

Supplementary Information

Broadly neutralizing antibodies isolated from HEV convalescents confer protective effects in human liver-chimeric mice

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Table of Contents:

Figure S1: Characterization of recombinant HEV P domains used in this study

Figure S2: FACS plots showing the gating strategy for the sorting of single HEV-specific memory B cells

Figure S3: Neutralization of HEV by single chain variable fragments (scFvs)

Figure S4: Kinetic binding analysis of bnAbs with pORF2 P-domain

Figure S5: Broad nAb reactivity with GT 3 strain 83-2

Figure S6: Immunofluorescence analysis using human α -pORF2 IgGs

Figure S7: Epitopes targeted by glycan-sensitive human bnAbs

Figure S8: Electron density maps for HEV bnAbs in complex with P domain

Figure S9: Semiquantitative analysis of glycan-sensitive Abs in patient sera

Figure S10: nAb p60.1 prevents fecal-oral HEV infection in human liver-chimeric mice

Figure S11: Amino acid alignment of P domain sequences showing conservation and residues involved in interaction with bnAbs p60.1 and p60.12

Table S1: Sequence information of HEV pORF2 P-domain-specific antibodies

Table S2: Kinetic interaction analysis between bnAbs and glycosylated and non-glycosylated GT 3 P domains

Table S3: Characteristics of patient samples used for ELISA and isolate neutralization in HLCs

Table S4: Kinetic interaction analysis between bnAbs and non-glycosylated GT 3 P domains of different strains

Table S5: Diffraction data collection and refinement statistics

Table S6: Accession numbers of virus strains and isolates used in this study

Table S7: HEV antigen detection by different ELISAs

Table S8: List of oligonucleotides used in this study

Source Data for Figure S1

Figure S1: Recombinant HEV P domains used in this study

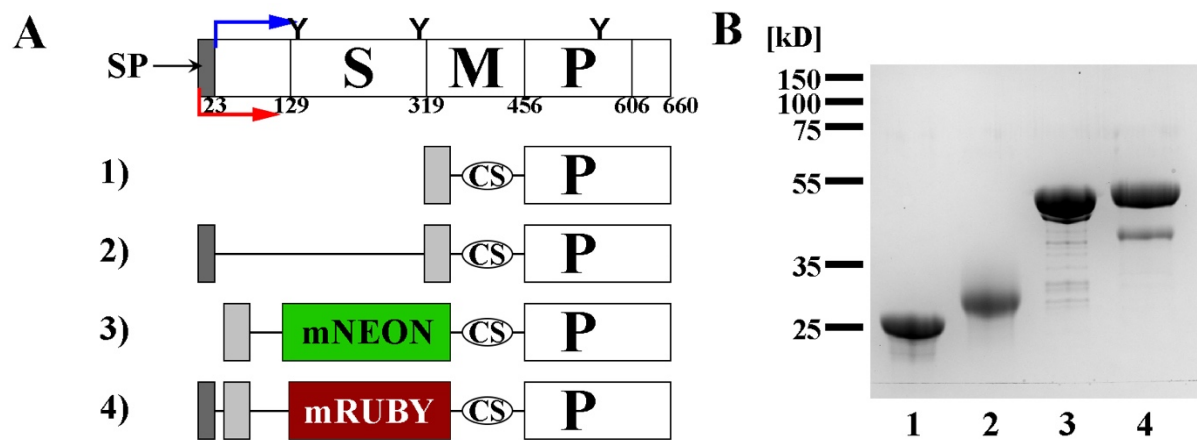
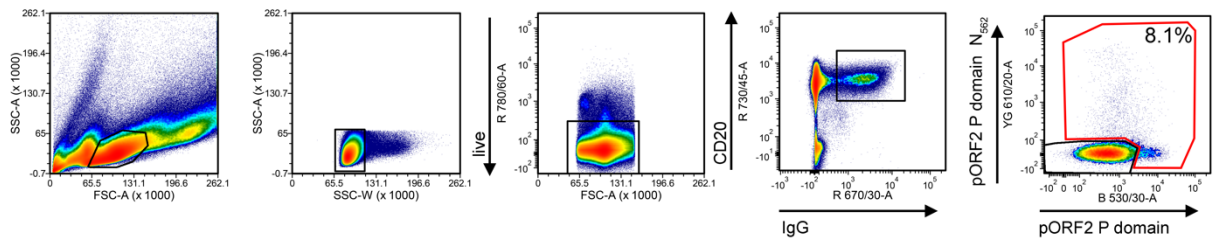


Figure S1: A) Schematic primary structure of ORF2, its individual domains (S/M/P), and the different expression constructs for the recombinant GT 3 P domains used in this study. Red and blue arrows indicate the described start codons for the glycosylated and non-glycosylated ORF2 in the presence and absence of a signal peptide, respectively. Numbers indicate amino acid positions within the GT 3 ORF2. SP, signal peptide (dark grey box); DST, double strep tag (light grey box); CS, proteolytic cleavage site for removal of affinity tag and fluorophore. Possible N-linked glycosylation sites are indicated. B) SDS-PAGE analysis followed by Coomassie staining reveals a slower migration of the glycosylated P domain (lanes 2 and 4) compared to the non-glycosylated P domain (lanes 1 and 3), likely caused by the attached glycan at position 562. Lanes 1-4 correspond to constructs 1) - 4) in panel A. Source data are provided within the Source Data file.

Figure S2: FACS plots showing the gating strategy for the sorting of single HEV-specific memory B cells

A



B

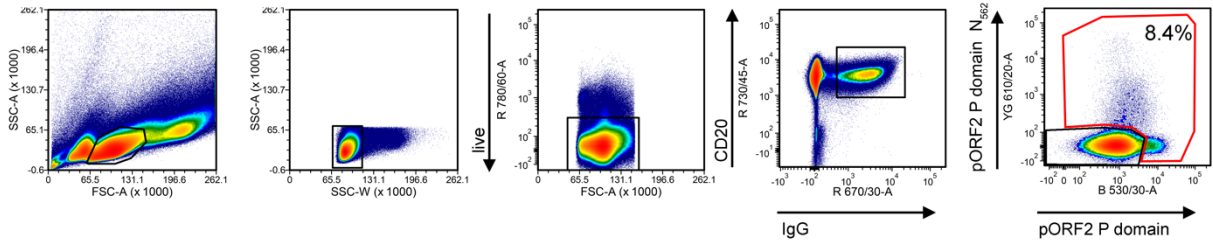


Figure S2: Representative FACS plots showing the gating strategy for sorting single pORF2 P-domain-specific memory B cells. (A) patient p60 and (B) patient p61. The red gate highlights the sorted cells.

Figure S3: Neutralization of HEV by single chain variable fragments (scFvs)

