



# Insights into garlic (*Allium Sativum*)'s nutrigenomics-associated fly-repellent potency in cattle

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## Abstract

Despite effective control of flies using synthetic pesticides, fly resistance and environmental contamination have led to the inadequacy of this strategy. The use of integrated pest management approaches has since been advocated in contemporary research to sustainably control fly populations. Recent studies have found garlic (*Allium Sativum*) and its derivative bioactive compounds to possess insect-repellent attributes among other key health and production enhancing properties. This highlights the potential of garlic as a botanical pesticide to control flies in cattle. Moreover, the ability of cattle to naturally repel flies is influenced by animal genetic predisposition. The dietary garlic supplementation and gene interaction in disease resistance could also be an influential factor in repelling flies in cattle. Transcriptomics has emerged as a valuable tool in animal breeding and genetics which allows identification of trait-associated genes and understanding of complex interactions between dietary nutrients and animal genome expression. This paper explores the nutrigenomic effects of garlic supplementation on cattle and its contribution towards fly repellence efficacy in cattle. It was concluded that garlic supplementation in cattle diets could offer a sustainable approach to managing fly infestations in cattle farming. These findings underscore the importance of further research to validate these assertions and optimise the use of garlic to control flies in cattle under different production systems.

**Keywords** Botanical pesticides · Cattle · Garlic supplementation · Gene expression · Fly infestation · Nutrigenomics

## Introduction

The repercussions of food and nutrition insecurity are manifold, often extending beyond the realms of mere access to food. In regions where livestock rearing is a crucial source

of nourishment and economic stability, the growing concern of fly infestation among cattle compounds the challenges (Ilemobade 2009; Saini et al. 2017; Moreki et al. 2022). Flies relentlessly attack and torment cattle inflicting a lot of stress to the animals that results in impaired productivity, hindering producers' ability to meet the demands of meat and dairy products (Taylor et al. 2012; Moreki et al. 2022). Consequently, this exacerbates the strain on already fragile food systems, intensifying food and nutrition insecurity for communities reliant on livestock. Obligate blood-sucking arthropods such as the stable fly (*Stomoxys calcitrans*) and the horn fly (*Haematobia irritans irritans*) can also act as mechanical and biological vectors of diseases, such as lumpy skin disease virus and bovine leukaemia virus (Panei et al. 2019; Sprygin et al. 2019; Makhahlela et al. 2022), respectively. In South Africa, the stable fly (*Stomoxys calcitrans*), the house fly (*Musca domestica*), and a few horse fly species (*Tabanidae*) are the most economically detrimental pests on beef cattle in feedlots (Evert 2014), causing an additional expense of chemical fly control (Evert and Van Hamburg

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2014; Makhahlela et al. 2022). For example, the annual economic losses associated with these flies account for over US\$ 2 billion in the United States (Taylor et al. 2012).

Filthy flies have over the years been effectively controlled with several chemical pesticides or larvicides; however, reliance on their use has become risky and inadequate due to the insecticidal resistance that has been amply reported (Foil and Hogsette 1994; Cook 2020). This is mostly due to the under- and over-application of insecticides over time, which led to flies developing immunity against currently used insecticides (Iqbal et al. 2014; Siddiqui et al. 2023). Furthermore, these synthetic chemicals' persistent residues can contaminate food and the environment (Carvalho 2006). Owing to an increase in the incidence of insecticide resistance, and environmental contamination concerns, it is thus imperative to investigate alternative mitigation strategies (Iqbal et al. 2014; Mohammed et al. 2016). The need for synthetic insecticide application to control pest flies can be reduced by employing integrated pest management (IPM) approaches, using biological, cultural, physical/mechanical, and chemical management tools. This IPM approach is one of the effective methods for controlling the incidence of flies in livestock production (Cook 2020; Nisar et al. 2021; Moreki et al. 2022).

In recent years, researchers have grown interest in harnessing the potential of locally available botanicals or phytochemicals to combat insect pests (Leesombun et al. 2022; Divekar 2023). Phytochemical compounds extracted from botanical plants such as *Melinis minutiflora* grass, moringa (*Moringa oleifera*), black pepper (*Piper nigrum*), soybean (*Glycine max*), chaste tree seeds (*Vitex agnus castus*), oregano (*Plectranthus amboinicus*), and garlic (*Allium sativum*) have been investigated for their effectiveness in controlling dipteran insect populations, including flies and mosquitoes (Showler 2017; Cook 2020; Leesombun et al. 2022). Exploiting defense mechanism of these plants against pests is one possibly cost-effective and sustainable fly control approach that can complement the existing IPM approach. Garlic supplementation has been a topic of interest in livestock production for its potential to control pests (El-Saber Batiha et al. 2020; Ding et al. 2023; Abd El-Ghany 2024), improve animal health (Banerjee and Maulik 2002; Shokrollahi et al. 2016) and growth performance (Ikyume et al. 2017). Garlic's repellent and biocidal properties makes it a potential sustainable biopesticide.

Selecting fly-resistant traits in cattle breeding programs, particularly through genetic predisposition and dietary interventions such as garlic supplementation could offer a sustainable solution to mitigate the impacts of fly infestation, safeguarding both animal welfare and food and nutrition security in vulnerable regions. This dual strategy could significantly reduce reliance on synthetic pesticides, lower production costs, and minimise environmental impact. Literature indicates that fly resistance is a heritable trait, with heritability

estimates around 0.25 (Basiel et al. 2021), suggesting that selective breeding can effectively enhance this trait in cattle populations. Cattle producers can identify and select individuals that exhibit natural resistance to flies, resulting in herds that are less affected by pest flies over time. Integrating garlic into cattle diets offers additional benefits. Garlic possesses natural pesticidal properties and may enhance the expression of genes related to immunity and stress tolerance through nutrigenomic interactions (Charron et al. 2016). This dietary approach can complement genetic resistance strategies, potentially leading to cattle that not only resist fly infestations but also exhibit improved feed efficiency (FE) and growth. By focusing on the interactions between diet and genetic predispositions, researchers can develop targeted strategies that maximise the synergistic effects of both genetic resistance and dietary enhancements. The current scoping review explores the intricate mechanisms underpinning garlic supplementation in livestock production, specifically focusing on its efficacy in repelling ectoparasites, nutrigenomic effects on gene expression, modulating immune responses, and impacting metabolic pathways. Identifying key gene-diet interactions could lead to the development of cattle breeds that are not only naturally resistant to flies but also more resilient in various environmental conditions. By doing so, the review sets the stage for future research aimed at refining these strategies and validating their effectiveness in diverse livestock production systems.

