ORIGINAL RESEARCH

The Role of Infiltrated T Lymphocyte in Oral Squamous Cell Carcinoma: Insights into Clinicopathological Characteristics and Prognosis

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Background: To compare and analyze the presence of CD4+ and CD8 + lymphocyte infiltrates in Oral squamous cell carcinoma (OSCC) tissue versus adjacent tissue and their clinical significance.

Methods: We enrolled a total of 152 patients diagnosed with OSCC, all of whom had confirmed diagnoses through pathological reports. Clinical and demographics data were extracted from medical records. Tissue microarrays were constructed and immunohistochemical staining for CD4 and CD8 was performed.

Findings: The average number of infiltrating CD4+ T cells in OSCC tumor tissue was 1026.22 ± 1163.36 cells/mm², which did not significantly differ from the count in adjacent tissue, which was 1163.36 ± 1013.23 cells/mm². However, the number of CD8+ T cell infiltration in tumor tissue was significantly higher than in adjacent tissue (655.25 ± 705.70 vs 504.56 ± 659.26 cells/mm2, p = 0.026). We observed that, among patients who consumed alcohol, the CD4+ T cell infiltration in tumor tissue being significantly lower than that in adjacent tissue (P=0.036). Moreover, the CD8+ T cell infiltration in cancer tissue was significantly higher than in adjacent tissue for T1-2 patients (p=0.005). Patients with higher CD8+ T cell in tumor tissue exhibited significantly improved overall survival (p = 0.043). Multivariate analyses revealed that alcohol consumption had a significant impact on the number of CD4+T lymphocytes in tumor tissue (OR = 0.403, P = 0.033) while T stage was the independent factor affecting CD8+ T lymphocyte infiltration in tumor tissue (OR = 0.459, P = 0.031).

Interpretation: OSCC patients with a higher number of CD8+ T lymphocyte infiltration in tumor tissue exhibited an improved prognosis.

Keywords: microenvironment, OSCC, CD8 T cells, CD4 T cells, prognosis

Introduction

Oral squamous cell carcinoma (OSCC) is the primary form of oral cancer.^{1,2} While the treatment of OSCC has evolved into comprehensive multimodal approaches, encompassing surgery, radiotherapy, chemotherapy, targeted therapy, and immunotherapy, the 5-year survival rate remains low at approximately 50%.^{3,4}

OSCC is marked by a multi-stage progressive malignant transformation, characterized by a unique tumor microenvironment.⁵ This microenvironment comprises diverse cellular and non-cellular elements. The cellular constituents encompass immune cells, fibroblasts, endothelial cells, adipocytes. Among immune cells, there are including T lymphocytes,

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B lymphocytes, neutrophils, dendritic cells, macrophages, natural killer cells and others. The interplay between OSCC tumor cells and the microenvironment elements creates a complex tumor ecosystem.⁶ These components influence tumor cells or engage in interactions via cytokines, therefore establishing a complicated regulatory network pivotal in the onset, progress, metastasis, recurrence, and treatment of OSCC.

Nonetheless, owing to the varied causes of OSCC and the intricacies microenvironment of the oral cavity, different OSCCs may present different tumor microenvironments.^{7,8} Notably, the infiltration of T lymphocytes within tumor tissue differs from that in adjacent tumor regions. Gannot et al reported an elevation in CD4+ CD8+ T lymphocytes and B cells in cases of moderate and severe dysplasia, as well as in OSCC compared to hyperkeratotic lesions, implying an enhanced antitumor immune response during OSCC development.⁹ Strauss et al demonstrated a transition toward an infiltrating CD25+FoxP3+ CD4+ cells in patients with head and neck squamous cell carcinoma.¹⁰ Ratio of CD8+ T lymphocytes to CD4+ cells was observed to increase following dysplastic changes, potentially indicating an immune system response.¹¹ Taken together, these findings may suggest that immune system play a crucial role in OSCC progression. But, however, the connection between T lymphocyte infiltration and its impact on the clinicopathological characteristics and prognosis of OSCC remains unclear.

Consequently, this study incorporated a serial of samples of OSCC tissues, aiming to investigate their potential relationship with the clinicopathological characteristics and prognosis. Our aim to compare and analyze the presence of CD4+ and CD8 + lymphocyte infiltrates in cancer tissue versus adjacent tissue and their potential clinical significance.

