



Cloning and Molecular Characterization of Porcine β -casein Gene (CNS2)

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ABSTRACT : The production of therapeutic proteins from transgenic animals is one of the most important successes of animal biotechnology. Milk is presently the most mature system for production of therapeutic proteins from a transgenic animal. Specifically, β -casein is a major component of cow, goat and sheep milk, and its promoter has been used to regulate the expression of transgenic genes in the mammary gland of transgenic animals. Here, we cloned the porcine β -casein gene and analyzed the transcriptional activity of the promoter and intron 1 region of the porcine β -casein gene. Sequence inspection of the 5'-flanking region revealed potential DNA elements including SRY, CdxA, AML-a, GATA-3, GATA-1 and C/EBP β . In addition, the first intron of the porcine β -casein gene contained the transcriptional enhancers Oct-1, SRY, YY1, C/EBP β , and AP-1, as well as the retroviral TATA box. We estimated the transcriptional activity for the 5'-proximal region with or without intron 1 of the porcine β -casein gene in HC11 cells stimulated with lactogenic hormones. High transcriptional activity was obtained for the 5'-proximal region with intron 1 of the porcine β -casein gene. The β -casein gene containing the mutant TATA box (CATAAAA) was also cloned from another individual pig. Promoter activity of the luciferase vector containing the mutant TATA box was weaker than the same vector containing the normal TATA box. Taken together, these findings suggest that the transcription of porcine β -casein gene is regulated by lactogenic hormone via intron 1 and promoter containing a mutant TATA box (CATAAAA) has poor porcine β -casein gene activity. (**Key Words** : Lactogenic Hormones, HC11 Cells, TATA Box, Milk Protein, Porcine Casein Gene)

INTRODUCTION

The milk of transgenic animals has the potential for use as a bioreactor to product useful recombinant proteins or bioactive substances (Clark, 1998; Houdebine, 2009). Casein, which is one of the major components of milk proteins, includes calcium-sensitive caseins (α S1-, β -, and α -S2-casein) and κ -casein (Rosen et al., 1999; Rijnkels, 2001). Casein genes expressed in the mammary gland during lactation and are regulated by a variety of factors including steroid hormones, peptides, cell-cell and cell-extracellular matrix interactions (Doppler et al., 1989; Boudreau et al., 1995; Rosen et al., 1999). β -casein is a major component of cow, goat and sheep milk and its promoter has been used to regulate the expression of transgenic genes in the mammary gland of transgenic animals (Clark, 1998). β -casein is an abundant milk protein expressed in mammary epithelial cells during lactation. In

mammary epithelial cell lines, expression of β -casein depends on the synergistic action of insulin, glucocorticoid and prolactin (Doppler et al., 1989). Since β -casein promoter is regulated by the lactogenic hormone during lactation, the β -casein promoter has often been used for the generation of transgenic animals that produce proteins with mammary gland specific expression (Chang et al., 2010). The production of therapeutic proteins using transgenic pig is one of the most important and significant successes of animal biotechnology because pig has some advantage such as a short gestation and large litter size (Wheeler and Walters, 2001).

Bovine β -casein cDNA has been identified and found to consist of a 672 nucleotide coding region (Baev et al., 1987). β -casein is the major casein in bovine milk, accounting for as much as 30% of its total protein mass. The porcine β -casein cDNA sequence consists of a 696-bp open reading frame and encodes a protein of 232 amino acids in length (Alexander and Beattie, 1992). The bovine β -casein gene spans at least 8.5 kb and encodes 9 exons (Gorodetsky et al., 1988). Expression of the β -casein milk

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protein gene has been shown to be regulated by the addition of glucocorticoid and prolactin hormones in the murine mammary epithelial cell line, HC11 (Schmitt-Ney et al., 1991). The gene regulatory DNA sequences recognized by transcription factors are conserved in the promoters of different casein genes from several species (Mercier and Viloite, 1993; Rosen et al., 1999). However, transcriptional enhancer elements present in the intron increase the basal activity of the β -casein promoter. In addition, these intronic enhancer elements augment the induction of β -casein promoter by lactogenic hormones (Kolb, 2003). Casein cDNA and genomic DNA of many species have been isolated and characterized by molecular cloning. However, porcine β -casein genomic DNA has not yet been isolated or characterized.