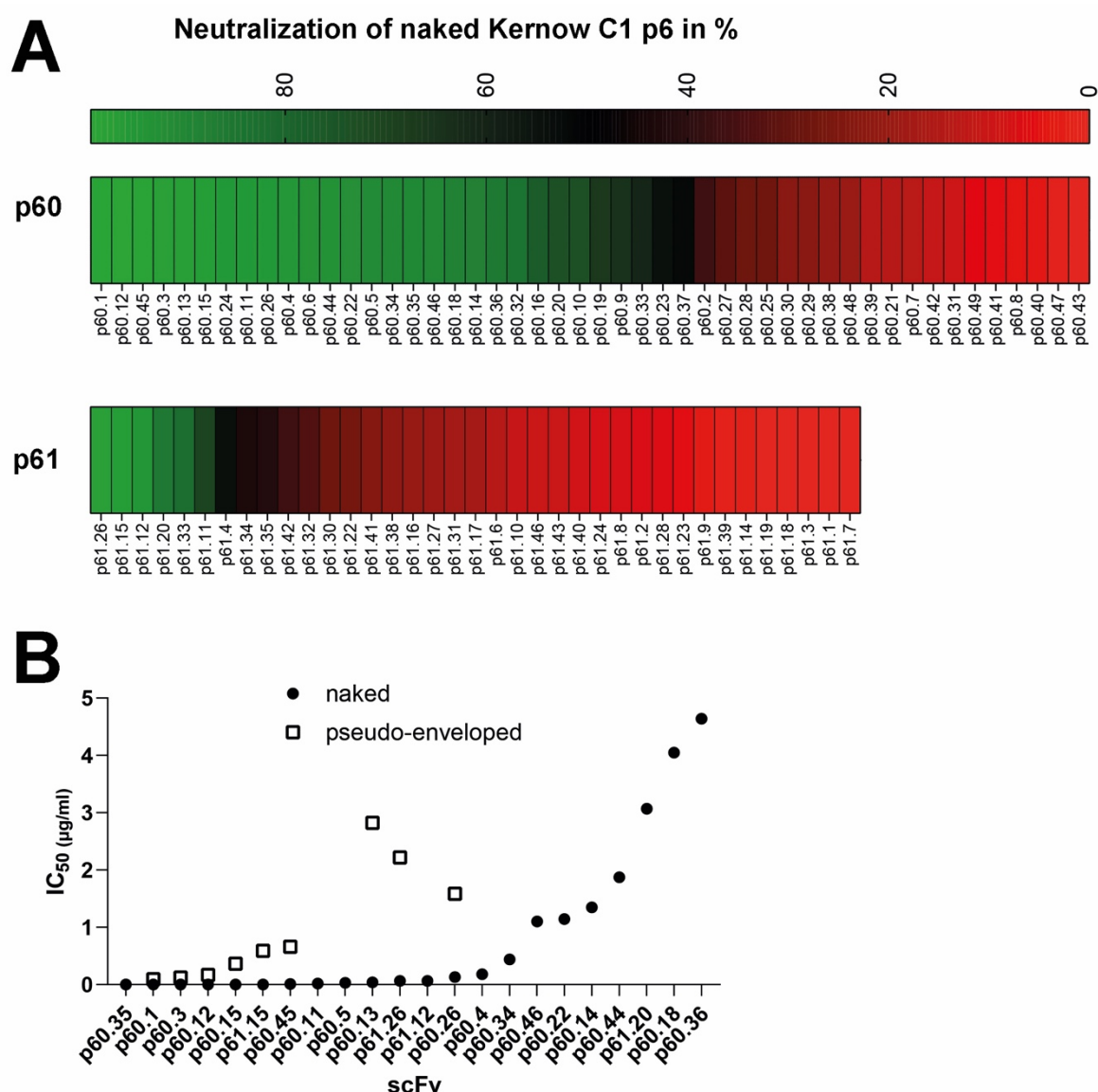


Figure S3: (A) 85 scFv expressed from patients 60 (p60) and 61 (p61) ranked according to their neutralizing capacity at a concentration of 10 µg/ml, with samples colored green showing at least 50% neutralization. Values are normalized to the control in the absence of scFv (n= 2 biological replicates) (B) The best neutralizers from (A) were further tested in serial dilution at concentrations of 5, 0.5, 0.05, and 0.005 µg/ml against naked HEV (dots) and at concentrations of 10, 5, 0.5, and 0.05 µg/ml against pseudo-enveloped HEV (boxes) to calculate the IC₅₀ using GraphPad Prism 9. Samples are ranked for the IC₅₀ against naked HEV. Neutralization of pseudo-enveloped HEV was not observed for all tested scFvs. Source data are provided within the Source Data file.

Figure S4: Kinetic binding analysis of bnAbs with pORF2 P-domain

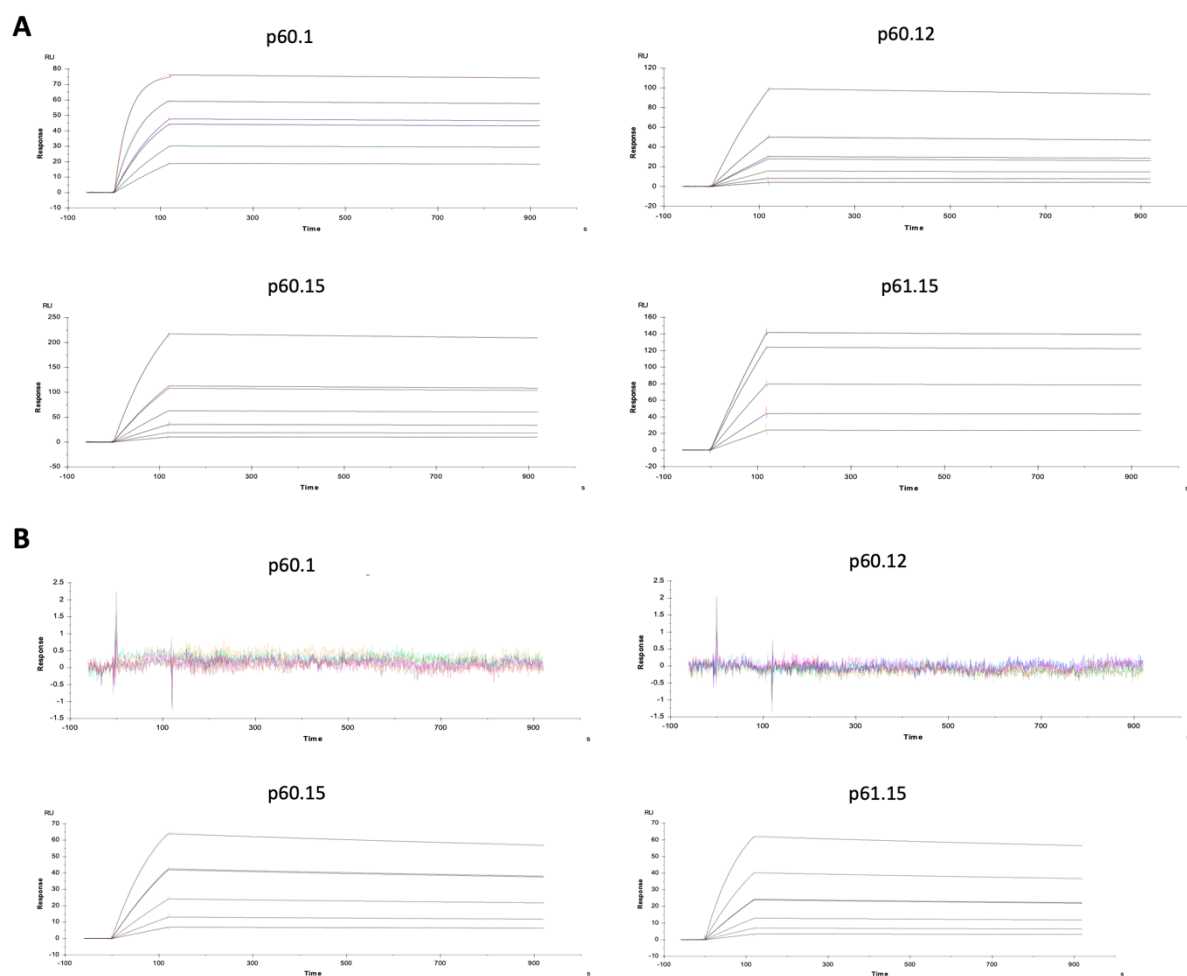


Figure S4: SPR analysis of nAbs p60.1, p60.12, p60.15, and p61.15 at 25, 12.5, 6.25, 3.125, and 1.5625 nM to the immobilized non-glycosylated (A) or glycosylated (B) HEV GT 3 P domain. The colored lines represent a 1:1 kinetic model fit. The kinetic binding parameters are shown in Table S2.

Figure S5: Broad nAb reactivity with GT 3 strain 83-2

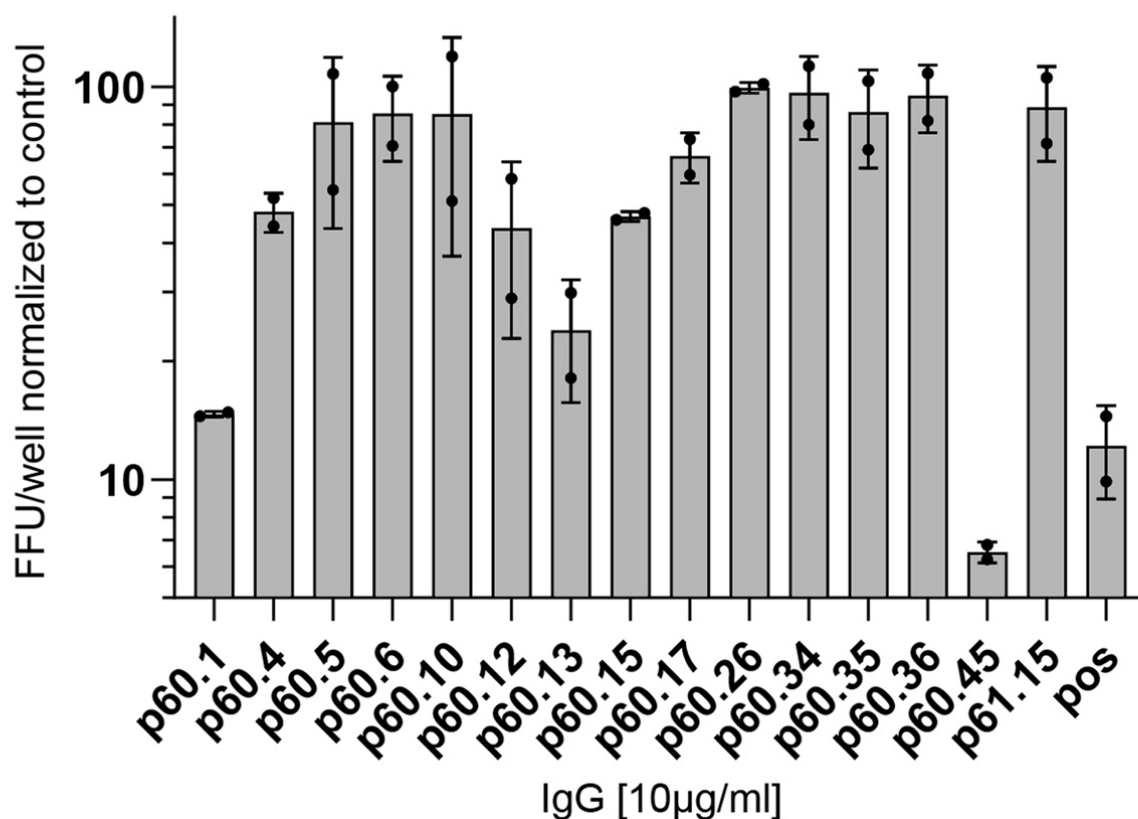


Figure S5: Percent of focus forming units per well of HepG2/C3A cells infected with HEV pUC83-2-27 neutralized with the indicated antibodies at a concentration of 10 µg/ml. The values are normalized to the control, to which no antibody was added. A human anti-HEV positive serum was used as positive control (pos) (n= 2 biological replicates). Source data are provided within the Source Data file.