Subjects and Methods

Patient Enrollment

We enrolled a total of 152 patients diagnosed with OSCC, all of whom had confirmed diagnoses through pathological reports. Detailed data were collected, including lifestyle and clinicopathological information. Inclusion criteria including complete medical records, demographics and clinicopathological data. Patients were excluded if they met any of the following conditions: 1) Inadequate tissue samples or incomplete medical records, 2) A primary malignancy other than OSCC confirmed. The study was ethically approved by the Ethics Committee of Sun Yat-sen University Cancer Hospital (Approval No. SL-B2023-374-01), and informed consent was obtained from all participants.

Tissue Microarray Construction, Immunohistochemistry, and Pathological Scoring

Tissue microarrays were constructed using pairs of OSCC and adjacent tissue specimens. Immunohistochemical staining for CD4 and CD8 (CD4 antibody, 1:2000, ab133616, Abcam; CD8 antibody, 1:500, 10,980-R081, SinoBiological) was performed. As shown in <u>Supplementary Figure 1</u>. The quantification of tissue-infiltrating T lymphocytes was carried out by clinical pathologists from the Department of Pathology at Sun Yat-sen University Cancer Center.

Statistical Analysis

Normally distributed data were presented as mean \pm standard deviation (SD), while categorical data were expressed as percentages. Comparative analyses were conducted using *t*-tests and Chi-square as appropriate. We employed Pearson correlation and logistic regression for data analysis. Statistical analyses were carried out using SPSS software (version 13.0), with significance levels set at p < 0.05.

Results

Association Between Clinicopathological Data and Infiltration of CD4+ T Cells

In our study comprising 152 OSCC patients, as detailed in Table 1, patients were categorized into two groups: one with a higher number of CD4+ T cells infiltrating the tumor tissue compared to adjacent tissue (CD4 N<T group), and the other with an equal or lower number (CD4 N \ge T group). Most clinical data between these patient groups were similar, except for the prevalence of alcohol drinking habits. Notably, a higher proportion of patients in CD4 N \ge T group had a history of drinking (p = 0.033).

Baseline Variable	OSCC Patients CD4+ Lyr	P value	
	N≥T	N <t< th=""><th></th></t<>	
Sample Size	77	75	-
Age, years	57.11 ± 11.96	56.88 ± 10.89	0.904
Sex			0.091
Male	58 (75.3%)	47 (62.7%)	
Female	19 (24.7%)	28 (37.3%)	
pl6 status			0.288
Positive	14 (18.2%)	9 (12.0%)	
Negative	63 (81.8%)	66 (88.0%)	
Cell Differentiation			0.284
Moderate-Well	68 (88.3%)	70 (93.3%)	
Undifferentiation-Low	9 (11.7%)	5 (6.7%)	
Tobacco smoking			0.317
Yes	37 (48.1%)	30 (40.0%)	
No	40 (51.9%)	45 (60.0%)	
Alcohol consumption			0.033
Yes	21 (27.3%)	10 (13.3%)	
No	56 (72.7%)	65 (86.7%)	
T stage			0.255
T1-2	51 (66.2%)	56 (74.7%)	
Т3-4	26 (33.8%)	19 (25.3%)	
N stage			0.683
N0	51 (66.2%)	52 (69.3%)	
NI+	26 (33.8%)	23 (30.7%)	
M stage			NS
M0	77 (100.0%)	75 (100.0%)	
МІ	0 (0.0%)	0 (0.0%)	
AJCC stage			0.741
I–II	39 (50.6%)	40 (53.3%)	
III–IV	38 (49.4%)	35 (46.7%)	

Table I	Clinical Data	of OSCC	Patient with	Different	Infiltrating	CD4+	Cells
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Association Between Clinicopathological Data and Infiltration of CD8+ T Cells

Similarly, the patients were divided into two groups based on the number of CD8+ T cells infiltrating tumor tissue relative to adjacent tissue, as presented in Table 2. Most clinicopathological characteristics were comparable between these groups, except for the T classification. Patients in CD8 N<T group had a higher proportion of early T stage (p = 0.029).

Analysis of T Lymphocyte Infiltration in OSCC Tumor and Adjacent Tissue

Further analysis of the relationship between T lymphocyte infiltration and clinical pathology is illustrated in Figure 1. The average number of infiltrating CD4+ T cells in OSCC tumor tissue was 1026.22 ± 1163.36 cells/mm², which did not significantly differ from the count in para-cancerous tissue, which was 1163.36 ± 1013.23 cells/mm². However, the number of CD8+ T cell infiltration in tumor tissue (655.25 ± 705.70 cells/mm²) was significantly higher than in adjacent tissue (504.56 ± 659.26 cells/mm², p = 0.026).