In the present study, we describe the exon/intron organization of the porcine β -casein gene. We also analyzed the transcription unit of the porcine β -casein gene in the mouse mammary epithelial cell line, HC11. We demonstrated that the first intron of the porcine β -casein gene contains a transcriptional promoter/enhancer sequence and that lactogenic expression of the porcine β -casein gene was probably regulated by the first intron. Moreover, we found that porcine β -casein genes promoters containing a mutant TATA box (CATAAAA) have poor promoter activity.

MATERIALS AND METHODS

Cloning of porcine β -casein gene by long PCR

Porcine genomic DNA was extracted from the ear tissues of three-way crossbreds (LWD) obtained at the slaughterhouse. LA-PCR (Long and Accurate PCR) was conducted to identify the porcine β -casein genomic DNA as follows using primers prepared based on the porcine β -casein promoter region (Accession No. AY452035) reported to the NCBI and the result of cDNA analysis. The primers for PCR were as follows; porcine β -casein promoter sense primer (CCCACTATTTCTGATTCTTGA TTAAC TTT), exon 6 antisense primer (TGTTGTTCTT CCCGCTTTAGCTTCTCAATT), exon 2 sense primer (GACTTGATCGCCATGAAGCTCCTCATCCTT) and exon 9 antisense primer (GCCTAAGGATTAATTTATTG AAATGACTGG). PCR was conducted using 100 ng of the genomic DNA extracted from porcine ear tissues with 10 pmol of sense and antisense primers, 0.5 U i-Max II DNA polymerase (Intron, Korea), 1 \times PCR buffer and 200 μ M dNTP mixture as follows: 2 min of initial denaturation at 94°C followed by 30 cycles of 94°C for 30 s, 63°C for 30 seconds, 72°C for 7 min and 30 s, followed by a final elongation step at 72°C for 15 min. The PCR products were confirmed by electrophoresis on 0.8% agarose gel, after which they were cloned into the pGEM T-easy vector

(Promega, USA). Sequencing was conducted using an ABI PRISM 377 sequencer. The nucleotide sequence was analyzed by Genetyx-win (version 4.0).

Construction of luciferase vector

To test for promoter activity, the 5'-flanking region containing promoter with and without intron 1 was fused to the firefly luciferase gene in pGV-B2 (TOYO INK MFG, Japan) as follows. For construction of luciferase vector containing only promoter (-2,657 to +39) of porcine β -casein gene, PCR was conducted using porcine β -casein 5' arm DNA containing normal TATA or mutant TATA box as a template. A sense primer (CCCACTATTTCTGATTCTT GATTAACTTT) and antisense primer (CTCGAGTCC AAAGTGGAGGACAAGAAGTGA) containing the *Xho*I restriction enzyme site were designed for ligation in the multi cloning site of pGV-B2 vector. PCR was conducted using 10 pg of DNA and 10 pmol of sense and antisense primers, 1 \times PCR buffer, 0.5 unit of i-MaxII polymerase (Intron, Korea) and 200 μ M of dNTP mixture. PCR was conducted under the following conditions: 2 min of initial denaturation at 94°C, followed by 35 cycles of 94°C for 30 s, 62°C for 30 s, 72°C for 4 min and a final elongation step at 72°C for 15 min. PCR products were ligated with pGEM T-easy vector to generate pGEM- β -Promoter plasmid (Promega, USA). Inserts were prepared by digestion of pGEM- β -Promoter plasmid with *Eco*RV and *Xho*I restriction enzyme. pGV-B2 vector was prepared by digestion of the *Sma*I and *Xho*I restriction sites, after which it was ligated with an insert containing normal TATA or mutant TATA box from pGEM- β -Promoter plasmid (denoted p β CE1TATA and p β CE1mTATA, respectively). For construction of luciferase vector containing promoter and intron 1 (-2,657 to +2,450) of porcine β -casein, PCR was conducted using porcine β -casein 5' arm DNA containing normal TATA or mutant TATA box as a template. For cloning in pGV-B2 vector, sense primer (CCCACTA TTTCTGATTCTT GATTAACTTT) and antisense primer (CCATGGCGATCAAGTCTGTGAATGGGGAA) containing the *Nco*I restriction enzyme site were designed. PCR was conducted with 10 pg of DNA using 10 pmol of sense and antisense primers, 1 \times PCR buffer, 0.5 unit of i-MaxII polymerase (Intron, Korea) and 200 μ M of dNTP mixture. PCR was conducted under the following conditions: 2 min of initial denaturation at 94°C followed by 35 cycles of 94°C for 30 s, 62°C for 30 s, and 72°C for 5 min, with a final elongation step at 72°C for 15 min. PCR products were ligated with pGEM T-easy vector to generate pGEM- β -intron1 plasmid (Promega, USA). Inserts were prepared by digestion of pGEM- β -intron1 plasmid with *Sma*I and *Nco*I restriction enzyme. Inserts were subcloned into the vector pGV-B2 to generate p β CE2TATA and