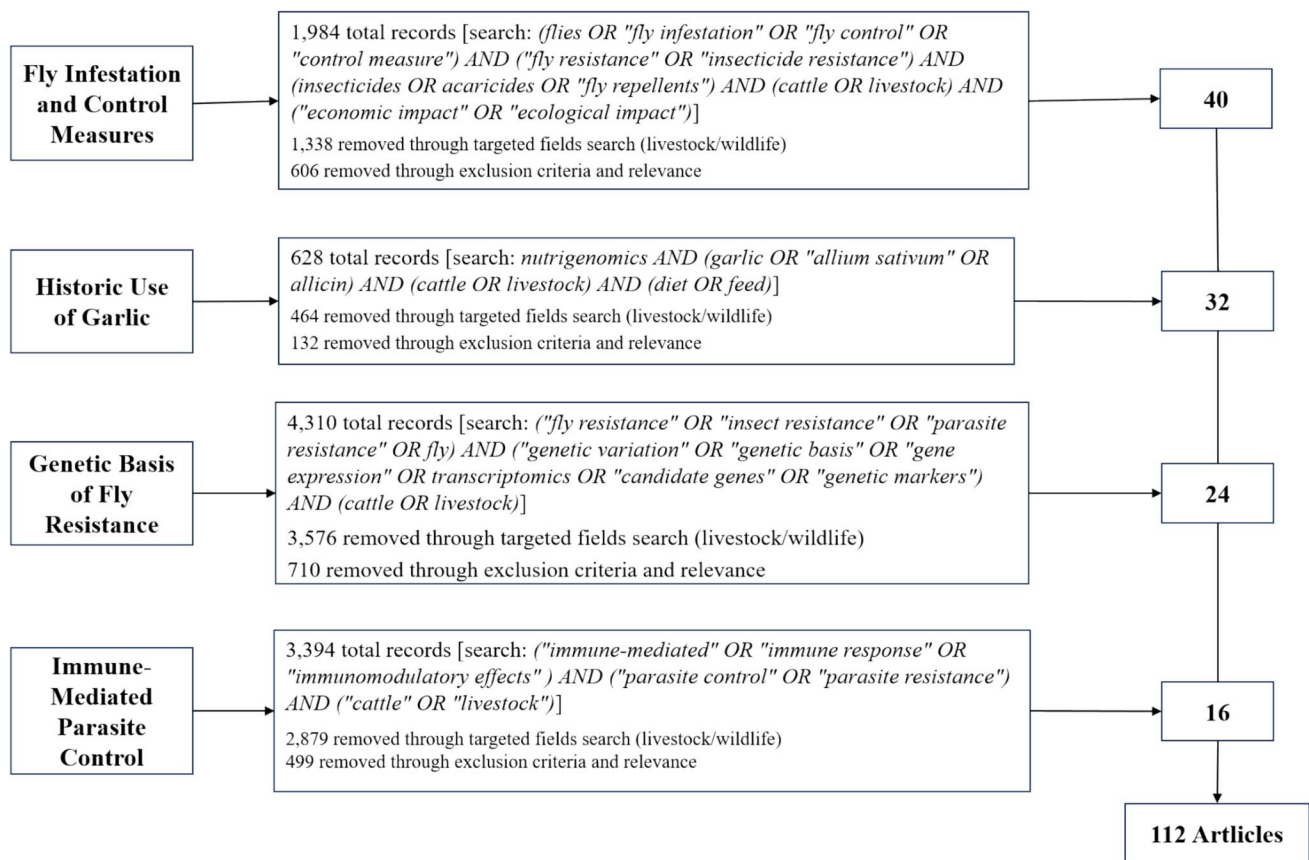
## Review methodology

A scoping search of peer-reviewed literature across multiple academic and grey literature databases, including PubMed, Scopus, Web of Science, ScienceDirect, SpringerLink, Google Scholar, Google, and Core, with a particular focus on fly abundance in cattle, control measures, and the impact of garlic supplementation, focusing on nutrigenomics and the genetic basis of fly resistance in cattle. This was accomplished using broad Boolean search strings, as indicated in Fig. 1. Where necessary, ResearchGate was used to access pre-print articles. Table 1 provides more information about the inclusion and exclusion criteria of the reviewed literature.

## Fly infestation in cattle and control measures

### Key filth flies, distributions, and their implications in Cattle Production

The global population of known fly species is estimated to be approximately 160,000, and they are characterized by



**Fig. 1** Scoping search strategy and literature selection

**Table 1** Inclusion and exclusion criteria of the reviewed literature

Criteria	Inclusion	Exclusion
Focus	Studies or reports on themes on fly abundance in cattle, control measures, and the impact of garlic supplementation, focusing on nutrigenomics and the genetic basis of fly resistance in cattle	
Search	Restricted to peer-reviewed papers from established journals and papers/ reports published within an organization (ex. pre-print, etc.)	Non-peer-reviewed; non-reputable organization-related reports and predatory journals
Language	English	Non-English
Publication year	Eligible paper from all years	None
Context	Globally, with a specific interest in South Africa for certain sections	None
Fly infestation	Livestock (ruminants, particularly cattle) and wildlife	Plants and fruits

having only two wings and belonging to the order Diptera, which translates to "two wings" in Latin. In the Afrotropical Region, a total of 108 families of Diptera are present, accounting for approximately 38% of the 130 extant families that exist worldwide (Kirk-Spriggs and Marshall 2017). The list of the top ten most well-represented families in the region, each with over 500 described species, includes Asilidae (1,685), Bombyliidae (1,384), Chironomidae (604), Culicidae (780), Dolichopodidae (770), Limoniidae and

Tipulidae (approximately 1,045), Muscidae (1,035), Syrphidae (about 600), Tabanidae (around 800), Tachinidae (1,126), and Tephritidae (approximately 1,000). Among the vast array of species, only a select few are classified as "filth flies" because of their close affiliation with animal excrement, decomposing food, or carrion (Machtinger et al. 2021), hence the focus of the present review.

Some of these flies, such as the house fly, stable fly, face fly (*Musca autumnalis* De Geer), horn fly (*Haematobia*

*irritans*), and little or lesser house fly (*Fannia canicularis*) pose significant concerns for animal production. These filth flies are often closely associated with animals and their excreta, which they preferentially or completely utilise for developing larvae. Adult flies of these species also obtain their nourishment from animals, such as blood, exudates, or other bodily secretions (Machtinger et al. 2021). These flies are not only a nuisance but carriers of disease-causing organisms that can affect both humans and livestock (Geden et al. 2021; Nisar et al. 2021). Furthermore, filth flies play a critical role in the transmission of pathogens such as fungi, bacteria, and viruses that can cause illness (Machtinger et al. 2021). The influence of fly species' distribution is intricate and remains an enigma despite extensive scientific investigation. The precise causes that restrain fly species from expanding their range are yet to be established. Considering that the distribution of their hosts does not confine most fly species, it is deducible that climate serves as the predominant inhibitor for the growth of fly species ranges (Goulson et al. 2005; Gilles et al. 2008).

### Economic and ecological impact of fly infestation in cattle production

The parasitic relationship that exists between cattle and flies leads to significant financial losses experienced in beef operations (Ling et al. 2020; Warner et al. 2022). For instance, flies decrease feed intake and feed conversion efficacy (FCE) and increase energy demand (Mohammed et al. 2016). This subsequently reduces weight gain and milk production in cattle (Durunna and Lardner 2021; Makhahlela et al. 2022). These effects collectively diminish the profitability of cattle production, with estimates suggesting that the economic threshold of stable fly infestation in cattle is approximately 15 flies per animal (Rochon et al. 2021). However, horn flies generally require intervention when their fly count reach around 200 flies per animal (Brewer et al. 2021). Additionally, the financial losses extend beyond direct production impacts. For instance, the horn fly, which feeds between 24–38 times a day and draws 11–21 mg of blood per feeding (Parra et al. 2013), not only present itself as an annoyance and irritant to the host (Mohammed et al. 2016), but also leads to increased costs for fly control measures and veterinary care to manage the health issues caused by these pests.