Given our previous observation of a potential link between alcohol consumption and CD4+ T cell infiltration, we conducted a comprehensive analysis. As demonstrated in Figure 1C and D, no significant differences were found in CD4+ T cell infiltration between drinkers or in non-drinkers both in adjacent and tumor tissue (P=0.171 and P=0.347, respectively). However, among the group of patients who consumed alcohol, a noteworthy distinction emerged, with CD4+ T cell infiltration in tumor tissue being significantly lower than that in adjacent tissue (P=0.036).

Baseline Variable	OSCC Patients CD8+ Lyn	P value		
	N≥T	N <t< th=""><th colspan="2"></th></t<>		
Sample Size	64	88	_	
Age, years	58.20 ± 11.29	55.85 ± 11.65	0.215	
Sex			0.525	
Male	46 (71.9%)	59 (67.0%)		
Female	18 (28.1%)	29 (33.0%)		
pl6 status			0.288	
Positive	12 (18.8%)	11 (12.5%)		
Negative	52 (81.3%)	77 (87.5%)		
Cell Differentiation			0.530	
Moderate-Well	57 (89.1%)	81 (92.0%)		
Undifferentiation-Low	7 (10.9%)	7 (8.0%)		
Tobacco smoking			0.465	
Yes	26 (40.6%)	41 (46.6%)		
Νο	38 (59.4%)	47 (53.4%)		
Alcohol consumption			0.699	
Yes	14 (21.9%)	17 (19.3%)		
Νο	50 (78.1%)	71 (80.7%)		
T stage			0.029	
ТІ-2	39 (60.9%)	68 (77.3%)		
Т3-4	25 (39.1%)	20 (22.7%)		
N stage			0.168	
N0	39 (60.9%)	63 (71.6%)		
NI+	25 (39.1%)	25 (28.4%)		
M stage			NS	
M0	64 (100.0%)	88 (100.0%)		
MI	0 (0.0%)	0 (0.0%)		
AJCC stage			0.161	
I–II	29 (45.3%)	50 (56.8%)		
III-IV	35 (54.7%)	38 (43.2%)		

Table 2 Clinical Data of OSCC Patient with Different Infiltrating CD8+ Cells

Similarly, patients were divided into T1-2 and T3-4 groups based on T stage. We observed no differences in the number of infiltrating CD8+ T cells in adjacent tissue between these two groups. However, in the tumor tissue, patients in the T1-2 group exhibited significantly higher CD8+ T cell infiltration than those in the T3-4 group. Moreover, by paired analysis, it was evident that CD8+ T cell infiltration in cancer tissue was significantly higher than in adjacent tissue for T1-2 patients. While there were no significant difference in CD8+ T cell infiltration between adjacent and cancer tissue in the T3-4 group.

Correlation Between Tissue Infiltrating T Lymphocytes and Prognosis

We examined the association between overall survival in the CD4 N \ge T group and the CD4 N<T group. As depicted in Figure 2A, we found no statistically significant difference in overall survival between these two groups. However, in patients of CD8 N<T group and of the CD8 N \ge T group, we observed a significant difference. Patients in the CD 8N<T group exhibited significantly improved overall survival compared to those in the CD8 N \ge T group, as shown in Figure 2B (p = 0.043).

Next, we analyzed whether T lymphocytes infiltrating adjacent tissue have an impact on prognosis. We categorized patients into high adjacent tissue CD4+T lymphocyte infiltration and low adjacent tissue CD4+T lymphocyte infiltration groups, based on the median value of CD4+T lymphocytes infiltrating in adjacent tissue. We did not find any different in