p β CE2mTATA luciferase vector.

DNA transfection

For luciferase vector transfection, pCMV β -galactosidase plasmid (Clontech, USA) was used for normalization. Transfection was conducted as follows. HC11 cells were inoculated on a 96 well plate at a density of 1×10^4 cells and the culture medium was replaced with fresh medium the next day. The cells were then transfected using jetPEI transfection reagent (Polyplus-transfection, France) according to the manufacturer's instruction. Luciferase vector (0.05 pmol) and pCMV- β -gal plasmid (0.02 pmol) were used. In the transfection experiment, parallel plates of cells were transfected with pGVB2 and pGV-P2, which served as negative and positive controls, respectively. The plasmid pGV-B2 lacks a eukaryotic promoter; therefore, no luciferase activity was detected in cells transfected with pGV-B2. The plasmid pGV-P2 contains the SV40 early promoter, which drives expression of the luciferase gene. The transfection efficiency was assayed by co-transfection with the β -galactosidase reporter, pCMV β -gal.

Lactogenic hormonal induction of HC11 cells

After the transfection, cells were cultured for 24 h. The culture medium was replaced with serum free medium for 48 hours with hormone treatment. Hormone treatment was conducted using 5 μ g/ml insulin (Sigma, USA), 1 μ M dexamethasone (Sigma, USA), and 5 μ g/ml prolactin (Sigma, USA) in serum free RPMI 1640 (HyClone, USA) for 48 hours. The negative control was not treated with hormones.

Firefly luciferase assays

After hormone treatment for 48 hours, the cells were washed two times with $1 \times$ PBS and then lysis buffer (50 μ l) was added into cells. The cell lysates were transferred to 1.5 ml tubes. After collection, cells were immediately put into the liquid nitrogen. Frozen cells were then thawed at 37°C and centrifuged at 12,000 rpm and 4°C for 2 min, after

which the supernatant was collected. For measurement of the luciferase activity, 20 μ l supernatant and 50 μ l of Luciferase Assay System with Reporter Lysis Buffer (Promega, USA) were added to a 96 well plate and analyzed using a GloMax 96 microplate Luminometer with single reagent injector (Promega, USA) according to manufacturer's instructions. The transfection efficiency was determined by co-transfection with the β -galactosidase reporter, pCMV β -gal. β -galactosidase activity was measured using a chemiluminescence assay kit (Promega, USA) according to the manufacturer's instructions. The relative light units were calculated by normalizing the firefly luciferase activity against the β -galactosidase activity to evaluate the transfection activity.

RESULTS

Cloning and molecular characterization of porcine β -casein gene

To identify the β -casein gene in porcine cells, LA-PCR was conducted using two pairs of β -casein specific primers and porcine genomic DNA as template. PCR products containing promoter-exon 6 and exons 2-9 were amplified with 6.8 kb and 5.5 kb (data not shown). Figure 1 summarizes the genomic organization of the porcine β -casein gene. Exon 1 encoded the 5'-UTR, while exon 2 contained the translational initiator codon AUG. Exon 8 contained the translation termination codon and exon 9 contained the 3'-UTR. The sequence reported in this paper has been deposited in the GenBank database under accession no. EU242520.

