasmodium falciparum* and *Plasmodium vivax* parasitemia in travelers: a meta-analysis and systematic review. *Travel Med Infect Dis* 2017; 17:5–18.
48. Arnold J, Alving AS, Hockwald RS *et al.* The antimalarial action of primaquine against the blood and tissue stages of falciparum malaria (Panama P-F-6 strain). *J Lab Clin Med* 1955; 46:391–7.
49. Flaherty GT, Walden LJ, Townend M. Travel medicine physician adherence to guidelines for the emergency self-treatment of malaria. *J Travel Med* 2016; 23(5). doi:10.1093/jtm/taw036.
50. Behrens R. Standby emergency treatment of malaria for travelers to low transmission destinations: does it make sense or save lives? *J Travel Med* 2017; 24(5). doi:10.1093/jtm/tax034.
51. Boubaker R, Harard Fossati A, Meige P *et al.* Malaria prevention strategies and recommendations from chemoprophylaxis to stand-by emergency treatment: a 10-year prospective study in a Swiss travel clinic. *J Travel Med* 2017; 24(5). doi:10.1093/jtm/tax043.
52. Hwang J, Cullen CA, Kachur SP, Arguin PM, Baird JK. Severe morbidity and mortality risk from malaria in the United States, 1985–2011. *Open Forum Infect Dis* 2014; 1:ofu034.
53. Barber BE, Grigg MJ, Piers KA *et al.* Intravascular haemolysis in severe *Plasmodium knowlesi* malaria: association with endothelial activation, microvascular dysfunction, and acute kidney injury. *Emerg Microbes Infect* 2018; 7:106.
54. Anstey NM, Douglas NM, Poespoprodjo JR, Price RN. *Plasmodium vivax*: clinical spectrum, risk factors and pathogenesis. *Adv Parasitol* 2012; 80:151–202.
55. Baird JK. Evidence and implications of mortality in acute *Plasmodium vivax* malaria. *Clin Microbiol Rev* 2013; 26:36–57.
56. Naing C, Whittaker MA, Wai VN, Mak JW. Is *Plasmodium vivax* malaria a severe malaria? A systematic review and meta-analysis. *PLoS Negl Trop Dis* 2014; 8:e3071.
57. Quispe AM, Pozo E, Guerrero E *et al.* *Plasmodium vivax* hospitalizations in a monoendemic malaria region: severe vivax malaria? *Am J Trop Med Hyg* 2014; 91:11–7.
58. Douglas NM, Pontororing GJ, Lampah DA *et al.* Mortality attributable to *Plasmodium vivax* malaria: a clinical audit from Papua, Indonesia. *BMC Med* 2014; 12:217.
59. Siqueira AM, Lacerda MVG, Magalhaes BML *et al.* Characterization of *Plasmodium vivax*-associated admissions to reference hospitals in Brazil and India. *BMC Med* 2015; 13:57.
60. Mace KE, Arguin PM, Tan KR. Malaria surveillance – United States, 2015. *MMWR Surveill Summ* 2018; 67:1–28.
61. Tatem AJ, Jia P, Ordanovich D *et al.* The geography of imported malaria to non-endemic countries: a meta-analysis of nationally reported statistics. *Lancet Infect Dis* 2017; 17:98–107.
62. Ryan JR, Stoute JA, Amon J *et al.* Evidence for transmission of *Plasmodium vivax* among a Duffy antigen negative population in Western Kenya. *Am J Trop Med Hyg* 2006; 75:575–81.
63. Howes RE, Reiner RC Jr, Battle KE *et al.* *Plasmodium vivax* transmission in Africa. *PLoS Negl Trop Dis* 2015; 9:e0004222.
64. Brazeau NF, Whitesell A, Doctor SM *et al.* *Plasmodium vivax* infections in Duffy-negative individuals in the Democratic Republic of the Congo. *Am J Trop Med Hyg* 2018. doi:10.4269/ajtmh.18-0277.
65. Gething PW, Elyazar IR, Moyes CL *et al.* A long neglected world map: *Plasmodium vivax* endemicity in 2010. *PLoS Negl Trop Dis* 2012; 6:e1814.