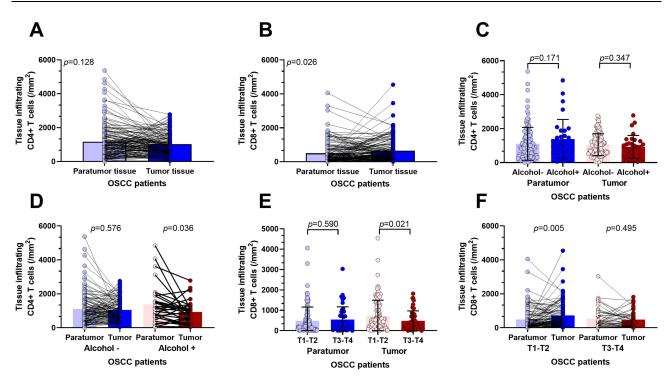


Figure I Analysis of T lymphocyte Infiltration in OSCC Tumor and Adjacent Tissue. (**A**). Number of CD4 cells infiltrating adjacent tissue averaged 1163.36±1013.23 cells/ mm², which did not significantly differ from the number in tumor tissue (1026.22±649.65 cells/mm²). (**B**). Number of CD8 cells infiltrating the adjacent tissue averaged 504.56±659.26 cells/mm², significantly lower than in the tumor tissue (655.25±705.71 cells/mm²) (P=0.026). (**C**). When comparing non-drinking patients to drinking patients, the CD4 cell in tumor tissue were 1054.09±642.43 and 930.79±672.70 cells/mm², respectively. CD4 cell in adjacent tissue were 1105.93±969.46 and 1385.66±1157.94 cells/ mm², respectively. (**D**). Among non-drinking patients, no statistically significant difference in CD4 levels between adjacent cancer tissues and cancer tissues. Among patients who alcohol consumption, the CD4 cell in tumor tissue were significantly lower than those in adjacent tissue (p=0.036). (**E**). For patients with T1-2 and T3-4 stages, the CD8 cell in the adjacent tissue were 485.78±674.97 and 549.21±625.39 cells/mm², respectively, while the CD8 cell in the cancer tissue were 727.64±769.49 and 483.14 ±489.46 cells/mm², respectively (p=0.021). (**F**). In patients with stage T1-2, CD8 cell in tumor tissue were significantly higher than in adjacent tissue (p=0.005), while there was no statistical difference in CD8 cell counts between adjacent tissue.

prognosis between these two groups. Similarly, when patients were divided into high adjacent tissue CD8+T lymphocyte and low adjacent tissue CD8+T lymphocyte groups, there were no significant differences in prognosis between these groups, as indicated in Figure 2C and D.

Regarding tumor recurrence, our analysis revealed that neither CD4+ T lymphocyte infiltration nor CD8+ T cell lymphocyte infiltration in tumor tissue had any impact on tumor recurrence. Similarly, the presence of CD4+ T lymphocytes and CD8+ T lymphocytes infiltrated in adjacent tissue did not influence tumor recurrence (Figure 2E–H).

Factors Related to Tumor-Infiltrating T Lymphocytes

We further explored variables linked to the number of T lymphocytes infiltrating tumor tissue. We observed a positive correlation between the numbers of CD4+ T lymphocytes and CD8+ T lymphocytes infiltrated in tumor tissue (p < 0.001, Figure 3A). Notably, data also revealed that increased CD4+ T lymphocyte infiltration into adjacent tissue was associated with a greater number of CD4+ T lymphocytes infiltrating tumor tissue (p = 0.027, Figure 3B). Similarly, CD8+ T lymphocyte infiltration into adjacent tissue corresponded to higher CD8+ T lymphocyte infiltration into the tumor tissue (p = 0.001, Figure 3C). And the increased number of CD4+T cell infiltrating cells in adjacent tissue correlated with a greater number of CD8+T cell infiltrating cells in the adjacent tissue (p < 0.001, Figure 3D).

Multivariable Analysis of T Lymphocyte Infiltration in Tumor Tissue

To access the independent factors affecting the numbers of CD4+ T lymphocyte infiltrations in tumor tissue, we conducted univariate and multivariate analyses. Our findings revealed that only alcohol consumption had a significant impact on the number of CD4+T lymphocytes in tumor tissue (OR = 0.403, P = 0.033), as shown in Table 3.

