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Chronic Inflammation in the Context of Everyday Life: Dietary Changes as Mitigating Factors

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Abstract: The lifestyle adopted by most people in Western societies has an important impact on the propensity to metabolic disorders (e.g., diabetes, cancer, cardiovascular disease, neurodegenerative diseases). This is often accompanied by chronic low-grade inflammation, driven by the activation of various molecular pathways such as STAT3 (signal transducer and activator of transcription 3), IKK (IkB kinase), MMP9 (matrix metallopeptidase 9), MAPK (mitogen-activated protein kinases), COX2 (cyclooxigenase 2), and NF-K β (nuclear factor kappa-light-chain-enhancer of activated B cells). Multiple intervention studies have demonstrated that lifestyle changes can lead to reduced inflammation and improved health. This can be linked to the concept of real-life risk simulation, since humans are continuously exposed to dietary factors in small doses and complex combinations (e.g., polyphenols, fibers, polyunsaturated fatty acids, etc.). Inflammation biomarkers improve in patients who consume a certain amount of fiber per day; some even losing weight. Fasting in combination with calorie restriction modulates molecular mechanisms such as m-TOR, FOXO, NRF2, AMPK, and sirtuins, ultimately leads to significantly reduced inflammatory marker levels, as well as improved metabolic markers. Moving toward healthier dietary habits at the individual level and in publicly-funded institutions, such as schools or hospitals, could help improving public health, reducing healthcare costs and improving community resilience to epidemics (such as COVID-19), which predominantly affects individuals with metabolic diseases.

Keywords: diet; polyphenols; inflammation; real-life risk simulation; fasting; COVID-19

1. Introduction

Chronic inflammation is a central process involved in a high number of metabolic disorders (e.g., obesity, metabolic syndrome, diabetes, dyslipidemia, etc.), including neurodegenerative (Alzheimer), malignant diseases, and autoimmune diseases. In most if not all chronic inflammatory conditions, there is an extensively failed resolution of inflammation with high influx of leukocytes, which in their effort to resolve inflammation stimulate the synthesis of pro-inflammatory molecules and establish a highly inflammatory micro-environment, leading to extensive fibrosis and tissue damage [1]. Chronic low-grade inflammation has been shown to either induce or aggravate metabolic disturbances, including insulin resistance and dyslipidemia, which contributes to the development of other complications [2]. There is accumulating evidence that, in the case of autoimmune diseases, when the immune system loses self-tolerance and attacks the body's cells and tissues, metabolic disturbances are key contributors to disease progression. Results from the type 1 diabetes mellitus (T1DM) prediction and prevention studies on T1DM showed that metabolic disturbances mellitus [3,4].

Many chronic inflammatory diseases originate or have their development promoted by an unbalanced diet. Although the exact mechanism remains unclear, De Rosa et al. suggest that metabolic pressure, as a result of increased caloric intake, leads to an altered adipose tissue homeostasis. This results in the synthesis of adipokines and facilitates the overactivation of nutrient-sensing mechanisms, altering the balance between pro-inflammatory and regulatory T-cells, ultimately resulting in the loss of immunotolerance [5–7]. In addition, dietary components have the ability to influence the immune response through the modulation of gut bacteria metabolism, impacting the risk of developing chronic diseases either directly in the gastrointestinal tract, or in other more distant organs that impact general metabolism [8–11]. Recent studies have investigated long term exposure to low doses of chemical mixtures that can be a part of modern lifestyles, such as pesticides, food additives, or additives contained in food coating materials, proving that different disturbances appeared from minor biochemical disturbances. These early alterations are generally followed by oxidative stress induction and organ damage depending on the period of exposure [12–17]. Recently, it has been shown that long term exposure to stressors might also have a positive association with increased vulnerability of the population to the microbial and viral infections [18].

Metabolomics are an emerging biological field that allow for the identification and simultaneous measurement of a large number of small molecules called metabolites in biological matrixes. It has become the most accurate method to detect metabolic imbalances and is useful for prevention and early detection of diseases. Moreover, metabolomics have vast applications in clinical practice [19]. Targeted metabolomic analysis provides insights regarding the normal function of endogenous metabolism, dietary intake, microbiota, drug metabolism, and nutrient adequacy [20]. The challenge of chronic inflammatory diseases with respect to early diagnosis can be tackled with metabolomics through the identification of biomarkers that can discriminate high-risk populations. In a group of autoimmune patients, it was found that their fatty acid-based metabolic profile and lifestyle factors including physical activity and alcohol consumption were valuable predictive markers of autoimmune diseases [21].

Humans are exposed to a large number of substances from food, water, cosmetics, air, and so forth, each at low levels of exposure, and are able to induce cumulative/synergistic effects. Many studies have focused on the effects induced by administering a single substance at medium-high doses to laboratory animals. Recently, the concept of real-life risk simulation has emerged, since there is growing evidence that the effects of chemical mixtures at concentrations for which individual components failed to elicit have adverse effects when tested individually [14]. The concept of real-life risk simulation can also incorporate dietary interventions because, in our diets, we expose the human body to myriad substances in diverse doses [14,22,23].

The discovery of inflammation regulators opened a new window in therapeutics to clear low-grade chronic inflammation. A large number of physiological processes promote the physiological process of regulating inflammation. The development of such an approach targets the stimulation of endogenous

processes that naturally occur during inflammation, which are hampered mainly by the lack of suitable human models and the heterogeneity of inflammatory disorders. Another limitation includes the lack of sensitive measurements able to capture the different stages of inflammation and metabolites [24–28].

The present paper aims to evaluate the impact diet might have on immune response, with special attention as to how lifestyle changes can help mitigate low-grade inflammation. real-life risk simulation (RLRS) concept. This analysis can be highly relevant in the context of the present viral spread of SARS-COV-2, since the inflammation is once again in the front line of an acute pathological response. Identifying strategies to modulate the immune response might prove useful for reducing the virus's impact on the respiratory tract and thus diminishing its impact on each patient, as well as on the general medical system.

2. Food Frequency Questionnaires, Dietary Inflammation Index, and Metabolomics

The majority of studies that assess dietary habits, metabolism, and nutrient intake are based on food frequency questionnaires. However, food frequency questionnaires (FFQ) have several limitations, including inconsistent responses on food choices, mostly because answers depend on responders' memory. Moreover, FFQ that are filled out by the responder instead of a trained healthcare professional only provide an overview of the macronutrients' intake while not fully capturing the micronutrient status of the person [29]. Micronutrient deficiencies are common in both developing and affluent countries, affecting two billion people worldwide, according to the World Health Organization (WHO) [30]. The primary cause of micronutrient deficiencies or "hidden hunger" is poor dietary intake of micronutrients while other socioeconomic factors play an important role as well [31]. Several diet and nutrients assessment tools have been developed to evaluate inflammation status. The dietary inflammation index (DII) is based on literature data and aims to evaluate if a responder follows a proor anti-inflammatory diet. Since its development, there has been an increasing interest in DII, although other indexes are being developed with similar efficacy [32]. An important limitation of these indexes is the lack of causality and direct association to a person's symptoms. Thus, the application of these indexes in clinical practice is hampered.

A novel empirical, close-ended, and self-administered questionnaire developed by the European Institute Of Nutritional Medicine provides an inflammation status score that captures the interaction between the autonomic nervous system and inflammation [33]. There is growing evidence that imbalances in the autonomic nervous system reflect local or systemic inflammation found in various diseases and that diet and lifestyle factors can act as regulators of sympathetic and parasympathetic activity [34–37]. Through a 23-question series, responders provide data on the presence/absence and status of inflammatory response in different body systems. Overall, the nutritional medicine exam (Numex) consists of 118 questions and assesses the nutritional deficiencies status in seven categories: inflammation, nutrition, perceived stress, oxidation, sugar metabolism, amino acids metabolism, and gut microbiome. Designed by medical doctors and nutritionists, the aim of this empirical test is to assist the individual and the healthcare professional to evaluate the overall inflammatory status based on autonomic nervous system changes and track its progression after targeted lifestyle changes. At a molecular level, metabolomics is the only method that can capture small, time-dependent fluctuations in the metabolism, thus indicating NCD-related metabolic imbalances [21]. Overall, traditional and well-established diet and nutrient assessment methods including FFQ and DII have provided valuable information on the role of specific foods on health and disease, as discussed in the present review. With the advent of advanced tools, metabolomics is complementary to the standard approach to provide tailor-made recommendations depending on an individual's specific needs at a given time.

3. Changing Lifestyle as a Way to Mitigate Inflammation

Recent literature considers BMI cut-off values to not fully depict metabolic disturbances associated with obesity. The BMI is a mathematical approximation and does not reflect the percentage of total body fat between body fat and total body muscle or bone mass. As such, BMI does not reflect cardiometabolic

risk. A more comprehensive classification describes four phenotypes for obese individuals: normal weight obese (NWO), metabolically obese normal weight (MONW), metabolically healthy obese (MHO), and metabolically unhealthy obese (MUO), or "at risk" obese with MS. This classification takes into account BMI, fat mass, and waist circumference, but also general biochemical parameters (e.g., fasting plasma glucose, total cholesterol, LDL, HDL, triglycerides) [38–43]. All four classes are characterized by impairments of different severity of inflammatory pathways [38,40].

Lifestyle and nutrition are modifiable factors that interact with genetics in regulating chronic inflammation, leading to aforementioned complications. The changes in nutritional patterns in Western societies—caused by a high intake of fat and energy-dense, processed foods, as well as a low intake of fibers, fruits, and vegetables—are associated with a rising prevalence of asthma, allergies, and autoimmune diseases involving inflammatory mechanisms [44,45]. High fat diets determine, among other things: intestinal inflammation, favoring lipopolysaccharides (LPS) absorption from gram-negative gut bacteria, and increasing lipoperoxidation that induces insulin resistance and inflammation. Saturated fatty acids and LPS activate toll-like receptor 4 (TLR4) signaling pathways further contribute to promoting systemic inflammation and consequent metabolic disorders [46–52] (Figure 1).

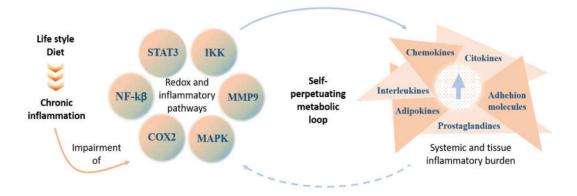


Figure 1. Impairment of metabolic pathways in lifestyle-associated low-grade chronic inflammation. STAT3 (signal transducer and activator of transcription 3), IKK (IκB kinase), MMP9 (matrix metallopeptidase 9), MAPK (mitogen-activated protein kinases), COX2 (cyclooxigenase 2), and NF-Kβ (nuclear factor kappa-light-chain-enhancer of activated B cells).

Lifestyle- and diet-induced inflammation affects several cellular pathways, which stimulates the synthesis and secretion of various pro-inflammatory molecules. This ultimately maintains the low-grade inflammation state.

Interestingly, populations that consume a diet rich in fruits, vegetables, and fibers have lower incidences of inflammatory diseases compared to Western populations [53,54]. The Mediterranean diet—based on olive oil, fish, vegetables, and fruits, in addition to incorporating myriad beneficial phytochemicals—discourages cardiovascular diseases [55–58]. Sourcing food from organic agriculture could further improve the beneficial health effects of a Mediterranean diet, as suggested in a study comparing an organic and nonorganic Mediterranean diet on male patients with chronic kidney disease [59]. This was hypothesized to be due to a decreased exposure to pesticides, since animal studies have repeatedly found that exposure to pesticide mixtures can be a source of toxicity [16]. However, although most studies have found that organic food consumers are healthier, it is not clear whether health benefits can be attributed to a decreased exposure to synthetic pesticides [60].

Dietary changes that include specific metabolites can modulate gene expression via epigenetic modifications, such as DNA methylation or chromatin remodeling (e.g., histone acetylation or deacetylation). For example, a diet rich in folate and methionine can shape the host epigenome with a direct impact on molecular pathways associated with obesity-related inflammation. Moreover, global

DNA hypermethylation in adipocytes derived from obese subjects is correlated with the expression of genes involved in proinflammatory interactions [61,62]. For example, hypermethylation at 1 kb upstream of the adiponectin gene's promoter site was observed in adipocytes of obese mice fed a high-fat diet, but also in human adipocytes. DNA methyltransferase 1 (DNMT1) expression is correlated with the methylation of the adiponectin gene, resulting in decreased expression of adiponectin in obese mice and increased expression in healthy mice. Studies on human adipocytes show a correlation between DNMT1 expression and BMI, suggesting that obesity is a cause or cofactor of hypermethylation of adiponectin gene [63,64]. Another factor inducing epigenetic changes is ROS overproduction in expanded adipose tissue, influencing histone acetylation/deacetylation equilibrium, thus inducing NF κ B activation [65–67].

On the other hand, nutrient restriction decreases AKT (protein kinase B) activity and stimulates FOXO (a forkhead box O transcription factor) activity, thus stimulating the expression of proteins involved in cell metabolism, autophagy, and stress-response, contributing to the resolution of inflammation [61,62]. Fasting regimens are correlated with increased insulin sensitivity, improvement of blood pressure, and inflammatory status, regardless if they are associated with weight reduction. For example, 15 days of intermittent fasting induced an increase of glucose uptake rates and a significant increase of anti-inflammatory adiponectin in lean young men (BMI of 25 kg/m²) without a significant decrease in body weight. These results were consistent with data from animal studies [68,69].

An important causal factor for low grade inflammation influenced mainly by lifestyle is the impairment of gut microbiota. Bacteroidetes and firmicutes constitute approximately 90% of the intestinal population, but the equilibrium is fundamentally changing with ageing and depending on diet composition. A decline of microbiota diversity occurs during ageing and obese individuals. Gut dysbiosis has been found in several inflammatory pathologies such as obesity, diabetes, cardiovascular, and neurodegenerative diseases. This can be connected to the induction of chronic low-grade inflammation since the gut microbiome is intimately connected to innate immune responses [70–72]. The relationship between gut microbiome and the host immune system are influenced by lifestyle interventions. For instance, secretory IgA levels increase after periodic fasting. This can be linked to changes in gut microbiome composition [14,73], with proteobacteria modulating the adaptive humoral local response.