Ecologically, fly infestations compromise animal welfare and pose health risks to both humans and animals. Flies serve as vectors for the transmission of contagious diseases and contribute to the prevalence of conditions such as myiasis and mastitis in animals (Mohammed et al. 2016; Makhahlela et al. 2022). Stable flies have been implicated in the transmission of lumpy skin disease (Rochon et al. 2021). The overall economic losses due to reduced productivity and increased management costs, coupled with the

ecological impacts on animal welfare and health, can be substantial, further exacerbating the economic burden on cattle producers.

### Conventional and contemporary fly control methods and their limitations

Over a century of research has led to the development and refinement of numerous methodologies to control pest flies, particularly to protect cattle and minimise production losses, employing a variety of chemical, biological, physical, and cultural methods (Cook 2020). Such methods include back rubbers, spray-on, pour-on, insecticide-impregnated ear tags, larvicide oral treatment, biological control, and vacuum fly traps have also been used as methods to control livestock flies (Kienitz et al. 2018; Brewer et al. 2021). However, insecticides act as a temporary management tool, since reliance on their use and long-term efficacy has proven inadequate (Ling et al. 2020). Hence, effectively controlling pests is necessary to increase profitability through improved animal health and performance (Durunna and Lardner 2021).

Botanical repellents sticky traps are frequently used by organic dairy farmers to control fly infestations; however, these methods often have limited effectiveness (Sorge et al. 2015). This limitation is partly due to the rapid reproduction and spread of flies, for instance, horn flies, which can quickly migrate across neighbouring pastures. The productivity of a herd is determined by the difference between the speed at which traps can eradicate and eliminate flies from cattle and the rate at which they naturally emerge in the pastures (Kaufman et al. 2005). A proficient IPM program for house flies and stable flies on dairy farms involves regular removal of bedding, judicious choice of bedding material, utilization of less-toxic insecticides, release and preservation of biological control agents, and deployment of physical controls like traps (Geden and Hogsette 1994; Kaufman et al. 2005). There is currently limited information available on the use of botanical pesticides in conjunction with integrated pest management (IPM) to combat pest flies in cattle production. Therefore, it is crucial to increase awareness of the potential benefits of incorporating botanical pesticides into IPM programs for fly control in cattle production. By promoting the integration of botanical pesticides into IPM programs and raising awareness of their potential, livestock producers can effectively manage fly populations while minimizing the negative environmental and health impacts associated with the use of synthetic pesticides.

### Fly resistance to existing conventional insecticides

Over 100 years ago, Melander (1914) documented the first case of insecticide resistance, specifically in scale insects that displayed resistance to an inorganic insecticide.

Eleven more cases of resistance to inorganic insecticides were reported thereafter. Insecticide resistance occurs when an insect population becomes less sensitive to an insecticide, causing the insecticide to fail to control the insects despite proper product storage, application, and normal environmental conditions ('Insecticide Resistance Action Committee, IRAC'; Siddiqui et al. 2023). The use of organic insecticides, such as Dichlorodiphenyltrichloroethane (DDT) was adopted to mitigate the issue of insecticide resistance (Carvalho 2006). However, housefly resistance to DDT was documented by the year 1947. The emergence of resistance to insecticides has been a recurring phenomenon with the introduction of each new class of insecticides, with the timeframe ranging from 2 to 20 years. Insecticide resistance is most likely attributed to the overapplication of synthetic chemicals (Siddiqui et al. 2023). Insects develop insecticide resistance through a variety of mechanisms, the most important of which are behavioural resistance, fitness cost, reduced penetration, target resistance, and metabolic resistance (Siddiqui et al. 2023).

Insects may reduce their exposure to pesticides by changing their behaviours, such as avoiding the toxin or ceasing feeding upon encountering certain insecticides (Khan et al. 2020; Siddiqui et al. 2023; Gul et al. 2023). Resistant insects may absorb the toxin more slowly or develop outer cuticles that slow the penetration of the insecticide (IRAC). The specific binding site of an insecticide in the insect may be genetically modified to prevent the insecticide from binding, rendering the target site incompatible (Khan et al. 2020). Resistant insects may detoxify or destroy the toxin faster or rapidly eliminate it from their bodies. They may possess higher levels or more efficient forms of enzymes that break down insecticides, and these enzyme systems may have broad activity (IRAC). These mechanisms can occur independently or in combination, making effective control of insect populations challenging.

### The use of odour-based fly repellents

Flies use a combination of odour and visual cues for host location, with odour playing a significant role in attracting them from a distance and the latter become crucial for the landing (Borst and Heisenberg 1982). Through exploitation of this host-seeking behaviour, researchers have created colourful traps and targets, incorporating host attractants, to combat the issue of fly abundance. As part of the IPM program, studies have explored the use of odour-repellents from non-preferred hosts such as zebra and waterbuck skins as alternative approach to control pest fly abundance in livestock (Saini et al. 2017; Olaide et al. 2019; Ogolla et al. 2023). Mireji et al. (2022) recently reviewed odour-based control strategies for tsetse flies in Africa, concluding that

artificial bait technologies leveraging long-range olfactory responses to host cues and synthetic blends have effectively reduced certain tsetse fly populations using repellent-odour-based "push" tactics.

Garlic and its bioactive compounds, particularly allicin (diallyl-thiosulfinate), have been shown to possess insect-repellent properties, making them a promising botanical solution for managing pest infestations in livestock (Mohammed et al. 2016; Plata-Rueda et al. 2017; Hagg et al. 2024). The repellent properties of garlic are linked to its sulfur-containing compounds, such as allicin, ajoene, and allyl mercaptan, which are released through the skin and breath after ingestion (Rahman 2007; Zeng et al. 2017). These odorous compounds disrupt fly host-seeking behaviour, potentially reducing fly burden on cattle.

Although several studies have reported garlic's effectiveness in repelling flies, inconsistencies in fly reduction percentages have been observed. Mohammed et al. (2016) and Durunna and Lardner (2021) found a notable reductions in fly abundance and defensive behaviours in cattle treated with a garlic-based pour-on and those supplemented with garlic, respectively. However, Durunna and Lardner (2021) reported inconsistencies in their second-year findings, where garlic supplementation did not significantly reduce fly numbers. These discrepancies may stem from genetic differences in cattle, environmental conditions, variations in garlic processing methods and inconsistencies in the bioactive sulfur compounds responsible for fly repellence. Factors such as dosage, delivery methods (e.g., feed inclusion vs. topical application), and interactions with other dietary components may also influence garlic's effectiveness. Future research should focus on quantifying these compounds and optimising supplementation protocols for consistency.