66. Schlagenhauf P, Petersen E. Malaria chemoprophylaxis: strategies for risk group. *Clin Microbiol Rev* 2008; 21:466–72.
67. Davlantes EA, Tan KR, Arguin PM. Quantifying malaria risk in travelers: a quixotic pursuit. *J Travel Med* 2017; 24(6). doi:10.1093/jtm/tax066.
68. Luzzatto L, Seneca E. G6PD deficiency: a classic example of pharmacogenetics with on-going clinical implications. *Br J Haematol* 2014; 164:469–80.
69. Howes RE, Piel FB, Patil AP *et al.* G6PD deficiency prevalence estimates of affected populations in malaria endemic countries: a geostatistical model-based map. *PLoS Med* 2012; 9:e1001339.
70. World Health Organization Evidence Review Group. Point-of-care testing to support safe use of primaquine for the treatment of vivax malaria. Malaria Policy Advisory Committee Meeting, 5–7 March, 2015, Geneva, Switzerland. WHO/HTM/GMP/MPAC/2015.6. <http://www.who.int/malaria/mpac/mpac-march2015-erg-g6pd.pdf> accessed on 15 September 2018.
71. Howes RE, Dewi M, Piel FB *et al.* Spatial distribution of G6PD deficiency variants across malaria-endemic regions. *Malar J* 2013; 12:418.
72. Rueangweerayut R, Bancone G, Harrell EJ *et al.* Hemolytic potential of tafenoquine in female volunteers heterozygous for glucose-6-phosphate dehydrogenase (G6PD) deficiency (G6PD Mahidol variant) versus G6PD-normal volunteers. *Am J Trop Med Hyg* 2017; 97:702–11.
73. Chu CS, Bancone G, Moore KA *et al.* Haemolysis in G6PD heterozygous females treated with primaquine for *Plasmodium vivax* malaria: a nested cohort in a trial of radical curative regimens. *PLoS Med* 2017; 14:e1002224.
74. Arakoda™ package insert. 60 Degrees Pharmaceuticals LLC, Washington, DC; 2018. https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/210607lbl.pdf
75. Baird JK, Dewi M, Subekti D, Elyazar I, Satyagraha AW. Noninferiority of glucose-6-phosphate dehydrogenase deficiency diagnosis by a point-of-care rapid test vs the laboratory fluorescent spot test demonstrated by copper inhibition in normal human red blood cells. *Transl Res* 2015; 165:677–88.
76. Chu CS, Bancone G, Nosten F, White NJ, Luzzatto L. Primaquine-induced haemolysis in females heterozygous for G6PD deficiency. *Malar J* 2018; 17:101.
77. Recht J, Ashley EA, White NJ. Use of primaquine and glucose-6-phosphate dehydrogenase deficiency testing: divergent policies and practices in endemic countries. *PLoS Negl Trop Dis* 2018; 12:e0006230.
78. Roca-Feltrer A, Khim N, Kim S *et al.* Field trial evaluation of the performance of point-of-care tests for screening G6PD deficiency in Cambodia. *PLoS One* 2014; 9:e116143.
79. Bancone G, Chu CS, Chowwiwat N *et al.* Suitability of capillary blood for quantitative assessment of G6PD activity and performances of G6PD point-of-care tests. *Am J Trop Med Hyg* 2015; 92:818–24.
80. Oo NN, Bancone G, Maw LZ *et al.* Validation of G6PD point-of-care tests among healthy volunteers in Yangon, Myanmar. *PLoS One* 2016; 11:e0152304.
81. Ley B, Bancone G, von Seidlein L *et al.* Methods for the field evaluation of quantitative G6PD diagnostics: a review. *Malar J* 2017; 16:361.
82. Marcisisin SR, Reichard G, Pybus BS. Primaquine pharmacology in the context of CYP 2D6 pharmacogenomics: current state of the art. *Pharmacol Ther* 2016; 16:1–10.

83. Bennett JW, Pybus BS, Yadava A *et al.* Primaquine failure and cytochrome P-450 2D6 in *Plasmodium vivax* malaria. *N Engl J Med* 2013; 369:1381–2.
