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High phylogenetic diversity is preserved in species-poor high-elevation temperate moth assemblages

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Understanding the diversity and composition of species assemblages and identifying underlying biotic and abiotic determinants represent great ecological challenges. Addressing some of these issues, we investigated the α -diversity and phylogenetic composition of species-rich geometrid moth (Lepidoptera: Geometridae) assemblages in the mature temperate forest on Changbai Mountain. A total of 9285 geometrid moths representing 131 species were collected, with many species displaying wide elevational distribution ranges. Moth α -diversity decreased monotonously, while the standardized effect size of mean pairwise phylogenetic distances (MPD) and phylogenetic diversity (PD) increased significantly with increasing elevation. At high elevations, the insect assemblages consisted largely of habitat generalists that were individually more phylogenetically distinct from co-occurring species than species in assemblages at lower altitudes. This could hint at higher speciation rates in more favourable low-elevation environments generating a species-rich geometrid assemblage, while exclusion of phylogenetically closely related species becomes increasingly important in shaping moth assemblages at higher elevations. Overall, it appears likely that high-elevation temperate moth assemblages are strongly resilient to environmental change, and that they contain a much larger proportion of the genetic diversity encountered at low-elevation assemblages in comparison to tropical geometrid communities.

Identifying biotic and abiotic determinants that shape the diversity patterns and composition of species assemblages represents one of the most challenging ecological issues of our time^{1,2}. While the prevailing decrease in species richness with increasing latitude across most taxonomic groups is well-established^{3,4}, the underlying causes for this and other diversity response patterns to environmental changes are poorly understood. Sanders and Rahbek⁵ suggest that detailed investigations of altitudinal species distribution patterns are the best approach to establish the ecological basis for species responses to environmental gradients. A monotonic decrease in biodiversity similar to that recorded for increasing latitudes has regularly been observed with increasing altitude. Nonetheless, other patterns such as hump-shaped responses and diversity plateaus have also been reported^{6–9}.

While species surveys along altitudinal gradients provide us with information on current spatial species distribution patterns, a complementary phylogenetic analysis of species assemblages allows further key insights into spatio-temporal evolutionary pathways, as well as into phylogenetic and ecological traits^{10–12}. Such an analysis enables us to gain important information about the factors underlying the current structure of species assemblages^{13–15}. Evolving species and clades tend to retain some of their functional traits and niches, a phenomenon termed ‘niche conservatism’¹. Such functional traits in turn form a strong base for understanding how assemblage structures are shaped by environmental factors¹⁶. Phylogenetic niche conservatism, when integrated into the general concept of niche conservatism, suggests that closely related species might share more similar ecological traits than species with more dissimilar phylogenies¹.

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Overall, studies into phylogenetic assemblage structures along altitudinal gradients, particularly at the level of individual families, are rare. Several studies have reported an increase in phylogenetic relatedness with increasing elevation. These include studies of trees¹⁷, butterflies¹⁸ and soil bacteria¹⁹. However, they focussed on high taxonomic levels, therefore including a wide variety of different families forming distinctive inherent phylogenetic clusters. As niche-differentiations and related ecological traits between different families are likely to be highly conservative over time²⁰, a phylogenetic analysis combining numerous different families might underestimate the effects of environmental controls. An investigation of a species-rich family in this context may provide a different perspective in identifying the mechanisms shaping diversity patterns along environmental gradients.

In this study, we investigated the altitude-dependent species composition and phylogenetic structure in one of the most species-rich insect families, geometrid moths (Lepidoptera: Geometridae), as our target taxon. Geometrid moths are taxonomically well-known²¹, sensitive to environmental change²², and they can be surveyed readily on a range of different light sources^{23–25}. Apart from a very wide range of plant species, the extensive range of food sources used by geometrid caterpillars even includes other insects²⁶, and the wide ecological amplitude occupied by members of this family leads to their widespread distribution that regularly includes high latitude and altitude environments^{27–31}.

The altitudinal diversity patterns of geometrid moths have been studied intensively in tropical forest ecosystems of South America^{30–33}, Southeast Asia^{34–36} and Africa^{27,37}. In contrast, altitudinal transect studies from temperate regions particularly in Asia are generally rare. Our study was carried out in the largest remaining mature temperate forest area of Northern China inside the Changbaishan Nature Reserve (CNR). Established in the 1960s, this reserve contains one of the best-protected mature temperate forests in Asia^{38,39}.

Similar to results from other taxa, previous studies of altitudinal α -diversity gradients in geometrid moths provide mixed outcomes, with a monotonous decrease^{40,41}, hump-shaped distributions⁴² and other, more complicated distribution patterns^{27,43,44} being reported. In addition, Brehm, *et al.*³³ recorded a clustering trend in phylogenetic relatedness with increasing elevation in geometrid assemblages from a South American tropical rainforest. This trend resulted in an overall decrease in phylogenetic diversity for a standardized species richness with increasing elevation, indicating that only a small group of strongly interrelated geometrid species occupied the high-elevation environments. Results from tropical regions might nonetheless not be easily transferable to temperate regions, not least since temperate species are commonly believed to have wider environmental tolerance ranges^{45,46}.

Given the large prevailing gaps in knowledge and understanding of the diversity patterns in the highly species-rich insect taxa of temperate Asia, this study aimed to establish the spatial distribution of α -diversity for geometrid moths in the temperate forests of CNR along an altitudinal gradient, and to identify the drivers of the observed diversity changes. In response to environmental conditions becoming increasingly less favourable, we hypothesized that α -diversity in geometrid moths would decrease monotonously with increasing elevation. Our second aim was to examine patterns of phylogenetic relatedness in geometrid moth assemblages along the altitudinal gradient to obtain insights into underlying evolutionary and ecological processes. In this context, we hypothesized that geometrid species at high elevations would show strong phylogenetic relatedness and traits allowing them to adapt to the less favourable, more extreme environmental conditions in comparison to conditions experienced by species at lower elevations. We therefore hypothesized that the phylogenetic diversity for a standardized species richness would decrease with increasing elevation.

