PROKARYOTES



Genome Sequence of *"Candidatus* Carsonella ruddii" Strain BC, a Nutritional Endosymbiont of *Bactericera cockerelli*

gen@meAnnouncements™

Alex B. Riley, Dohyup Kim, Allison K. Hansen

AMERICAN SOCIETY FOR MICROBIOLOGY

Department of Entomology, University of Illinois at Urbana-Champaign, Urbana, Illinois, USA

ABSTRACT Here, we report the genome of *"Candidatus* Carsonella ruddii" strain BC, a nutritional endosymbiont of the tomato psyllid *Bactericera cockerelli*. The 173,802-bp genome contains 198 protein-coding genes, with a G+C content of 14.8%.

nsects in the family Psyllidae are vectors for a number of plant pathogens, making them agriculturally and economically important (1–3). Because they feed on nutrient poor phloem sap, psyllids depend on their obligate bacterial endosymbiont "*Candidatus* Carsonella ruddii" as a source of essential amino acids (4). Each species of psyllid harbors a specific strain of "*Ca.* Carsonella ruddii," which is maternally transmitted. The genomes of "*Ca.* Carsonella ruddii" strains are highly reduced (158 kb to 174 kb) (4–6), making them biologically interesting organisms. Here, we present the sequenced genome of "*Ca.* Carsonella ruddii" BC, which is associated with the tomato psyllid *Bactericera cockerelli.*

Bacteriocytes, specialized insect cells that harbor "*Ca.* Carsonella," from 10 tomato psyllids were dissected and pooled, and DNA was extracted. The extracted DNA was prepared using the Nextera DNA library preparation kit (Illumina, San Diego, CA) and sequenced within a 1/4 of a lane on an Illumina HiSeq 2500 (Illumina) using TruSeq SBS sequencing chemistry (Illumina). Fastq files were generated with the software Casava 1.8.2 (Illumina). The initial assembly and quality trimming were performed using a customized A5ud pipeline, which was modified from the A5 genome assembly pipeline (7). Further assembly was done using GapFiller (8) and SEQuel version 1.0.2 (9). Due to the high content of insect DNA in the sequencing reads, the contigs and scaffolds were filtered by G+C content and contig lengths. In order to complete the genome assembly, filtered reads and contigs were reassembled into gapped regions using Mimicking Intelligent Read Assembly (MIRA) version 4.0.2 (10). The remaining gaps were closed using Sanger sequencing of PCR products. The assembled genome was annotated using both the Rapid Annotation using Subsystem Technology (RAST) (11) and the Bacterial Annotation System (BASys) servers (12).

The complete circular genome of "*Ca*. Carsonella ruddii" BC is 173,802 bp long, with 198 protein-coding genes and a G+C content of 14.8%. Genes involved in essential amino acid biosynthesis that are present and absent in all other "*Ca*. Carsonella" strains sequenced to date have the same pattern of presence and absence in "*Ca*. Carsonella ruddii" BC (4, 5, 13). One exception, however, is that *pheA*, a gene conserved in all other sequenced "*Ca*. Carsonella" genomes to date and that is involved in phenylalanine biosynthesis, is missing from "*Ca*. Carsonella ruddii" BC's genome. Potentially, a homolog of *pheA* that is microbe in origin has been horizontally transferred to *Bactericera cockerelli*'s genome to complement "*Ca*. Carsonella ruddii" BC's essential amino acid pathways, analogous to mealybug, whitefly, and psyllid obligate symbiont symbioses (13–15). In sum, our genome sequence supports the role

Received 28 February 2017 Accepted 7 March 2017 Published 27 April 2017

Citation Riley AB, Kim D, Hansen AK. 2017. Genome sequence of "Candidatus Carsonella ruddii" strain BC, a nutritional endosymbiont of *Bactericera cockerelli*. Genome Announc 5: e00236-17. https://doi.org/10.1128/genomeA .00236-17.

Copyright © 2017 Riley et al. This is an openaccess article distributed under the terms of the Creative Commons Attribution 4.0 International license.

Address correspondence to Allison K. Hansen, akh@illinois.edu.

Riley et al.

of "Ca. Carsonella ruddii" BC for the biosynthesis of essential amino acids for its insect host, *Bactericera cockerelli*.

Accession number(s). The complete genome sequence of "*Ca.* Carsonella ruddii" BC and its annotation are deposited at GenBank, NCBI, under accession number CP019943.

ACKNOWLEDGMENTS

We thank David Crowder for kindly sending us psyllid samples. We also thank the undergraduates Yihui Zhu, Aejin Shon, and Joshua Newman for their help with this project.

This project was supported by University of Illinois, Urbana-Champaign (UIUC).