Some studies showed that microbiota composition and diversity has a great impact on a population's general health status. For example, when comparing the fecal microbiota of European and rural African children (Burkina Faso), a higher proportion of Prevotella and Xylanibacter (involved in the digestion of fibers and generation of short chain fatty acids (SCFAs)) was found in the latter group, which lacked European subjects. These observations could be correlated with the higher prevalence of inflammatory diseases in European populations compared to rural African ones [74,75].

Chronic exposure to environmental pollutants or food additives could also predispose one to chronic pathologies, which promotes inflammation [76]. Xenobiotics promotes chronic inflammation, which is thought to be the generation of lipotoxic conditions, i.e., in the development non-alcoholic fatty liver disease [77]. This can be mitigated by lifestyle interventions such as periodic fasting [78]. The exposure to xenobiotics such as heavy metals, pesticides, nanoparticles, polycyclic aromatic hydrocarbons, dioxins, furans, polychlorinated biphenyls, or non-caloric artificial sweeteners can also promote chronic inflammation by disturbing the gut microbiota [77,79,80].

Decreasing inflammatory burden is more important than ever during the COVID-19 pandemic. This can be accomplished through everyday actions (e.g., lifestyle, diet, smoking cessation, weight decrease, sport, etc.). There is a lot of information available in the scientific community regarding the risk of COVID-19 complications; even the likelihood of death is highly increased by some chronic diseases, mostly associated with an impaired inflammatory profile (e.g., obesity, type II diabetes, hypertension, chronic pulmonary disease, etc.) [81,82]. The literature data shows that people without comorbidities have a much lower risk of severe symptoms as a result of the SARS-COV-2 infection [83]. On the other hand, increased levels of inflammatory markers cytokines with pro-inflammatory outcomes

constitute predictors of adverse outcome in COVID-19 patients [84]. Evidence proves that some dietary elements such as zinc or vitamin D might provide protective effects against viral load [84]. As such, this reduces the inflammatory burden through a healthy diet, associating (based on RLRS principles) several protective components (e.g., fiber, polyphenols, PUFAs, vitamins, etc.) that constantly increase our chance of being better protected against different immune challenges.

4. The Importance of Dietary Fiber for Reducing Inflammation

Fermentable dietary fiber are not enzymatically digested in the small intestine; they pass into the colon and are transformed by gut bacteria into SCFAs [85,86]. The systemic distribution and generation of SCFAs—acetate, propionate, and butyrate (the most abundant)—in the distal colon is important to inhibit inflammatory signals. Germ-free animal models were characterized by inflammatory flairs, due to the absence of tissue/blood SCFAs [87–89].

Butyrate is a representative member of SCFAs and has a high affinity for different G-protein-coupled receptors (GPCRs) found throughout the body: GPR41 is found in adipose tissues and immune cells and GPR43 is found in immune cells. However, GPR109A is present in colonic cells and GPR41 and GPR43 are activated by butyrate, which favors the production of peptide YY (PYY). This contributes to gastric emptying and intestinal transit inhibition, which thereby reduces appetite and promotes glucagon-like peptide 1 (GLP-1). These outcomes indirectly stimulate insulin secretion. GPR109A activates the inflammation-associated pathway in colonic macrophages and dendritic cells, inducing the differentiation of IL-10-producing T-cells and release of IL-18 from intestinal epithelial cells [85,90,91].

The presence of fiber in the diet is extremely important (Table 1) as it generates SCFAs and promotes the proliferation of commensal bacteria, which limits the access of pathogenic bacteria to the gut epithelium. Moreover, SCFAs favor epithelial mucus secretion that increases the protective effect on the intestinal surface and the proper maintenance of the barrier function [92,93]. SCFAs have anti-inflammatory effects that bind to the nuclear transcription factor PPAR γ (peroxisome proliferator-activated receptor γ) and, consequently, inhibit the NF-kB pathway [94,95]. This ultimately lowers the expression of VCAM-1 (vascular adhesion molecule 1) and ICAM-1 (intracellular adhesion molecule 1), as well as the synthesis of TNF α , IL-6, and IFN- γ (interferon γ) [96].

The main physiological role of histones is to interact with DNA and stabilize its structure. When they are acetylated, this loosens the contact between histones and DNA, uncoiling the DNA structure that thus becomes transcriptionally active. The histone acetylation process is a result of the balance between the induction of histone acetyl transferases (HATs) and the inhibition of histone deacetylase (HDACs). The same acetylation process causes DNA to bind to transcription factors, such as STAT3 (signal transducer and activator of transcription 3), NF-K β (nuclear factor kappa-light-chain-enhancer of activated B cells, and FoxP3. Consequently, this regulates gene expression, including inflammation proteins [97,98]. SCFAs (with butyrate and acetate being the most and least effective, respectively) act as inhibitors for histone deacetylase (HDACs), thus contributing to the inhibition of the transcription for inflammatory proteins [99,100].

Design	Population	Dietary Intervention	Outcome	Reference
Randomized cross-over trial	50 Danish subjects with high risk of metabolic syndrome	two eight-week dietary intervention periods of whole grain intake (179 \pm 50 g/day) and refined grain period (maximum 13 \pm 10 g/day of whole grain), separated by a washout period of \geq 6 weeks.	↓ body weight, serum inflammatory markers (IL-6, CRP)	[101]
Double-blind, randomized, placebo-controlled, crossover study	45 overweight adults with metabolic syndrome risk factors	galactooligosaccharide mixture intervention to increase dietary fiber content, with a 4-week wash-out period between interventions	↓ fecal calprotectin, CRP	[102]
Randomized controlled trial	143 individuals with metabolic syndrome	12 weeks of rye and whole wheat was compared with a diet containing the equivalent amount of refined cereal foods	no significant effects on the expression of inflammatory markers' genes or insulin sensitivity	[103]
Randomized crossover study	19 adults with metabolic syndrome	4-week interventions diet enriched with arabinoxylan and resistant starch compared to a low-fiber Western-style diet	↓ fecal calprotectin, IL-23A and NF-κB	[104]
Crossover intervention study	25 hypercholesterolemic subjects	5-week intervention using high fiber (HF) and low fiber (LF) diet, separated by a 3-week washout.	↓ CRP and fibrinogen	[105]
Randomized controlled trial	68 overweight with prediabetes	12 weeks of 45 g/d of high-amylose maize (RS2) versus an isocaloric amount of amylopectin (control)	\downarrow TNF-α, no change in insulin sensitivity	[106]
Randomized controlled trial	166 subjects with features of metabolic syndrome	4-week using healthy diet (whole-grain products, berries, fruits and vegetables, rapeseed oil, three fish meals per week) compared to an average Nordic diet	Control diet: ↑ IL-1 Ra (versus healthy diet group)	[107]
Crossover study	10 healthy subjects	Subjects received either 910-calorie high-fat/high-carbohydrate meal or an American Heart Association (AHA) meal (fruit and fiber) during the first visit and the other meal during the second visit	\uparrow oxidative stress (plasma concentrations of TBARS, FFA, and LPS) and proinflammatory markers (TNFα, and IL-1β)	[108]

Table 1. Clinical studies regarding the effect of high fiber intake on inflammatory markers in obesity and associated pathology.

Design	Population	Dietary Intervention	Outcome	Reference
Randomized controlled trial	28 T2DM patients	Subjects received brown rice $(n = 14)$ or white rice $(n = 14)$ diet for 8 weeks	↓ CRP in brown rice group	[109]
Parallel design, dietary intervention trial	104 subjects with metabolic syndrome risk	Subjects received Healthy Diet (n = 44), a whole-grain-enriched diet (n = 42) or a control (n = 45) diet,	Healthy Diet group: ↓ E-selectin Healthy Diet and whole grain group: ↓ CRP	[110]
Randomized, double-blind, placebo-controlled, cross-over study	12 overweight and obese subjects	Subjects received 20 g/day of inulin-propionate ester, a high-fermentable fiber control (inulin) and a low-fermentable fiber control (cellulose) for 42 days	IPE: ↓ IL-8 levels (versus cellulose) Inulin: no effect on the inflammatory markers	[111]
Crossover clinical study	18 subjects at low-to-moderate cardiometabolic risk	Subjects received breakfast rich in saturated fatty acids (SFA), the other in unsaturated fatty acids unSFA) and fiber for 4 weeks SFA: \uparrow IL-1 β unSFA: \downarrow IL-6		[112]
Interventional diet study	21 overweight/obese children	Subjects were placed on a regimen of ad libitum, high-fiber, low-fat diet, and daily exercise regimen for 2 weeks \downarrow IL-6, IL-8, TNF α , PAI-1, amylin, leptin, insulin, an \uparrow adiponectin		[113]
Randomized, placebo-controlled study	31 hemodialysis patients	Patients received either resistant starch or placebo supplementation for 4 weeks \downarrow IL-6 and TBARS		[114]
Randomized controlled clinical trial	55 women with T2DM	Subjects received a daily supplement of 10 g resistant dextrin or a similar amount of maltodextrin for 8 weeks	↓ IL-6, TNF-α and MDA ↑ Insulin sensitivity	[115]
Randomized cross-over double-blind placebo-controlled trial	17 obese knee osteoarthritis patients	Patients received freeze-dried strawberries or placebo for 2 periods of 12 weeks with 2 weeks of wash-out	\downarrow TNF- α and 4-HNE	[116]
Randomized study	59 T2DM patients	Patients received metformin, acarbose, and either a high fiber or a low fiber diet intervention for 8 weeks	Low fiber group: \downarrow IL-18	[117]

Table 1. Cont.

Design	Population	Dietary Intervention	Outcome	Reference
Crossover study	33 healthy, middle-aged adults	Patients received either high or low in in wholegrain intervention for 6-week periods, separated by a 4-week washout. Whole grain: a slight decrease IL-10 and CRP		[118]
Observational study	8 subjects with impaired fasting glucose	subjects received (1) high-fiber formula; (2) high-monounsaturated fatty acid formula or (3) control formula	High fiber group: \downarrow NF- κ B in PBMCs	[119]
Randomized controlled clinical trial	60 females with T2DM	Patients received 10 g/d resistant starch or placebo for 8 weeks, respectively	\downarrow TNF- α , no effect on IL-6 or CRP	[120]
Crossover clinical trial	80 overweight subjects	Subjects received two isocaloric breakfast interventions -one rich in saturated fat and one in unsaturated fatty acids and fibers for 4 weeks with a 2-weeks washout.	Fiber group: \downarrow IF- γ and TNF- α	[112]
Observational study	49 T2DM females	Patients received either 10 g/day inulin or maltodextrin/day for 8 weeks	Inulin: \downarrow CRP, TNF- α and LPS	[121]
Randomized controlled clinical trial	52 overweight/obese women with T2DM	Patients received either 10 g/d of oligofructose-enriched inulin or maltodextrin (control) for 8 weeks	oligofructose-enriched-Inulin: \downarrow CRP, TNF- α and LPS	[122]
Randomized crossover clinical trial	44 overweight/obese girls 8–15 years old	Subjects received either whole-grain or control for 2 periods of 6 weeks with 4-week washout period	Whole grain: ↓ CRP, ICAM-1 and leptin	[123]

Table 1. Cont.

IL-6, IL-1β, IL-6, IL-8, IL-18, IL-10—interleukin 6, 1β, 6, 8, 18, 10; IL-1 Ra—interleukin 1 receptor agonist; CRP—C reactive protein; NF-κB—nuclear factor kappa B; TNF-α—tumor necrosis factor α; TBARS—thiobarbituric acid reactive substances; FFA—free fatty acids; LPS—lipopolysaccharide; MDA—malondialdehyde; 4-HNE—4-hydroxynonenal; IF-γ—interferon γ; ICAM-1—intercellular adhesion molecule 1.

5. Anti-Inflammatory Effects of Fasting

Fasting is a process that has been known for thousands of years. It was quite frequent in ancient times, because access to food was difficult and as a result, individuals were obliged to survive without food until it was available again [124].

Fasting is a survival mechanism in both animals [125,126] and humans, especially in countries where food conservation is not widespread [127]. In the rest of the world, fasting has been employed either due to religious convictions or in wellness centers. Since 1960, one of the methods used to address morbid obesity and related diseases has been the "zero calorie diet", thus translating into clinical practice the scientific data generated by centuries of fasting.

Fasting can be divided into three broad categories:

- Periodic fasting, which lasts from 2 days to a few weeks;
- Intermittent fasting, which lasts from 16 to 20 h and can be done daily or every second day or twice a week, and
- Fasting-mimicking diet, the diet that mimics fasting to achieve its beneficial effects, in which restriction of calories and specific foods is necessary (e.g., fat) [128–130].

There is a lot of research that shows the beneficial effects of fasting on health and also on different pathological conditions. Fasting increased lifespan in prokaryotic organisms such as yeast S. Cerevisiae and nematode C. Elegans [128,131–134], but also on animal models that performed fasting for long periods (e.g., the royal penguin) [135]. Other models have shown better brain function [136,137], increased lifespan and longevity [138–140], and improved maintenance of muscle mass after fasting [141].

Studies on animal models reveal the beneficial effect fasting has on cancer as a complementary disease management strategy in concert with drug treatments [142–145]. Studies on animal models show an improvement in neurodegenerative diseases after fasting, while other studies prove that intermittent fasting diets boost the levels of antioxidant defense, neurotrophic factors (BDNFS, H-70 and FGF2), proteins involved in adaptive response (HSP-70 and GRP-78), and reduce pro-inflammatory cytokines levels (TNFa, IL-1β, and IL-6) [146–150].

It has been found that intermittent fasting can prevent and reverse all aspects of metabolic syndrome in rodents: body fat, inflammation, and blood pressure are reduced; insulin sensitivity is increased; and the functional capacity of the neuromuscular and cardiovascular systems are improved [151–153]. An intermittent fasting diet has also been found to improve hyper-glycaemia in diabetic rodent models [154] and in myocardial infarction models, as the heart is protected from ischemic damage by this type of regimen [155]. Elevated leptin levels usually predict a pre-inflammatory condition, while adiponectin and ghrelin may suppress inflammation and increase insulin sensitivity [150,156]. Fasting can reverse every major abnormality caused by metabolic syndrome, by increasing insulin and leptin sensitivity, suppressing inflammation and stimulating autophagy [157,158].