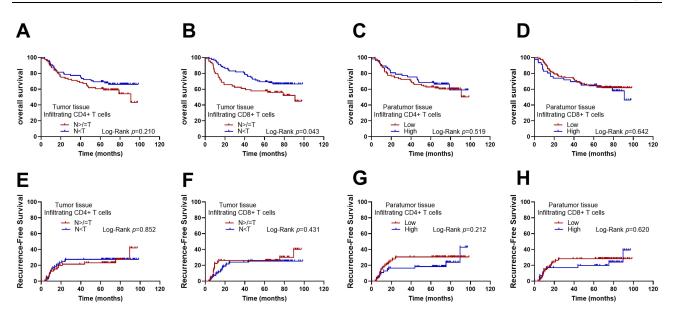


Figure 2 Correlation Between Tissue Infiltrating T Lymphocytes and Prognosis. (A). There is no statistically significant difference in overall survival between patients with CD4 cells in tumor tissue N \geq T group and other patients. (B). However, the overall survival of patients in the tumor tissue CD8 cell N<T group was significantly better than that of other patients (p=0.043). (C). No statistical relationship was found between CD4 cell infiltration in adjacent tissue and overall survival. (D). No statistical relationship was observed between CD8 cell infiltration in adjacent tissue and overall survival. (E). The number of CD4 cell infiltrates in tumor tissue did not have a statistically significant relationship with recurrence. (F). The number of CD8 cell infiltrates in tumor tissue did not have a statistically significant relationship between the number of CD4 cell infiltrates in adjacent tissue and recurrence. (H). Similarly, there was no statistically significant relationship between the number of CD8 cell infiltrates in adjacent tissue and recurrence.

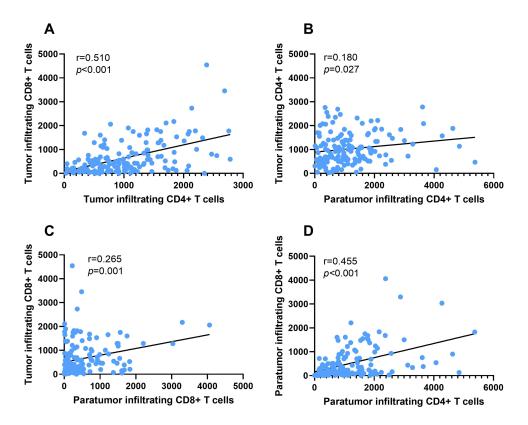


Figure 3 Factors Related to Tumor-Infiltrating T Lymphocytes. (A). In OSCC tumor tissue, a positive correlation between the number of CD4 cell infiltrations and the number of CD8 cell infiltrations. (B). The number of adjacent tissues CD4 cell infiltrations exhibits a positive correlation with the number of CD4 cells infiltrations in tumor tissue. (C). The number of CD8 cells in adjacent tissue shows a positive correlation with the number of CD8 cells infiltrating tumor tissue. (D). In OSCC adjacent tissue, the number of CD4 cells infiltrated was positively correlated with the number of CD8 cells.

Variables	Univariate Analysis			Mu	tivariate Ana	lysis
	OR	95% CI	Р	OR	95% CI	Р
Age, years	0.998	0.971-1.027	0.903			
Sex	1.787	0.888-3.595	0.103			
Tobacco Smoking	0.703	0.369-1.339	0.283			
Alcohol Consumption	0.403	0.175-0.928	0.033	0.403	0.175-0.928	0.033
T stage	0.652	0.323-1.319	0.234			
N stage	0.851	0.430-1.683	0.642			
AJCC stage	0.922	0.487-1.747	0.804			
Cell Differentiation	0.532	0.169–1.668	0.279			
p16 status	0.604	0.244–1.495	0.275			

 Table 3 Univariate and Multivariate Analyses for High Infiltrating CD4+ T Cells

Table 4 Univariate and Multivariate Analyses for High Infiltrating CD8+ T Cells

OR 0.982	95% CI	Р	OR	95% CI	Р
0.982				7378 CI	r
	0.955-1.010	0.215			
1.256	0.622-2.538	0.525			
1.275	0.665-2.446	0.465			
0.855	0.386-1.893	0.699			
0.459	0.226-0.931	0.031	0.459	0.226-0.931	0.031
0.619	0.313-1.226	0.169			
0.630	0.329-1.204	0.162			
0.704	0.234–2.116	0.532			
0.619	0.254-1.508	0.291			
	1.256 1.275 0.855 0.459 0.619 0.630 0.704	1.256 0.622–2.538 1.275 0.665–2.446 0.855 0.386–1.893 0.459 0.226–0.931 0.619 0.313–1.226 0.630 0.329–1.204 0.704 0.234–2.116	1.256 0.622–2.538 0.525 1.275 0.665–2.446 0.465 0.855 0.386–1.893 0.699 0.459 0.226–0.931 0.031 0.619 0.313–1.226 0.169 0.630 0.329–1.204 0.162 0.704 0.234–2.116 0.532	1.256 0.622-2.538 0.525 1.275 0.665-2.446 0.465 0.855 0.386-1.893 0.699 0.459 0.226-0.931 0.031 0.459 0.619 0.313-1.226 0.169 0.329-1.204 0.162 0.704 0.234-2.116 0.532 0.525 0.459	1.256 0.622-2.538 0.525 1.275 0.665-2.446 0.465 0.855 0.386-1.893 0.699 0.459 0.226-0.931 0.031 0.459 0.619 0.313-1.226 0.169 0.630 0.329-1.204 0.162 0.704 0.234-2.116 0.532

Similarly, for CD8+ T lymphocyte infiltration in tumor tissue, the multivariable analyses, as presented in Table 4, indicated that only T stage was the independent factor affecting CD8+ T lymphocyte infiltration in tumor tissue (OR = 0.459, P = 0.031).