### Historic use of garlic in livestock production

Garlic has been used for centuries across various civilisations for its medicinal and performance-enhancing properties (Rivlin 2001; Rahman 2007). In livestock production, garlic and its bioactive compound, allicin, have been explored for their potential to improve animal health, performance, and product quality. Studies have demonstrated that garlic supplementation in animal diets enhances meat quality, growth performance, and immune response. For example, garlic powder improved meat marbling, firmness in pigs (Chen et al. 2008) and enhanced rumen fermentation and health status in lambs infected with gastrointestinal nematodes (Zhong et al. 2019). Given its reported bioactivity, further in vivo trials are essential to validate garlic's efficacy in controlling flies and improving livestock production (Ding et al. 2023).



## Chemical profiles of garlic and its by-products

Garlic (Fig. 2) contains a diverse array of bioactive compounds with potential benefits in livestock production (Bose et al. 2014). Key sulfur-based compounds such as alliin, diallyl sulfide (DAS), diallyl disulfide (DADS), and diallyl trisulfide (DATS) are responsible for its insecticidal, antimicrobial, and immune-modulatory properties (El-Saber Batiha et al. 2020; Hardiansyah et al. 2020). Alliin, (Fig. 3) which is enzymatically produced by alliinase from alliin when garlic is crushed (Rahman 2007; Chen et al. 2021), is highly unstable and rapidly degrades into more stable sulfur-containing metabolites such as DAS, DADS, DATS, and/or sulfur dioxide (Melguizo-Rodríguez et al. 2022). These stable sulfur compounds contribute to the overall bioactivity and therapeutic effects of garlic.

Despite the instability of alliin, these compounds retain their efficacy in promoting health and combating diseases (Reinhart et al. 2009; Hardiansyah et al. 2020), owing to the presence of these stable sulfur compounds, although their relative proportions may change over time. For instance, garlic essential oil has been found to have insecticidal activity against various pests, including the mealworm beetle and grain moth (Zhao et al. 2013; Plata-Rueda et al. 2017). It is crucial to understand the chemical profiles and stability of sulfur-based compounds in garlic products to optimize their

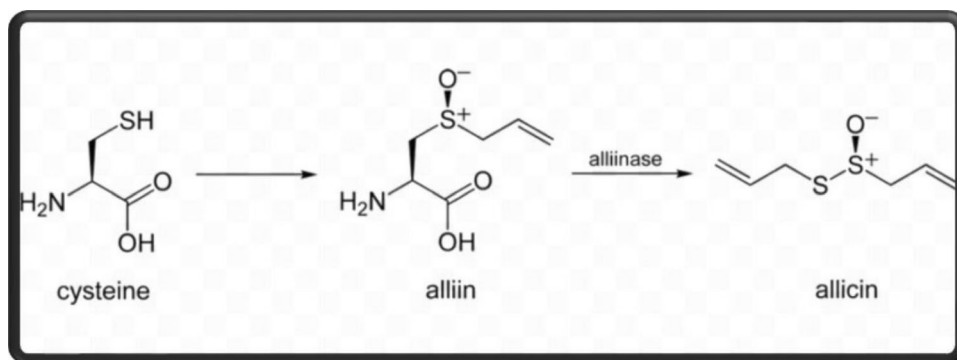
therapeutic potential and ensure consistent quality in both pesticidal, medicinal and culinary applications.

## Garlic as a feed additive in cattle

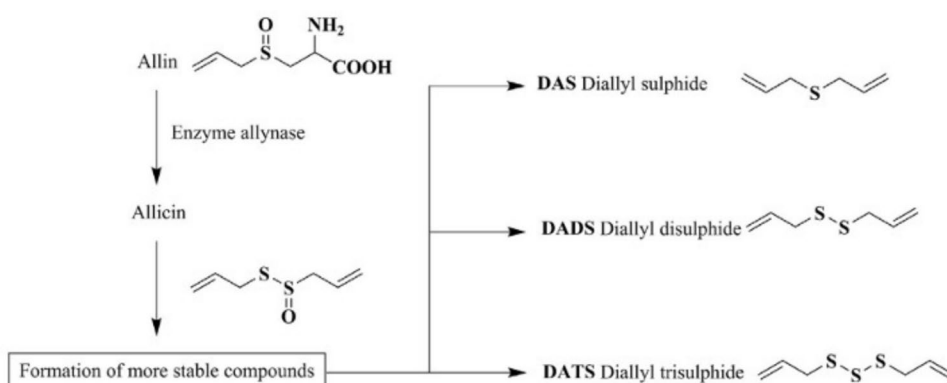
Garlic has been widely explored as a feed supplement (Table 2) due to its potential to enhance animal health and productivity (Durunna and Lardner 2021). Various forms of garlic, including dried flakes, essential oils, and powders, have been studied for their effects on rumen fermentation, nutrient utilisation, and immune function (Ikyume et al. 2017; Saastamoinen et al. 2019; Durunna and Lardner 2021). These garlic products can be administered to animals via feed inclusion, mineral and salt supplement. Martin and Chaudhry (2024) used a meta-analysis approach to evaluate the impact of various garlic-based feed additives on ruminal fermentability and ruminant performance. Their findings suggested that garlic supplementation positively influences volatile fatty acid production, microbial activity, and overall feed efficiency, further supporting its potential as a functional feed ingredient.

In cattle, garlic supplementation has shown promising results in improving feed conversion efficiency (FCE), weight gain, and fly repellence (Mohammed et al. 2016; Rossi et al. 2018; Durunna and Lardner 2021). However, the precise metabolic pathways through which garlic exerts its

**Fig. 2** Chemical structure of garlic. Adopted from Abd El-Ghany (2024)



**Fig. 3** Alliin transformation to more stable compounds. Adapted from Morales-González et al. (2019)



**Table 2** Studies exploring the use of garlic in ruminant livestock

References	Species studies	Inclusion level	Measurement	Effect
(Mohammed et al. 2016)	Cattle	500 g chopped garlic clove / 200 mL paraffin oil	Flies' attack rate and animal's defensive behaviour	Reduced fly count
(Hasan et al. 2015)	Goat	25 mL and 50 mL of 10% water solution of garlic	Egg per gram (EPG) count for gastrointestinal parasites, live weight, and hematological parameters	Decreased EPG count, increased weight gain, significant changes in the hematological parameters
(Zhong et al. 2019)	Sheep	50 g/kg garlic powder	Growth performance, rumen fermentation, and the health status of lambs	Increased lambs' average daily gain, digestibility of dry matter, and crude protein
(Rabee et al. 2025)	Sheep	2% of garlic powder	Growth performance, blood metabolites, immunity, rumen fermentation, and bacteria community in lambs	Improved the performance and immune status of growing lambs
(Xu et al. 2024)	Sheep	80 g/kg dry matter garlic skin	Growth performance, rumen and fecal microbiota, serum and urine metabolism, and transcriptomics of rumen epithelial cells in fattening sheep	Improved the energy metabolism and immune function of fattening sheep

fly-repellent effects remain unclear, requiring further research to determine optimal inclusion levels, delivery methods, and long-term benefits in cattle production. To effectively implement garlic supplementation for fly control in cattle, it is essential to establish standardised protocols regarding dosage, formulation, and delivery methods. Optimal dosages should be determined to ensure effective fly repellence while maintaining feed palatability and avoiding potential negative effects on animal health. Previous studies suggest that inclusion levels ranging from 0.2% to 8% of the diet can improve growth performance and immune function as shown in Table 2. However, fly repellence-specific dosages require further evaluation. Addressing these factors would provide necessary information to cattle farmers for practical application and provide a viable alternative to synthetic pesticides.