84. Ingram RJ, Crenna-Darusallam C, Soebianto S, Noviyanti R, Baird JK. The clinical and public health problem of relapse despite primaquine therapy: case review of repeated relapses of *Plasmodium vivax* acquired in Papua New Guinea. *Malar J* 2014; 13:488.
85. Nelwan EJ, Ekawati LL, Tjahjono B *et al.* Randomized trial of primaquine hypnozoitocidal efficacy when administered with artemisinin-combined blood schizontocides for radical cure of *Plasmodium vivax* in Indonesia. *BMC Med* 2015; 13:294.
86. Baird JK, Louisa M, Noviyanti R *et al.* Association of impaired cytochrome P-450 2D6 activity genotype and phenotype with therapeutic efficacy of primaquine treatment for latent *Plasmodium vivax* malaria. *JAMA Open Network* 2018; 1: e181449.
87. Marcisisin SR, Sousa JC, Reichard GA *et al.* Tafenoquine and NPC-1161B require CYP2D metabolism for antimalarial activity: implications for the 8-aminoquinoline class of antimalarial compounds. *Malar J* 2014; 13:2.
88. St Jean PL, Zue Z, Carter N *et al.* Tafenoquine treatment of *Plasmodium vivax* malaria: suggestive evidence that CYP2D6 reduced metabolism is not associated with relapse in the Phase 2b DETECTIVE trial. *Malar J* 2016; 15:97.
89. Hill DR, Baird JK, Parise ME, Lewis LS, Ryan T, Magill AJ. Primaquine: report from CDC expert meeting on malaria chemoprophylaxis I. *Am J Trop Med Hyg* 2006; 75:402–15.
90. Nasveld PE, Edstein MD, Brennan RM *et al.* Randomized, double-blind study of the safety, tolerability, and efficacy of tafenoquine versus mefloquine for malaria prophylaxis in nonimmune subjects. *Antimicrob Agents Chemother* 2010; 54:792–8.
91. Shanks GD, Oloo AJ, Aleman GM *et al.* A new primaquine analog, tafenoquine (WR 238605), for prophylaxis against *Plasmodium falciparum* malaria. *Clin Infect Dis* 2001; 33:1968–74.
92. Hale BR, Owusu-Agyei S, Fryauff DJ *et al.* A randomized, double-blind, placebo-controlled, dose-ranging trial of tafenoquine for weekly prophylaxis against *Plasmodium falciparum*. *Clin Infect Dis* 2003; 36:541–9.
93. Dow GS, McCarthy WF, Reid M, Smith B, Tang D, Shanks GD. A retrospective analysis of the protective efficacy of tafenoquine and mefloquine as prophylactic antimalarials in non-immune individuals during deployment to a malaria-endemic area. *Malar J* 2014; 14:49.
94. Dow GS, Liu J, Lin G *et al.* Summary of antimalarial prophylactic efficacy of tafenoquine from three placebo-controlled studies of residents of malaria-endemic areas. *Malar J* 2015; 14:473.
95. Shanks D. The conundrum of malaria chemoprophylaxis. *J Travel Med* 2016; 23(6). pii: taw065. PMID: 27694470.
96. Brueckner RP, Coster T, Wesche DL, Shmuklarsky M, Schuster B. Prophylaxis of *Plasmodium falciparum* infection in a human challenge model with WR238605, a new 8-aminoquinoline compound. *Antimicrob Agents Chemother* 1998; 42:1293–4.
97. Dow G, Smith B. The blood schizontocidal activity of tafenoquine makes an essential contribution to its prophylactic efficacy in non-immune subjects at the intended dose (200mg). *Malar J* 2017; 16:209.
98. Lell B, Faucher J-P, Missinou MA *et al.* Malaria chemoprophylaxis with tafenoquine: a randomized study. *Lancet* 2000; 355:2041–5.
99. Thakkar N, Green JA, Koh GC, Duparc S, Tenero D, Goyal N. Population pharmacokinetics of tafenoquine, a novel antimalarial. *Antimicrob Agents Chemother* 2018. doi:10.1128/AAC.07111-18.
100. Novitt-Moreno A, Ransom J, Dow G, Smith B, Read LT, Toovey S. Tafenoquine for malaria prophylaxis in adults: an integrated safety analysis. *Travel Med Infect Dis* 2017; 17:19–27.