Discussion

Lymphocyte infiltration into tumor tissue has a strong association between immune response, tumor characteristics, and overall prognosis.^{12–14} In this study, we noted that patients with a higher number of CD8+ T lymphocyte infiltration in tumor tissue exhibited an improved prognosis. This finding aligns with the results of previous research studies.^{15,16} We also found that the number of CD4+ T lymphocytes in tumor tissue or the level of T lymphocyte infiltration in adjacent tissue did not significantly correlate with prognosis. Furthermore, we identified an association between alcohol consumption and the infiltration of CD4+ T lymphocytes in tumor tissue, as well as a relationship between T stage and the infiltration of CD8+ T lymphocytes in tumor tissue. However, the underlying mechanisms behind these associations remain unclear.

Previous studies have found that the migration of T lymphocyte into tumor tissue is influenced by numerous factors, including vascular formation, cytokine expression, and tumor antigens.^{17,18} Moreover, the level of T lymphocyte infiltration in tumor tissue may have implications for immune checkpoint therapy.^{19–21} In this study, we observed that patients who consumed alcohol exhibited lower levels of tumor-infiltrating CD4+ T lymphocytes. In contrast, patients with early T-stage OSCC presented with increased infiltration of CD8+ T lymphocytes. This result may implied that early-stage OSCC patients may benefit more from immunotherapy. Conversely, for advanced T-stage OSCC patients, it may advisable to implement closer monitoring and follow-up after immunotherapy. However, these hypotheses warrant further research for validation.

Interactions between different immune lymphocytes also influence the infiltration of T lymphocytes into tumor tissue. For instance, CD4+ T lymphocytes are known to participate in the activation and proliferation of CD8+ T lymphocytes, and CD4+ T lymphocytes can regulate the inflammatory microenvironment, facilitating the recruitment of CD8+ T lymphocytes.^{22,23} Wong et al²⁴ reported that intratumoral CD4+ T lymphocytes promote the recruitment of CD8+ T lymphocytes. Other studies have suggested that CD4+ T lymphocytes in tumors can stimulate the expression of chemokines, such as CXCL9 and CXCL10, via IFN- γ production, thereby enhancing the infiltration of CD8+ T lymphocytes and CD8+ T lymphocytes in tumor tissue. Targeting CD4+ T lymphocytes to enhance the infiltration of CD8+ T lymphocytes may offer a potential therapeutic strategy to improve treatment response rates of immune checkpoint inhibitor and prognosis in patients.

The underlying reasons for the diminished infiltration of CD4+ T lymphocytes into tumor tissue among patients who consume alcohol raise intriguing questions. Although we found no significant difference in the number of CD4+ T lymphocytes infiltrating adjacent cancer tissues, regardless of alcohol consumption, it is possible that alcohol diminishes the migratory capacity of CD4+ T lymphocytes into tumor tissue. Existing research suggests that alcohol toxicity can impact CD4+ T cell function.²⁵ It may potentially reduce their infiltration into tumor tissue. In our study, we did not further classify the subsets of CD4+ T lymphocytes. However, we observed that patients with drinking habits exhibited a decreased number of CD4+ T lymphocytes infiltrating tumor tissue.

Patients with early T stage presented with increased CD8+ T lymphocyte infiltration in tumor tissue also raises questions. Previous research has indicated that the number of lymphocyte infiltrations changed as the disease progresses, reflecting immune evasion by tumor lymphocytes.²⁶ Factors such as changes in tumor antigens, modifications in cytokine expression profiles, and other elements might contribute to dynamic change of infiltrated T lymphocytes. In our study, we did not further evaluate the functional aspects of CD8+ T lymphocytes infiltrating tumor tissue. It may be possible that the function of these tumor-infiltrating CD8+ T lymphocytes might also undergo alterations as the T stage advances. Therefore, it's worth investigating whether the functionality of CD8+ T lymphocytes in tumor tissue is compromised as the T stage increases.