Figure S6: Immunofluorescence analysis using human α -pORF2 IgGs

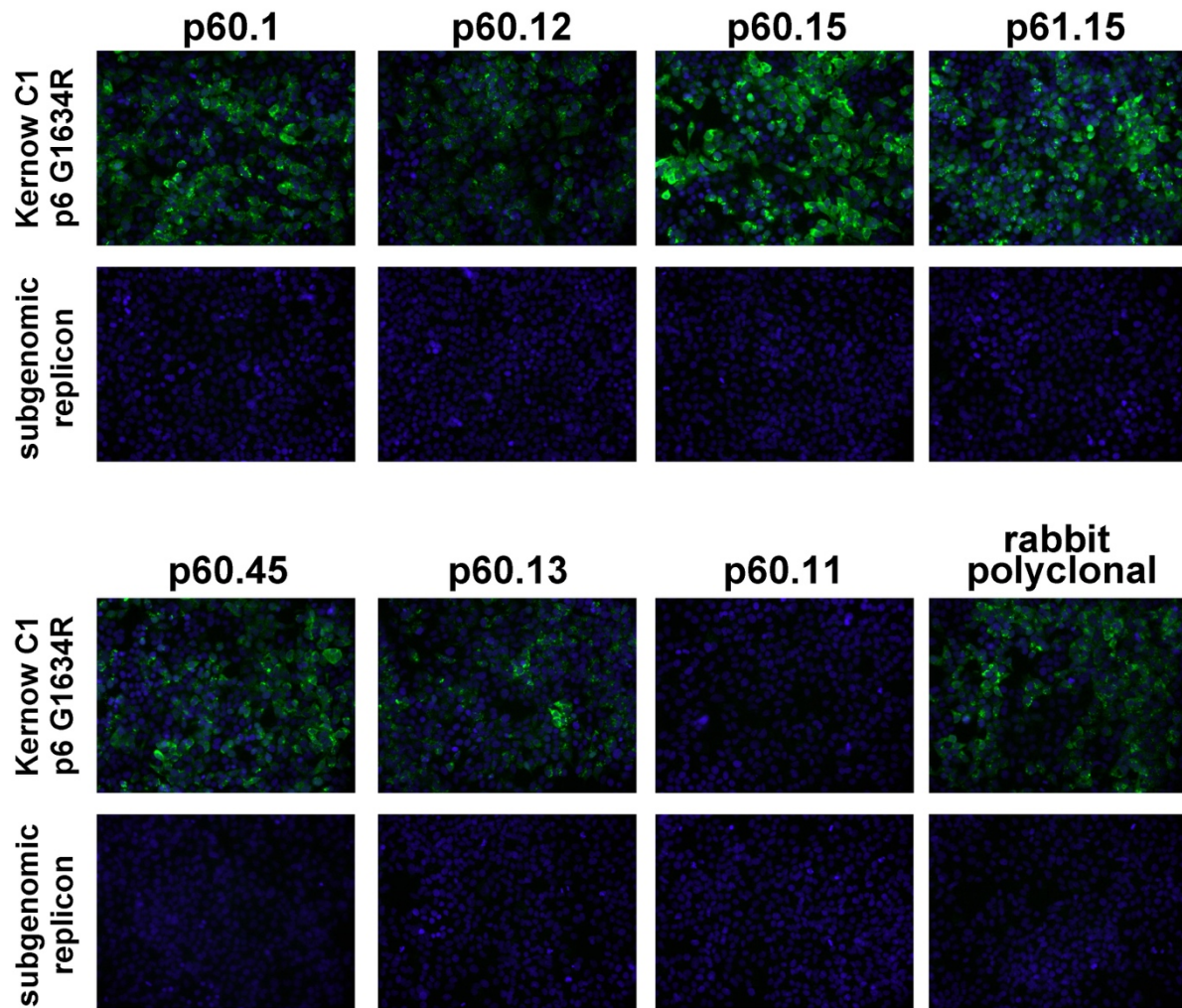


Figure S6: Immunofluorescence images depict HepG2 cells transfected with the viral genome (Kernow C1 p6 G1634R) or a subgenomic replicon lacking ORF-2 as control (subgenomic replicon). Four days after transfection, cells were washed and fixed using 3% PFA and permeabilized with Triton X-100 prior to staining with 0.25 μ g/ml of the indicated primary antibodies. As control, a rabbit polyclonal anti-P domain antibody was used. Bound nAbs were detected using fluorescently labelled anti-human or anti-rabbit secondary antibodies. Images were taken at 20x magnification, and nuclei stained with DAPI. The experiments were repeated three times with similar results.

Figure S7: Glycan-sensitive antibodies bind to a conserved epitope at the tip of the P domain dimer

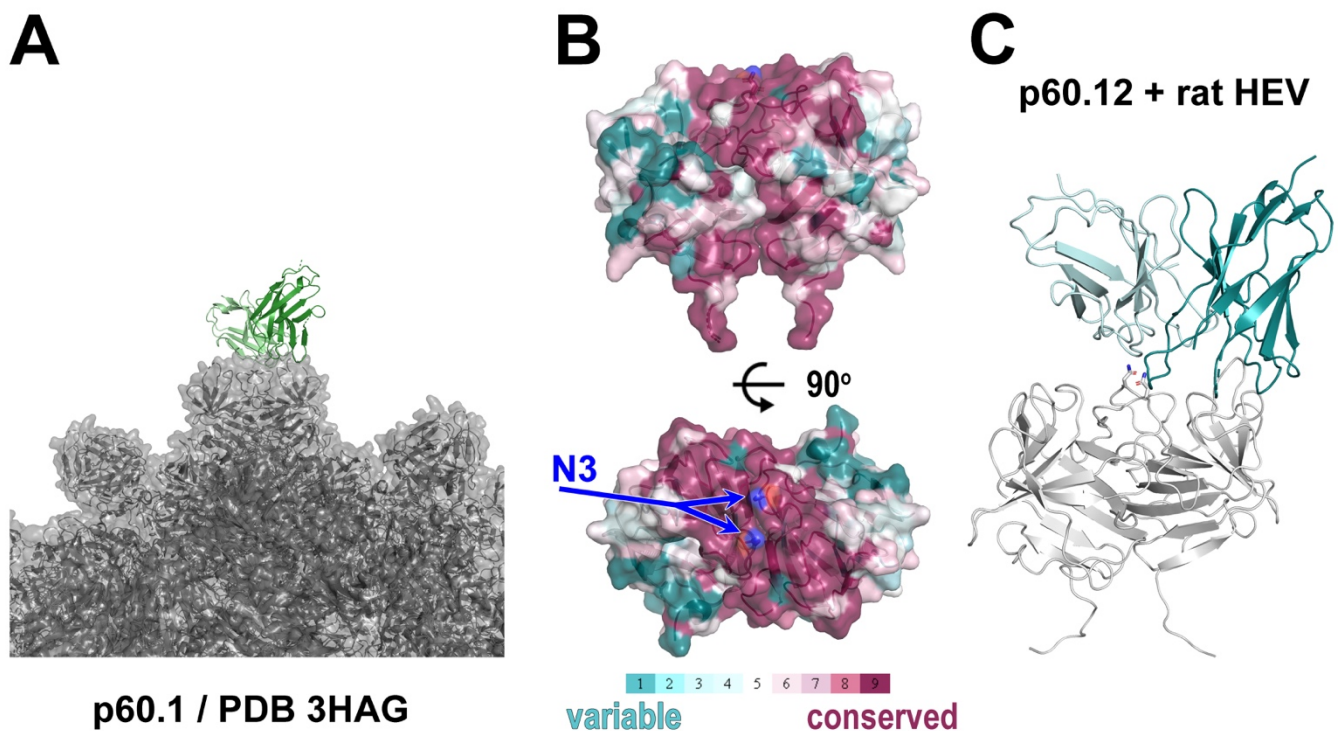


Figure S7: A) Superposition of the crystal structure of the P domain in complex with bnAb p60.1 to the crystal structure of HEV virus-like particles (PDB 3HAG). The VLP is shown as surface representation in grey, while p60.1 is shown as a green-coloured cartoon. B) Amino acid conservation of different HEV genotypes used in this study (Table S4) mapped onto the P domain crystal structure. Sidechain oxygen and nitrogen atoms of N562 are colored in red and blue, respectively, and a blue arrow indicates the position of N3. Amino acid sequence alignments were performed using Clustal Omega (EBI), and the conservation was mapped using the ConSurfDB server (<https://consurfdb.tau.ac.il/>). C) Cartoon view of the complex crystal structure of the rat HEV P domain (grey) and bnAb p60.12 (cyan). Sidechain oxygen and nitrogen atoms of N₅₆₂ are colored in red and blue, respectively, to indicate the position of N3.