Results

In total, we caught 9285 geometrid moths. Of these, 1072 specimens (11.5%) were so badly damaged due chiefly to rainfall events in sampling nights that identification was deemed impossible. The remaining 8213 individuals were initially divided into 156 morpho-species. Apart from 1 rare species (2 individuals), all remaining 155 morphospecies formed the basis for DNA sequencing, and the pre-estimated species number was subsequently reduced to 130 molecular operational taxonomic units (MOTUs). Overall, the 131 putative species present in our samples represented five subfamilies; Ennominae, Geometrinae, Larentiinae, Orthostixinae and Sterrhinae (note: the validity of Orthostixinae at subfamily level is controversial, it was supported in the molecular phylogenetic analyses of Yamamoto and Sota⁴⁷ and Sihvonen, *et al.*⁴⁸, but questioned by Sihvonen, *et al.*⁴⁹).

The phylogenetic trees (Fig. 1) show that a large number of species occurred across a wide altitudinal range, with 44 species recorded in the entire forest area covered by the altitudinal transect. Furthermore, species with more limited distribution ranges were distributed across the entire taxonomic tree.

In relation to diversity patterns of individual subfamilies, Ennominae dominated at low elevations, while Larentiinae became increasingly dominant above 1600 m. Orthostixinae and Sterrhinae only occurred at elevations below 1100 m (Fig. 2). Along with the increase in elevation, the proportion of both Ennominae and Geometridae decreased in relation to both, overall abundance (Pearson correlation: $r = -0.82$ and $r = -0.66$, $P < 0.001$ for both case) and species richness ($r = -0.48$, $P = 0.013$, and $r = -0.57$, $P = 0.003$, respectively), while Larentiinae showed a significant increase in their proportional representation across the samples ($r = 0.86$ for abundance and $r = 0.75$ for richness, both at $P < 0.001$).

The overall rarefied species number of geometrid moths decreased significantly with increasing altitude (Pearson correlation, $r = -0.81$, $P < 0.001$, Fig. 3a). The overall estimated PD of each plot was linearly correlated with the total number of recorded species (Pearson correlation, $r = 0.99$, $P < 0.001$), hence also showing a significant decrease with increasing elevation ($r = -0.42$, $P = 0.034$, Fig. 3b). In contrast, the standardized effect size of MPD increased significantly with increasing elevation over the transect ($r = 0.66$, $P < 0.001$, Fig. 3c). Species showed a trend to form phylogenetic clusters (mean value = -1.74) at elevations below 1000 m asl., a mixed pattern between 1000 m and 1500 m (mean value = 0.28), and a trend to over-dispersion above 1500 m (mean value = 0.70). When standardizing the PD values for the minimum number of recorded species across the plots