This study has some limitations, including a relatively small sample size, a single-center cohort, and an absence of analysis pertaining to further T lymphocyte subpopulations. These limitations need to be addressed and validated by large-sample multi-center studies.

In conclusion, this study unveiled the relationship between lymphocyte infiltration in OSCC tumor tissue. There is a positive correlation between the number of CD8+ T lymphocytes and CD4+ T lymphocytes infiltrating tumor tissue. Our study also revealed novel findings regarding the impact of alcohol drinking on the number of CD4+ T cells infiltrating tumor tissue and the influence of T stage on the number of CD8+ T cells infiltrating tumor tissue. OSCC patients who consume alcohol exhibit reduced infiltration of CD4+ T lymphocytes while patients with advanced T stage experience decreased infiltration of CD8+ T lymphocytes. A higher count of infiltrated CD8+ T cells within the tumor tissue is associated with better patient prognosis. These results offer insights the potential mechanism of tumor immune evasion and contribute to the development of precise and personalized immune checkpoint-based treatments.

Data Sharing Statement

Authors can confirm all relevant data are included in the article, materials are available on reasonable request from the authors and the data was deposited in the <u>Research Data Deposit respository</u>(NO. RDDA2024478017)

Ethics Approval and Consent to Participate

The study was performed in accordance with the Declaration of Helsinki and was approved by the Ethics Committee of Sun Yat-sen University Cancer Hospital endorsed the study (Approval No. SL-B2023-374-01). All patient provided written informed consent, agreed to follow the protocol and take specimens, and was willing to anonymously publish details of the medical record.

Acknowledgments

The authors would like to thank all the patients included in this study, and the nurses who assisted in patient management and collection of samples.

Funding

This study was supported by the Guangdong Basic and Applied Basic Research Fund (Natural Science Foundation of Guangdong Province) with No. 2023A1515010464, 2022A1515012631 and 2024A1515013244 and Guangdong Province Health Technology Promotion Project No. 202107021915135539.

Disclosure

None of the authors have competing interests to disclose.