All intron/exon splice junctions conformed to the GT/AG rule. As shown in Table 1, exon 1 was 45 bp, exon 2 was 63 bp, exon 3 was 27 bp, exon 4 was 27 bp, exon 5 was 24 bp, exon 6 was 45 bp, exon 7 was 519 bp, exon 8 was 42 bp, and exon 9 was 306 bp. With the exception of exon 7 and exon 9, all other exons very short. Additionally, intron 1 was 2,398 bp, intron 2 was 735 bp, intron 3 was 118 bp, intron 4 was 1,325 bp, intron 5 was 94 bp, intron 6

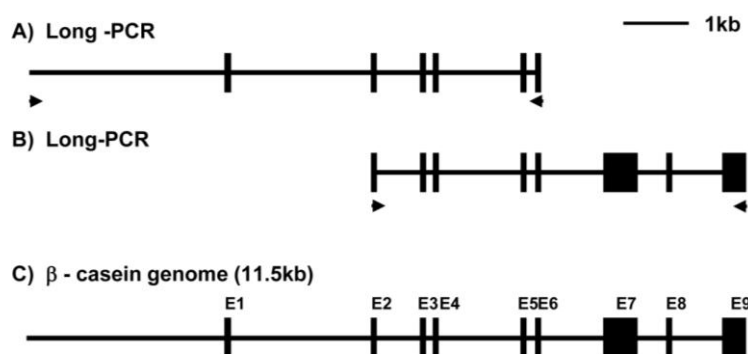


Figure 1. Schematic diagram of overlapping PCR products encoding the porcine β -casein gene. Exons are indicated by numbers and closed boxes. Arrows indicate PCR primer.

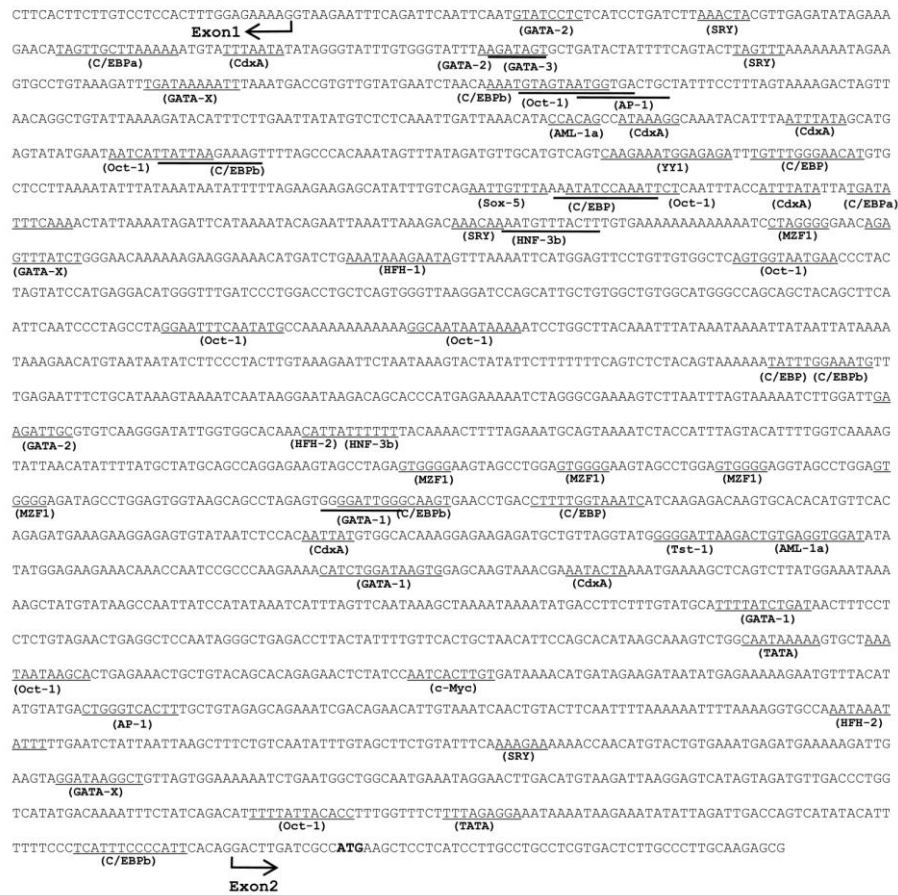


Figure 4. Nucleotide sequence of intron 1 of the porcine β -casein gene. Potential binding sites for transcription factors (>90% cutoff score) are underlined. Sequence information is available under GenBank Accession number EU242520.