There are several studies that show an increased use of fat and ketone bodies for energy [159,160], as well as an increase of growth hormone and glucagon secretion [161–163], with a decrease in blood sugar, insulin, and IGF-1 levels. After intermittent fasting, total fat, abdominal fat, and blood pressure are decreased, while glucose metabolism is improved in obese individuals [69,164–167]. In addition, periodic fasting significantly changes the composition of the human gut microbiota [73]. Finally, studies of utmost importance show the effect of intermittent fasting mainly in the fight against cancer as a supplement along with the classic treatment, with promising results [168,169]. Below, we analyze the effects of the interrelation between fasting and inflammation as well as the relevant molecular mechanisms.

5.1. Molecular Mechanism of Fasting

Fasting not only results in weight loss but it is a survival mechanism that impacts many metabolic pathways [128,161,170]. Fasting's many benefits are related to the regulation of key molecular pathways.

Initially, during fasting, the downregulation of insulin-like growth-factor-1 (IGF-1) and mammalian target of rapamycin (mTOR) occur. These pathways are upregulated in the presence of food excess as they sense nutrients and therefore activate anabolic metabolism. When there is a lack of food for several hours, catabolic processes are activated.

Aging appears modulated by changes in the insulin-like growth-factor-1 receptor signaling system, as longevity is enhanced by a decrease in IGF-1 signaling [171–173]. The IGF-1 signal induces mTOR activation. Reduced mTOR activity is related to extended lifespan in different organisms [174], as mTOR induces activation of FoxO proteins. FoxO proteins are transported to the nucleus and activate genes associated with autophagy [175], which emphasizes the link between autophagy and FoxO proteins.

When the AMP/ATP ratio is high, the AMPK path is activated [176]; this results in increased energy production and reduced ATP utilization. In addition, the mitochondrial biogenesis and mitophagy repair and replace damaged mitochondria. As a result, the cells have "younger" and more efficient mitochondria. In addition to the aforementioned condition, activation of this path has been associated with increased lifespan in various studies in both C. Elegans and Drosophila melanogaster [177,178]. In mammals, fasting does not appear to affect AMPK activation, but further studies are needed to be able to draw surer conclusions [179].

Like AMPK, sirtuins are associated with life [180] and autophagy [181]. Some sirtuins are found in the cytoplasm (SIRT2), some (SIRT1) in the nucleus having DNA repair action, and others in mitochondria. In general, sirtuins are associated with mitochondrial biogenesis and mitophagy for damaged mitochondria, thereby enhancing mitochondrial cells without problems and are thus more efficient in energy production [132,182]. SIRT1 is modulated by NAD⁺ level and is increased in energy depletion states (such as fasting or exercise) for which NAD⁺ is a sensor, which contributes to the reduction of inflammation through NF-K β down-regulation and related transcription factors [132,182,183].

A study on rats showed that inflammation decreased with fasting [184]. Other work has shown NF-K β inhibition and the modulation of Nrf2, sirtuins, SOD2, and increased lifespan [185–188]. In a 2017 study, intermittent fasting appears to significantly reduce corticosterone (CORT), interleukin 6 (IL-6), and tumor necrosis factor-alpha (TNF- α) levels [189].

Nrf2 plays a key role in oxidative stress and toxicity; the right balance in ROS levels is very important so that mitochondrial and all other pathways can function properly. The absence of ROS, however, does not activate Nrf2, which in turn does not activate ARE (antioxidant response). Thus, a critical amount of ROS ("Hormesis hypothesis") is necessary for the upregulation of ARE, which allows cells and mitochondria to be able to deal with oxidative stress and different kinds of toxins [190], which consequently increases their lifespan [191] (Figure 2).

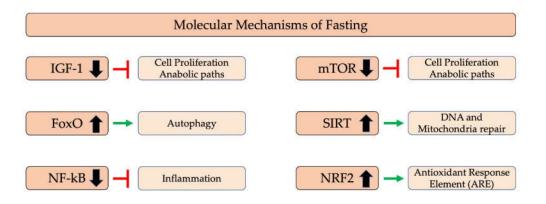


Figure 2. Molecular mechanisms of fasting. IGF-1, insulin-like growth factor 1; FoxO, forkhead box Proteins; NF-K β , nuclear factor-K β ; mTOR, mammalian target of rapamycin; SIRT, sirtuins; NRF2, nuclear factor erythroid 2-related factor 2.

5.2. Effects of Fasting on Humans

During the last decades, many studies have been conducted on the effect of fasting on several markers related to metabolism. Most of them determine the effect of fasting on weight. However, there are several studies that identify changes in lipid and carbohydrate metabolism as well as key hormones that affect the above (e.g., insulin). Recently, some studies have focused on fasting's effect on inflammatory markers such as TNF- α , interleukins, CRP, and BDNF, as well as the hormones adiponectin and leptin.

The largest study on fasting's effects is an observational study including 1422 subjects that describes metabolic changes after a 4- to 21-day fasting period [159,170]. All the participants fasted according to the Buchinger Wilhelmi fasting guidelines, which include a daily caloric intake of 200–250 kcal together with a variety of lifestyle changes (e.g., dietary advices, physical exercise). A beneficial modulating effect of fasting was observed on blood lipids, glucoregulation, and altogether general health-related blood parameters. Additionally, it was associated with a reduction in weight, abdominal circumference, and blood pressure. In another study, which used the same fasting guidelines, improved metabolic markers were observed after periodic fasting, including a decrease in blood glucose levels associated with changes in gut microbiome composition [73]. In this study, the analysis of the gut microbiome after 10 days of periodic fasting showed that fasting caused a decrease in the abundance of bacteria known to degrade dietary polysaccharides such as Lachnospiraceae and Ruminococcaceae, concomitant to an increase in Bacteroidetes and proteobacteria known to use host-derived energy substrates.

A study of eight healthy non-obese men discovered that in 15 days of fasting every other day, with 20 h fasting on fasting days, adiponectin increased and leptin decreased, while no changes in IL-6 or TNF- α were observed. Protocol allowed them to maintain normal exercise but also to consume food in order to keep their weight stable [69].

Redman et al., in a two-year study of 34 people who followed a reduced calorie intake diet (15%) observed a reduction in leptin. These individuals lost an average of 8.7 kg while the control group gained an average of 1.8 kg in the same period [192].

Another study including eight women and two men, all overweight with asthma, showed that fasting every other day and reducing calories to less than 20% of their normal intake on the days of fasting, for eight weeks, resulted in a reduction in TNF- α and BDNF, but no change in CRP. In this study, patients lost 8% of their initial weight during the study. Asthma symptoms also improved as well as some indicators of oxidative stress (8-protein carbonyls, isoprostane, nitrotyrosine, and 4-hydroxynonenal adducts) [193].

In 2013, another study [194] found that 12 weeks of reduced calorie intake every other day resulted in reduced CRP levels, increased adiponectin levels, and reduced leptin levels in 30 adults who were either overweight or normal weight. On fasting days, they consumed only 25% of the calories they normally consumed each day. In addition, their weight was significantly reduced by 3.6 ± 0.7 kg and the coronary heart disease risk was improved as the concentration of TG reduced.

Another study [195] found reduced high sensitivity-C-reactive protein (hs-CRP) of 27 women with polycystic ovary syndrome (PCOS) during the Ramadan period in Iran and in which participants aged 18 to 40 years old with an average of age 27.5, followed everyday 16.5 h fasting, isocaloric diet, for 29 days.

An important study involving 34 men (resistance-trained) who fasted every day 16 h, followed an isocaloric diet for eight weeks, and consumed 100% of their energy needs in the 8-h eating window, showed that there was an increase in adiponectin and a reduction of leptin of IL-6 and IL-1 β [196].

Faris' study on 50 healthy volunteers (21 men and 29 women) fasting for 14–15 h each day for 21 days, showed a reduction in IL-6, IL-1 β , TNF- α , total leukocytes, granulocytes, lymphocytes, and monocytes [197].

Another study [198] involving 42 patients aged 20 to 50 years old with nonalcoholic fatty liver disease, following Ramadan fasting (every day 16-h fasting for 29 days), showed a reduction in IL-6

and hs-CRP, compared to 41 volunteers who did not fast. These and other studies are presented in detail in Table 2.

Collectively, an increasing number of studies show that fasting has numerous health benefits and could be used to prevent or manage the development of cardiometabolic disorders, metabolic diseases, and immune diseases. Although extended periods of fasting can be challenging without medical advices, recent studies also showed that time-restricted eating can be practiced safely as a routine. For instance, a recent study showed that 10 h of time-restricted eating for 12 weeks improved cardiometabolic health for patients with metabolic syndrome [199].

Intervention Duration and Type of Fasting	Population	Comparison Group or Condition	Effects	Reference
15 days: alternate day fasting (20 h fasting intervals)	8 males non-obese adults	None	↑ adiponectin, ↓ leptin ↓ glucose, NS insulin, NS IL-6, NS TNF-α	[69]
29 days: every day 16.5 h fasting, isocaloric diet	27 females, polycystic ovary syndrome (PCOS)	None	NS insulin, NS LDL, NS HDL, NS TG, \downarrow hs-CRP ($p = 0,072$), \uparrow GSH, NS MDA, NS TAC	[195]
22 days: no caloric intake every other day (36 h fasting intervals)	8 females, 8 males, non-obese adults	None	NS glucose, \downarrow insulin, \downarrow weight	[167]
8weeks: every day 16 h fasting, isocaloric diet	34 resistance-trained males	2 groups time-restricted feeding (TRF) or normal diet group (ND).	NS Weight, ↓Fat mass, ↑Adiponectin, ↓ Leptin, ↓IL-6, ↓IL-1β, NS insulin, NS T3, NS Glucose, NS Cholesterol, NS Cortisol, NS HDL, NS LDL, TG↓,	[196]
1 day: water only (28 h fasting interval)	20 females, 10 males Healthy adults	None	↓ glucose, ↓ insulin, ↓ weight, \uparrow LDL \uparrow HDL, ↓ TG, NS CRP, NS adiponectin	[200]
29 days: every day 16 h fasting	83 patients with NAFLD	42 fasted, 41 control, comparison between groups	↓Weight, ↓BMI, NS BF, ↓Glucose, ↓insulin, ↓IL-6, ↓hs-CRP	[198]
20 weeks: 1 day per week fast OR 5-day consecutive fasts every 5 weeks (400–600 kcals on fasting days)	31 females, 23 males Overweight or obese diabetics	1200–1500 kcal weight loss diet	↓ weight, NS glucose, NS insulin, NS LDL, NS HDL	[201]
21 days: every day 14–15 h fasting	Fifty (21 men and 29 women) healthy volunteers	7 days before fasting vs. 21 days after fasting	\downarrow Weight, \downarrow BMI, \downarrow BF%, \downarrow SBP, \downarrow DPB, \downarrow IL-6, \downarrow IL-1 β , \downarrow TNF- α , \downarrow Total \downarrow leukocytes, \downarrow Granulosytes, \downarrow Lymphocytes, \downarrow Monocytes	[197]
8 weeks: <20% of usual intake on alternate days. Ad libitum diet on non-fasting days.	8 females, 2 males Overweight adults with asthma	None	↓ TNF-α, ↓ BDNF, ↓ weight, NS glucose, NS insulin, NS LDL, ↑ HDL, ↓ TGs, NS CRP, NS leptin, ↓ Protein Carbonyls, ↓ Nitrotyrosine, ↓ 8-isoprostane	[193]
2 years caloric restriction	34 CR (15%) and 19 control for 2 years	comparison between groups	\downarrow Weight, \downarrow BMI, \downarrow BF, \downarrow Leptin ns, Insulin NS, \downarrow 2,3-dinor-iPF(2 α)-III	[192]

Table 2. Recent reports concernation	erning fasting and inflammation.
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Intervention Duration and Type of Fasting	Population	Comparison Group or Condition	Effects	Reference
8 weeks: weight loss diet with alternate day modified fasting (~25% of total energy needs)	12 females, 8 males Obese adults	None	\downarrow weight, \downarrow LDL, NS HDL, \downarrow TGs	[165]
6 months: 25% energy restriction 2 days per week	107 females Young, overweight or obese adults	25% energy restriction 7 days per week	NS CRP, NS adiponectin, NS leptin NS BDNF NS glucose,↓insulin NS glucose,↓insulin	[166]
12 weeks: 25% of energy needs alternating with ad libitum intake	39 females, 2 males Obese adults	Control group	↓ weight, NS CRP, NS glucose NS insulin, NS LDL, NS HDL, NS TG	[202]
6±3 years caloric restriction	18 CR and 18 control	comparison between groups	↓Weight, ↓BMI, ↓ BF, ↓ LDL, ↓HDL, ↓TG, ↓ Cholesterol, ↓insulin, ↓glucose, ↓hs-CRP, ↓SBP, ↓DBP	[203]
6 weeks: 25–30% energy needs on Sat. Mon, Wed; ad libitum other days	15 females Overweight or obese	None	↓ weight, NS LDL, NS HDL, NS TG	[204]
12 weeks: 25% energy restriction 2 consecutive days per week	37 females Overweight or obese women	25% energy restriction all days of week	NS adiponectin, NS leptin, NS IL-6, NS TNF-α, NS glucose, NS HbA1c, ↓ insulin, NS LDL NS HDL, NS TG, NS weight	[205]
10 weeks intermittent fasting (IF)-calorie restriction (CR) regimen (with or without liquid meals)	54 were randomized to either the IFCR-liquid (IFCR-L) or IFCR-food based (IFCR-F) diet	comparison between 3rd and 10rd week in each group	In IFCR-L: ↓Leptin, ↓IL-6, ↓TNF-α, NS CRP, ↓IGF-1, In IFCR-F: ↓Leptin, NS IL-6, NS TNF-α, NS CRP, NS IGF-1,	[206]
12 weeks: weight loss diet with alternate day modified fasting (~25% of energy needs)	22 females, 8 males Normal and overweight adults	Control group	↓ CRP, ↑ adiponectin, ↓ leptin NS LDL, NS HDL, ↓ TG, ↓ weight	[194]

Table 2. Cont.