Figure S8: Electron density maps for HEV bnAbs in complex with P domain

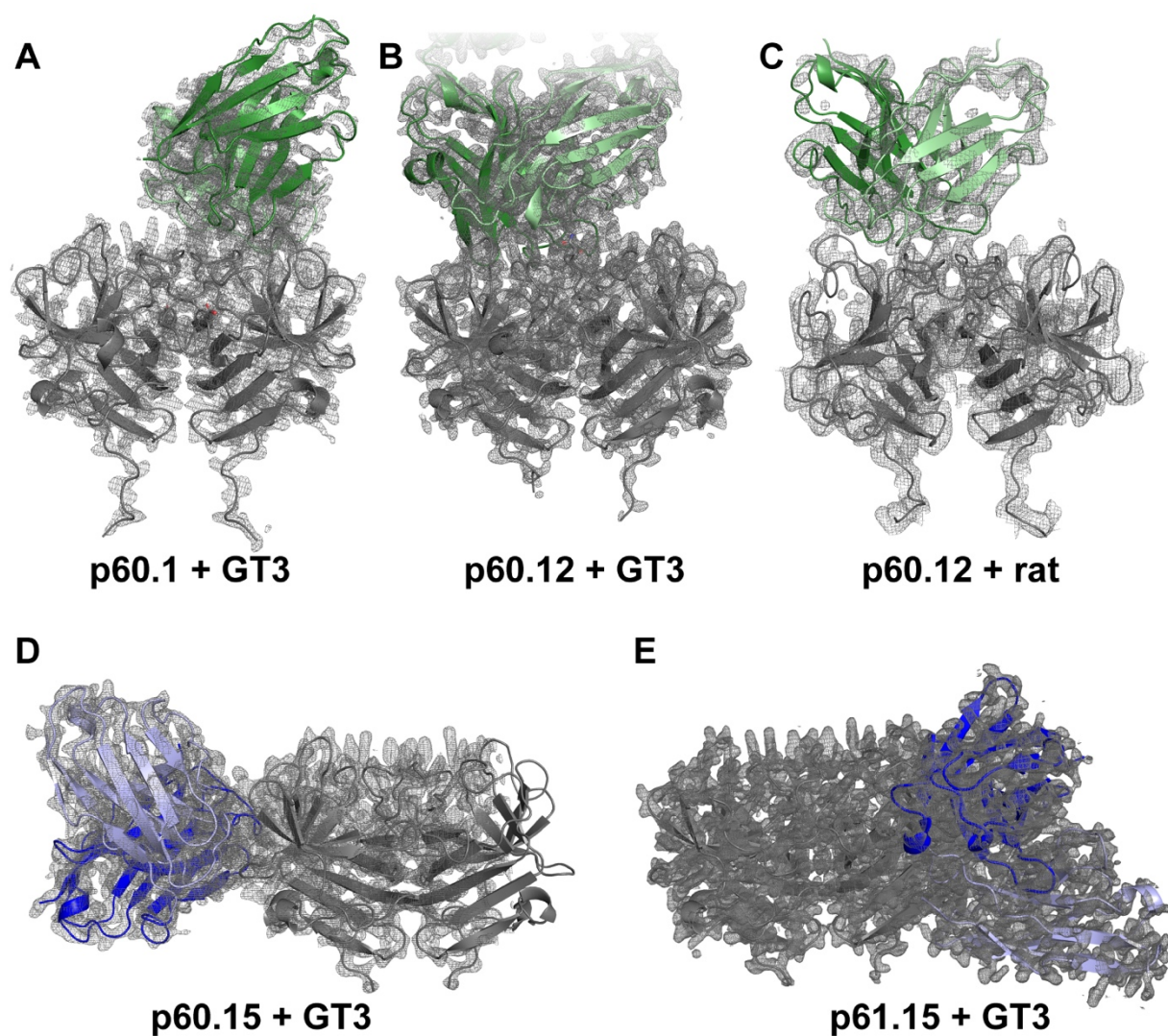


Figure S8: 2Fo-Fc electron density maps contoured at 1.5σ are superimposed on the corresponding refined atomic models shown as cartoon. For clarity, one P domain dimer is shown in each case. A GT 3 strain P domain was crystallized in complex with (A) p60.1, (B) p60.12, (D) p60.15, and (E) p61.15. (C) A rat HEV P domain was crystallized in complex with p60.12.

Figure S9: Glycan-sensitive Abs are induced during acute HEV infection

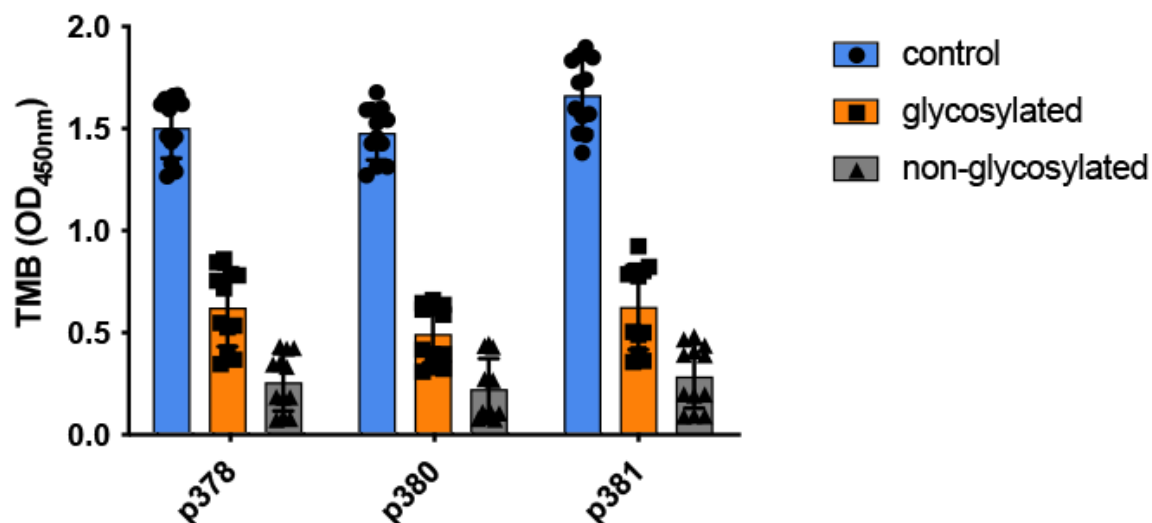


Figure S9: Sera from three patients, who underwent an acute HEV infection (p378, p380, and p381) were depleted in parallel with either glycosylated (orange), non-glycosylated (grey) or no (control, blue) GT3 P domain and subsequently subjected to a direct ELISA against the non-glycosylated P domain. The signal difference between the two differentially depleted serum samples is likely caused by steric interference of the glycan chain attached to N₅₆₂ with antibodies recognizing a juxtaposed epitope. The chart shows the mean of four independent biological replicates, error bars indicate the standard deviation. Source data are provided within the Source Data file.

Figure S10: nAb p60.1 prevents fecal-oral HEV infection in human liver-chimeric mice

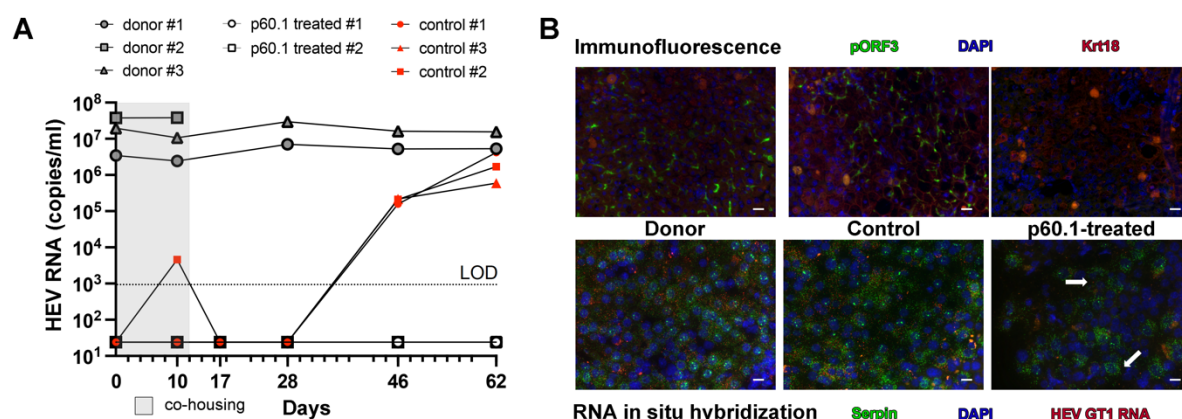


Figure S10: (A) Viral titers in the serum (copies/ml) of three donor mice (grey symbols), two antibody-treated mice (red symbols), and three control antibody-treated mice (empty symbols) were monitored over 62 days. At specific time points, blood was taken retrobulbar, viral RNA was isolated and measured by quantitative RT-PCR. LOD = 1000 copies/ml, light grey box = co-housing phase. Source data are provided within the Source Data file. (B) Visualization of HEV GT1 ORF3 protein (pORF3) in the liver of infected mice. Representative immunofluorescent staining of pORF3 (green) in liver sections from donor mice, control antibody-treated mice, and p60.1-treated mice. Human hepatocytes are stained with anti-Krt18 antibodies (red), and nuclei are visualized with DAPI (blue). Scale bars represent 20 μ m.