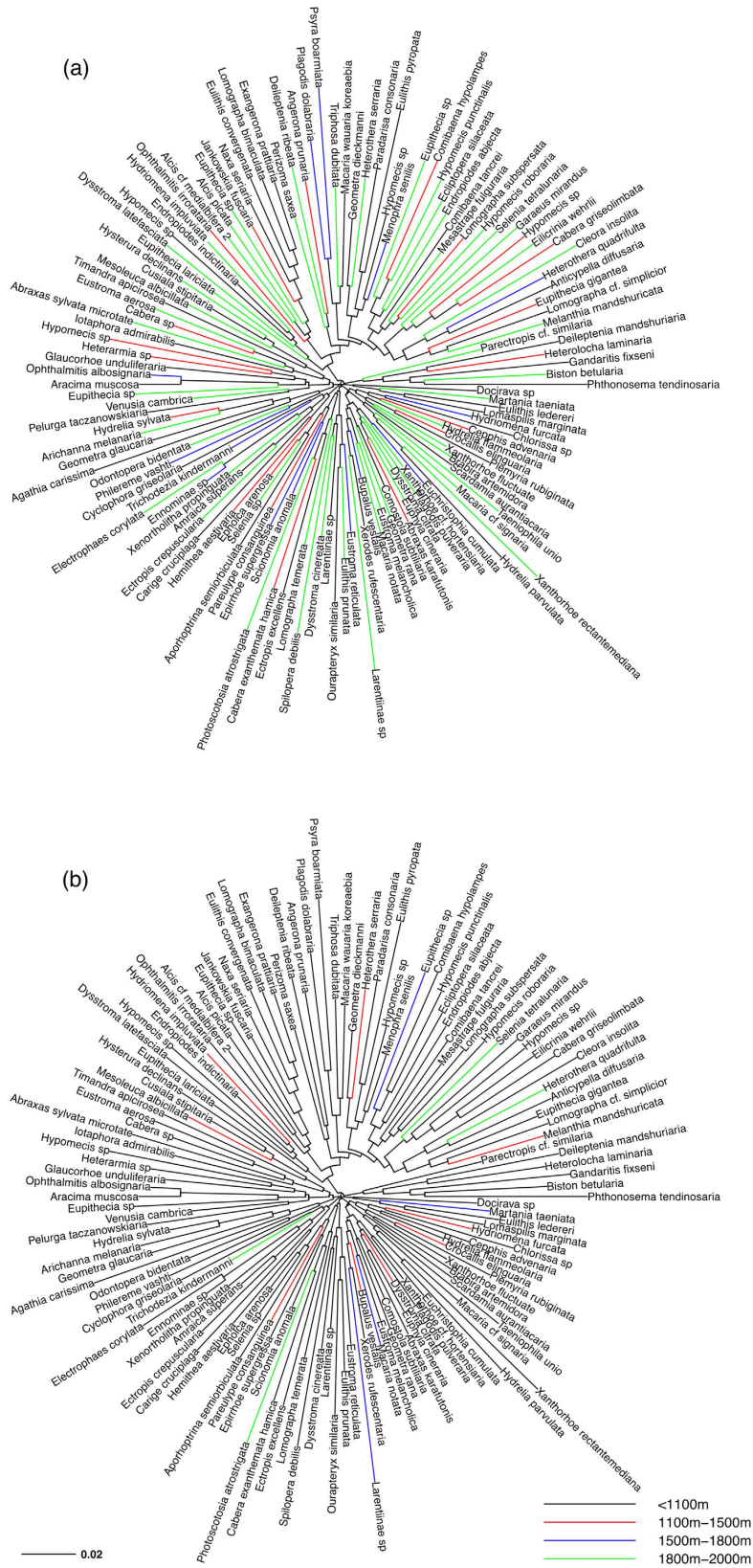


Figure 1. Neighbour-joining trees (COI 5' data, based on K2P distance) of geometrids on Changbai Mountain. Different colours of branch tips illustrate where species reach their maximum (a) and minimum (b) altitudinal limit.

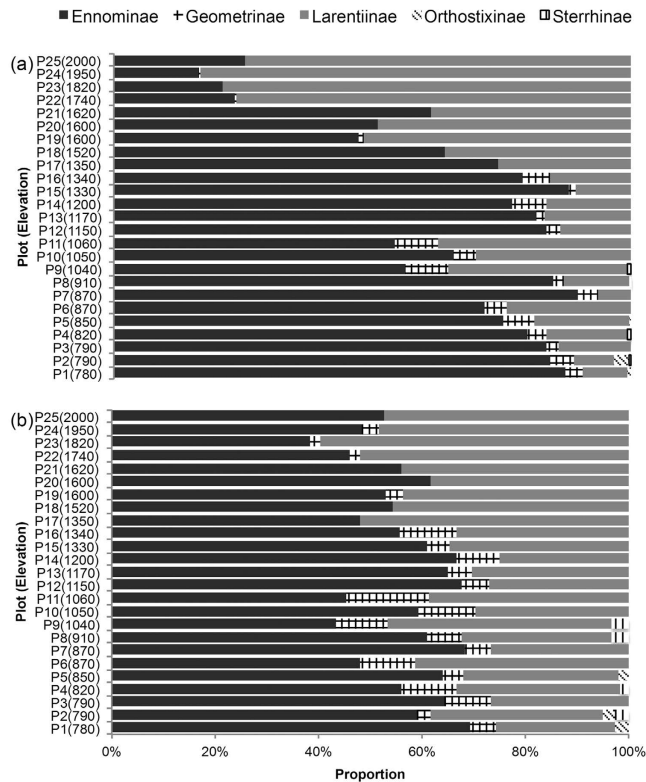


Figure 2. Proportion of subfamilies abundance (a) and species richness (b) at each sampling plot, ordered by elevation.

($n = 18$), a significant positive correlation with elevation was established ($r = 0.47$, $P = 0.016$, Fig. 3d), again indicating an overall positive correlation between the degree of over-dispersion in moth assemblages and altitude.

Discussion

In accordance with our first hypothesis, the α -diversity of geometrid moths decreased significantly and monotonously with increasing elevation across the entire altitudinal gradient. This result is in line with previous observations in the CNR area of geometrid moths by Chen *et al.*⁴¹, and of other insect taxa such as noctuid moths⁴⁰ and ground beetles⁵⁰, as well as for plant species¹⁷. The results suggest that only relatively few geometrid species can live in the cold conditions occurring at high altitude forests on Changbai Mountain. The decreasing geometrid species richness can also be related to the increased limitation of available resources linked to the decrease both in the overall plant diversity and in the area of altitudinal bands from the base to the top of the mountain^{51,52}.

The observed proportional decrease in Ennominae and increase in Larentiinae along the altitudinal gradient are consistent with previous studies from South America³³, Southeast Asia⁴² and East Africa²⁷. The increased proportion of Larentiinae with increased elevation reflects particularly strong adaptations to cooler and wetter environmental conditions in comparison to other geometrid subfamilies⁵³. Members of the Sterrhinae in this study showed a clear elevational boundary well below the forest line, with members of this family not encountered above 1100 m. This again mirrors findings by Axmacher, *et al.*²⁷, who reported that no Sterrhinae were present above 2600 m in the afro-tropical forests of Mt Kilimanjaro, indicating that Sterrhinae are particularly sensitive to the severe climatic conditions encountered at high-mountain environments.

In direct contradiction to our second hypothesis, a surprising key finding of this study is that both the average phylogenetic diversity and the phylogenetic diversity for a standardized species number increase significantly with increasing elevation. The species in the smaller species pool encountered at high elevations are therefore genetically significantly more distinct than species in assemblages at lower elevations, indicating an aggregating of phylogenetic lineages in low and an over-dispersion in high-elevation forest moth assemblages. This result strongly contrasts observations of tropical geometrid assemblages by Brehm, *et al.*³³, who reported a trend for over-dispersion at low elevations and a trend for phylogenetic clustering at high elevations. Given the much higher overall species richness in tropical Andean forests and the more complex α -diversity patterns with changing elevation⁴³, the contrast might be partly explained by the general differences between tropical and temperate geometrid assemblages and species pools. Due to the much greater uniformity of environmental conditions in tropical forests, species in these regions can have much narrower distribution ranges associated with narrower niche tolerances in comparison to species in temperate regions⁴⁶. Our results indeed support this assumption, given the large number of species with a wide altitudinal distribution range recorded from our study area. Most of these species must be assumed to be habitat and host-plant generalists, given their wide distribution across the different forest zones with their distinct differences not only in the tree cover, but also in the undergrowth