NS—non-significant; hs-CRP—highly sensitive CRP; LDL—low density lipoproteins; HDL—high density lipoproteins; GSH—glutathione; TAC—total antioxidant capacity; T3—triiodotironine; TG—triglycerides; BMI—body mass index; BF—body fat; SBP—systolic blood pressure; DBP—diastolic blood pressure; 2,3-dinor-iPF(2 α)-III –2,3-dinor-8-iso Prostaglandin F 2 α ; HbA1C—glycated hemoglobin; IGF-1—insulin growth factor 1.

6. Conclusions

Given that a large part of the global population suffers from various metabolic disorders, it is important to look for non-pharmacological ways to deal with these conditions. Targeted changes in lifestyle and especially diet can be economical tools to mitigate the development of metabolic disorders when they are at an early stage. These changes include increased fiber and polyphenol intake compared to the current western diets, but also well-structured, personalized fasting protocols, which can reduce the risk of metabolic disorders (Figure 3).

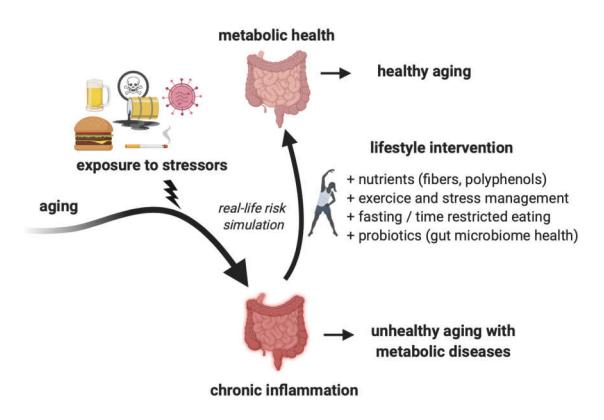


Figure 3. Lifestyle changes can promote healthy aging by mitigating the deleterious effects of stressors. Exposure to a variety of stressors (processed food, overfeeding, environmental toxicants, infectious agents, or drugs) during life course can promote chronic inflammation and the development of metabolic disorders. A large number of real-life risk simulation studies showed that this can be mitigated by lifestyle interventions.

This could be implemented in various institutions by improving the nutritional quality of foods served in schools, hospitals, prisons, government buildings, or senior centers. Moving toward healthier food options in hospitals might ultimately help improve patient health and reduce healthcare costs.

Improving diet can have a positive effect on the immune response as a hallmark of RLRS. Since inflammation is associated with the acute pathological response to COVID-19 and other infectious diseases, improvement of the immune response and inflammatory markers may lead to an improved physiological resilience to disturbances by infectious agents such as viruses and bacteria, and possibly milder symptoms.

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References

- Libby, P. Inflammation in atherosclerosis. *Arterioscler. Thromb Vasc. Biol.* 2012, 32, 2045–2051. [CrossRef] [PubMed]
- Reddy, P.; Lent-Schochet, D.; Ramakrishnan, N.; McLaughlin, M.; Jialal, I. Metabolic syndrome is an inflammatory disorder: A conspiracy between adipose tissue and phagocytes. *Clin. Chim. Acta* 2019, 496, 35–44. [CrossRef] [PubMed]
- Oresic, M.; Simell, S.; Sysi-Aho, M.; Nanto-Salonen, K.; Seppanen-Laakso, T.; Parikka, V.; Katajamaa, M.; Hekkala, A.; Mattila, I.; Keskinen, P.; et al. Dysregulation of lipid and amino acid metabolism precedes islet autoimmunity in children who later progress to type 1 diabetes. *J. Exp. Med.* 2008, 205, 2975–2984. [CrossRef] [PubMed]
- 4. Lamichhane, S.; Kemppainen, E.; Trost, K.; Siljander, H.; Hyoty, H.; Ilonen, J.; Toppari, J.; Veijola, R.; Hyotylainen, T.; Knip, M.; et al. Circulating metabolites in progression to islet autoimmunity and type 1 diabetes. *Diabetologia* **2019**, *62*, 2287–2297. [CrossRef] [PubMed]
- 5. De Rosa, V.; La Cava, A.; Matarese, G. Metabolic pressure and the breach of immunological self-tolerance. *Nat. Immunol.* **2017**, *18*, 1190–1196. [CrossRef] [PubMed]
- 6. Gradinaru, D.; Khaddour, H.; Margina, D.; Ungurianu, A.; Borsa, C.; Ionescu, C.; Prada, G.I.; Usher, J.; Elshimali, Y. Insulin-Leptin Axis, Cardiometabolic Risk and Oxidative Stress in Elderly with Metabolic Syndrome. *Exp. Clin. Endocrinol. Diabetes* **2018**. [CrossRef] [PubMed]
- Gradinaru, D.; Margina, D.; Borsa, C.; Ionescu, C.; Ilie, M.; Costache, M.; Dinischiotu, A.; Prada, G.I. Adiponectin: Possible link between metabolic stress and oxidative stress in the elderly. *Aging Clin. Exp. Res.* 2017, 29, 621–629. [CrossRef] [PubMed]
- 8. Galland, L. Diet and inflammation. Nutr. Clin. Pract. 2010, 25, 634–640. [CrossRef] [PubMed]
- 9. Lewis, J.D.; Chen, E.Z.; Baldassano, R.N.; Otley, A.R.; Griffiths, A.M.; Lee, D.; Bittinger, K.; Bailey, A.; Friedman, E.S.; Hoffmann, C.; et al. Inflammation, Antibiotics, and Diet as Environmental Stressors of the Gut Microbiome in Pediatric Crohn's Disease. *Cell Host Microbe* **2015**, *18*, 489–500. [CrossRef] [PubMed]
- 10. Hills, R.D., Jr.; Pontefract, B.A.; Mishcon, H.R.; Black, C.A.; Sutton, S.C.; Theberge, C.R. Gut Microbiome: Profound Implications for Diet and Disease. *Nutrients* **2019**, *11*, 1613. [CrossRef] [PubMed]
- 11. Gachon, F.; Yeung, J.; Naef, F. Cross-regulatory circuits linking inflammation, high-fat diet, and the circadian clock. *Genes Dev.* **2018**, *32*, 1359–1360. [CrossRef] [PubMed]
- 12. Fountoucidou, P.; Veskoukis, A.S.; Kerasioti, E.; Docea, A.O.; Taitzoglou, I.A.; Liesivuori, J.; Tsatsakis, A.; Kouretas, D. A mixture of routinely encountered xenobiotics induces both redox adaptations and perturbations in blood and tissues of rats after a long-term low-dose exposure regimen: The time and dose issue. *Toxicol. Lett.* **2019**, *317*, 24–44. [CrossRef] [PubMed]
- 13. Petrakis, D.; Vassilopoulou, L.; Mamoulakis, C.; Psycharakis, C.; Anifantaki, A.; Sifakis, S.; Docea, A.O.; Tsiaoussis, J.; Makrigiannakis, A.; Tsatsakis, A.M. Endocrine Disruptors Leading to Obesity and Related Diseases. *Int. J. Environ. Res. Public Health* **2017**, *14*, 1282. [CrossRef] [PubMed]
- 14. Margina, D.; Nitulescu, G.M.; Ungurianu, A.; Mesnage, R.; Goumenou, M.; Sarigiannis, D.A.; Aschner, M.; Spandidos, D.A.; Renieri, E.A.; Hernandez, A.F.; et al. Overview of the effects of chemical mixtures with endocrine disrupting activity in the context of real-life risk simulation: An integrative approach (Review). *World Acad. Sci. J.* **2019**, *1*, 157–164. [CrossRef] [PubMed]
- Tsatsakis, A.; Docea, A.O.; Constantin, C.; Calina, D.; Zlatian, O.; Nikolouzakis, T.K.; Stivaktakis, P.D.; Kalogeraki, A.; Liesivuori, J.; Tzanakakis, G.; et al. Genotoxic, cytotoxic, and cytopathological effects in rats exposed for 18 months to a mixture of 13 chemicals in doses below NOAEL levels. *Toxicol. Lett.* 2019, 316, 154–170. [CrossRef] [PubMed]
- 16. Docea, A.O.; Gofita, E.; Goumenou, M.; Calina, D.; Rogoveanu, O.; Varut, M.; Olaru, C.; Kerasioti, E.; Fountoucidou, P.; Taitzoglou, I.; et al. Six months exposure to a real life mixture of 13 chemicals' below individual NOAELs induced non monotonic sex-dependent biochemical and redox status changes in rats. *Food Chem. Toxicol.* **2018**, *115*, 470–481. [CrossRef] [PubMed]
- Tsatsakis, A.; Petrakis, D.; Nikolouzakis, T.; Docea, A.; Calina, D.; Vinceti, M.; Goumenou, M.; Kostoff, R.; Mamoulakis, C.; Aschne, R.M.; et al. COVID-19, an opportunity to reevaluate the correlation between long-term effects of anthropogenic pollutants on viral epidemic/pandemic events and prevalence. *Food Chem. Toxicol.* 2020. [CrossRef] [PubMed]

- Petrakis, D.; Margina, D.; Tsarouhas, K.; Tekos, F.; Stan, M.; Nikitovic, D.; Kouretas, D.; Spandidos, D.A.; Tsatsakis, A. Obesity a risk factor for increased COVID 19 prevalence, severity and lethality (Review). *Mol. Med. Rep.* 2020. [CrossRef] [PubMed]
- 19. Clish, C.B. Metabolomics: An emerging but powerful tool for precision medicine. *Cold Spring Harb. Mol. Case Stud.* **2015**, *1*, a000588. [CrossRef] [PubMed]
- 20. Tsoukalas, D.; Alegakis, A.; Fragkiadaki, P.; Papakonstantinou, E.; Nikitovic, D.; Karataraki, A.; Nosyrev, A.E.; Papadakis, E.G.; Spandidos, D.A.; Drakoulis, N.; et al. Application of metabolomics: Focus on the quantification of organic acids in healthy adults. *Int. J. Mol. Med.* **2017**, *40*, 112–120. [CrossRef] [PubMed]
- 21. Tsoukalas, D.; Fragoulakis, V.; Sarandi, E.; Docea, A.O.; Papakonstaninou, E.; Tsilimidos, G.; Anamaterou, C.; Fragkiadaki, P.; Aschner, M.; Tsatsakis, A.; et al. Targeted Metabolomic Analysis of Serum Fatty Acids for the Prediction of Autoimmune Diseases. *Front. Mol. Biosci.* **2019**, *6*, 120. [CrossRef] [PubMed]
- 22. Tsatsakis, A.; Tyshko, N.V.; Docea, A.O.; Shestakova, S.I.; Sidorova, Y.S.; Petrov, N.A.; Zlatian, O.; Mach, M.; Hartung, T.; Tutelyan, V.A. The effect of chronic vitamin deficiency and long term very low dose exposure to 6 pesticides mixture on neurological outcomes—A real-life risk simulation approach. *Toxicol. Lett.* **2019**, *315*, 96–106. [CrossRef] [PubMed]
- 23. Hernandez, A.F.; Docea, A.O.; Goumenou, M.; Sarigiannis, D.; Aschner, M.; Tsatsakis, A. Application of novel technologies and mechanistic data for risk assessment under the real-life risk simulation (RLRS) approach. *Food Chem. Toxicol.* **2020**, *137*, 111123. [CrossRef] [PubMed]
- 24. Fullerton, J.N.; Gilroy, D.W. Resolution of inflammation: A new therapeutic frontier. *Nat. Rev. Drug Discov.* **2016**, *15*, 551–567. [CrossRef] [PubMed]
- 25. Serhan, C.N.; Chiang, N. Endogenous pro-resolving and anti-inflammatory lipid mediators: A new pharmacologic genus. *Br. J. Pharmacol.* **2008**, *153*, S200–S215. [CrossRef] [PubMed]
- 26. Serhan, C.N.; Chiang, N.; Van Dyke, T.E. Resolving inflammation: Dual anti-inflammatory and pro-resolution lipid mediators. *Nat Rev. Immunol.* **2008**, *8*, 349–361. [CrossRef] [PubMed]
- 27. Nimigean, V.; Poll, A.; Minculescu, C.A.; Nimigean, V.R.; Moraru, S.A.; Vîrlan, M.J.R.; Bălăceanu, R.A.; Păun, D.L. Immunohistochemical evaluation of autogenous mandibular bone grafts integration: An experimental study. *Rom Biotechnol. Lett.* **2019**, *24*, 229–235. [CrossRef]
- Nimigean, V.; Poll, A.; Nimigean, V.; Moraru, S.; Badita, D.; Paun, D. The Routine and Specialised Staining for the Histologic Evaluation of Autogenous Mandibular Bone Grafts An experimental study. *Rev. Chim.* 2018, 69, 1106–1109. [CrossRef]
- 29. Sedlmeier, A.; Kluttig, A.; Giegling, I.; Prehn, C.; Adamski, J.; Kastenmuller, G.; Lacruz, M.E. The human metabolic profile reflects macro-and micronutrient intake distinctly according to fasting time. *Sci. Rep.* **2018**, *8*, 12262. [CrossRef] [PubMed]
- 30. WHO/WFP/UNICEF. Preventing and Controlling Micronutrient Deficiencies in Populations Affected by an Emergency. Available online: https://www.who.int/docs/default-source/nutritionlibrary/preventing-and-controlling-micronutrient-deficiencies-in-populations-affected-by-an-emergency.pdf?sfvrsn=e17f6dff_2 (accessed on 25 May 2020).
- Bailey, R.L.; West, K.P., Jr.; Black, R.E. The epidemiology of global micronutrient deficiencies. *Ann. Nutr. Metab.* 2015, *66*, 22–33. [CrossRef] [PubMed]
- 32. Tabung, F.K.; Smith-Warner, S.A.; Chavarro, J.E.; Fung, T.T.; Hu, F.B.; Willett, W.C.; Giovannucci, E.L. An Empirical Dietary Inflammatory Pattern Score Enhances Prediction of Circulating Inflammatory Biomarkers in Adults. *J. Nutr.* 2017, *147*, 1567–1577. [CrossRef] [PubMed]
- 33. EINM. Test Numex. Available online: https://www.einum.org/numex-test/ (accessed on 26 May 2020).
- 34. Pongratz, G.; Straub, R.H. The sympathetic nervous response in inflammation. *Arthritis Res. Ther.* **2014**, *16*, 504. [CrossRef] [PubMed]
- 35. Marvar, P.J.H.; Harrison, D.G. Inflammation, Immunity and the Autonomic Nervous System. In *Primer on the Autonomic Nervous System*; Robertson, D., Ed.; Academic Press: Cambridge, MA, USA, 2012; pp. 325–329. [CrossRef]
- Luyer, M.D.; Habes, Q.; van Hak, R.; Buurman, W. Nutritional stimulation of the autonomic nervous system. World J. Gastroenterol. 2011, 17, 3859–3863. [CrossRef] [PubMed]
- 37. Costa, J.; Moreira, A.; Moreira, P.; Delgado, L.; Silva, D. Effects of weight changes in the autonomic nervous system: A systematic review and meta-analysis. *Clin. Nutr.* **2019**, *38*, 110–126. [CrossRef] [PubMed]