[illegible]

Figure S11: Amino acid alignment of the P domain sequences used in this study and the Hecolin® sequence. The secondary structure elements of the P domain are shown below the alignment. Residues involved in interaction with bnAb p60.1 (A) are colored in green, residues interacting with p60.12 (B) in cyan. Dots represent conserved amino acid residues. Amino acids numbered according to HEV genotype 3 (UniProt accession number C4B4T9)

Table S1: Sequence information of HEV pORF2 P-domain-specific antibodies

Ab code	Donor	VH germline gene	CDRH1	CDRH2	CDRH3	VL germline gene	CDRL1	CDRL2	CDRL3
p60.1	p60	IGHV 6-1	GDSVSTINAA	TYRSKWYH	CARDRGPTPFDFW	IGKV 1-39	QSIIRS	AAS	CQQSFVTPFTF
p60.2	p60	IGHV 6-1	GDSVSTINAA	TYRSKWYH	CARDRGPTPFDFW	IGLV3-21	NIGSKS	DDS	CQVWSSSDQGVF
p60.3	p60	IGHV 6-1	DVSNINVA	TYRDKWYT	CASHGSDPYAFDIW	IGLV 1-40	RSNIGTRY	GNI	CQSYDLSLGVVF
p60.4	p60	IGHV 1-2	GYTFTGIS	INPKTGGT	CARLDLLWSGELTGPKIYYYYGMDVW	IGKV 1D-39	QISINY	AAS	CQQSDSTPFTF
p60.5	p60	IGHV 3-21	GFTFSYSY	ISSSSSYI	CARGGGLMITFGVGMGMDVW	IGKV 2-28	QSLHNSNGYNY	LGS	CMQALQTPYTF
p60.6	p60	IGHV 3-23	GFTFSYIT	MSGSGDNT	CATRRLYCTTSCVMYFDDW	IGLV 2-8	STDVGGYNY	DVN	CSSYAASDTLLF
p60.7	p60	IGHV3-30	GFPFSTYG	ISYDGNK	CAKDDYASGPFDFYW	IGKV1-9	QGISY	AAS	CQQLNTYPTF
p60.8	p60	IGHV4-30-2	GSSIRSGDYI	IYSGTT	CARAGRKTIVTPDLDYW	IGLV1-44	WTNIGDNY	KHS	CASWDDSLRGWVF
p60.9	p60	IGHV5-51	GYSTYSYW	IYPGDSIT	CARSRDRGVGFDPW	IGKV3-20	QSVSSTY	GAS	CQHYGSSPHTF
p60.10	p60	IGHV 6-1	GDSVSSNSAA	TYRSKWYN	CASGSYQHFDFW	IGLV 2-23	SSDVGSYNL	EVS	CCSYAGSSYYVF
p60.11	p60	IGHV 1-46	GYTFTSYI	INPSAGHT	CARAAARRREILDYW	IGLV 3-21	NIGSR5	DGS	CQMWDTSSDRGVF
p60.12	p60	IGHV 1-69	GDTFTSYV	IPIIGTA	CASNVLQRRGNWFDPW	IGLV 2-14	SSDIGDYNF	DVT	CCSYTNTNTPVVF
p60.13	p60	IGHV 3-48	GF5FRSYE	ITSGSGGT	CATVIRRDGYN5DW	IGLV 1-40	TSNIGAPYD	GNT	CQSYDNL5G5VF
p60.14	p60	IGHV 4-4	GG5ISSNNW	IHH5GSI	CAREAIYNGMDVW	IGKV 3-20	QRISSY	GAS	CQQYASSPKTF
p60.15	p60	IGHV 5-51	GDTFANYW	IYPRDSOI	CARHLRLRLGLFPFDYW	IGKV 1-9	QDIFSY	AAS	CQQLNRYPPAF
p60.16	p60	IGHV1-18	GYSTYSYG	ISGNGNPN	CARHLDYDSSGYPWPYDFW	IGLV3-21	SIGGKN	DDS	CQVWDTDTDHVIF
p60.18	p60	IGHV 1-18	GFPTNYG	ITPYNRNT	CASLKRHGYNLPYFDYW	IGKV 1-12	QGVNSW	AAS	CQQANSPYTF
p60.19	p60	IGHV 1-18	GYTFTSYG	ISVHNGNI	CARDGGWNSRLFDYW	IGLV1-44	NSNVGTIA	SNN	CATWNSDLNDWVF
p60.20	p60	IGHV1-18	GYTFTSYG	ISAYNGNT	CARDRGSFISGSVGADEFDNDW	IGKV2-30	QGLVYIDGNTY	KVS	CMQGTWHPYTF
p60.21	p60	IGHV 1-18	GYTFTSYG	ISGYNNGT	CAREGSSGWHRLIDYW	IGLV1-44	SSNIGTKT	TNN	CAAWDDSLNGWVF
p60.22	p60	IGHV 1-18	GYTFATYD	ISTYSGNT	CARDDKWRSITPSYSHGYAPLDYW	IGKV 1-5	QSISSW	KAS	CQSYSSPTVF
p60.23	p60	IGHV1-18	GYTFTAYG	INGYNGNT	CARVGTDTSGYVWLW	IGKV1-8	QDIGSF	AAS	CQQYD5VPRTF
p60.24	p60	IGHV 1-18	GYTFSSYA	ISAYNGNT	CARVYYYGWGDFDPW	IGKV 3-20	QSVSSSY	GAS	CQQYSSSPYTF
p60.25	p60	IGHV 1-18	GYTFSSYG	ISAYNGDS	CARDRHSYDRLGHYHYAMDVW	IGLV3-19	SLRSY	GEN	CNSRDS5GNGLLGIF
p60.26	p60	IGHV 1-18	GYNFTTYS	ISNYNGKA	CARDPIYDSGSYIAHFQYIGDYW	IGLV 3-25	ALPKQY	KDS	CQ5ADSSSGWVF
p60.27	p60	IGHV 1-18	GYFTFSYG	ISTYNGNT	CARDVKPRSGHRAHYHMEVW	IGLV2-8	SSDVGGYNY	EVS	CCSY5NNNLVF
p60.28	p60	IGHV 1-18	GYTFSSYG	ITPYNGNT	CAGSVEVERDYNGMDVW	IGKV1D-39	QISITY	AAS	CQQSYSTPPFTF
p60.29	p60	IGHV1-18	GYNFHNYG	INTFTGNR	CARDVRCSGDCSDSYHFYGMDDW	IGKV3D-15	QSISSN	GAS	CQQYNNWPPWTF
p60.30	p60	IGHV1-18	GYTFTSYG	IRAVTGENVY	CARDQKEIVVPAEYGMDDW	IGLV1-51	SSNIGNHF	DNN	CGTWDDSLRIVVF
p60.31	p60	IGHV1-2	GVRTGTGY	IDPDSGGT	CARGQDHVSVDVSLGDHW	IGLV1-51	STNIGDNY	ENN	CGTWDRSLSAWVF
p60.32	p60	IGHV1-2	GYSTFTGY	INPNSGGT	CAGGCVTTISDYW	IGLV2-11	SSDVGGYNY	DVT	CCSYAG5WVF
p60.33	p60	IGHV1-2	GFTFTGYI	ISPN5GGT	CARGRVVVPAILYW	IGLV6-57	SGDISSNY	EDN	CQSYD5TPWVF
p60.34	p60	IGHV 1-2	GYTFTGY	INPNSGGT	CLRGHYVDSGLW	IGLV 4-69	SGHSSYA	LNSDGS	CQTWGTGTGVF
p60.35	p60	IGHV 1-2	GYTFTGYI	INPNSGVT	CAKRAPIIPWVDPW	IGLV 6-57	SGSIASNY	EDD	CQSYDSDTVVF
p60.36	p60	IGHV 1-2	GYNFIGYI	INPNRGST	CARDITG5NSNDWFDPW	IGLV 3-10	ALPKKF	EDT	CYSTDS5GNGGGVF
p60.37	p60	IGHV 1-2	GYTFTAYY	INPNSGGT	CARWVVPGLYSMDVW	IGLV3-21	NIGSKR	DES	CQVWDTDTDPVVF
p60.38	p60	IGHV1-2	GYPNVHYI	ISPNTGGT	CAREGRGLTMSRGTGYGMDVW	IGKV3-11	QSVSDY	DAS	CQQR5NWP1TF
p60.39	p60	IGHV1-24	GYTLTELS	FDPEDEGR	CSTASLAPIAWPFDYW	IGKV1D-39	QSI5RH	GAD	CQQR5NP1WTF
p60.40	p60	IGHV1-24	GHTLPDLF	FDHEDGER	CATGIVEDIPGGLDYGLD5W	IGLV1-44	SSNVEFNS	AND	CSVWDDNLH5VF