References

- 1. Gharat SA, Momin M, Bhavsar C. Oral squamous cell carcinoma: current treatment strategies and nanotechnology-based approaches for prevention and therapy. *Crit Rev Ther Drug.* 2016;33(4):363–400. doi:10.1615/CritRevTherDrugCarrierSyst.2016016272
- Bugshan A, Farooq I. Oral squamous cell carcinoma: metastasis, potentially associated malignant disorders, etiology and recent advancements in diagnosis. *F1000Res*. 2020;9:229. doi:10.12688/f1000research.22941.1
- 3. Sasahira T, Kirita T. Hallmarks of Cancer-Related Newly Prognostic Factors of Oral Squamous Cell Carcinoma. Int J Mol Sci. 2018;19(8). doi:10.3390/ijms19082413
- 4. Jiang M, Li B. STAT3 and its targeting inhibitors in oral squamous cell carcinoma. Cells-Basel. 2022;11:19.
- 5. Mughees M, Sengupta A, Khowal S, Wajid S. Mechanism of tumour microenvironment in the progression and development of oral cancer. *Mol Biol Rep.* 2021;48(2):1773–1786. doi:10.1007/s11033-020-06054-6
- 6. Pignatelli P, Romei FM, Bondi D, Giuliani M, Piattelli A, Curia MC. Microbiota and oral cancer as a complex and dynamic microenvironment: a narrative review from etiology to prognosis. *Int J Mol Sci.* 2022;23(15):8323. doi:10.3390/ijms23158323
- 7. Ai R, Tao Y, Hao Y, et al. Microenvironmental regulation of the progression of oral potentially malignant disorders towards malignancy. *Oncotarget*. 2017;8(46):81617–81635. doi:10.18632/oncotarget.20312
- Eckert AW, Wickenhauser C, Salins PC, Kappler M, Bukur J, Seliger B. Clinical relevance of the tumor microenvironment and immune escape of oral squamous cell carcinoma. J Transl Med. 2016;14(1):85. doi:10.1186/s12967-016-0828-6
- 9. Gannot G, Gannot I, Vered H, Buchner A, Keisari Y. Increase in immune cell infiltration with progression of oral epithelium from hyperkeratosis to dysplasia and carcinoma. *Br J Cancer*. 2002;86(9):1444–1448. doi:10.1038/sj.bjc.6600282
- Strauss L, Bergmann C, Szczepanski M, Gooding W, Johnson JT, Whiteside TL. A unique subset of CD4+CD25highFoxp3+ T cells secreting interleukin-10 and transforming growth factor-beta1 mediates suppression in the tumor microenvironment. *Clin Cancer Res.* 2007;13(15 Pt 1):4345–4354. doi:10.1158/1078-0432.CCR-07-0472
- 11. Flores-Hidalgo A, Murrah V, Fedoriw Y, Padilla RJ. Relationship of infiltrating intraepithelial T lymphocytes in the diagnosis of oral lichen planus versus oral epithelial dysplasia: a pilot study. Or Surg or Med or Pa. 2019;127(6):e123-e135. doi:10.1016/j.oooo.2019.02.004
- 12. Stanton SE, Disis ML. Clinical significance of tumor-infiltrating lymphocytes in breast cancer. J Immunother Cancer. 2016;4(1):59. doi:10.1186/s40425-016-0165-6
- 13. van der Leun AM, Thommen DS, Schumacher TN. CD8(+) T cell states in human cancer: insights from single-cell analysis. *Nat Rev Cancer*. 2020;20(4):218–232. doi:10.1038/s41568-019-0235-4
- 14. Farhood B, Najafi M, Mortezaee K. CD8(+) cytotoxic T lymphocytes in cancer immunotherapy: a review. J Cell Physiol. 2019;234(6):8509–8521. doi:10.1002/jcp.27782
- Zhang Y, Li L, Zheng W, Zhang L, Yao N. CD8(+) T-cell exhaustion in the tumor microenvironment of head and neck squamous cell carcinoma determines poor prognosis. Ann Transl Med. 2022;10(6):273. doi:10.21037/atm-22-867
- Yamaguchi T, Kinoshita J, Saito H, et al. High CD8/CD33 ratio in peritoneal metastatic lesions is associated with favorable prognosis in gastric cancer. Cancer Rep Us. 2021;4(5):e1389. doi:10.1002/cnr2.1389
- 17. Rahir G, Moser M. Tumor microenvironment and lymphocyte infiltration. Cancer Immunol Immun. 2012;61(6):751-759. doi:10.1007/s00262-012-1253-1
- Paijens ST, Vledder A, de Bruyn M, Nijman HW. Tumor-infiltrating lymphocytes in the immunotherapy era. Cell Mol Immunol. 2021;18 (4):842–859. doi:10.1038/s41423-020-00565-9
- Gibney GT, Weiner LM, Atkins MB. Predictive biomarkers for checkpoint inhibitor-based immunotherapy. *Lancet Oncol.* 2016;17(12):e542–e551. doi:10.1016/S1470-2045(16)30406-5
- Baba Y, Nomoto D, Okadome K, et al. Tumor immune microenvironment and immune checkpoint inhibitors in esophageal squamous cell carcinoma. *Cancer Sci.* 2020;111(9):3132–3141. doi:10.1111/cas.14541
- 21. Tay C, Tanaka A, Sakaguchi S. Tumor-infiltrating regulatory T cells as targets of cancer immunotherapy. *Cancer Cell*. 2023;41(3):450–465. doi:10.1016/j.ccell.2023.02.014
- 22. Nakanishi Y, Lu B, Gerard C, Iwasaki A. CD8(+) T lymphocyte mobilization to virus-infected tissue requires CD4(+) T-cell help. *Nature*. 2009;462 (7272):510–513. doi:10.1038/nature08511
- Castellino F, Huang AY, Altan-Bonnet G, Stoll S, Scheinecker C, Germain RN. Chemokines enhance immunity by guiding naive CD8+ T cells to sites of CD4+ T cell-dendritic cell interaction. *Nature*. 2006;440(7086):890–895. doi:10.1038/nature04651
- 24. Wong SB, Bos R, Sherman LA. Tumor-specific CD4+ T cells render the tumor environment permissive for infiltration by low-avidity CD8+ T cells. *J Immunol.* 2008;180(5):3122–3131. doi:10.4049/jimmunol.180.5.3122
- 25. Barve SS, Kelkar SV, Gobejishvilli L, Joshi-Barve S, McClain CJ. Mechanisms of alcohol-mediated CD4+ T lymphocyte death: relevance to HIV and HCV pathogenesis. *Front Biosci.* 2002;7:d1689–d1696. doi:10.2741/A872
- 26. Hu S, Lu H, Xie W, et al. TDO2+ myofibroblasts mediate immune suppression in malignant transformation of squamous cell carcinoma. J Clin Invest. 2022;132(19). doi:10.1172/JCI157649

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