### Garlic implications on metabolic pathways

Once ingested, allicin undergoes rapid metabolism in the gastrointestinal tract, converting into stable sulfur metabolites such as allyl mercaptan and S-allyl mercapto cysteine (SAMC), which may modulate immune and metabolic functions (Rahman 2007; Ansary et al. 2020). Some studies suggest that garlic-derived metabolites can influence red blood cell permeability, immune responses, and antioxidant activity (Miron et al. 2000; Rao et al. 2015; Miekus et al. 2020), potentially contributing to fly resistance in cattle. However, given that allicin is short-lived and rapidly transforms into more stable compounds such as DAS, DADS and DATS (Rao et al. 2015; Miekus et al. 2020), future studies should focus on nutrigenomic evaluations to determine how garlic supplementation regulates immune-related and insect-repellent pathways in cattle, which could provide valuable insights for sustainable pest control strategies.

### Potential immune-mediated fly control mechanisms of garlic in cattle

Flies have evolved sophisticated mechanisms to evade and suppress the host's immune system, ensuring their survival and completion of their life cycle (Vilcinskas 2013). These mechanisms include molecular mimicry, immune suppression by regulatory T cells, antigenic camouflage, and modulation of host immunological factors (Caljon et al. 2006; Bezie 2014). For instance, tsetse fly saliva contains Gloss2, an immunoregulatory peptide that inhibits the secretion of key pro-inflammatory cytokines such as TNF, IFN- $\gamma$ , and IL-6, impairing the host's ability to mount an effective immune response (Lackie 1988; Caljon et al. 2006). This suppression allows flies to evade early detection and elimination, thereby enhancing their chances of survival and increasing the disease transmission.

The primary immune response to fly bites occurs at the skin, involving the activation of immune cells such as macrophages, dendritic cells, monocytes, and lymphocytes, as well as the production of antibodies and cellular immune responses (Gomes and Oliveira 2012; Abdeladhim et al. 2014). Rapid immune activation at the bite site is often associated with a higher level of resistance to flies (Naessens et al. 2003). However, flies' ability to suppress immune responses compromises this defense mechanism. Given this, strategies that enhance cattle's natural immune response could provide a sustainable means of fly control.

Garlic has been identified as a natural immunomodulator with potential in improving fly resistance in cattle (Durunna and Lardner 2021). Although there is limited literature specifically addressing garlic's influence on immune responses towards fly repellence, its effects on enhancing immune function suggest a promising role (Arreola et al. 2015). Studies have shown that garlic compounds, particularly S-allyl cysteine, enhance antioxidant enzyme activity and stimulates the immune response of ruminants by increasing the levels of immunoglobulins, such as immunoglobulin A, G, and immunoglobulin M (Kekana et al. 2020; Redoy et al. 2020; Kewan et al. 2021). Furthermore, garlic supplementation has been linked to increases activation of immune cell types, including macrophages, lymphocytes, natural killer cells, dendritic cells, and eosinophils, through mechanisms such as modulation of cytokine secretion, immunoglobulin production, phagocytosis, macrophage activation, cellular co-receptor expression, class switching, lymphocyte expression, and histamine release (Mahima et al. 2012; Arreola et al. 2015; Melguizo-Rodríguez et al. 2022). These immunomodulatory effects suggest that garlic could enhance cattle's ability to mount a stronger immune response against fly bites, potentially reducing fly burden and mitigating disease transmission. Moreover, diet-driven immune responses have been shown to influence gene expression, affecting an animal's resilience to environmental stressors, including parasite infestations (Le Coz et al. 2021; Brouklogiannis et al. 2023). While research on garlic's direct impact on immune-mediated fly resistance in cattle is still limited, its effects on immune function and inflammation modulation warrant further investigation.

## Genetic basis of fly resistance in cattle

Fly resistance in livestock, particularly in cattle, can be influenced by various factors, including genetic traits and gene expression. Studies have shown that some cattle are naturally resistant to flies, and both nutrition and genetic factors influence this resistance. For instance, genetic analyses have demonstrated significant genetic variation in fly resistance traits in cattle, with heritability estimates

ranging from 10 to 80% (Brown et al. 1992; Basiel et al. 2021). This resistance can be observed as the difference in the number of flies on some animals compared to their herd-mates and is influenced by factors such as breed, size of the animals, and individual differences (Brown et al. 1992). There are differences among breeds of cattle in resistance to parasites, with Zebu (*Bos indicus*) cattle breeds being most adaptable and acquiring fly resistance relatively faster than the European (*Bos taurus*) and some African indigenous (*Sanga*) cattle breeds (Mattioli et al. 2000; Steelman et al. 2003; Mwai et al. 2015).

The *Bos indicus* breeds of cattle, domesticated in harsh environmental conditions, are believed to have developed superior natural resistance to external stressors and parasites (Machado et al. 2010). Studies have shown that Zebu cattle exhibit better resistance to various pests and diseases, including trypanosomiasis transmitted by the tsetse fly (Mwai et al. 2015; Pal and Chakravarty 2020). Selective breeding for fly resistance has been proposed as a sustainable management method to control fly infestation (Steelman et al. 1993; Oyarzún et al. 2008). This suggests that the genetic foundation for fly resistance in cattle, particularly *Bos indicus* breeds, can be leveraged to enhance their ability to withstand fly infestations and other environmental stressors. Individual variation within and across breeds of beef cattle in resistance to horn flies has also been observed, with some cattle consistently hosting fewer horn flies, indicating a genetic basis for fly resistance (Steelman et al. 1993; Guglielmone et al. 2000). Identifying the mechanisms by which resistant cattle prevent high fly infestation is an essential step for the development of predictive markers for resistance. Although some work on the heritability and dynamics of host resistance, and the factors that can influence it has been undertaken, the mechanisms by which the resistant host prevents the pest fly infestation on an individual remain to be elucidated.

Gene expression, the essential biological process through which information encoded within a gene is transcribed into functional gene products, including proteins and non-coding RNAs, plays a crucial role in livestock production. The expression of genes in livestock is influenced by various factors, including genetic variation, environmental conditions including nutrients, bioactive compounds, and management practices (Berry et al. 2011; Mierziak et al. 2021). For example, certain genes may be expressed more or less strongly in response to changes in diet, temperature. Weyrich et al. (2019) investigated the transmission of DNA methylation changes to male offspring following paternal exposure to either a diet with reduced protein content or an increase in temperature. Through selective breeding, gene expression can be modified to enhance desirable traits in livestock (Kiplagat et al. 2012). Equally important, phytochemical diets can also stimulate expression of genes (Mierziak et al. 2021) involved in fly resistance.