p60.41	p60	IGHV1-3	GYMFS5HS	INTDTGNT	CARDRYGSGHYNYFDYW	IGKV3-20	QADSKY	GIS	CQQYSS5PWT
p60.42	p60	IGHV1-3	GYTETTYS	INAGNGNT	CAREMPRIVGATGWFDPW	IGLV3-19	SLRSY	GKN	CCSRD55NHPVLF
p60.43	p60	IGHV1-3	GYNLTDYA	VNGDNGDT	CAREGGDCTGTCYSGYMDVW	IGLV3-25	ALPKQY	KDS	CQSPD5SATYVF
p60.44	p60	IGHV 1-3	GY5FTNYA	INAGNGDT	CARVYSTSKTPRRDYSYGMDDW	IGLV 1-40	SSNIGAGYD	GNN	CQSFDD5SL5G5VF
p60.45	p60	IGHV 1-46	GYTFTSYF	INPSAGST	CARDLKTSGWQRQFDSW	IGLV 3-21	NIGGER	DDS	CQVWSSSDQGVF
p60.46	p60	IGHV 1-46	GYTFTNYI	INPSGGST	CARDQAGSGYFDW	IGKV 1-5	QSISSW	KAS	CQQFNSYPPWTF
p60.47	p60	IGHV1-46	GYTFASYY	ISPGDNST	CAR5PPPSEISYHLHFDYW	IGLV1-40	SNIGAGYH	DND	CQSYD5SL5G5VF
p60.48	p60	IGHV1-46	GYTFTSYI	INPSGGST	CARGNIWVPAASNNWFDPW	IGLV2-14	SSDIGAYNY	DVI	CCSYTTRTLVF
p60.49	p60	IGHV1-58	GPLNLNSA	IIVGSGYT	CAADVPITIFGVFS5W	IGKV3-11	QNVHSF	DAS	CQQR5WP5RAF
p61.1	p61	IGHV1-2	GHTFTGYI	INPN5GGT	CARLYRDAFDIW	IGLV2-8	SSDVGGYNY	EVK	CASYAG55YVF
p61.2	p61	IGHV4-59	GG5FTDYI	INHKGNT	CARHFSYSALEFLTVISRMGAFD5W	IGLV3-19	SLRKYY	GKD	CN5GD5NTYVF
p61.3	p61	IGHV2-5	GFSLRNSGLG	IYWDDDK	CAHSRRTVAGAFAPW	IGKV2-28	QSLHSDGYNV	LGS	CMQJQLQPOTF
p61.4	p61	IGHV3-23	GF5FS5YA	ITGSGDIT	CAKVVDLMSVAIYYAMDVW	IGKV1D-39	QSISSY	AAS	CQQ5FRT5SYTF
p61.6	p61	IGHV3-74	GFTFRSDW	IRDDG5TS	CAREAPGRGNWYDLW	IGLV3-25	ALPNEY	KDT	CQ5AD55SATVVF
p61.7	p61	IGHV1-2	GYTFTAYF	INAH5GGT	CARGEFC5GGLCYFDSW	IGKV1-5	QRIRDY	KAS	CQQYDVP5ISF
p61.8	p61	IGHV2-5	GFSL5TYGEG	I5WDD5K	CAHRRVIGPYPDPFDIW	IGLV3-19	SLRHFS	DEN	CM5RDPTDHIH5F
p61.9	p61	IGHV3-23	GFTFS5YA	ISGSGDST	CAKAWTYEYFDYW	IGKV1-5	QSISSR	KVS	CQQYD5Y5SF
p61.10	p61	IGHV4-59	GG5ISSFY	VDYSGST	CARDASIAPIVQD5NWFDPW	IGKV1D-39	QTI5RT	AAS	CQQ555PITF
p61.11	p61	IGHV4-61	GASIGSGGYI	IFHSGST	CARRDCSSAC5FYDW	IGLV3-21	DIVRRS	DDS	CQVWDTSDHPVVF
p61.12	p61	IGHV 3-23	GFTFS5YA	IDASAFGT	CAK5AMLRGH5NDYW	IGKV 1-16	QGINNY	AAS	CQQYRSYPTVF
p61.14	p61	IGHV4-34	SGFFTGGF	IDRGGTS	CARGRRIRGFD5W	IGKV1-17	QGRIND	SAS	CLQYND5PYTF
p61.15	p61	IGHV 1-46	GYTFTNYF	FNPRGGST	CARMG5VTGPVGRWRLDPW	IGKV 3-11	Q5VDTY	DAS	CQQR5NWP5ITF
p61.16	p61	IGHV3-7	GFG5NF5W	IKDDG5EK	CMRGH55NDW	IGKV1-27	QDIRH	DAS	CQKYN5VPWTF
p61.17	p61	IGHV4-39	GS5IRTHFYI	IFYSGKT	CARHPAAEIVGW	IGKV3-11	Q5GVTY	DAS	CQQR5NWL5F5F
p61.18	p61	IGHV1-2	GY5FG5YI	INP5SGDT	CARLT5T5GNF5DRGWFDPW	IGLV1-47	TSNIAQYN	KTN	CASFD5SL5G5WVF
p61.19	p61	IGHV1-46	GDTFASYN	FNP5NPLIT	CAREETS5GWVQW	IGLV6-57	SGNID5NF	EDD	CQ5YD5GANQWVF
p61.20	p61	IGHV 1-8	GY5TFNFE	MNP5NTGNT	CAREVGS5YVWFDPW	IGLV 3-19	SLRSY	GKN	CCSRD5TSGHYWVF
p61.22	p61	IGHV3-11	GRF5SDYI	ISERGHTI	CARTEEYGTNSRRGAFD5W	IGLV3-1	YLGNY	QNY	CQAWDITQYVF
p61.23	p61	IGHV3-15	GFTFS5AW	IK5KT5DGGTT	CTTGAQWLTFDYW	IGKV1D-39	QITITY	AAS	CQQYSS5PPYTF
p61.24	p61	IGHV3-30	GFTFTSYG	T5PDGTYN	CVKEAFYDGYDIW	IGLV2-14	SSDVGGHNY	GVS	CCSYT5GLYVF
p61.26	p61	IGHV 3-33	GFNFTRYA	LSYDGS5K	CARDPRLLPMFYFDW	IGKV 3-20	Q5T5TY	DVS	CQHYG55FLTF
p61.27	p61	IGHV3-66	GFTV5TNY	NYSG5SI	CARAVVQLQSHYFDYW	IGKV3-20	Q5VSS5Y	GAS	CQQY555PATF
p61.28	p61	IGHV3-74	GFTFSGYW	INIDG5ST	CVRD5N5W5YW	IGKV2D-29	QSLH5NSGKTY	EVS	CMQ5IQLPLTF
p61.30	p61	IGHV4-34	GG5FSGDY	IDHTG5T	CARGGDFD5T5YHLFTW	IGLV3-21	DIGTKT	DDS	CQVWDT55AQVVF
p61.31	p61	IGHV4-59	SG5ISR5YH	IYD5D5GT	CARVTGQKFFRAGMDYW	IGKV1D-39	QYIMKY	GAS	CQQ5YSTPDTF
p61.32	p61	IGHV4-59	GG5ISNYI	IYTR5T	CATSATY5MEHW	IGKV1-9	HDISTY	LAS	CQQLR5YPLTF
p61.33	p61	IGHV5-51	GYTRD5YW	IFPGD5DT	CARRFY55GVKYFDLW	IGLV3-21	DVGSKA	DDN	CQVWDD5N5H5VF
p61.34	p61	IGHV1-18	GYFTSYG	ISEYNGNT	CARDWGVAPLVPDYW	IGKV1-17	QGRIND	AAS	CLQHTNYPLTF
p61.35	p61	IGHV1-2	GYTIGYI	INPN5SGAT	CARDPVDTAIYPHYHYGLD5W	IGLV2-14	SSDVGGYNY	DVS	CCSYG55TLVVF
p61.38	p61	IGHV2-5	GFSLAS5GVG	IYWDD5K	CAHRDYD5DALQYW	IGLV1-51	DSNIGNNF	DNN	CGTWDD5SLTAGVF
p61.39	p61	IGHV2-5	GF5FSL5GVG	IYWDDDK	CGHHD5GVVGH5W	IGKV3-20	Q5V5TY	GAT	CQQYD55RRLTF
p61.40	p61	IGHV3-30	GFTFRNYG	LISYD55NR	CAKDKAYYD55GWTFDYW	IGLV3-21	QGINNA	DAS	CQQ555PYTF
p61.41	p61	IGHV3-33	GFTFS5YA	ISYD5GNNQ	CAREMYD5FWSAYYAMDVW	IGKV1-27	HDISNY	AAS	CQKYN5APPLTF
p61.42	p61	IGHV3-49	GFTAD5YI	IR55KTYGAAT	CSRAGDFTNFFD5W	IGLV2-14	SGDIG5VDY	DVT	CT5Y5LDTAPYIF
p61.43	p61	IGHV3-66	G5SVTGYI	IY5SGTT	CARGTYTYGYD5IW	IGKV3-20	Q5V55YI	RAS	CQQY5IS5PFTF
p61.46	p61	IGHV4-34	GG5FSGYI	INQLGNT	CARTINRVGRYFDLW	IGKV1-12	QGLNSW	TTS	CQQAN55PRTF