101. Leary KJ, Riel MA, Roy MJ *et al.* A randomized, double-blind, safety and tolerability study to assess the ophthalmic and renal effects of tafenoquine 200mg weekly versus placebo for 6 months in healthy volunteers. *Am J Trop Med Hyg* 2009; 81:356–62.
102. Fukuda M, Krudsood S, Mohamed K *et al.* A randomized, double-blind, active-control trial to evaluate the efficacy and safety of a three day course of tafenoquine monotherapy for the treatment of *Plasmodium vivax* malaria. *PLoS One* 2017; 12:e0187376.
103. Baird JK, Hoffman SL. Primaquine therapy for malaria. *Clin Infect Dis* 2004; 39:1336–45.
104. Chu CS, White NJ. Management of relapsing *Plasmodium vivax* malaria. *Expert Rev Anti Infect Ther* 2016; 14:885–900.
105. Baird JK, Valecha N, Duparc S, White NJ, Price RN. Diagnosis and treatment of *Plasmodium vivax* malaria. *Am J Trop Med Hyg* 2016; 95:35–51.
106. Ashley EA, Recht J, White NJ. Primaquine: the risks and benefits. *Malar J* 2014; 13:418.
107. Baird JK. Resistance to therapies by *Plasmodium vivax*. *Clin Microbiol Rev* 2009; 22:508–34.
108. White NJ, Imwong M. Relapse. *Adv Parasitol* 2012; 80:113–42.
109. Sutanto I, Tjahjono B, Basri H *et al.* Randomized, open-label trial of primaquine against vivax malaria relapse in Indonesia. *Antimicrob Agents Chemother* 2013; 57:1128–35.
110. Llanos-Cuentas A, Lacerda MV, Rueangweerawat R *et al.* Tafenoquine plus chloroquine for the treatment and relapse prevention of *Plasmodium vivax* malaria (DETECTIVE): a multicenter, double-blind, randomized, phase 2b dose-selection study. *Lancet* 2014; 383:1049–58.
111. Krintafel™ package insert. Washington, DC: GlaxoSmithKline Pharmaceuticals; 2018. Reference ID: 4294835. https://www.accessdata.fda.gov/drugsatfda_docs/label/2018/210795s000lbl.pdf
112. Baird JK. Resistance to chloroquine unhinges vivax malaria therapeutics. *Antimicrob Agents Chemother* 2011; 55:1827–30.
113. Dow GS, Gettayacamin M, Hansukjariya P *et al.* Radical curative efficacy of tafenoquine combination regimens in *Plasmodium cynomolgi*-infected Rhesus monkeys (*Macaca mulatta*). *Malar J* 2011; 10:212.
114. Alving AS, Arnold J, Hockwald RS *et al.* Potentiation of the curative action of primaquine in vivax malaria by quinine and chloroquine. *J Lab Clin Med* 1955; 46:301–6.
115. Miller AK, Harrell E, Ye L *et al.* Pharmacokinetic interactions and safety evaluations of coadministered tafenoquine and chloroquine in healthy subjects. *Br J Clin Pharmacol* 2013; 76:858–67.
116. Green JA, Mohamed K, Goyal N *et al.* Pharmacokinetic interactions between tafenoquine and dihydroartemisinin-piperaquine or artemether-lumefantrine in healthy adult subjects. *Antimicrob Agents Chemother* 2016; 60:7321–32.
117. Clyde DF. Clinical problems associated with use of primaquine as a tissue schizontocidal and gametocytocidal drug. *Bull World Health Organ* 1981; 59:391–5.
118. Recht J, Ashley EA, White N. *Safety of 8-aminoquinoline antimalarial medicines*. Geneva: World Health Organization, 2014, 222. ISBN 978 92 4 150697 7.
119. Carson PE, Hohl R, Nora MV *et al.* Toxicology of the 8-aminoquinolines and genetic factors associated with their toxicity in man. *Bull World Health Organ* 1981; 59:427–37.
120. Gething PW, Patil AP, Smith DL *et al.* A new world malaria map: *Plasmodium falciparum* endemicity in 2010. *Malar J* 2011; 10:378.