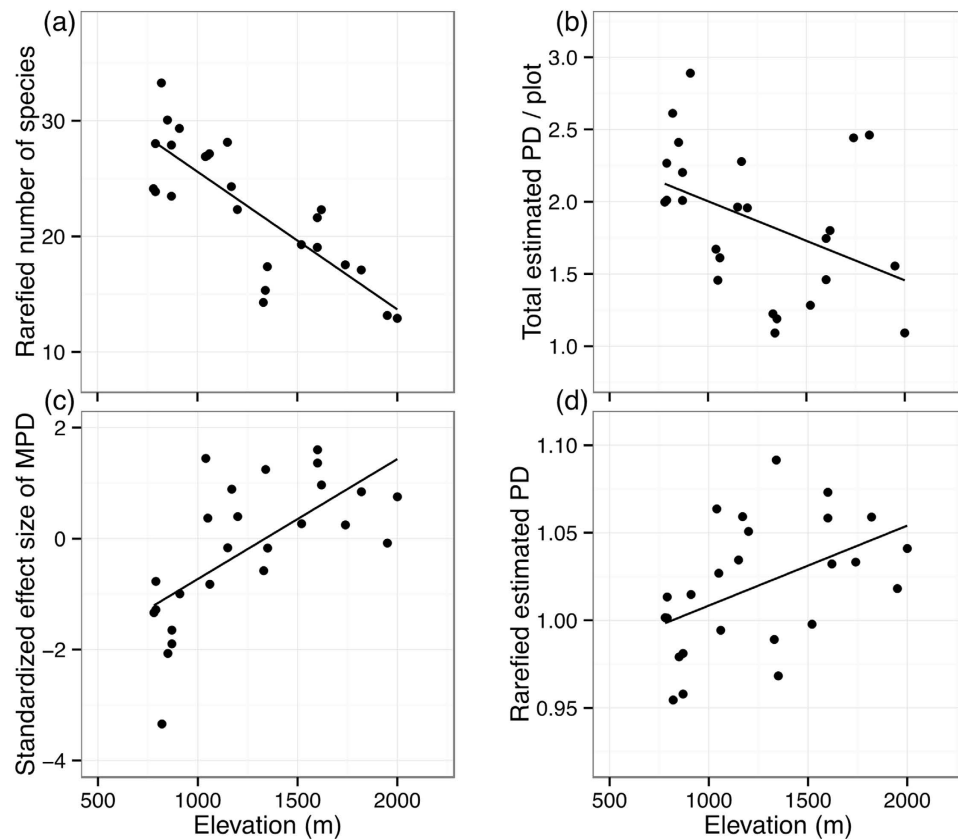


Figure 3. Rarefied number of species (a), total estimated phylogenetic diversity (b), standardized effect size of mean pairwise phylogenetic distance (MPD) (c) and rarefied estimated phylogenetic diversity ($n = 18$) (d) for geometrid moths with changing elevation.

vegetation⁵⁴. As our study area is covered in continuous forest vegetation and lacks strong physical barrier, these forest habitat and host-plant generalist moth species can easily disperse across the entire mountain range. Furthermore, our results indicate that cold tolerance in geometrid moths on Changbai mountain did not originate from a small number of basal lineages as observed in the tropics. In contrast, it appears that widely unrelated sets of habitat generalists have obtained this ability independently throughout their evolutionary past.

Our finding also stands in contrast to the increased phylogenetic relatedness with increasing elevation observed in angiosperms from our study region¹⁷. This might at first seem surprising, given that moths are closely linked to plants as herbivores and pollinators. Nonetheless, since Qian *et al.*¹⁷ focussed on a higher taxonomic level where phylogenetic patterns and associated traits are expected to be differentiated much more strongly than within species of a single family, results are not directly comparable. Shorter life cycles, higher fecundity and mobility in insects may also result in general differences between the phylogenetic patterns of insects and plants along spatial and environmental gradients. While niche theory suggests strong positive links between the α -diversity of herbivorous geometrid moths and plants, the existence of such positive links has furthermore rarely been recorded, with weak, non-significant or even significantly negative correlations reported from a number of field studies^{27,44}. The underlying lack of similarities in spatial α -diversity patterns of moths and plants was also observed from our study area (Zou, unpublished data). Overall, our results indicate that host-plant generalists dominate our temperate geometrid moth assemblages.

Local species pools are generally directly determined by the processes of speciation, extinction and dispersal^{55–57}. These determinants are strongly related to inter- and intra-specific interactions as well as to environmental conditions. Under natural conditions, the actual composition of species assemblages is widely determined by both environmental filtering and interspecific repulsion^{19,58,59}. In cases where local species pools are dominated by clusters of phylogenetically closely related species, their sharing of similar environmental niches suggests that repulsion in these clusters is only a minor determinant for the assemblage composition. In these cases, evolution will have favoured the specific traits present in these clusters that render them well-adapted to the local environmental settings. At low elevations, warm temperatures and a large amount and diversity of available resources are associated with an increased availability of niche space. Our results indicate that in combination with an intense intra-specific competition, these environmental conditions have resulted in higher speciation rates that have led to the existence of a greater diversity in phylogenetically closely related geometrid species in comparison to assemblages at higher elevations^{60,61}. Overall, it must be remembered that a large proportion of the regional geometrid species pool consists of environmental generalists that are able to survive across the environmental gradient, but with distinctly more low-elevation than high-elevation specialists being present in the samples. With

increasing elevation, exclusive competition appears to become increasingly important for closely related sets of geometrid moth species. This pattern can be linked to the combination of increasingly harsh environmental conditions, a reduction in available resources and the decrease in overall available area, and it explains the observed formation of high-elevation assemblages consisting of fewer, phylogenetically more distinct species.

Despite the decrease in species richness and the associated overall phylogenetic information contained in the moth assemblages with increasing elevation in this temperate forest ecosystem, their increased phylogenetic distinctiveness actually indicates a high diversity of the assemblages from a genetic perspective. This is likely linked to a strong resilience of the overall high-elevation assemblage towards environmental changes, an assumption further supported by the large proportion (69%) of environmental generalist species in this assemblage that were recorded over the entire environmental gradient. Of the 54 geometrid species encountered between 1800 and 2000 m, only four were restricted to this elevational band. Our results also provide support to the assumption that species in temperate geometrid moth assemblages are adapted to much wider ranges of environmental conditions throughout their phylogenetic lineages in comparison to tropical assemblages where the ability to survive in high-altitude environments appears much more restricted to a limited number of phylogenetic lineages.