- 38. Goossens, G.H. The Metabolic Phenotype in Obesity: Fat Mass, Body Fat Distribution, and Adipose Tissue Function. *Obes. Facts* **2017**, *10*, 207–215. [CrossRef] [PubMed]
- 39. De Lorenzo, A.; Martinoli, R.; Vaia, F.; Di Renzo, L. Normal weight obese (NWO) women: An evaluation of a candidate new syndrome. *Nutr. Metab. Cardiovasc. Dis.* **2006**, *16*, 513–523. [CrossRef] [PubMed]
- De Lorenzo, A.; Soldati, L.; Sarlo, F.; Calvani, M.; Di Lorenzo, N.; Di Renzo, L. New obesity classification criteria as a tool for bariatric surgery indication. *World J. Gastroenterol.* 2016, 22, 681–703. [CrossRef] [PubMed]
- Karelis, A.D.; Faraj, M.; Bastard, J.P.; St-Pierre, D.H.; Brochu, M.; Prud'homme, D.; Rabasa-Lhoret, R. The metabolically healthy but obese individual presents a favorable inflammation profile. *J. Clin. Endocrinol. Metab.* 2005, *90*, 4145–4150. [CrossRef] [PubMed]
- Karelis, A.D.; St-Pierre, D.H.; Conus, F.; Rabasa-Lhoret, R.; Poehlman, E.T. Metabolic and body composition factors in subgroups of obesity: What do we know? *J. Clin. Endocrinol. Metab.* 2004, *89*, 2569–2575. [CrossRef] [PubMed]
- 43. WHO. Obesity: Preventing and managing the global epidemic. Report of a WHO consultation. *World Health Organ. Tech. Rep. Ser.* **2000**, *894*, 1–253.
- 44. Manzel, A.; Muller, D.N.; Hafler, D.A.; Erdman, S.E.; Linker, R.A.; Kleinewietfeld, M. Role of "Western diet' in inflammatory autoimmune diseases. *Curr. Allergy Asthma Rep.* **2014**, *14*, 404. [CrossRef] [PubMed]
- 45. Statovci, D.; Aguilera, M.; MacSharry, J.; Melgar, S. The Impact of Western Diet and Nutrients on the Microbiota and Immune Response at Mucosal Interfaces. *Front. Immunol.* **2017**, *8*, 838. [CrossRef] [PubMed]
- 46. Brun, P.; Castagliuolo, I.; Di Leo, V.; Buda, A.; Pinzani, M.; Palu, G.; Martines, D. Increased intestinal permeability in obese mice: New evidence in the pathogenesis of nonalcoholic steatohepatitis. *Am. J. Physiol. Gastrointest. Liver Physiol.* **2007**, 292, G518–G525. [CrossRef] [PubMed]
- Chaurasia, B.; Summers, S.A. Ceramides—Lipotoxic Inducers of Metabolic Disorders. *Trends Endocrinol. Metab.* 2015, 26, 538–550. [CrossRef] [PubMed]
- 48. Ding, S.; Lund, P.K. Role of intestinal inflammation as an early event in obesity and insulin resistance. *Curr. Opin. Clin. Nutr. Metab. Care* **2011**, *14*, 328–333. [CrossRef] [PubMed]
- 49. Ghoshal, S.; Witta, J.; Zhong, J.; de Villiers, W.; Eckhardt, E. Chylomicrons promote intestinal absorption of lipopolysaccharides. *J. Lipid Res.* **2009**, *50*, 90–97. [CrossRef] [PubMed]
- 50. Lu, Y.C.; Yeh, W.C.; Ohashi, P.S. LPS/TLR4 signal transduction pathway. *Cytokine* **2008**, 42, 145–151. [CrossRef] [PubMed]
- Ungurianu, A.; Seremet, O.; Gagniuc, E.; Olaru, O.T.; Gutu, C.; Gradinaru, D.; Ionescu-Tirgoviste, C.; Margina, D.; Danciulescu-Miulescu, R. Preclinical and clinical results regarding the effects of a plant-based antidiabetic formulation versus well established antidiabetic molecules. *Pharmacol. Res.* 2019, 150, 104522. [CrossRef] [PubMed]
- 52. Ungurianu, A.; Margina, D.; Gradinaru, D.; Bacanu, C.; Ilie, M.; Tsitsimpikou, C.; Tsarouhas, K.; Spandidos, D.A.; Tsatsakis, A.M. Lipoprotein redox status evaluation as a marker of cardiovascular disease risk in patients with inflammatory disease. *Mol. Med. Rep.* **2017**, *15*, 256–262. [CrossRef] [PubMed]
- 53. Eder, W.; Ege, M.J.; von Mutius, E. The asthma epidemic. *N. Engl. J. Med.* **2006**, 355, 2226–2235. [CrossRef] [PubMed]
- 54. Park, Y.; Subar, A.F.; Hollenbeck, A.; Schatzkin, A. Dietary fiber intake and mortality in the NIH-AARP diet and health study. *Arch. Intern. Med.* **2011**, *171*, 1061–1068. [CrossRef] [PubMed]
- Berthon, B.S.; Macdonald-Wicks, L.K.; Gibson, P.G.; Wood, L.G. Investigation of the association between dietary intake, disease severity and airway inflammation in asthma. *Respirology* 2013, *18*, 447–454. [CrossRef] [PubMed]
- 56. Estruch, R.; Ros, E.; Salas-Salvado, J.; Covas, M.I.; Corella, D.; Aros, F.; Gomez-Gracia, E.; Ruiz-Gutierrez, V.; Fiol, M.; Lapetra, J.; et al. Primary prevention of cardiovascular disease with a Mediterranean diet. *N. Engl. J. Med.* 2013, 368, 1279–1290. [CrossRef] [PubMed]
- 57. Nagel, G.; Weinmayr, G.; Kleiner, A.; Garcia-Marcos, L.; Strachan, D.P.; Group, I.P.T.S. Effect of diet on asthma and allergic sensitisation in the International Study on Allergies and Asthma in Childhood (ISAAC) Phase Two. *Thorax* **2010**, *65*, 516–522. [CrossRef] [PubMed]
- Margina, D.; Ilie, M.; Gradinaru, D. Quercetin and epigallocatechin gallate induce in vitro a dose-dependent stiffening and hyperpolarizing effect on the cell membrane of human mononuclear blood cells. *Int. J. Mol. Sci.* 2012, *13*, 4839–4859. [CrossRef] [PubMed]

- De Lorenzo, A.; Noce, A.; Bigioni, M.; Calabrese, V.; Della Rocca, D.G.; Di Daniele, N.; Tozzo, C.; Di Renzo, L. The effects of Italian Mediterranean organic diet (IMOD) on health status. *Curr. Pharm. Des.* 2010, 16, 814–824. [CrossRef] [PubMed]
- 60. Mesnage, R.; Tsakiris, I.N.; Antoniou, M.N.; Tsatsakis, A. Limitations in the evidential basis supporting health benefits from a decreased exposure to pesticides through organic food consumption. *Curr. Opin. Toxicol.* **2020**, *19*, 50–55. [CrossRef]
- 61. Liu, J.J.; Ward, R.L. Folate and one-carbon metabolism and its impact on aberrant DNA methylation in cancer. *Adv. Genet.* **2010**, *71*, 79–121. [CrossRef] [PubMed]
- Naidoo, V.; Naidoo, M.; Ghai, M. Cell- and tissue-specific epigenetic changes associated with chronic inflammation in insulin resistance and type 2 diabetes mellitus. *Scand. J. Immunol.* 2018, *88*, e12723. [CrossRef] [PubMed]
- 63. Kim, A.Y.; Park, Y.J.; Pan, X.; Shin, K.C.; Kwak, S.H.; Bassas, A.F.; Sallam, R.M.; Park, K.S.; Alfadda, A.A.; Xu, A.; et al. Obesity-induced DNA hypermethylation of the adiponectin gene mediates insulin resistance. *Nat. Commun.* **2015**, *6*, 7585. [CrossRef] [PubMed]
- 64. Furukawa, S.; Fujita, T.; Shimabukuro, M.; Iwaki, M.; Yamada, Y.; Nakajima, Y.; Nakayama, O.; Makishima, M.; Matsuda, M.; Shimomura, I. Increased oxidative stress in obesity and its impact on metabolic syndrome. *J. Clin. Invest.* **2004**, *114*, 1752–1761. [CrossRef] [PubMed]
- 65. Rahman, I. Dietary polyphenols mediated regulation of oxidative stress and chromatin remodeling in inflammation. *Nutr. Rev.* **2008**, *66*, S42–S45. [CrossRef] [PubMed]
- 66. Kasinska, M.A.; Drzewoski, J.; Sliwinska, A. Epigenetic modifications in adipose tissue—Relation to obesity and diabetes. *Arch. Med. Sci.* 2016, *12*, 1293–1301. [CrossRef] [PubMed]
- 67. Bronte, V.; Pittet, M.J. The spleen in local and systemic regulation of immunity. *Immunity* **2013**, *39*, 806–818. [CrossRef] [PubMed]
- 68. Anson, R.M.; Guo, Z.; de Cabo, R.; Iyun, T.; Rios, M.; Hagepanos, A.; Ingram, D.K.; Lane, M.A.; Mattson, M.P. Intermittent fasting dissociates beneficial effects of dietary restriction on glucose metabolism and neuronal resistance to injury from calorie intake. *Proc. Natl. Acad. Sci. USA* **2003**, *100*, 6216–6220. [CrossRef] [PubMed]
- Halberg, N.; Henriksen, M.; Soderhamn, N.; Stallknecht, B.; Ploug, T.; Schjerling, P.; Dela, F. Effect of intermittent fasting and refeeding on insulin action in healthy men. *J. Appl. Physiol.* 1985 2005, 99, 2128–2136. [CrossRef] [PubMed]
- Westfall, S.; Lomis, N.; Kahouli, I.; Dia, S.Y.; Singh, S.P.; Prakash, S. Microbiome, probiotics and neurodegenerative diseases: Deciphering the gut brain axis. *Cell Mol. Life Sci.* 2017, 74, 3769–3787. [CrossRef]
- 71. Martins, R.; Lithgow, G.J.; Link, W. Long live FOXO: Unraveling the role of FOXO proteins in aging and longevity. *Aging Cell* **2016**, *15*, 196–207. [CrossRef] [PubMed]
- 72. Power, S.E.; O'Toole, P.W.; Stanton, C.; Ross, R.P.; Fitzgerald, G.F. Intestinal microbiota, diet and health. *Br. J. Nutr.* **2014**, *111*, 387–402. [CrossRef] [PubMed]
- 73. Mesnage, R.; Grundler, F.; Schwiertz, A.; Le Maho, Y.; Wilhelmi de Toledo, F. Changes in human gut microbiota composition are linked to the energy metabolic switch during 10 d of Buchinger fasting. *J. Nutr. Sci.* **2019**, *8*, e36. [CrossRef] [PubMed]
- 74. Ou, J.; Carbonero, F.; Zoetendal, E.G.; DeLany, J.P.; Wang, M.; Newton, K.; Gaskins, H.R.; O'Keefe, S.J. Diet, microbiota, and microbial metabolites in colon cancer risk in rural Africans and African Americans. *Am. J. Clin. Nutr.* 2013, *98*, 111–120. [CrossRef] [PubMed]
- 75. De Filippo, C.; Cavalieri, D.; Di Paola, M.; Ramazzotti, M.; Poullet, J.B.; Massart, S.; Collini, S.; Pieraccini, G.; Lionetti, P. Impact of diet in shaping gut microbiota revealed by a comparative study in children from Europe and rural Africa. *Proc. Natl. Acad. Sci. USA* **2010**, *107*, 14691–14696. [CrossRef] [PubMed]
- 76. Tsatsakis, A.; Goumenou, M.; Liesivuori, J.; Dekant, W.; Hernandez, A.F. Toxicology for real-life risk simulation—Editorial preface to this special issue. *Toxicol. Lett.* **2019**, *309*, 33–34. [CrossRef] [PubMed]
- 77. Mesnage, R.; Biserni, M.; Balu, S.; Frainay, C.; Poupin, N.; Jourdan, F.; Wozniak, E.; Xenakis, T.; Mein, C.A.; Antoniou, M.N. Integrated transcriptomics and metabolomics reveal signatures of lipid metabolism dysregulation in HepaRG liver cells exposed to PCB 126. *Arch. Toxicol.* 2018, *92*, 2533–2547. [CrossRef] [PubMed]