Caseins are major milk proteins that account for nearly 80% of the proteins in milk. Milk contains several caseins, α s1-, α s2-, β - and κ -casein, which are encoded by independent genes (Doppler et al., 1989; Rosen et al., 1999). Transcription of the casein gene in mammary epithelial cells is regulated by the lactogenic hormones, insulin, glucocorticoids, and prolactin. The synergistic action of glucocorticoid and prolactin hormones on the rat and pig β -casein promoter has been examined in the murine mammary epithelial cell line, HC11 (Doppler et al., 1989; Boudreau et al., 1995; Rosen et al., 1999; Lee et al. 2008).

Functional analysis of the 5'-flanking region of the porcine β -casein gene was conducted in HC11 cells using the genomic DNA fragment containing the 5'-flanking region -2,657 to +39 and -2,657 to +2,450, which includes intron 1 in the β -casein gene containing mutant TATA and normal TATA. In the β -casein gene containing the normal TATA box, the luciferase activity of the 5'-flanking region containing intron 1 (-2,657 to +2,450) by lactogenic hormone treatment was 15 fold higher than that of the 5'-flanking region not containing intron 1 (-2,657 to +39). Our results are consistent with previous data, which suggests

that expression of the β -casein gene is regulated by lactogenic signals via intron 1 (Kolb, 2003; Lee et al., 2008). Moreover, the luciferase activity of the 5'-flanking region containing the mutant TATA box in the promoter region was lower than that of the 5'-flanking region containing the normal TATA box. It was previously reported that the TATA box in the vasopressin and β -globin gene have a deviant CATAAAA box (Xu et al., 1991; Ho and Murphy, 2002). These mutant TATA boxes have also been shown to elicit relatively weak promoter activity (Xu et al., 1991; Ho and Murphy, 2002). The results of the present study indicate that the TATA box in the promoter region plays an important role in expression of the β -casein gene.

The 5' flanking region of the porcine β -casein gene also contained DNA motifs for SRY, CdxA, AML-a, GATA-3, GATA-1, and C/EBP α . However, the GR, AP-1, STAT5 and YY-1 DNA motifs were not found in this region of the porcine β -casein gene. The first intron of the porcine β -casein gene contained the putative Oct-1, SRY, YY1, C/EBP β , AP-1, and retroviral TATA box.

The glucocorticoid receptor (GR), AP-1 (fos/jun dimer), STAT5 and YY-1 (Ying Yang-1) have been implicated in

β -casein gene expression in mice and humans (Mercier and Viloite, 1993; Rosen et al., 1999). However, these DNA motifs were not found in the 5'-flanking region of the porcine β -casein gene. It has previously been reported that the first intron of the mouse and bovine β -casein gene contains enhancer elements (Kang et al., 1998; Kolb, 2003). The first intron of the mouse β -casein gene contains putative GR, YY1, C/EBP, Oct-1, AP-1 and TATA box (Kolb, 2003). In this study, the first intron of the porcine β -casein gene contained the transcriptional enhancers Oct-1, SRY, YY1, C/EBP β , and AP-1, and the retroviral TATA box. However, the DNA motif of the glucocorticoid receptor was not found in the first intron of the β -casein gene. These potential sites may be required for basal transcription and regulation of the porcine β -casein gene by the lactogenic hormone.

Here, we demonstrated that the first intron of the porcine β -casein gene contains a transcriptional promoter/enhancer sequence and that lactogenic expression of the porcine β -casein gene was probably regulated by the first intron. However, the mutant TATA box (CATAAAA) in the porcine β -casein gene has weak promoter activity. Further studies are required to elucidate the actual nature and regulation of the porcine β -casein gene. Thus, these data can serve as a foundation for mammary gland-specific expression of foreign genes in gene targeting pigs.

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