Methods

Study area and sampling plots. The study was conducted on the northern slopes of Changbai Mountain (E127°43′–128°16′; N41°41′–42°51′), Jilin Province, Northeast China. The natural vegetation in this area can be classified as cool-temperate moist forest based on Holdridge's⁶² life zone system⁶³. A linear decrease in temperature with increasing elevation is accompanied by a general increase in precipitation along the entire mountain range¹⁷. Along the altitudinal gradient, distinct forest zones can be distinguished: mixed coniferous and broad-leaved forests at the mountain base (700 m–1100 m), followed with increasing elevation by mixed coniferous forests (1100 m–1500 m) and sub-alpine mixed coniferous forests (1500 m–1800 m), with birch forests (1800 m–2100 m) forming the upper forest boundary^{54,64,65}.

Sampling of geometrid moths. We selected 25 sampling plots located in the aforementioned forest types along an altitudinal gradient ranging from 700 m to 2000 m. Geometrid moths were sampled in automatic light traps. Sampling was conducted between 19:30 and 22:30, when geometrid moths are most active, with each plot sampled once each month in July and August in 2011 and in June 2012.

All specimens were first sorted to morpho-species. Subsequently, samples of the majority of morpho-species were sent to the Canadian Centre for DNA Barcoding to analyse the interspecific genetic dissimilarity as a proxy for phylogenetic distances.

Data analysis. As the number of observed species usually provides only a poor representation of α -diversity in mobile insects⁶⁶, we chose Hurlbert rarefaction⁶⁷ for the comparison of the α -diversity in geometrid moth assemblages between sampling plots. These rarefaction techniques have proven robust in comparing diversity values in a wide range of ecological studies^{68–70}.

For species identification, the first step was based on distinguishing morpho-species based on their wing patterns. In a second step, 152 selected specimens representing these morpho-species were submitted to DNA barcoding, which allowed discrimination to molecular operational taxonomic units ('MOTUs') based on a sequence divergence threshold of 2%^{71,72} using the Kimura 2-parameter (K2P) distance⁷³. This also roughly corresponds to the BIN system implemented on the Barcode of Life Data System (BOLD) database⁷⁴. If individuals of a species varied below the 2% threshold, the sequence with the shortest distance to the other respective species was included in the subsequent analysis³³. DNA barcoding allowed species identification and attribution to a Linnean binomen in 127 of the 152 MOTUs. These were supplemented by 3 MOTUs of species that we identified, but where we then used data already stored on BOLD for our subsequent analysis rather than sending specimens for barcoding ourselves. Although we acknowledge that some distinct species may share very similar COI sequences below the 2% threshold, we believe that this effect will be negligible for our limited samples from one small study region. Species were then further identified based on the comparison of MOTU information from BOLD database, where all our sequence data were also deposited.

Based on the K2P distance, the expected phylogenetic diversity (PD, sum of total phylogenetic branch length) for all species recorded at individual plots was then estimated using Faith's index⁷⁵. Additionally, the standardized effect size of the mean pairwise phylogenetic distance (MPD) was calculated to evaluate whether the respective species assemblage is clustered or over-dispersed across the phylogenetic tree. In this approach, a low and negative value indicates clustering on the phylogenetic tree, whereas high and positive values indicate an evenly or over-dispersed phylogenetic pattern⁷⁶. The value of this index represents the reverse of the relatedness index^{77–79}. It was calculated based on 1000 random iterations drawn from the recorded geometrid species pool.

All calculations and statistical analysis were conducted in R⁸⁰. We used the 'vegan' package⁸¹ to calculate rarefactions, 'Picante'⁷⁷ for the calculation of Faith's PD and MPD and 'ape'⁸² for calculations of phylogenetic distances.

References

- Wiens, J. J. *et al.* Niche conservatism as an emerging principle in ecology and conservation biology. *Ecol. Lett.* **13**, 1310–1324, doi: 10.1111/j.1461-0248.2010.01515.x (2010).
- Lomolino, M. Elevation gradients of species-density: historical and prospective views. *Global Ecol. Biogeogr.* **10**, 3–13, doi: 10.1046/j.1466-822x.2001.00229.x (2008).
- Mora, C. & Robertson, D. R. Causes of latitudinal gradients in species richness: a test with fishes of the Tropical Eastern Pacific. *Ecology* **86**, 1771–1782, doi: 10.1890/04-0883 (2005).
- Willig, M. R., Kaufman, D. M. & Stevens, R. D. Latitudinal gradients of biodiversity: pattern, process, scale, and synthesis. *Annu. Rev. Ecol. Syst.* **34**, 273–309, doi: 10.1146/annurev.ecolsys.34.012103.144032 (2003).