- Drinda, S.; Grundler, F.; Neumann, T.; Lehmann, T.; Steckhan, N.; Michalsen, A.; Wilhelmi de Toledo, F. Effects of Periodic Fasting on Fatty Liver Index-A Prospective Observational Study. *Nutrients* 2019, *11*, 2601. [CrossRef] [PubMed]
- 79. Tsiaoussis, J.; Antoniou, M.N.; Koliarakis, I.; Mesnage, R.; Vardavas, C.I.; Izotov, B.N.; Psaroulaki, A.; Tsatsakis, A. Effects of single and combined toxic exposures on the gut microbiome: Current knowledge and future directions. *Toxicol. Lett.* **2019**, *312*, 72–97. [CrossRef] [PubMed]
- 80. Lobionda, S.; Sittipo, P.; Kwon, H.Y.; Lee, Y.K. The Role of Gut Microbiota in Intestinal Inflammation with Respect to Diet and Extrinsic Stressors. *Microorganisms* **2019**, *7*, 271. [CrossRef] [PubMed]
- Farsalinos, K.; Niaura, R.; Le Houezec, J.; Barboun, I.A.; Tsatsakis, A.; Kouretas, D.; Vantarakis, A.; Poulas, K. Nicotine and SARS-CoV-2: COVID-19 may be a disease of the nicotinic cholinergic system. *Toxicol. Rep.* 2020. [CrossRef] [PubMed]
- Docea, A.O.; Tsatsakis, A.; Albulescu, D.; Cristea, O.; Zlatian, O.; Vinceti, M.; Moschos, S.A.; Tsoukalas, D.; Goumenou, M.; Drakoulis, N.; et al. A new threat from an old enemy: Reemergence of coronavirus (Review). *Int. J. Mol. Med.* 2020. [CrossRef] [PubMed]
- 83. Goumenou, M.; Sarigiannis, D.; Tsatsakis, A.; Anesti, O.; Docea, A.O.; Petrakis, D.; Tsoukalas, D.; Kostoff, R.; Rakitskii, V.; Spandidos, D.A.; et al. COVID19 in Northern Italy: An integrative overview of factors possibly influencing the sharp increase of the outbreak (Review). *Mol. Med. Rep.* **2020**. [CrossRef]
- 84. Skalny, A.V.; Rink, L.; Ajsuvakova, O.P.; Aschner, M.; Gritsenko, V.A.; Alekseenko, S.I.; Svistunov, A.A.; Petrakis, D.; Spandidos, D.A.; Aaseth, J.; et al. Zinc and respiratory tract infections: Perspectives for COVID19 (Review). *Int. J. Mol. Med.* **2020**. [CrossRef] [PubMed]
- 85. Bach Knudsen, K.E.; Laerke, H.N.; Hedemann, M.S.; Nielsen, T.S.; Ingerslev, A.K.; Gundelund Nielsen, D.S.; Theil, P.K.; Purup, S.; Hald, S.; Schioldan, A.G.; et al. Impact of Diet-Modulated Butyrate Production on Intestinal Barrier Function and Inflammation. *Nutrients* **2018**, *10*, 1499. [CrossRef]
- 86. Flint, H.J. The impact of nutrition on the human microbiome. Nutr. Rev. 2012, 70, S10–S13. [CrossRef]
- 87. Herbst, T.; Sichelstiel, A.; Schar, C.; Yadava, K.; Burki, K.; Cahenzli, J.; McCoy, K.; Marsland, B.J.; Harris, N.L. Dysregulation of allergic airway inflammation in the absence of microbial colonization. *Am. J. Respir. Crit. Care Med.* **2011**, *184*, 198–205. [CrossRef] [PubMed]
- Maslowski, K.M.; Vieira, A.T.; Ng, A.; Kranich, J.; Sierro, F.; Yu, D.; Schilter, H.C.; Rolph, M.S.; Mackay, F.; Artis, D.; et al. Regulation of inflammatory responses by gut microbiota and chemoattractant receptor GPR43. *Nature* 2009, 461, 1282–1286. [CrossRef]
- 89. Tan, J.; McKenzie, C.; Potamitis, M.; Thorburn, A.N.; Mackay, C.R.; Macia, L. The role of short-chain fatty acids in health and disease. *Adv. Immunol.* **2014**, *121*, 91–119. [CrossRef] [PubMed]
- 90. Bindels, L.B.; Dewulf, E.M.; Delzenne, N.M. GPR43/FFA2: Physiopathological relevance and therapeutic prospects. *Trends Pharmacol. Sci.* 2013, 34, 226–232. [CrossRef] [PubMed]
- 91. Macia, L.; Tan, J.; Vieira, A.T.; Leach, K.; Stanley, D.; Luong, S.; Maruya, M.; Ian McKenzie, C.; Hijikata, A.; Wong, C.; et al. Metabolite-sensing receptors GPR43 and GPR109A facilitate dietary fibre-induced gut homeostasis through regulation of the inflammasome. *Nat. Commun.* **2015**, *6*, 6734. [CrossRef] [PubMed]
- 92. Shan, M.; Gentile, M.; Yeiser, J.R.; Walland, A.C.; Bornstein, V.U.; Chen, K.; He, B.; Cassis, L.; Bigas, A.; Cols, M.; et al. Mucus enhances gut homeostasis and oral tolerance by delivering immunoregulatory signals. *Science* 2013, 342, 447–453. [CrossRef] [PubMed]
- 93. Thorburn, A.N.; Macia, L.; Mackay, C.R. Diet, metabolites, and "western-lifestyle" inflammatory diseases. *Immunity* **2014**, 40, 833–842. [CrossRef]
- Alex, S.; Lange, K.; Amolo, T.; Grinstead, J.S.; Haakonsson, A.K.; Szalowska, E.; Koppen, A.; Mudde, K.; Haenen, D.; Al-Lahham, S.; et al. Short-chain fatty acids stimulate angiopoietin-like 4 synthesis in human colon adenocarcinoma cells by activating peroxisome proliferator-activated receptor gamma. *Mol. Cell Biol.* 2013, 33, 1303–1316. [CrossRef] [PubMed]
- 95. Kinoshita, M.; Suzuki, Y.; Saito, Y. Butyrate reduces colonic paracellular permeability by enhancing PPARgamma activation. *Biochem. Biophys. Res. Commun.* **2002**, *293*, 827–831. [CrossRef]
- 96. Vinolo, M.A.; Rodrigues, H.G.; Nachbar, R.T.; Curi, R. Regulation of inflammation by short chain fatty acids. *Nutrients* **2011**, *3*, 858–876. [CrossRef] [PubMed]
- 97. Wang, C.C.; Goalstone, M.L.; Draznin, B. Molecular mechanisms of insulin resistance that impact cardiovascular biology. *Diabetes* **2004**, *53*, 2735–2740. [CrossRef] [PubMed]

- Wang, L.; de Zoeten, E.F.; Greene, M.I.; Hancock, W.W. Immunomodulatory effects of deacetylase inhibitors: Therapeutic targeting of FOXP3+ regulatory T cells. *Nat. Rev. Drug Discov.* 2009, *8*, 969–981. [CrossRef] [PubMed]
- Chang, P.V.; Hao, L.; Offermanns, S.; Medzhitov, R. The microbial metabolite butyrate regulates intestinal macrophage function via histone deacetylase inhibition. *Proc. Natl. Acad. Sci. USA* 2014, 111, 2247–2252. [CrossRef] [PubMed]
- 100. Arpaia, N.; Campbell, C.; Fan, X.; Dikiy, S.; van der Veeken, J.; deRoos, P.; Liu, H.; Cross, J.R.; Pfeffer, K.; Coffer, P.J.; et al. Metabolites produced by commensal bacteria promote peripheral regulatory T-cell generation. *Nature* 2013, 504, 451–455. [CrossRef]
- 101. Roager, H.M.; Vogt, J.K.; Kristensen, M.; Hansen, L.B.S.; Ibrugger, S.; Maerkedahl, R.B.; Bahl, M.I.; Lind, M.V.; Nielsen, R.L.; Frokiaer, H.; et al. Whole grain-rich diet reduces body weight and systemic low-grade inflammation without inducing major changes of the gut microbiome: A randomised cross-over trial. *Gut* 2019, *68*, 83–93. [CrossRef] [PubMed]
- Vulevic, J.; Juric, A.; Tzortzis, G.; Gibson, G.R. A mixture of trans-galactooligosaccharides reduces markers of metabolic syndrome and modulates the fecal microbiota and immune function of overweight adults. *J. Nutr.* 2013, 143, 324–331. [CrossRef]
- 103. Giacco, R.; Lappi, J.; Costabile, G.; Kolehmainen, M.; Schwab, U.; Landberg, R.; Uusitupa, M.; Poutanen, K.; Pacini, G.; Rivellese, A.A.; et al. Effects of rye and whole wheat versus refined cereal foods on metabolic risk factors: A randomised controlled two-centre intervention study. *Clin. Nutr.* 2013, *32*, 941–949. [CrossRef] [PubMed]
- 104. Hald, S.; Schioldan, A.G.; Moore, M.E.; Dige, A.; Laerke, H.N.; Agnholt, J.; Bach Knudsen, K.E.; Hermansen, K.; Marco, M.L.; Gregersen, S.; et al. Effects of Arabinoxylan and Resistant Starch on Intestinal Microbiota and Short-Chain Fatty Acids in Subjects with Metabolic Syndrome: A Randomised Crossover Study. *PLoS ONE* 2016, 11, e0159223. [CrossRef] [PubMed]
- 105. Johansson-Persson, A.; Ulmius, M.; Cloetens, L.; Karhu, T.; Herzig, K.H.; Onning, G. A high intake of dietary fiber influences C-reactive protein and fibrinogen, but not glucose and lipid metabolism, in mildly hypercholesterolemic subjects. *Eur. J. Nutr.* 2014, 53, 39–48. [CrossRef] [PubMed]
- 106. Peterson, C.M.; Beyl, R.A.; Marlatt, K.L.; Martin, C.K.; Aryana, K.J.; Marco, M.L.; Martin, R.J.; Keenan, M.J.; Ravussin, E. Effect of 12 wk of resistant starch supplementation on cardiometabolic risk factors in adults with prediabetes: A randomized controlled trial. *Am. J. Clin. Nutr.* 2018, 108, 492–501. [CrossRef] [PubMed]
- 107. Uusitupa, M.; Hermansen, K.; Savolainen, M.J.; Schwab, U.; Kolehmainen, M.; Brader, L.; Mortensen, L.S.; Cloetens, L.; Johansson-Persson, A.; Onning, G.; et al. Effects of an isocaloric healthy Nordic diet on insulin sensitivity, lipid profile and inflammation markers in metabolic syndrome—A randomized study (SYSDIET). J. Intern. Med. 2013, 274, 52–66. [CrossRef] [PubMed]
- 108. Dandona, P.; Ghanim, H.; Abuaysheh, S.; Green, K.; Batra, M.; Dhindsa, S.; Makdissi, A.; Patel, R.; Chaudhuri, A. Decreased insulin secretion and incretin concentrations and increased glucagon concentrations after a high-fat meal when compared with a high-fruit and -fiber meal. *Am. J. Physiol. Endocrinol. Metab.* 2015, 308, E185–E191. [CrossRef] [PubMed]
- 109. Kondo, K.; Morino, K.; Nishio, Y.; Ishikado, A.; Arima, H.; Nakao, K.; Nakagawa, F.; Nikami, F.; Sekine, O.; Nemoto, K.I.; et al. Fiber-rich diet with brown rice improves endothelial function in type 2 diabetes mellitus: A randomized controlled trial. *PLoS ONE* 2017, 12, e0179869. [CrossRef] [PubMed]
- 110. De Mello, V.D.; Schwab, U.; Kolehmainen, M.; Koenig, W.; Siloaho, M.; Poutanen, K.; Mykkanen, H.; Uusitupa, M. A diet high in fatty fish, bilberries and wholegrain products improves markers of endothelial function and inflammation in individuals with impaired glucose metabolism in a randomised controlled trial: The Sysdimet study. *Diabetologia* 2011, *54*, 2755–2767. [CrossRef] [PubMed]
- 111. Chambers, E.S.; Byrne, C.S.; Morrison, D.J.; Murphy, K.G.; Preston, T.; Tedford, C.; Garcia-Perez, I.; Fountana, S.; Serrano-Contreras, J.I.; Holmes, E.; et al. Dietary supplementation with inulin-propionate ester or inulin improves insulin sensitivity in adults with overweight and obesity with distinct effects on the gut microbiota, plasma metabolome and systemic inflammatory responses: A randomised cross-over trial. *Gut* **2019**, *68*, 1430–1438. [CrossRef] [PubMed]
- Monfort-Pires, M.; Crisma, A.R.; Bordin, S.; Ferreira, S.R.G. Greater expression of postprandial inflammatory genes in humans after intervention with saturated when compared to unsaturated fatty acids. *Eur. J. Nutr.* 2018, 57, 2887–2895. [CrossRef] [PubMed]