REFERENCES

- Halbert SE, Manjunath KL. 2004. Asian citrus psyllids (Sternorrhyncha: Psyllidae) and greening disease of citrus: a literature review and assessment of risk in Florida. Fla Entomol 87:330–353. https://doi.org/10.1653/ 0015-4040(2004)087[0330:ACPSPA]2.0.CO;2.
- Hansen AK, Trumble JT, Stouthamer R, Paine TD. 2008. A new huanglongbing species, "Candidatus Liberibacter psyllaurous," found to infect tomato and potato, is vectored by the psyllid Bactericera cockerelli (Sulc). Appl Environ Microbiol 74:5862–5865. https://doi.org/10.1128/AEM .01268-08.
- 3. Weintraub PG, Beanland L. 2006. Insect vectors of phytoplasmas. Annu Rev Entomol 51:91–111. https://doi.org/10.1146/annurev.ento.51 .110104.151039.
- Nakabachi A, Yamashita A, Toh H, Ishikawa H, Dunbar HE, Moran NA, Hattori M. 2006. The 160-kilobase genome of the bacterial endosymbiont *Carsonella*. Science 314:267. https://doi.org/10.1126/science .1134196.
- Nakabachi A, Ueoka R, Oshima K, Teta R, Mangoni A, Gurgui M, Oldham NJ, Van Echten-Deckert G, Okamura K, Yamamoto K, Inoue H, Ohkuma M, Hongoh Y, Miyagishima SY, Hattori M, Piel J, Fukatsu T. 2013. Defensive bacteriome symbiont with a drastically reduced genome. Curr Biol 23:1478–1484. https://doi.org/10.1016/j.cub.2013.06.027.
- Sloan DB, Moran NA. 2012. Genome reduction and co-evolution between the primary and secondary bacterial symbionts of psyllids. Mol Biol Evol 29:3781–3792. https://doi.org/10.1093/molbev/mss180.
- Tritt A, Eisen JA, Facciotti MT, Darling AE. 2012. An integrated pipeline for *de novo* assembly of microbial genomes. PLoS One 7:e42304 https:// doi.org/10.1371/journal.pone.0042304.
- Nadalin F, Vezzi F, Policriti A. 2012. GapFiller: a *de novo* assembly approach to fill the gap within paired reads. BMC Bioinformatics 13:S8. https://doi.org/10.1186/1471-2105-13-S14-S8.

- Ronen R, Boucher C, Chitsaz H, Pevzner P. 2012. SEQuel: improving the accuracy of genome assemblies. Bioinformatics 28:i188–i196. https://doi .org/10.1093/bioinformatics/bts219.
- Chevreux B, Pfisterer T, Drescher B, Driesel AJ, Müller WE, Wetter T, Suhai S. 2004. Using the miraEST assembler for reliable and automated mRNA transcript assembly and SNP detection in sequenced ESTs Genome Res 14:1147–1159. https://doi.org/10.1101/gr.1917404.
- Aziz RK, Bartels D, Best AA, DeJongh M, Disz T, Edwards RA, Formsma K, Gerdes S, Glass EM, Kubal M, Meyer F, Olsen GJ, Olson R, Osterman AL, Overbeek RA, McNeil LK, Paarmann D, Paczian T, Parrello B, Pusch GD, Reich C, Stevens R, Vassieva O, Vonstein V, Wilke A, Zagnitko O. 2008. The RAST server: rapid annotations using subsystems technology. BMC Genomics 9:75. https://doi.org/10.1186/1471-2164-9-75.
- Van Domselaar GH, Stothard P, Shrivastava S, Cruz JA, Guo AC, Dong X, Lu P, Szafron D, Greiner R, Wishart DS. 2005. BASys: a Web server for automated bacterial genome annotation. Nucleic Acids Res 33: W455–W459. https://doi.org/10.1093/nar/gki593.
- Sloan DB, Nakabachi A, Richards S, Qu J, Murali SC, Gibbs RA, Moran NA. 2014. Parallel histories of horizontal gene transfer facilitated extreme reduction of endosymbiont genomes in sap-feeding insects. Mol Biol Evol 31:857–871. https://doi.org/10.1093/molbev/msu004.
- Husnik F, Nikoh N, Koga R, Ross L, Duncan RP, Fujie M, Tanaka M, Satoh N, Bachtrog D, Wilson ACC, von Dohlen CD, Fukatsu T, McCutcheon JP. 2013. Horizontal gene transfer from diverse bacteria to an insect genome enables a tripartite nested mealybug symbiosis. Cell 153: 1567–1578. https://doi.org/10.1016/j.cell.2013.05.040.
- Luan JB, Chen W, Hasegawa DK, Simmons AM, Wintermantel WM, Ling KS, Fei Z, Liu SS, Douglas AE. 2015. Metabolic coevolution in the bacterial symbiosis of whiteflies and related plant sap-feeding insects. Genome Biol Evol 7:2635–2647. https://doi.org/10.1093/gbe/evv170.