5. Sanders, N. J. & Rahbek, C. The patterns and causes of elevational diversity gradients. *Ecography* **35**, 1–3, doi: 10.1111/j.1600-0587.2011.07338.x (2012).
6. McCain, C. M. & Grytnes, J.-A. Elevational Gradients in Species Richness In *Encyclopedia of Life Sciences* doi: 10.1002/9780470015902.a0022548 (eLS, John Wiley & Sons, Ltd, 2010).
7. García-López, A., Micó, E. & Galante, E. From lowlands to highlands: searching for elevational patterns of species richness and distribution of scarab beetles in Costa Rica. *Divers. Distrib.* **18**, 543–553, doi: 10.1111/j.1472-4642.2011.00846.x (2011).
8. Rahbek, C. The elevational gradient of species richness: a uniform pattern? *Ecography* **18**, 200–205, doi: 10.1111/j.1600-0587.1995.tb00341.x (1995).
9. Rahbek, C. The role of spatial scale and the perception of large-scale species-richness patterns. *Ecol. Lett.* **8**, 224–239, doi: 10.1111/j.1461-0248.2004.00701.x (2005).
10. Webb, C. O., Ackerly, D. D., McPeck, M. A. & Donoghue, M. J. Phylogenies and community ecology. *Annu. Rev. Ecol. Syst.* **33**, 475–505, doi: 10.2307/3069271 (2002).
11. Du, Y. *et al.* Phylogenetic constraints and trait correlates of flowering phenology in the angiosperm flora of China. *Global Ecol. Biogeogr.* **24**, 928–938, doi: 10.1111/geb.12303 (2015).
12. Pyron, R. A. & Burbrink, F. T. Ecological and evolutionary determinants of species richness and phylogenetic diversity for island snakes. *Global Ecol. Biogeogr.* **23**, 848–856, doi: 10.1111/geb.12162 (2014).
13. Cavender-Bares, J., Kozak, K. H., Fine, P. V. A. & Kembel, S. W. The merging of community ecology and phylogenetic biology. *Ecol. Lett.* **12**, 693–715, doi: 10.1111/j.1461-0248.2009.01314.x (2009).
14. Smith, M. A., Hallwachs, W. & Janzen, D. H. Diversity and phylogenetic community structure of ants along a Costa Rican elevational gradient. *Ecography* **37**, 720–731, doi: 10.1111/j.1600-0587.2013.00631.x (2014).
15. Chalmardrier, L., Münkemüller, T., Lavergne, S. & Thuiller, W. Effects of species' similarity and dominance on the functional and phylogenetic structure of a plant meta-community. *Ecology* **96**, 143–153, doi: 10.1890/13-2153.1 (2014).
16. De Bello, F. *et al.* Partitioning of functional diversity reveals the scale and extent of trait convergence and divergence. *J. Veg. Sci.* **20**, 475–486, doi: 10.1111/j.1654-1103.2009.01042.x (2009).
17. Qian, H., Hao, Z. Q. & Zhang, J. Phylogenetic structure and phylogenetic diversity of angiosperm assemblages in forests along an elevational gradient in Changbaishan, China. *J. Plant Ecol.* **7**, 154–165, doi: 10.1093/jpe/rtt072 (2014).
18. Pellissier, L. *et al.* Phylogenetic alpha and beta diversities of butterfly communities correlate with climate in the western Swiss Alps. *Ecography* **36**, 541–550, doi: 10.1111/j.1600-0587.2012.07716.x (2013).
19. Bryant, J. A. *et al.* Microbes on mountainsides: Contrasting elevational patterns of bacterial and plant diversity. *P. Natl. Acad. Sci. USA* **105**, 11505–11511, doi: 10.1073/pnas.0801920105 (2008).
20. Hadly, E. A., Spaeth, P. A. & Li, C. Niche conservatism above the species level. *P. Natl. Acad. Sci. USA* **106**, 19707–19714, doi: 10.1073/pnas.0901648106 (2009).
21. Scoble, M. J. *Geometrid moths of the World - A catalogue (Lepidoptera: Geometridae)*. (CSIRO Publishing, 1999).
22. Holloway, J. D. Moth as indicator organisms for categorizing rain-forest and monitoring changes and regeneration processes. In *Tropical rain-forest: the Leeds Symposium* (eds A. C. Chadwick & S. L. Sutton) 235–242 (Philosophical and Literary Society, 1985).
23. Muirhead-Thomson, R. C. *Trap Responses of Flying Insects. The Influence of Trap Design on Capture Efficiency*. (Academic Press, 1991).
24. Intachat, J. & Woiwod, I. P. Trap design for monitoring moth biodiversity in tropical rainforests. *Bull. Entomol. Res.* **89**, 153–163, doi: 10.1017/S0007485399000243 (1999).
25. Axmacher, J. C. & Fiedler, K. Manual versus automatic moth sampling at equal light sources—a comparison of catches from Mt. Kilimanjaro. *J. Lepidopt Soc* **58**, 196–202 (2004).