- 113. Izadpanah, A.; Barnard, R.J.; Almeda, A.J.; Baldwin, G.C.; Bridges, S.A.; Shellman, E.R.; Burant, C.F.; Roberts, C.K. A short-term diet and exercise intervention ameliorates inflammation and markers of metabolic health in overweight/obese children. *Am. J. Physiol. Endocrinol. Metab.* 2012, 303, E542–E550. [CrossRef] [PubMed]
- 114. Esgalhado, M.; Kemp, J.A.; Azevedo, R.; Paiva, B.R.; Stockler-Pinto, M.B.; Dolenga, C.J.; Borges, N.A.; Nakao, L.S.; Mafra, D. Could resistant starch supplementation improve inflammatory and oxidative stress biomarkers and uremic toxins levels in hemodialysis patients? A pilot randomized controlled trial. *Food Funct.* 2018, *9*, 6508–6516. [CrossRef] [PubMed]
- 115. Aliasgharzadeh, A.; Dehghan, P.; Gargari, B.P.; Asghari-Jafarabadi, M. Resistant dextrin, as a prebiotic, improves insulin resistance and inflammation in women with type 2 diabetes: A randomised controlled clinical trial. *Br. J. Nutr.* **2015**, *113*, 321–330. [CrossRef] [PubMed]
- 116. Basu, A.; Kurien, B.T.; Tran, H.; Byrd, B.; Maher, J.; Schell, J.; Masek, E.; Barrett, J.R.; Lyons, T.J.; Betts, N.M.; et al. Strawberries decrease circulating levels of tumor necrosis factor and lipid peroxides in obese adults with knee osteoarthritis. *Food Funct.* **2018**, *9*, 6218–6226. [CrossRef] [PubMed]
- 117. Nowotny, B.; Zahiragic, L.; Bierwagen, A.; Kabisch, S.; Groener, J.B.; Nowotny, P.J.; Fleitmann, A.K.; Herder, C.; Pacini, G.; Erlund, I.; et al. Low-energy diets differing in fibre, red meat and coffee intake equally improve insulin sensitivity in type 2 diabetes: A randomised feasibility trial. *Diabetologia* 2015, 58, 255–264. [CrossRef] [PubMed]
- 118. Ampatzoglou, A.; Williams, C.L.; Atwal, K.K.; Maidens, C.M.; Ross, A.B.; Thielecke, F.; Jonnalagadda, S.S.; Kennedy, O.B.; Yaqoob, P. Effects of increased wholegrain consumption on immune and inflammatory markers in healthy low habitual wholegrain consumers. *Eur. J. Nutr.* **2016**, *55*, 183–195. [CrossRef] [PubMed]
- Kim, J.Y.; Kwon, H.Y.; Kim, K.S.; Kim, M.K.; Kwon, O. Postprandial glucose and NF-kappaB responses are regulated differently by monounsaturated fatty acid and dietary fiber in impaired fasting glucose subjects. *J. Med. Food* 2013, *16*, 1168–1171. [CrossRef] [PubMed]
- Gargari, B.P.; Namazi, N.; Khalili, M.; Sarmadi, B.; Jafarabadi, M.A.; Dehghan, P. Is there any place for resistant starch, as alimentary prebiotic, for patients with type 2 diabetes? *Complement. Ther. Med.* 2015, 23, 810–815. [CrossRef] [PubMed]
- 121. Dehghan, P.; Gargari, B.P.; Jafar-Abadi, M.A.; Aliasgharzadeh, A. Inulin controls inflammation and metabolic endotoxemia in women with type 2 diabetes mellitus: A randomized-controlled clinical trial. *Int. J. Food Sci. Nutr.* 2014, 65, 117–123. [CrossRef] [PubMed]
- 122. Dehghan, P.; Pourghassem Gargari, B.; Asghari Jafar-abadi, M. Oligofructose-enriched inulin improves some inflammatory markers and metabolic endotoxemia in women with type 2 diabetes mellitus: A randomized controlled clinical trial. *Nutrition* **2014**, *30*, 418–423. [CrossRef] [PubMed]
- 123. Hajihashemi, P.; Azadbakht, L.; Hashemipor, M.; Kelishadi, R.; Esmaillzadeh, A. Whole-grain intake favorably affects markers of systemic inflammation in obese children: A randomized controlled crossover clinical trial. *Mol. Nutr. Food Res.* 2014, 58, 1301–1308. [CrossRef] [PubMed]
- 124. Di Francesco, A.; Di Germanio, C.; Bernier, M.; de Cabo, R. A time to fast. *Science* 2018, 362, 770–775. [CrossRef] [PubMed]
- 125. Bertile, F.; Fouillen, L.; Wasselin, T.; Maes, P.; Le Maho, Y.; Van Dorsselaer, A.; Raclot, T. The Safety Limits of an Extended Fast: Lessons from a Non-Model Organism. *Sci. Rep.* **2016**, *6*, 39008. [CrossRef] [PubMed]
- 126. McCue, M.D. Comparative Physiology of Fasting, Starvation, and Food Limitation; Springer: Berlin/Heidelberg, Germany, 2012.
- 127. Prentice, A.M.; Whitehead, R.G.; Roberts, S.B.; Paul, A.A. Long-term energy balance in child-bearing Gambian women. *Am. J. Clin. Nutr.* **1981**, *34*, 2790–2799. [CrossRef] [PubMed]
- Longo, V.D.; Mattson, M.P. Fasting: Molecular mechanisms and clinical applications. *Cell Metab.* 2014, 19, 181–192. [CrossRef] [PubMed]
- De Cabo, R.; Mattson, M.P. Effects of Intermittent Fasting on Health, Aging, and Disease. N. Engl. J. Med. 2019, 381, 2541–2551. [CrossRef] [PubMed]
- 130. Mattson, M.P.; Longo, V.D.; Harvie, M. Impact of intermittent fasting on health and disease processes. *Ageing Res. Rev.* **2017**, *39*, 46–58. [CrossRef] [PubMed]
- Longo, V.D.; Ellerby, L.M.; Bredesen, D.E.; Valentine, J.S.; Gralla, E.B. Human Bcl-2 reverses survival defects in yeast lacking superoxide dismutase and delays death of wild-type yeast. *J. Cell Biol.* 1997, 137, 1581–1588. [CrossRef] [PubMed]

- Kaeberlein, M.; McVey, M.; Guarente, L. The SIR2/3/4 complex and SIR2 alone promote longevity in Saccharomyces cerevisiae by two different mechanisms. *Genes Dev.* 1999, 13, 2570–2580. [CrossRef] [PubMed]
- Longo, V.D.; Shadel, G.S.; Kaeberlein, M.; Kennedy, B. Replicative and chronological aging in Saccharomyces cerevisiae. *Cell Metab.* 2012, *16*, 18–31. [CrossRef] [PubMed]
- 134. Lee, G.D.; Wilson, M.A.; Zhu, M.; Wolkow, C.A.; de Cabo, R.; Ingram, D.K.; Zou, S. Dietary deprivation extends lifespan in Caenorhabditis elegans. *Aging Cell* **2006**, *5*, 515–524. [CrossRef] [PubMed]
- 135. Eichhorn, G.; Groscolas, R.; Le Glaunec, G.; Parisel, C.; Arnold, L.; Medina, P.; Handrich, Y. Heterothermy in growing king penguins. *Nat. Commun.* **2011**, *2*, 435. [CrossRef] [PubMed]
- 136. Fontan-Lozano, A.; Saez-Cassanelli, J.L.; Inda, M.C.; de los Santos-Arteaga, M.; Sierra-Dominguez, S.A.; Lopez-Lluch, G.; Delgado-Garcia, J.M.; Carrion, A.M. Caloric restriction increases learning consolidation and facilitates synaptic plasticity through mechanisms dependent on NR2B subunits of the NMDA receptor. J. Neurosci. 2007, 27, 10185–10195. [CrossRef] [PubMed]
- 137. Singh, R.; Lakhanpal, D.; Kumar, S.; Sharma, S.; Kataria, H.; Kaur, M.; Kaur, G. Late-onset intermittent fasting dietary restriction as a potential intervention to retard age-associated brain function impairments in male rats. *Age* 2012, *34*, 917–933. [CrossRef] [PubMed]
- Goodrick, C.L.; Ingram, D.K.; Reynolds, M.A.; Freeman, J.R.; Cider, N. Effects of intermittent feeding upon body weight and lifespan in inbred mice: Interaction of genotype and age. *Mech. Ageing Dev.* 1990, 55, 69–87. [CrossRef]
- 139. Carlson, A.J.; Hoelzel, F. Apparent prolongation of the life span of rats by intermittent fasting. *J. Nutr.* **1946**, *31*, 363–375. [CrossRef] [PubMed]
- 140. Kendrick, D.C. The effects of infantile stimulation and intermittent fasting and feeding on life span in the black-hooded rat. *Dev. Psychobiol.* **1973**, *6*, 225–234. [CrossRef] [PubMed]
- Sakamoto, K.; Grunewald, K.K. Beneficial effects of exercise on growth of rats during intermittent fasting. J. Nutr. 1987, 117, 390–395. [CrossRef] [PubMed]
- 142. Lee, C.; Raffaghello, L.; Brandhorst, S.; Safdie, F.M.; Bianchi, G.; Martin-Montalvo, A.; Pistoia, V.; Wei, M.; Hwang, S.; Merlino, A.; et al. Fasting cycles retard growth of tumors and sensitize a range of cancer cell types to chemotherapy. *Sci. Transl. Med.* **2012**, *4*, 124ra127. [CrossRef] [PubMed]
- 143. Shi, Y.; Felley-Bosco, E.; Marti, T.M.; Orlowski, K.; Pruschy, M.; Stahel, R.A. Starvation-induced activation of ATM/Chk2/p53 signaling sensitizes cancer cells to cisplatin. *BMC Cancer* **2012**, *12*, 571. [CrossRef] [PubMed]
- Descamps, O.; Riondel, J.; Ducros, V.; Roussel, A.M. Mitochondrial production of reactive oxygen species and incidence of age-associated lymphoma in OF1 mice: Effect of alternate-day fasting. *Mech. Ageing Dev.* 2005, 126, 1185–1191. [CrossRef] [PubMed]
- 145. Raffaghello, L.; Lee, C.; Safdie, F.M.; Wei, M.; Madia, F.; Bianchi, G.; Longo, V.D. Starvation-dependent differential stress resistance protects normal but not cancer cells against high-dose chemotherapy. *Proc. Natl. Acad. Sci. USA* 2008, 105, 8215–8220. [CrossRef] [PubMed]
- 146. Halagappa, V.K.; Guo, Z.; Pearson, M.; Matsuoka, Y.; Cutler, R.G.; Laferla, F.M.; Mattson, M.P. Intermittent fasting and caloric restriction ameliorate age-related behavioral deficits in the triple-transgenic mouse model of Alzheimer's disease. *Neurobiol. Dis.* **2007**, *26*, 212–220. [CrossRef] [PubMed]
- 147. Griffioen, K.J.; Wan, R.; Brown, T.R.; Okun, E.; Camandola, S.; Mughal, M.R.; Phillips, T.M.; Mattson, M.P. Aberrant heart rate and brainstem brain-derived neurotrophic factor (BDNF) signaling in a mouse model of Huntington's disease. *Neurobiol. Aging* **2012**, *33*, e1481–e1485. [CrossRef] [PubMed]
- 148. Duan, W.; Guo, Z.; Jiang, H.; Ware, M.; Li, X.J.; Mattson, M.P. Dietary restriction normalizes glucose metabolism and BDNF levels, slows disease progression, and increases survival in huntingtin mutant mice. *Proc. Natl. Acad. Sci. USA* 2003, 100, 2911–2916. [CrossRef] [PubMed]
- 149. Arumugam, T.V.; Phillips, T.M.; Cheng, A.; Morrell, C.H.; Mattson, M.P.; Wan, R. Age and energy intake interact to modify cell stress pathways and stroke outcome. *Ann. Neurol.* 2010, 67, 41–52. [CrossRef] [PubMed]
- 150. Yamauchi, T.; Kamon, J.; Waki, H.; Terauchi, Y.; Kubota, N.; Hara, K.; Mori, Y.; Ide, T.; Murakami, K.; Tsuboyama-Kasaoka, N.; et al. The fat-derived hormone adiponectin reverses insulin resistance associated with both lipoatrophy and obesity. *Nat. Med.* **2001**, *7*, 941–946. [CrossRef] [PubMed]
- 151. Speakman, J.R.; Mitchell, S.E. Caloric restriction. Mol. Aspects Med. 2011, 32, 159–221. [CrossRef] [PubMed]

- 152. Castello, L.; Froio, T.; Maina, M.; Cavallini, G.; Biasi, F.; Leonarduzzi, G.; Donati, A.; Bergamini, E.; Poli, G.; Chiarpotto, E. Alternate-day fasting protects the rat heart against age-induced inflammation and fibrosis by inhibiting oxidative damage and NF-kB activation. *Free Radic. Biol. Med.* 2010, 48, 47–54. [CrossRef] [PubMed]
- 153. Wan, R.; Camandola, S.; Mattson, M.P. Intermittent fasting and dietary supplementation with 2-deoxy-D-glucose improve functional and metabolic cardiovascular risk factors in rats. *FASEB J.* **2003**, *17*, 1133–1134. [CrossRef] [PubMed]
- 154. Pedersen, C.R.; Hagemann, I.; Bock, T.; Buschard, K. Intermittent feeding and fasting reduces diabetes incidence in BB rats. *Autoimmunity* **1999**, *30*, 243–250. [CrossRef] [PubMed]
- 155. Ahmet, I.; Wan, R.; Mattson, M.P.; Lakatta, E.G.; Talan, M. Cardioprotection by intermittent fasting in rats. *Circulation* **2005**, *112*, 3115–3121. [CrossRef] [PubMed]
- 156. Baatar, D.; Patel, K.; Taub, D.D. The effects of ghrelin on inflammation and the immune system. *Mol. Cell Endocrinol.* **2011**, *340*, 44–58. [CrossRef] [PubMed]
- 157. Singh, R.; Kaushik, S.; Wang, Y.; Xiang, Y.; Novak, I.; Komatsu, M.; Tanaka, K.; Cuervo, A.M.; Czaja, M.J. Autophagy regulates lipid metabolism. *Nature* **2009**, *458*, 1131–1135. [CrossRef] [PubMed]
- 158. Wan, R.; Ahmet, I.; Brown, M.; Cheng, A.; Kamimura, N.; Talan, M.; Mattson, M.P. Cardioprotective effect of intermittent fasting is associated with an elevation of adiponectin levels in rats. *J. Nutr. Biochem.* 2010, 21, 413–417. [CrossRef] [PubMed]
- 159. Wilhelmi de Toledo, F.; Buchinger, A.; Burggrabe, H.; Holz, G.; Kuhn, C.; Lischka, E.; Lischka, N.; Lutzner, H.; May, W.; Ritzmann-Widderich, M.; et al. Fasting therapy—An expert panel update of the 2002 consensus guidelines. *Forsch. Komplementmed.* 2013, 20, 434–443. [CrossRef] [PubMed]
- Antoni, R.; Johnston, K.L.; Collins, A.L.; Robertson, M.D. Effects of intermittent fasting on glucose and lipid metabolism. *Proc. Nutr. Soc.* 2017, *76*, 361–368. [CrossRef] [PubMed]
- Mattson, M.P.; Moehl, K.; Ghena, N.; Schmaedick, M.; Cheng, A. Intermittent metabolic switching, neuroplasticity and brain health. *Nat. Rev. Neurosci.* 2018, 19, 63–80. [CrossRef] [PubMed]
- 162. Anton, S.D.; Moehl, K.; Donahoo, W.T.; Marosi, K.; Lee, S.A.; Mainous, A.G., 3rd; Leeuwenburgh, C.; Mattson, M.P. Flipping the Metabolic Switch: Understanding and Applying the Health Benefits of Fasting. *Obes. Silver Spring* 2018, 26, 254–268. [CrossRef] [PubMed]
- 163. Kinouchi, K.; Magnan, C.; Ceglia, N.; Liu, Y.; Cervantes, M.; Pastore, N.; Huynh, T.; Ballabio, A.; Baldi, P.; Masri, S.; et al. Fasting Imparts a Switch to Alternative Daily Pathways in Liver and Muscle. *Cell Rep.* 2018, 25, 3299–3314. [CrossRef] [PubMed]
- 164. Klempel, M.C.; Kroeger, C.M.; Varady, K.A. Alternate day fasting (ADF) with a high-fat diet produces similar weight loss and cardio-protection as ADF with a low-fat diet. *Metabolism* 2013, 62, 137–143. [CrossRef] [PubMed]
- 165. Varady, K.A.; Bhutani, S.; Church, E.C.; Klempel, M.C. Short-term modified alternate-day fasting: A novel dietary strategy for weight loss and cardioprotection in obese adults. *Am. J. Clin. Nutr.* 2009, 90, 1138–1143. [CrossRef] [PubMed]
- 166. Harvie, M.N.; Pegington, M.; Mattson, M.P.; Frystyk, J.; Dillon, B.; Evans, G.; Cuzick, J.; Jebb, S.A.; Martin, B.; Cutler, R.G.; et al. The effects of intermittent or continuous energy restriction on weight loss and metabolic disease risk markers: A randomized trial in young overweight women. *Int. J. Obes. Lond.* 2011, 35, 714–727. [CrossRef] [PubMed]
- Heilbronn, L.K.; Smith, S.R.; Martin, C.K.; Anton, S.D.; Ravussin, E. Alternate-day fasting in nonobese subjects: Effects on body weight, body composition, and energy metabolism. *Am. J. Clin. Nutr.* 2005, *81*, 69–73. [CrossRef] [PubMed]
- 168. Safdie, F.M.; Dorff, T.; Quinn, D.; Fontana, L.; Wei, M.; Lee, C.; Cohen, P.; Longo, V.D. Fasting and cancer treatment in humans: A case series report. *Aging Albany NY* **2009**, *1*, 988–1007. [CrossRef] [PubMed]
- Nencioni, A.; Caffa, I.; Cortellino, S.; Longo, V.D. Fasting and cancer: Molecular mechanisms and clinical application. *Nat. Rev. Cancer* 2018, 18, 707–719. [CrossRef] [PubMed]
- 170. Wilhelmi de Toledo, F.; Grundler, F.; Bergouignan, A.; Drinda, S.; Michalsen, A. Safety, health improvement and well-being during a 4 to 21-day fasting period in an observational study including 1422 subjects. *PLoS ONE* 2019, 14, e0209353. [CrossRef] [PubMed]
- 171. Tatar, M.; Bartke, A.; Antebi, A. The endocrine regulation of aging by insulin-like signals. *Science* **2003**, 299, 1346–1351. [CrossRef] [PubMed]