## Potential nutrigenomic effects of garlic supplementation on fly repellence in cattle

In molecular animal nutrition, a subset of the broader field of nutrigenomics addresses the genetic basis of response to diet and, in parallel, the variations in dietary responsiveness among animals that are assignable to genotype (German et al. 2011). Utilizing different “omics” techniques from a variety of disciplines, (such as genomics, epigenomics, transcriptomics, proteomics, and metabolomics), nutrigenomics in practice examines animal cellular and molecular responses to different dietary nutrients, revealing the global influence of nutrients and phytochemicals on animal genomes, methylomes/epigenomes, transcriptomes, proteomes, and metabolomes, respectively (García-Cañas et al. 2010; Hasan et al. 2019; Haq et al. 2022). Transcriptomics has been instrumental in identifying genes associated with feed efficiency and metabolism in ruminants (Zhang et al. 2019; Lam et al. 2020; Lindholm-Perry et al. 2022). For instance, RNA-Seq technology has been used to identify functional candidate single nucleotide polymorphisms (SNPs) within genes associated with feed efficiency and metabolic pathways in cattle breeds (Higgins et al. 2019; Lam et al. 2020).

The influence of dietary bioactive compounds on gene expression regulation is significant in nutrigenetics and nutrigenomics. Dietary bioactive compounds, such as polyphenols, vitamins, flavonoids, carotenoids, glucosinolates, isothiocyanates, terpenes, fatty acids, allicin, DAS, DADS, DATS, S-allylcystine, and S-allylmercaptocysteine, transfer information from the external environment and influence gene expression (Takemura et al. 2013; Sheoran et al. 2017). Garlic contains most of these bioactive compounds, which can influence the expression of genes related to immune function and metabolism. Although there is lack of nutrigenomic evaluation of garlic in cattle, studies done on non-ruminants suggest that garlic can upregulate genes related to immunity, apoptosis, and xenobiotic metabolism (Charron et al. 2015; Sheoran et al. 2017). For instance, Sheoran et al. (2017) reported that garlic powder significantly enhanced the relative mRNA expression of toll-like receptor cell markers, indicating its potential to stimulate the immune responses in livestock. Recent studies, (Xu et al. 2024; Rabee et al. 2025) found that garlic supplementation influences gut microbiota composition and metabolic pathways, potentially affecting immune regulation in ruminants. Additionally, garlic extracts activate the nuclear factor erythroid-2 related factor 2-antioxidant response element pathway, enhancing expression of protective enzymes such as heme oxygenase-1 and glutamate-cysteine ligase modifier subunit (Mierziak et al. 2021).

Despite these findings, there is a significant gap in the literature regarding studies that directly investigated garlic's effects on gene expression related to fly resistance in cattle.

While garlic's potential to influence immune function and metabolism is well documented (Melguizo-Rodríguez et al. 2022; Xu et al. 2024; Rabee et al. 2025), little is known about how these changes could affect fly behaviour, feeding patterns, or reproduction. This knowledge gap highlights the importance for further research to assess garlic's nutrigenomic role in fly repellence, specifically in cattle. Future studies should use RNA-seq to identify differentially expressed genes associated with immune response and oxidative stress, and skin barrier function that may contribute to fly repellence. Furthermore, epigenetic studies may uncover regulatory modifications (such as DNA methylation or histone modifications) that influence gene expression related to fly resistance. Additionally genome-wide association studies (GWAS) could help identify genetic markers linked to fly repellence and assess the impact of garlic supplementation on these traits. Understanding the genetic basis of fly resistance and leveraging nutrigenomic strategies may enhance resilience, reducing reliance on synthetic pesticides while improving productivity and animal welfare.

## Conclusions

Garlic supplementation in livestock production presents a promising natural strategy for fly repellence, enhanced animal health, and improved meat production. Additionally, selecting fly-resistant cattle and integrating garlic-based phytochemical diets could offer a sustainable and environmentally friendly approach to mitigating fly infestations. The synergistic effects of genetic resistance and dietary interventions hold significant potential for reducing reliance on synthetic pesticides, and subsequent improvement of cattle productivity and welfare. As such, further research is important to determine the precise nutrigenomic mechanisms through which garlic influences fly resistance. This integrated approach could enhance resilience in intensive production systems, contributing to more sustainable and profitable livestock farming.