Grey: Abs isolated from patient 60; Orange: Abs isolated from patient 61

Table S2: Kinetic interaction analysis between nAbs and glycosylated and non-glycosylated GT 3 P domains

bnAb	non-glycosylated GT 3 P domain		
	ka (M x s ⁻¹)	kd (s ⁻¹)	KD (M)
p60.1	6.87x10 ⁵	3.3x10 ⁻⁵	4.8x10 ⁻¹¹
p60.12	8.17x10 ⁴	7.43x10 ⁻⁵	9.09x10 ⁻¹⁰
p60.15	3.12x10 ⁵	5.18x10 ⁻⁵	1.66x10 ⁻¹⁰
p61.15	3.14x10 ⁵	2.56x10 ⁻⁵	8.16x10 ⁻¹¹
	glycosylated GT 3 P domain		
p60.1	n.d.	n.d.	n.d.
p60.12	n.d.	n.d.	n.d.
p60.15	5.13x10 ⁵	1.63x10 ⁻⁴	3.18x10 ⁻¹⁰
p61.15	2.41x10 ⁵	1.21x10 ⁻⁴	5.03x10 ⁻¹⁰

n.d.: not determined

Table S3: Characteristics of patient samples used for ELISA and isolate neutralization in HLCs

Patient	Sex	Age range	HEV RNA in plasma [10 ⁵ IU/ml]	WANTAI Ag ELISA serum [OD _{450-630nm}]	WANTAI IgG ELISA serum [OD _{450-630nm}]	Samples used in this study
1	f	60-69	6	3.61	3.36	serum
2	f	30-39	1	3.76	2.89	serum
3	f	40-49	9	4.03	2.62	serum and stool
4	f	30-39	8	3.74	1.72	serum
5	m	50-59	200	3.69	1.93	serum and stool
6	m	70-79	6	2.53	3.58	serum
7	m	50-59	10	3.73	3.67	serum
8	m	50-59	3	3.97	2.19	serum
9	f	30-39	0.9	3.64	0.43	serum and stool
10	f	80-89	8	3.72	n.t.	serum
11	m	60-69	2	>ULOQ	3.29	serum
12	f	40-49	20	3.80	3.04	serum
13	m	40-49	30	3.23	0.01 (negative)	serum and stool
14	f	20-29	70	3.83	0.01 (negative)	serum and stool
60	m	40-49	n.d.	n.d.	3.90	peripheral blood mononuclear cells
61	m	70-79	n.d.	n.d.	>ULOQ	peripheral blood mononuclear cells
378	f	40-49	n.d.	n.d.	3.67	serum
380	f	50-59	n.d.	n.d.	3.83	serum
381	m	60-69	n.d.	n.d.	3.90	serum

IU=international unit; OD= optical density; >ULOQ= above upper limit of quantification

Table S4: Kinetic interaction analysis between nAbs and non-glycosylated P domains of different strains

	HEV GT 1 P domain		
bnAb	ka (M x s⁻¹)	kd (s⁻¹)	KD (M)
p60.1	1.08x10 ⁶	2.56x10 ⁻⁵	2.38x10 ⁻¹¹
p60.12	9.87x10 ⁴	3.25x10 ⁻⁴	3.29x10 ⁻⁹
p60.15	1.78x10 ⁶	6.79x10 ⁻⁵	3.81x10 ⁻¹¹
p61.15	4.0x10 ⁵	6.04x10 ⁻⁴	1.51x10 ⁻⁹
	HEV GT 2 P domain		
p60.1	6.36x10 ⁵	8.16x10 ⁻⁵	1.28x10 ⁻¹⁰
p60.12	5.83x10 ⁴	2.96x10 ⁻⁵	5.09x10 ⁻¹⁰
p60.15	1.32x10 ⁶	2.3x10 ⁻⁴	1.74x10 ⁻¹⁰
p61.15	6.53x10 ⁵	5.85x10 ⁻⁴	8.96x10 ⁻¹⁰
	HEV GT 4 P domain		
p60.1	1.35x10 ⁶	5.14x10 ⁻⁵	3.82x10 ⁻¹¹
p60.12	6.96x10 ⁴	6.09x10 ⁻⁴	8.75x10 ⁻⁹
p60.15	8.83x10 ⁵	1.04x10 ⁻⁴	1.18x10 ⁻¹⁰
p61.15	9.25x10 ⁵	1.42x10 ⁻⁴	1.54x10 ⁻¹⁰
	rat HEV P domain		
p60.1	9.24x10 ⁴	4.56x10 ⁻¹	4.93x10 ⁻⁶
p60.12	4.4x10 ⁴	2.46x10 ⁻¹	5.59x10 ⁻⁶
p60.15	n.d.	n.d.	n.d.
p61.15	n.d.	n.d.	n.d.

n.d.: not determined

Table S5: Diffraction data collection and refinement statistics

	scFv p60.1 + HEV GT3 P domain	Fab p60.12 + HEV GT3 P domain	scFv p60.15 + HEV GT3 P domain	scFv p61.15 + HEV GT3 P domain	Fab p60.12 + rat- HEV P domain
Data collection					
Space group	C 2 2 2 ₁	P 2 ₁	C 2 2 2 ₁	P 2 ₁ 2 ₁ 2	P 4 ₃ 2 ₁ 2
Resolution (Å)	43.48 -1.98 (2.05-1.98) ^a	47.24 -2.07 (2.15 -2.07) ^a	48.64 - 2.41 (2.49 - 2.41) ^a	47.85 -1.91 (1.98 -1.91) ^a	48.7 -3.89 (4.031 -3.89) ^a
Cell dimensions					
<i>a</i> , <i>b</i> , <i>c</i> (Å)	75.81 159.26 205.67	88.43 94.48 91.25	56.19 122.95 158.67	108.59 178.77 49.66	112.86 112.86 184.44
α , β , γ (°)	90.0 90.0 90.0	90.0 90.21 90.0	90.0 90.0 90.0	90.0 90.0 90.0	90.0 90.0 90.0
PDB code	8PMW	8PNO	8PMY	8PMZ	8PMX
Complexes in AU	2	2	1	1	1
R _{meas}	0.13 (1.345)	0.129 (1.653)	0.293 (2.45)	0.176 (1.50)	0.322 (4.798)
< I/ σ (I) >	12.43 (1.98)	5.3 (0.56)	7.43 (1.01)	11.23 (1.73)	7.68 (0.68)
CC _{1/2}	0.998 (0.77)	0.991 (0.193)	0.993 (0.482)	0.997 (0.906)	0.998 (0.401)
Completeness (%)	99.47 (94.93)	84.7 (38.81)	99.61 (99.34)	97.1 (90.4)	99.18 (93.03)
Redundancy	13.23 (21.4)	3.27 (3.8)	13.12 (19.16)	13.52 (25.2)	25.91 (42.45)
Refinement					
No. reflections	86310 (8192)	77316 (3527)	21738 (2116)	72647 (5777)	11444
R _{work} / R _{free}	0.191 / 0.211	0.214 / 0.244	0.1993 / 0.238	0.2065 / 0.2395	0.2806 / 0.3537
No. atoms					
Protein	1069	1461	2888	5804	5572
ligands	26	0	3	38	0
solvent	308	184	107	321	0

B-factor					
Protein	45.92	49.31	58.05	36.31	188.35
ligands	60.38		87.37	51.79	
solvent	46.30	35.24	60.94	33.55	
R.m.s deviations					
Bond lengths (Å)	0.011	0.011	0.011	0.011	0.010
Bond angles (°)	1.57	1.61	1.63	1.60	1.45
Ramachandran plot [§]					
Favored (%)	96.58	96.04	96.48	97.18	84.09
Allowed (%)	3.23	3.75	3.52	2.82	15.08
Outliers (%)	0.19	0.21	0.00	0.00	0.83
Rotamer outliers (%)	0.67	1.72	1.28	0.00	10.02
Clashscore	0.93	2.63	2.1	1.47	10.74

A single crystal was used to collect each of the individual diffraction data sets

^a Values in parentheses are for highest-resolution shell.

[§] Ramachandran statistics were calculated with MolProbity

Table S6: Accession numbers of virus strains and isolates used in this study

Synthetic gene	Origin	UniprotKB/ Genbank
HEV ORF2_GT1	isolate/human/Pakistan/Sar55	P33426
HEV ORF2_GT2	isolate/human/Mexico	Q03500
HEV ORF2_GT3	isolate/human/Japan	C4B4T9
HEV ORF2_GT4	isolate/human/China	Q9IVZ8
HEV ORF2_rat	R68/DEU/2009	E0XL23
G3-HEV 83-2-27	Shiota et al., 2013, J Virol	AB740232
Kernow_C1_p6 wt	Isolate/human/United Kingdom	JQ679013

Table S7: HEV antigen detection by different ELISAs

Patient 1, female, aged 60-69:

timepoint	HEV RNA (IU/ml)	non-glycosylated pORF2 (p60.1, S/CO)	all pORF2 (p60.15, S/CO)	WANTAI HEV Ag (S/CO)
pre-treatment	600000	28,62	30,60	16,45
treatment (TW16)	n. d.	0,75	36,76	19,05
12 months FU	n. d.	0,34	7,69	19,97

Patient 2, male, aged 20-29:

timepoint	HEV RNA (IU/ml)	non-glycosylated pORF2 (p60.1, S/CO)	all pORF2 (p60.15, S/CO)	WANTAI HEV Ag (S/CO)
pre-treatment	3000000	34,77	29,06	13,60
treatment (TW4)	n. d.	0,88	15,26	13,93
12 months FU	n. d.	0,32	0,57	0,20

Red: HEV detection positive; Green: HEV detection negative; n.d.: not detected; Source data are provided within the Source Data file.