- 172. Selman, C.; Lingard, S.; Choudhury, A.I.; Batterham, R.L.; Claret, M.; Clements, M.; Ramadani, F.; Okkenhaug, K.; Schuster, E.; Blanc, E.; et al. Evidence for lifespan extension and delayed age-related biomarkers in insulin receptor substrate 1 null mice. *FASEB J.* **2008**, *22*, 807–818. [CrossRef] [PubMed]
- 173. Kenyon, C. The first long-lived mutants: Discovery of the insulin/IGF-1 pathway for ageing. *Philos. Trans. R. Soc. Lond. B Biol. Sci.* **2011**, *366*, 9–16. [CrossRef] [PubMed]
- 174. Johnson, S.C.; Rabinovitch, P.S.; Kaeberlein, M. mTOR is a key modulator of ageing and age-related disease. *Nature* **2013**, *493*, 338–345. [CrossRef] [PubMed]
- 175. Kenyon, C.J. The genetics of ageing. Nature 2010, 464, 504–512. [CrossRef] [PubMed]
- Hardie, D.G. Sensing of energy and nutrients by AMP-activated protein kinase. *Am. J. Clin. Nutr.* 2011, 93, 891S–896S. [CrossRef] [PubMed]
- 177. Apfeld, J.; O'Connor, G.; McDonagh, T.; DiStefano, P.S.; Curtis, R. The AMP-activated protein kinase AAK-2 links energy levels and insulin-like signals to lifespan in C. elegans. *Genes Dev.* 2004, 18, 3004–3009. [CrossRef] [PubMed]
- Tohyama, D.; Yamaguchi, A. A critical role of SNF1A/dAMPKalpha (Drosophila AMP-activated protein kinase alpha) in muscle on longevity and stress resistance in Drosophila melanogaster. *Biochem. Biophys. Res. Commun.* 2010, 394, 112–118. [CrossRef] [PubMed]
- 179. Gonzalez, A.A.; Kumar, R.; Mulligan, J.D.; Davis, A.J.; Weindruch, R.; Saupe, K.W. Metabolic adaptations to fasting and chronic caloric restriction in heart, muscle, and liver do not include changes in AMPK activity. *Am. J. Physiol. Endocrinol. Metab.* 2004, 287, E1032–E1037. [CrossRef] [PubMed]
- Longo, V.D. Linking sirtuins, IGF-I signaling, and starvation. *Exp. Gerontol.* 2009, 44, 70–74. [CrossRef]
 [PubMed]
- 181. Huang, R.; Xu, Y.; Wan, W.; Shou, X.; Qian, J.; You, Z.; Liu, B.; Chang, C.; Zhou, T.; Lippincott-Schwartz, J.; et al. Deacetylation of nuclear LC3 drives autophagy initiation under starvation. *Mol. Cell* 2015, 57, 456–466. [CrossRef] [PubMed]
- 182. Nakagawa, T.; Guarente, L. Sirtuins at a glance. J. Cell Sci. 2011, 124, 833–838. [CrossRef] [PubMed]
- 183. Fernandez-Marcos, P.J.; Auwerx, J. Regulation of PGC-1alpha, a nodal regulator of mitochondrial biogenesis. *Am. J. Clin. Nutr.* **2011**, *93*, 884S–890S. [CrossRef]
- 184. Horrillo, D.; Sierra, J.; Arribas, C.; Garcia-San Frutos, M.; Carrascosa, J.M.; Lauzurica, N.; Fernandez-Agullo, T.; Ros, M. Age-associated development of inflammation in Wistar rats: Effects of caloric restriction. *Arch. Physiol. Biochem.* 2011, 117, 140–150. [CrossRef] [PubMed]
- 185. Chung, H.Y.; Lee, E.K.; Choi, Y.J.; Kim, J.M.; Kim, D.H.; Zou, Y.; Kim, C.H.; Lee, J.; Kim, H.S.; Kim, N.D.; et al. Molecular inflammation as an underlying mechanism of the aging process and age-related diseases. *J. Dent. Res.* 2011, *90*, 830–840. [CrossRef] [PubMed]
- Fontana, L.; Partridge, L.; Longo, V.D. Extending healthy life span-from yeast to humans. *Science* 2010, 328, 321–326. [CrossRef] [PubMed]
- Lopez-Otin, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The hallmarks of aging. *Cell* 2013, 153, 1194–1217. [CrossRef] [PubMed]
- 188. Madkour, M.I.; El-Serafi, A.T.; Jahrami, H.A.; Sherif, N.M.; Hassan, R.E.; Awadallah, S.; Faris, M.A.E. Ramadan diurnal intermittent fasting modulates SOD2, TFAM, Nrf2, and sirtuins (SIRT1, SIRT3) gene expressions in subjects with overweight and obesity. *Diabetes. Res. Clin. Pract.* 2019, 155, 107801. [CrossRef] [PubMed]
- 189. Shojaie, M.; Ghanbari, F.; Shojaie, N. Intermittent fasting could ameliorate cognitive function against distress by regulation of inflammatory response pathway. *J. Adv. Res.* **2017**, *8*, 697–701. [CrossRef] [PubMed]
- 190. Ma, Q. Role of nrf2 in oxidative stress and toxicity. *Annu. Rev. Pharmacol. Toxicol.* **2013**, 53, 401–426. [CrossRef] [PubMed]
- Ristow, M.; Schmeisser, S. Extending life span by increasing oxidative stress. *Free Radic. Biol. Med.* 2011, 51, 327–336. [CrossRef] [PubMed]
- 192. Redman, L.M.; Smith, S.R.; Burton, J.H.; Martin, C.K.; Il'yasova, D.; Ravussin, E. Metabolic Slowing and Reduced Oxidative Damage with Sustained Caloric Restriction Support the Rate of Living and Oxidative Damage Theories of Aging. *Cell Metab.* 2018, 27, 805–815. [CrossRef] [PubMed]

- 193. Johnson, J.B.; Summer, W.; Cutler, R.G.; Martin, B.; Hyun, D.H.; Dixit, V.D.; Pearson, M.; Nassar, M.; Telljohann, R.; Maudsley, S.; et al. Alternate day calorie restriction improves clinical findings and reduces markers of oxidative stress and inflammation in overweight adults with moderate asthma. *Free Radic. Biol. Med.* 2007, 42, 665–674. [CrossRef] [PubMed]
- 194. Varady, K.A.; Bhutani, S.; Klempel, M.C.; Kroeger, C.M.; Trepanowski, J.F.; Haus, J.M.; Hoddy, K.K.; Calvo, Y. Alternate day fasting for weight loss in normal weight and overweight subjects: A randomized controlled trial. *Nutr. J.* **2013**, *12*, 146. [CrossRef] [PubMed]
- 195. Asemi, Z.; Samimi, M.; Taghizadeh, M.; Esmaillzadeh, A. Effects of Ramadan Fasting on Glucose Homeostasis, Lipid Profiles, Inflammation and Oxidative Stress in Women with Polycystic Ovary Syndrome in Kashan, Iran. Arch. Iran Med. 2015, 18, 806–810. [PubMed]
- 196. Moro, T.; Tinsley, G.; Bianco, A.; Marcolin, G.; Pacelli, Q.F.; Battaglia, G.; Palma, A.; Gentil, P.; Neri, M.; Paoli, A. Effects of eight weeks of time-restricted feeding (16/8) on basal metabolism, maximal strength, body composition, inflammation, and cardiovascular risk factors in resistance-trained males. *J. Transl. Med.* 2016, 14, 290. [CrossRef] [PubMed]
- 197. Faris, M.A.; Kacimi, S.; Al-Kurd, R.A.; Fararjeh, M.A.; Bustanji, Y.K.; Mohammad, M.K.; Salem, M.L. Intermittent fasting during Ramadan attenuates proinflammatory cytokines and immune cells in healthy subjects. *Nutr. Res.* 2012, *32*, 947–955. [CrossRef] [PubMed]
- 198. Aliasghari, F.; Izadi, A.; Gargari, B.P.; Ebrahimi, S. The Effects of Ramadan Fasting on Body Composition, Blood Pressure, Glucose Metabolism, and Markers of Inflammation in NAFLD Patients: An Observational Trial. J. Am. Coll. Nutr. 2017, 36, 640–645. [CrossRef] [PubMed]
- 199. Wilkinson, M.J.; Manoogian, E.N.C.; Zadourian, A.; Lo, H.; Fakhouri, S.; Shoghi, A.; Wang, X.; Fleischer, J.G.; Navlakha, S.; Panda, S.; et al. Ten-Hour Time-Restricted Eating Reduces Weight, Blood Pressure, and Atherogenic Lipids in Patients with Metabolic Syndrome. *Cell Metab.* 2020, *31*, 92–104. [CrossRef] [PubMed]
- 200. Horne, B.D.; Muhlestein, J.B.; Lappe, D.L.; May, H.T.; Carlquist, J.F.; Galenko, O.; Brunisholz, K.D.; Anderson, J.L. Randomized cross-over trial of short-term water-only fasting: Metabolic and cardiovascular consequences. *Nutr. Metab. Cardiovasc. Dis.* 2013, 23, 1050–1057. [CrossRef] [PubMed]
- 201. Williams, K.V.; Mullen, M.L.; Kelley, D.E.; Wing, R.R. The effect of short periods of caloric restriction on weight loss and glycemic control in type 2 diabetes. *Diabetes Care* **1998**, *21*, 2–8. [CrossRef] [PubMed]
- 202. Bhutani, S.; Klempel, M.C.; Kroeger, C.M.; Trepanowski, J.F.; Varady, K.A. Alternate day fasting and endurance exercise combine to reduce body weight and favorably alter plasma lipids in obese humans. *Obes. Silver Spring* **2013**, *21*, 1370–1379. [CrossRef] [PubMed]
- 203. Fontana, L.; Meyer, T.E.; Klein, S.; Holloszy, J.O. Long-term calorie restriction is highly effective in reducing the risk for atherosclerosis in humans. *Proc. Natl. Acad. Sci. USA* 2004, 101, 6659–6663. [CrossRef] [PubMed]
- 204. Eshghinia, S.; Mohammadzadeh, F. The effects of modified alternate-day fasting diet on weight loss and CAD risk factors in overweight and obese women. *J. Diabetes Metab. Disord.* **2013**, *12*, 4. [CrossRef] [PubMed]
- 205. Harvie, M.; Wright, C.; Pegington, M.; McMullan, D.; Mitchell, E.; Martin, B.; Cutler, R.G.; Evans, G.; Whiteside, S.; Maudsley, S.; et al. The effect of intermittent energy and carbohydrate restriction v. daily energy restriction on weight loss and metabolic disease risk markers in overweight women. *Br. J. Nutr.* 2013, 110, 1534–1547. [CrossRef] [PubMed]
- 206. Kroeger, C.M.; Klempel, M.C.; Bhutani, S.; Trepanowski, J.F.; Tangney, C.C.; Varady, K.A. Improvement in coronary heart disease risk factors during an intermittent fasting/calorie restriction regimen: Relationship to adipokine modulations. *Nutr. Metab. Lond.* 2012, *9*, 98. [CrossRef] [PubMed]



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