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## Declarations

**Conflict of interest** The authors declare no conflict of interest.

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## References

- Abd El-Ghany WA (2024) Potential effects of garlic (*Allium sativum* L.) on the performance, immunity, gut health, anti-oxidant status, blood parameters, and intestinal microbiota of poultry: an updated comprehensive review. *Animals* 14:498. <https://doi.org/10.3390/ani14030498>
- Abdeladhim M, Kanhawi S, Valenzuela JG (2014) What's behind a sand fly bite? The profound effect of sand fly saliva on host hemostasis, inflammation and immunity. *Infect Genet Evol* 28:691–703. <https://doi.org/10.1016/j.meegid.2014.07.028>
- Ansary J, Forbes-Hernández TY, Gil E, Cianciosi D, Zhang J, Elexpuru-Zabaleta M, Simal-Gandara J, Giampieri F, Battino M (2020) Potential health benefit of garlic based on human intervention studies: a brief overview. *Antioxidants* 9(7):619
- Arreola R, Quintero-Fabián S, López-Roa R, Flores-Gutiérrez E, Reyes-Grajeda J, Carrera-Quintanar L, Ortuño-Sahagún D (2015) Immunomodulation and anti-inflammatory effects of garlic compounds. *J Immunol Res* 2015:1–13
- Banerjee SK, Maulik SK (2002) Effect of garlic on cardiovascular disorders: a review. *Nutr J* 1:1–14. <https://doi.org/10.1186/1475-2891-1-4>
- Basiel BL, Hardie LC, Heins BJ, Dechow CD (2021) Genetic parameters and genomic regions associated with horn fly resistance in organic Holstein cattle. *J Dairy Sci* 104:12724–12740. <https://doi.org/10.3168/jds.2021-20366>
- Berry DP, Bermingham ML, Good M, More SJ (2011) Genetics of animal health and disease in cattle. *Ir Vet J* 64:5. <https://doi.org/10.1186/2046-0481-64-5>
- Bezie M (2014) African trypanosomes: virulence factors, pathogenicity and host responses. *J Vet Adv* 4:732. <https://doi.org/10.5455/jva.20141129012406>
- Borst A, Heisenberg M (1982) Osmotropotaxis in *Drosophila melanogaster*. *J Comp Physiol* 147:479–484
- Bose S, Laha B, Banerjee S (2014) Quantification of allicin by high performance liquid chromatography-ultraviolet analysis with effect of post-ultrasonic sound and microwave radiation on fresh garlic cloves. *Pharmacogn Mag* 10:S288–S293. <https://doi.org/10.4103/0973-1296.133279>
- Brewer GJ, Boxler DJ, Domingues LD, Trout Fryxell RT, Holderman C, Loftin KM, Machtinger E, Smythe B, Talley JL, Watson W (2021) Horn fly (Diptera: Muscidae) - biology, management, and future research directions. *J Integr Pest Manag* 12:42. <https://doi.org/10.1093/jipm/pmab019>
- Brouklogiannis IP, Anagnostopoulos EC, Griela E, Paraskeuas VV, Mountzouris KC (2023) Dietary phytochemical inclusion level affects production performance and expression of ovarian cytoprotective genes in laying hens. *Poult Sci* 102:102508. <https://doi.org/10.1016/j.psj.2023.102508>
- Brown AH, Steelman CD, Johnson ZB, Rosenkrans CF, Brasuell TM (1992) Estimates of repeatability and heritability of horn fly resistance in beef cattle. *J Anim Sci* 70:1375–1381. <https://doi.org/10.2527/1992.7051375x>
- Caljon G, Van Den Abbeele J, Sternberg JM, Coosemans M, De Baetselier P, Magez S (2006) Tsetse fly saliva biases the immune response to Th2 and induces anti-vector antibodies that are a useful tool for exposure assessment. *Int J Parasitol* 36:1025–1035. <https://doi.org/10.1016/j.ijpara.2006.05.002>
- Carvalho FP (2006) Agriculture, pesticides, food security and food safety. *Environ Sci Policy* 9:685–692. <https://doi.org/10.1016/j.envsci.2006.08.002>
- Charron CS, Dawson HD, Albaugh GP, Solverson PM, Vinyard BT, Solano-Aguilar GI, Molokin A, Novotny JA (2015) A single meal containing raw, crushed garlic influences expression of immunity- and cancer-related genes in whole blood of humans. *J Nutr* 145:2448–2455. <https://doi.org/10.3945/jn.115.215392>
- Charron CS, Dawson HD, Novotny JA (2016) Garlic influences gene expression in vivo and in vitro. *J Nutr* 146:444S–449S. <https://doi.org/10.3945/jn.114.202481>
- Chen YJ, Kim IH, Cho JH, Yoo JS, Wang Q, Wang Y, Huang Y (2008) Evaluation of dietary L-carnitine or garlic powder on growth performance, dry matter and nitrogen digestibilities, blood profiles and meat quality in finishing pigs. *Anim Feed Sci Technol* 141:141–152. <https://doi.org/10.1016/j.anifeedsci.2007.05.025>
- Chen J, Wang F, Yin Y, Ma X (2021) The nutritional applications of garlic (*Allium sativum*) as natural feed additives in animals. *PeerJ* 9:e11934. <https://doi.org/10.7717/peerj.11934>
- Cook D (2020) A historical review of management options used against the stable fly (Diptera: Muscidae). *Insects* 11:313. <https://doi.org/10.3390/insects11050313>
- Del Rayo Camacho M, Sanchez B, Quiroz H, Contreras JL, Mata R (1991) Pinocembrine: a bioactive flavanone from *Teloxys graveolens*. *J Ethnopharmacol* 31:383–389. [https://doi.org/10.1016/0378-8741\(91\)90022-6](https://doi.org/10.1016/0378-8741(91)90022-6)
- Ding H, Ao C, Zhang X (2023) Potential use of garlic products in ruminant feeding: a review. *Anim Nutr* 14:343. <https://doi.org/10.1016/j.aninu.2023.04.011>
- Divekar, P. 2023. Botanical pesticides: an eco-friendly approach for management of insect pests. *Acta Sci Agric* (ISSN 2581–365X) 7
- Durunna O, Lardner H (2021) Impact of garlic-infused salt supplement on fly abundance, salt intake, and defensive behaviors in grazing beef cows. *Sustain Agric Res* 10:54. <https://doi.org/10.5539/sar.v10n1p54>
- El-Saber Batiha G, MagdyBeshbishy A, Wasef LG, Elewa YHA, Al-Sagoff AA, Abd El-Hack ME, Taha AE, Abd-Elhakim YM, Devkota HP (2020) Chemical constituents and pharmacological activities of garlic (*Allium Sativum* L.): a review. *Nutrients* 12:872. <https://doi.org/10.3390/nu12030872>
- Evert M, Van Hamburg H (2014) Monitoring of feedlot associated flies fly populations at Karan Beef feedlot. *Suid-Afrikaanse Tydskrif Vir Natuurwetenskap En Tegnol* 33:4102. <https://doi.org/10.4102/satnt.v33i1.1261>
- Evert MM (2014) The temporal distribution and relative abundance of stable flies (*Stomoxys calcitrans*) (Diptera: Muscidae) in a feedlot near Heidelberg, Gauteng, South Africa / Maria Magdalena Evert
- Foil LD, Hogsette JA (1994) Biology and control of tabanids, stable flies and horn flies. *Rev Sci Tech* 13:1125–1158. <https://doi.org/10.20506/rst.13.4.821>
- García-Cañas V, Simó C, León C, Cifuentes A (2010) Advances in Nutrigenomics research: novel and future analytical approaches to investigate the biological activity of natural compounds and food functions. *J Pharm Biomed Anal* 51:290–304. <https://doi.org/10.1016/j.jpba.2009.04.019>
- Geden CJ, Nayduch D, Scott JG, Burgess ER, Gerry AC, Kaufman PE, Thomson J, Pickens V, Machtinger ET (2021) House fly (Diptera: Muscidae): biology, pest status, current management prospects,