Table S8: List of oligonucleotides used for cloning

P domain construct	Forward primer	Reverse primer
GT 3 P domain mNeon	CGCTGAAAACCTGTATTTTCAGGGCCCCGCCCCAAGCCGGCCC	GGGTTTAAACTCAGGGCCCTCACGATTCTCGCGTTTACCC
GT 3 P domain mRuby	GAGCTGTACAAGGACGATGACGATAAGCCCGCCCCAAGCCGGCCC	GGGTTTAAACTCAGGGCCCTCACGATTCTCGCGTTTACCC
GT 3 P domain non-glycosylated	TTTTCAGGGCGACGATGACGATAAGCCCGCCCCAAGCCGGCCC	GGGTTTAAACTCAGGGCCCTCACGATTCTCGCGTTTACCC
GT 3 P domain glycosylated	TTTTCAGGGCGACGATGACGATAAGCCCGCCCCAAGCCGGCCC	GGGTTTAAACTCAGGGCCCTCACGATTCTCGCGTTTACCC
GT 1 P domain non-glycosylated	TTTTCAGGGCGACGATGACGATAAGCCAGCCCCATCGCGTCCT	GGGTTTAAACTCAGGGCCCTCATAACTCCCGAGTTTACCCAC
GT 2 P domain non-glycosylated	TTTTCAGGGCGACGATGACGATAAGCCAGCCCCCAGCAGGCCA	GGGTTTAAACTCAGGGCCCTCAGAGCTCGCGGGTTTACC
GT 4 P domain non-glycosylated	TTTTCAGGGCGACGATGACGATAAGCCCGCCCCCAGTCGCCCCG	GGGTTTAAACTCAGGGCCCTCAGTATTCACGTGTCTTGCCCA
rat HEV P domain non-glycosylated	TTTTCAGGGCGACGATGACGATAAGCCCGCACCTGCCCCACCT	GGGTTTAAACTCAGGGCCCTCATACGCTATCGGCTGCGGC
IgG construct	Forward primer	Reverse primer
p60.1-HC	GCTGGTCGCTTCCTGCCTGGGCGAAGTTCAGCTGCAGCAGA	CCGATGGGCCCTTCGTA CTGGCCGA ACTAACAGTGACCAACG
p60.1-KC	GCTGGTCGCTTCCTGCCTGGGCATT CAGATGACACAAAGCCCT	CGGATGGGGCGGCCACGGTGCGCTTCAAATCGACTTTTGTTC
p60.4-HC	GCTGGTCGCTTCCTGCCTGGGCCAGGTACAGCTGGTACAAAGC	CCGATGGGCCCTTCGTA CTGGCACTCGATAACCGTTACTGTAGTTC
p60.4-KC	GCTGGTCGCTTCCTGCCTGGGCGATATACAAATGACACAGAGCCCCG	CGGATGGGGCGGCCACGGTGCGTTTTATCTCCACCTTAGTACCTCCT

p60.5-HC	GCTGGTCGCTTCCTGCCTGGGCGAAGTTCAACTCGTGGAAGTG	CCGATGGGCCCTTCGTA CTGGCACTACTACCGTCACTGTAGT
p60.5-KC	GCTGGTCGCTTCCTGCCTGGGCGATATTGTGATGACTCAGAGCCC	CGGATGGGGCGGCCACGGTGCGTTTAATTTTCGAGCTTGGTGCC
p60.6-HC	GCTGGTCGCTTCCTGCCTGGGCGAGGTTCAACTGCTGGAAGT	CCGATGGGCCCTTCGTA CTGGCGCTGGATACCGTTACCAAAGT
p60.6-LC	GCTGGTCGCTTCCTGCCTGGGCCAGTCCGCATTGACTCAAC	ATGGGGCGGCCCTTCGGCTGGCCCAGGACTGTGAGTTTGGTG C
p60.10-HC	GCTGGTCGCTTCCTGCCTGGGCCAGGTGCAACTCCAGCAA	CCGATGGGCCCTTCGTA CTGGCCGACGATACAGTTACCAAAGTG
p60.10-LC	GCTGGTCGCTTCCTGCCTGGGCCAAAGTGCTTTGACGCAACC	ATGGGGCGGCCCTTCGGCTGGCCCAGGACTGTCACCTTCGT
p60.12-HC	GCTGGTCGCTTCCTGCCTGGGCCAGGTACAGTTGGTTCAAAGTG	CCGATGGGCCCTTCGTA CTGGCGCTGCTGACCGTCACCAA
p60.12-LC	GCTGGTCGCTTCCTGCCTGGGCCAGAGTGCTCTCACTCAACC	ATGGGGCGGCCCTTCGGCTGGCCGAGAACAGTGAGCTTGGTG C
p60.13-HC	GCTGGTCGCTTCCTGCCTGGGCGAAGTGACAGCTCGTTGAGT	CCGATGGGCCCTTCGTA CTGGCCGAGGAGACTGTTACGAGAG
p60.13-LC	GCTGGTCGCTTCCTGCCTGGGCCAATCCGTTTTGACTCAACCC	ATGGGGCGGCCCTTCGGCTGGCCGAGAACAGTGAGCTTAGTTCCA
p60.15-HC	GCTGGTCGCTTCCTGCCTGGGCGAAGTCCAACCTCGTGCAATC	CCGATGGGCCCTTCGTA CTGGCCGAGCTAACAGTAACCAATGTTC
p60.15-KC	GCTGGTCGCTTCCTGCCTGGGCGCGATACAATTGACACAGAGTC	CGGATGGGGCGGCCACGGTGCGTTTGATATCGACCTTTGTTCCAGG
p60.17-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTCCAACCTCGTTCAGTCC	CCGATGGGCCCTTCGTA CTGGCCGA ACTAACCGTTACGAGCG
p60.17-LC	GCTGGTCGCTTCCTGCCTGGGCCAGTCGGCACTCACTCAA	ATGGGGCGGCCCTTCGGCTGGCCCAAGACGGTCAATTTCGTTCC
p60.26-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTGCAATTGGTGCAATCG	CCGATGGGCCCTTCGTA CTGGCGCTCGACACGGTTACGGT
p60.26-LC	GCTGGTCGCTTCCTGCCTGGGCTCCTATGAACTGACTCAACCAC	ATGGGGCGGCCCTTCGGCTGGCCCAGAACTGTCAGCTTGGTG C
p60.34-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTGCAATTGGTGCAAGT	CCGATGGGCCCTTCGTA CTGGCGCTGCTTACCGTTACCAGC
p60.34-LC	GCTGGTCGCTTCCTGCCTGGGCCAGCTGGTTCTCACGCAA	ATGGGGCGGCCCTTCGGCTGGCCCAAGACGGTCAGTTTGTGC
p60.35-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTACATTGGTGCAAGTCG	CCGATGGGCCCTTCGTA CTGGCGGAGGATATGGTCACGAGGG
p60.35-LC	GCTGGTCGCTTCCTGCCTGGGCAATTTTATGTTGACACAGCCTCATA	ATGGGGCGGCCCTTCGGCTGGCCCAGTACCGTGAGTTTGTGC
p60.36-HC	GCTGGTCGCTTCCTGCCTGGGCCAGGTGCAGTTGGTTCAAAG	CCGATGGGCCCTTCGTA CTGGCCGAGCTCACGGTTACCAA
p60.36-LC	GCTGGTCGCTTCCTGCCTGGGCAGCTATGAATTGACTCAACCACC	ATGGGGCGGCCCTTCGGCTGGCCCAGGACCGTCAGCTTGGT
p60.45-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTACAGCTCGTGCAAGT	CCGATGGGCCCTTCGTA CTGGCGCTGCTTACCGTAACCAATG
p60.45-LC	GCTGGTCGCTTCCTGCCTGGGCAGCTACGTTCTGACACAACC	ATGGGGCGGCCCTTCGGCTGGCCCAGTGCTGTGAGTTTAGTGCC
p61.15-HC	GCTGGTCGCTTCCTGCCTGGGCCAAGTGCAACTCGTGCAAT	CCGATGGGCCCTTCGTA CTGGCACTACTCACGGTAACGAGAGT
p61.15-KC	GCTGGTCGCTTCCTGCCTGGGCGAAATAGTTCTCACACAGTCCCC	CGGATGGGGCGGCCACGGTGCGTTTTTATTTTCGAGCCTTGTTCTTG
8G12-HC	GCTGGTCGCTTCCTGCCTGGGCGGACAGCTGCAGCAGAGT	CCGATGGGCCCTTCGTA CTGGCACTCGATACTGTAACCAACGTG
8G12-KC	GCTGGTCGCTTCCTGCCTGGGCGACATACAAATGACTCAGAGTCCT	CGGATGGGGCGGCCACGGTGCGCTTGATTTC AATTTGTACCGCT

Source Data for Fig. S1: HEV antigen detection by different ELISAs