26. Montgomery, S. L. Carnivorous caterpillars: the behavior, biogeography and conservation of Eupithecia (Lepidoptera: Geometridae) in the Hawaiian Islands. *GeoJournal* **7**, 549–556, doi: 10.1007/BF00218529 (1983).
27. Axmacher, J. C. *et al.* Diversity of geometrid moths (Lepidoptera: Geometridae) along an Afrotropical elevational rainforest transect. *Divers. Distrib.* **10**, 293–302, doi: 10.1111/j.1366-9516.2004.00101.x (2004).
28. Axmacher, J. C. *et al.* Determinants of diversity in afrotropical herbivorous insects (Lepidoptera: Geometridae): plant diversity, vegetation structure or abiotic factors? *J. Biogeogr.* **36**, 337–349, doi: 10.1111/j.1365-2699.2008.01997.x (2009).
29. Robinson, G. S., Ackery, P. R., Kitching, I. J., Beccaloni, G. W. & Hernández, L. M. *HOSTS - A Database of the World's Lepidopteran Hostplants* (2010) Available at <http://www.nhm.ac.uk/hosts> (Accessed: 28th October 2015).
30. Brehm, G., Homeier, J. & Fiedler, K. Beta diversity of geometrid moths (Lepidoptera: Geometridae) in an Andean montane rainforest. *Divers. Distrib.* **9**, 351–366, doi: 10.1046/j.1472-4642.2003.00023.x (2003).
31. Brehm, G. & Fiedler, K. Faunal composition of geometrid moths changes with altitude in an Andean montane rain forest. *J. Biogeogr.* **30**, 431–440, doi: 10.1046/j.1365-2699.2003.00832.x (2003).
32. Hilt, N., Brehm, G. & Fiedler, K. Diversity and ensemble composition of geometrid moths along a successional gradient in the Ecuadorian Andes. *J. Trop. Ecol.* **22**, 155–166, doi: 10.1017/S0266467405003056 (2006).
33. Brehm, G., Strutzenberger, P. & Fiedler, K. Phylogenetic diversity of geometrid moths decreases with elevation in the tropical Andes. *Ecography* **36**, 1247–1253, doi: 10.1111/j.1600-0587.2013.00030.x (2013).
34. Willott, S. J. The effects of selective logging on the distribution of moths in a Bornean rainforest. *Philos. T. Roy. Soc. B* **354**, 1783–1790, doi: 10.1098/rstb.1999.0520 (1999).
35. Beck, J., Schulze, C. H., Linsenmair, K. E. & Fiedler, K. From forest to farmland: diversity of geometrid moths along two habitat gradients on Borneo. *J. Trop. Ecol.* **18**, 33–51, doi: 10.1017/S026646740200202X (2002).
36. Chen, I.-C. *et al.* Elevation increases in moth assemblages over 42 years on a tropical mountain. *P. Natl. Acad. Sci. USA* **106**, 1479–1483, doi: 10.1073/pnas.0809320106 (2009).
37. Axmacher, J. C. & Fiedler, K. Habitat type modifies geometry of elevational diversity gradients in geometrid moths (Lepidoptera Geometridae) on Mt Kilimanjaro, Tanzania. *Trop. Zool.* **21**, 243–251 (2008).
38. Stone, R. A threatened nature reserve breaks down Asian borders. *Science* **313**, 1379–1380, doi: 10.1126/science.313.5792.1379 (2006).
39. Xue, D. & Tisdell, C. Valuing ecological functions of biodiversity in Changbaishan Mountain Biosphere Reserve in northeast China. *Biodivers. Conserv.* **10**, 467–481, doi: 10.1023/A:1016630825913 (2001).
40. Liu, S. D., Meng, Q. F. & Gao, W. T. Moth diversity in the northern slope of Changbai Mountain Nature Reserve. *Journal of Northeast Forestry University* **35**, 51–53 (2007).
41. Chen, Y. B. *et al.* On vertical distribution and fauna of the geometrid moth in Changbai Mountain. *Journal of Beihua University* **8**, 73–79 (2007).
42. Beck, J. & Chey, V. K. Explaining the elevational diversity pattern of geometrid moths from Borneo: a test of five hypotheses. *J. Biogeogr.* **35**, 1452–1464, doi: 10.1111/j.1365-2699.2008.01886.x (2008).
43. Brehm, G., Süssenbach, D. & Fiedler, K. Unique elevational diversity patterns of geometrid moths in an Andean montane rainforest. *Ecography* **26**, 456–466, doi: 10.1034/j.1600-0587.2003.03498.x (2003).
44. Axmacher, J. C., Liu, Y., Wang, C., Li, L. & Yu, Z. Spatial α -diversity patterns of diverse insect taxa in Northern China: Lessons for biodiversity conservation. *Biol. Conserv.* **144**, 2362–2368, doi: 10.1016/j.biocon.2011.06.016 (2011).
45. Olson, D. M. The distribution of leaf litter invertebrates along a Neotropical altitudinal gradient. *J. Trop. Ecol.* **10**, 129–150, doi: 10.1017/S0266467400007793 (1994).

46. Stevens, G. C. The latitudinal gradient in geographical range: how so many species coexist in the tropics. *Am. Nat.* **100**, 240–256, doi: 10.1086/284913 (1989).
47. Yamamoto, S. & Sota, T. Phylogeny of the Geometridae and the evolution of winter moths inferred from a simultaneous analysis of mitochondrial and nuclear genes. *Mol. Phylog. Evol.* **44**, 711–723, doi: 10.1016/j.ympev.2006.12.027 (2007).
48. Sihvonen, P., Staude, H. S. & Mutanen, M. Systematic position of the enigmatic African cycad moths: an integrative approach to a nearly century old problem (Lepidoptera: Geometridae, Diptychini). *Syst. Entomol.* **40**, 606–627, doi: 10.1111/syen.12125 (2015).
49. Sihvonen, P. *et al.* Comprehensive Molecular Sampling Yields a Robust Phylogeny for Geometrid Moths (Lepidoptera: Geometridae). *PLoS ONE* **6**, e20356, doi: 10.1371/journal.pone.0020356 (2011).
50. Zou, Y., Sang, W., Zhou, H., Huang, L. & Axmacher, J. C. Altitudinal diversity patterns of ground beetles (Coleoptera: Carabidae) in the forests of Changbai Mountain, Northeast China. *Insect Conserv. Divers.* **7**, 161–171, doi: 10.1111/icad.12039 (2014).
51. Rosenzweig, M. L. *Species Diversity in Space and Time*. (Cambridge University Press, 1995).
52. Price, P. W. *Insect Ecology*. (John Wiley and Sons, 1984).
53. Holloway, J. Macrolepidoptera diversity in the Indo-Australian tropics: geographic, biotopic and taxonomic variations. *Biol. J. Linn. Soc.* **30**, 325–341, doi: 10.1111/j.1095-8312.1987.tb00306.x (1987).
54. Bai, F., Sang, W. & Axmacher, J. C. Forest vegetation responses to climate and environmental change: A case study from Changbai Mountain, NE China. *Forest Ecol. Manag.* **262**, 2052–2060, doi: 10.1016/j.foreco.2011.08.046 (2011).
55. Condamine, F. L., Sperling, F. A. H., Wahlberg, N., Rasplus, J.-Y. & Kergoat, G. J. What causes latitudinal gradients in species diversity? Evolutionary processes and ecological constraints on swallowtail biodiversity. *Ecol. Lett.* **15**, 267–277, doi: 10.1111/j.1461-0248.2011.01737.x (2012).
56. Mittelbach, G. G. *et al.* Evolution and the latitudinal diversity gradient: speciation, extinction and biogeography. *Ecol. Lett.* **10**, 315–331, doi: 10.1111/j.1461-0248.2007.01020.x (2007).
57. Wiens, J. J. Global patterns of diversification and species richness in amphibians. *Am. Nat.* **170**, S86–S106, doi: 10.1086/519396 (2007).
58. Callaway, R. M. *et al.* Positive interactions among alpine plants increase with stress. *Nature* **417**, 844–848, doi: 10.1038/nature00812 (2002).
59. Bruno, J. F., Stachowicz, J. J. & Bertness, M. D. Inclusion of facilitation into ecological theory. *Trends Ecol. Evol.* **18**, 119–125, doi: 10.1016/S0169-5347(02)00045-9 (2003).
60. Culmsee, H. & Leuschner, C. Consistent patterns of elevational change in tree taxonomic and phylogenetic diversity across Malesian mountain forests. *J. Biogeogr.* **40**, 1997–2010, doi: 10.1111/jbi.12138 (2013).
61. Cavender-Bares, J., Ackerly, D. D., Baum, D. A. & Bazzaz, F. A. Phylogenetic overdispersion in Floridian oak communities. *Am. Nat.* **163**, 823–843, doi: 10.1086/386375 (2004).
62. Holdridge, L. R. *Life zone ecology*. (Tropical Science Center, 1967).
63. Zhang, X. A vegetation-climate classification system for global change studies in China. *Quaternary Sciences* **2**, 157–169 (1993).
64. Chen, L. Z., Bao, X. C. & Li, C. G. Major forests in various vertical zones on northern slope of Changbai Mountains of Jilin Province. *Acta Phytocologica et Geobotanica Sinica* **2**, 207–225 (1964).
65. Sang, W. & Bai, F. Vascular diversity patterns of forest ecosystem before and after a 43-year interval under changing climate conditions in the Changbaishan Nature Reserve, northeastern China. *Plant Ecol.* **201**, 115–130, doi: 10.1007/978-90-481-2795-5_10 (2009).
66. Gotelli, N. J. & Colwell, R. K. Quantifying biodiversity: procedures and pitfalls in the measurement and comparison of species richness. *Ecol. Lett.* **4**, 379–391, doi: 10.1046/j.1461-0248.2001.00230.x (2001).
67. Hurlbert, S. H. The nonconcept of species diversity: a critique and alternative parameters. *Ecology* **52**, 577–586, doi: 10.2307/1934145 (1971).
68. Hurlbert, A. H. Species–energy relationships and habitat complexity in bird communities. *Ecol. Lett.* **7**, 714–720, doi: 10.1111/j.1461-0248.2004.00630.x (2004).
69. Olszewski, T. D. A unified mathematical framework for the measurement of richness and evenness within and among multiple communities. *Oikos* **104**, 377–387, doi: 10.1111/j.0030-1299.2004.12519.x (2004).
70. Fiedler, K. & Truxa, C. Species richness measures fail in resolving diversity patterns of speciose forest moth assemblages. *Biodivers. Conserv.* **21**, 2499–2508, doi: 10.1007/s10531-012-0311-5 (2012).
71. Hausmann, A., Haszprunar, G. & Hebert, P. D. N. DNA barcoding the geometrid fauna of Bavaria (Lepidoptera): successes, surprises, and questions. *PLoS ONE* **6**, e17134, doi: 10.1371/journal.pone.0017134 (2011).
72. Strutzenberger, P., Brehm, G. & Fiedler, K. DNA barcoding-based species delimitation increases species count of Eois (Geometridae) moths in a well-studied tropical mountain forest by up to 50%. *Insect Sci.* **18**, 349–362, doi: 10.1111/j.1744-7917.2010.01366.x (2011).
73. Kimura, M. A simple method for estimating evolutionary rates of base substitutions through comparative studies of nucleotide sequences. *J. Mol. Evol.* **16**, 111–120, doi: 10.1007/BF01731581 (1980).
74. Ratnasingham, S. & Hebert, P. D. N. bold: The Barcode of Life Data System (<http://www.barcodinglife.org>). *Mol. Ecol. Notes* **7**, 355–364, doi: 10.1111/j.1471-8286.2007.01678.x (2007).
75. Faith, D. P. Conservation evaluation and phylogenetic diversity. *Biol. Conserv.* **61**, 1–10, doi: 10.1016/0006-3207(92)91201-3 (1992).
76. Kembel, S. W., Eisen, J. A., Pollard, K. S. & Green, J. L. The phylogenetic diversity of metagenomes. *PLoS ONE* **6**, e23214, doi: 10.1371/journal.pone.0023214 (2011).
77. Kembel, S. W. *et al.* Picante: R tools for integrating phylogenies and ecology. *Bioinformatics* **26**, 1463–1464, doi: 10.1093/bioinformatics/btq166 (2010).
78. Horner-Devine, M. C. & Bohannan, B. J. M. Phylogenetic clustering and overdispersion in bacterial communities. *Ecology* **87**, S100–S108, doi: 10.1890/0012-9658(2006)87[100:PCAOIB]2.0.CO;2 (2006).
79. Webb, C. O. Exploring the Phylogenetic Structure of Ecological Communities: An Example for Rain Forest Trees. *Am. Nat.* **156**, 145–155, doi: 10.1086/303378 (2000).
80. R Core Team (2011). *R: A language and environment for statistical computing*. R Foundation for Statistical Computing, Vienna, Austria. URL <http://www.R-project.org/>.
81. Oksanen, J. *et al.* *vegan: Community Ecology Package*. R package version 2.0-3. doi: 10.1111/j.1654-1103.2003.tb02228.x (2012).
82. Paradis, E., Claude, J. & Strimmer, K. APE: analyses of phylogenetics and evolution in R language. *Bioinformatics* **20**, 289–290, doi: 10.1093/bioinformatics/btg412 (2004).

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Author Contributions

Y.Z., J.C.A. and W.S. conceived and designed the experiments. Y.Z. performed the experiments. Y.Z., A.H. and J.C.A. analyzed the data. Y.Z., J.C.A., W.S. and A.H. wrote the manuscript.

Additional Information

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