- and research needs. *J Integr Pest Manag* 12:39–40. <https://doi.org/10.1093/jipm/pmaa021>
- Geden CJ, Hogsette JA (1994) Research and extension needs for integrated pest management for arthropods of veterinary importance - lincoln.pdf. Proceeding a work. Lincoln, Nebraska, 1–328
- German JB, Zivkovic AM, Dallas DC, Smilowitz JT (2011) Nutrigenomics and personalized diets: what will they mean for food? *Annu Rev Food Sci Technol* 2:97–123. <https://doi.org/10.1146/annurev.food.102308.124147>
- Gilles J, David JF, Duvallet G, Tillard E (2008) Potential impacts of climate change on stable flies, investigated along an altitudinal gradient. *Med Vet Entomol* 22:74–81. <https://doi.org/10.1111/j.1365-2915.2008.00717.x>
- Gomes R, Oliveira F (2012) The immune response to sand fly salivary proteins and its influence on Leishmania immunity. *Front Immunol* 3:1–8. <https://doi.org/10.3389/fimmu.2012.00110>
- Goulson D, Derwent LC, Hanley ME, Dunn DW, Abolins SR (2005) Predicting calyptate fly populations from the weather, and probable consequences of climate change. *J Appl Ecol* 42:795–804. <https://doi.org/10.1111/j.1365-2664.2005.01078.x>
- Guglielmone AA, Curto E, Anziani OS, Mangold AJ (2000) Cattle breed-variation in infestation by the horn fly *Haematobia irritans*. *Med Vet Entomol* 14:272–276. <https://doi.org/10.1046/j.1365-2915.2000.00235.x>
- Gul H, Gadratagi BG, Güncan A, Tyagi S, Ullah F, Desneux N, Liu X (2023) Fitness costs of resistance to insecticides in insects. *Front Physiol* 14:1238111. <https://doi.org/10.3389/fphys.2023.1238111>
- Hagg FM, Erasmus LJ, Stoltz WH (2024) The potential effect of Garlicum GEM HCTM as a tick control agent in cattle. *J S Afr Vet Assoc*. 95:1–6. <https://doi.org/10.36303/JSAVA.560>
- Haq Z ul, Saleem A, Khan AA, Dar MA, Ganaie AM, Beigh YA, Hamadani H, Ahmad SM (2022) Nutrigenomics in livestock sector and its human-animal interface-a review. *Vet Anim Sci* 17:100262. <https://doi.org/10.1016/j.vas.2022.100262>
- Hardiansyah MY, Al Ridho AF, Nurhidayat (2020) The effect of garlic (*Allium sativum*) extract pesticides in repelling rice eating bird pests. *Indones J Agric Res* 3:145–152. <https://doi.org/10.32734/injar.v3i3.3947>
- Hasan MMI, Begum S, Islam S, Rahman MM, Belal SA, Hossain MA, Akanda MR, Pal NC, Howlader MMR (2015) Effects of garlic supplementation on parasitic infestation, live weight, and hematological parameters in Black Bengal goat. *J Adv Vet Anim Res* 2:326–331. <https://doi.org/10.5455/javar.2015.b102>
- Hasan MS, Feugang JM, Liao SF (2019) A nutrigenomics approach using RNA sequencing technology to study nutrient-gene interactions in agricultural animals. *Curr Dev Nutr* 3:1–12. <https://doi.org/10.1093/cdn/nzz082>
- Higgins MG, Kenny DA, Fitzsimons C, Blackshields G, Coyle S, McKenna C, McGee M, Morris DW, Waters SM (2019) The effect of breed and diet type on the global transcriptome of hepatic tissue in beef cattle divergent for feed efficiency. *BMC Genom* 20:1–11. <https://doi.org/10.1186/s12864-019-5906-8>
- Ikyume TT, Sowande OS, Dele PA, Yusuf AO, Monday S, Egunjobi OK, Fatoba O (2017) Effect of varying levels of garlic (*Allium sativum*) powder on growth, apparent nutrient digestibility, rumen ecology, blood profile and cost analysis of feeding West African Dwarf. *Mal J Anim Sci* 20:61–74
- Ilemobade AA (2009) Tsetse and trypanosomosis in Africa: the challenges, the opportunities. *Onderstepoort J Vet Res* 76:35–40
- Insecticide Resistance Action Committee (n.d.) IRAC
- Iqbal W, Faheem MM, Kaleem SM, Iqra A, Iram N, Rashda A (2014) Role of housefly (*Musca domestica*, Diptera; Muscidae) as a disease vector; a review. *J Entomol Zool Stud* 2:159–163
- Kaufman PE, Rutz DA, Frisch S (2005) Large sticky traps for capturing house flies and stable flies in dairy calf greenhouse facilities. *J Dairy Sci* 88:176–181. [https://doi.org/10.3168/jds.S0022-0302\(05\)72676-X](https://doi.org/10.3168/jds.S0022-0302(05)72676-X)
- Kekana TW, Nherera-Chokuda VF, Baloyi JJ, Muya CM (2020) Immunoglobulin G response and performance in Holstein calves supplemented with garlic powder and probiotics. *S Afr J Anim Sci* 50:263–270. <https://doi.org/10.4314/SAJAS.V50I2.9>
- Kewan KZ, Ali MM, Ahmed BM, El-Kolty SA, Nayel UA (2021) The effect of yeast (*Saccharomyces cerevisiae*), garlic (*Allium sativum*) and their combination as feed additives in finishing diets on the performance, ruminal fermentation, and immune status of lambs. *Egypt J Nutr Feed* 24:55–76. <https://doi.org/10.21608/ejnf.2021.170304>
- Khan S, Uddin MN, Rizwan M, Khan W, Farooq M, Shah AS, Subhan F, Aziz F, Rahman KU, Khan A, Ali S, Muhammad. (2020) Mechanism of insecticide resistance in insects/pests. *Polish J Environ Stud* 29:2023–2030. <https://doi.org/10.15244/pjoes/108513>
- Kienitz MJ, Heins BJ, Moon RD (2018) Evaluation of a commercial vacuum fly trap for controlling flies on organic dairy farms. *J Dairy Sci* 101:4667–4675. <https://doi.org/10.3168/jds.2017-13367>
- Kiplagat SK, Limo MK, Kosgey IS (2012) Genetic improvement of livestock for milk production, Ch. 4 Chaiyabutr N, ed IntechOpen, Rijeka
- Kirk-Spriggs AH, Marshall SA (2017) Manual of Afrotropical Diptera, volume 1: introductory chapters and keys to Diptera families. Pretoria
- Lackie AM (1988) Immune mechanisms in insects. *Parasitol Today* 4:98–105. [https://doi.org/10.1016/0169-4758\(88\)90035-X](https://doi.org/10.1016/0169-4758(88)90035-X)
- Lam S, Zeidan J, Miglior F, Suárez-Vega A, Gómez-Redondo I, Fonseca PAS, Guan LL, Waters S, Cánovas A (2020) Development and comparison of RNA-sequencing pipelines for more accurate SNP identification: practical example of functional SNP detection associated with feed efficiency in Nellore beef cattle. *BMC Genomics* 21:1–17. <https://doi.org/10.1186/s12864-020-07107-7>
- Le Coz J, Ilic S, Fibi-Smetana S, Schatzmayr G, Zaunschirm M, Grenier B (2021) Exploring with transcriptomic approaches the underlying mechanisms of an essential oil-based phytogetic in the small intestine and liver of pigs. *Front Vet Sci* 8:1–11. <https://doi.org/10.3389/fvets.2021.650732>
- Leesombun A, Kavallieratos G, Sungpradit S, Boukouvala M, Boonmasawai S, Weluwanarak T, Klinsrithong S, Ruangsittichai J, Ampawong S, Masmeatathip R, Changbunjong T (2022) Insecticidal activity of *Plectranthus amboinicus* essential oil against the stable fly *Stomoxys calcitrans* (Diptera: Muscidae) and the horse fly *Tabanus megalops* (Diptera: Tabanidae). *Insects* 13:255. <https://doi.org/10.3390/insects13030255>
- Lindholm-Perry AK, Meyer AM, Kern-Lunbery RJ, Cunningham-Hollinger HC, Funk TH, Keel BN (2022) genes involved in feed efficiency identified in a meta-analysis of rumen tissue from two populations of beef steers. *Animals* 12:1514. <https://doi.org/10.3390/ani12121514>
- Ling A, Krause T, Heins B, Hinkle N, Pringle D, Aggrey SE, Rekaya R (2020) Genetic study of horn fly abundance in beef cattle. *J Anim Sci* 98:16–17. <https://doi.org/10.1093/jas/skaa278.031>
- Machado MA, Luisa A, Azevedo S, Teodoro RL, Pires MA, Gabriela M, Peixoto C, De Freitas C, Prata CA, Furlong J, Vinicius M, Da Silva G, Guimarães SE, Ca Regitano L, Coutinho LL, Gasparin G, Verneque RS (2010) Genome wide scan for quantitative trait loci affecting tick resistance in cattle (*Bos taurus* × *Bos indicus*). *BMC Genom* 11:1–1
- Machtinger ET, Gerry AC, Murillo AC, Talley JL (2021) Filth fly impacts to animal production in the united states and associated research and extension needs. *J. Integr. Pest Manag.* 12:41. <https://doi.org/10.1093/jipm/pmb026>
- Mahima Rahal A, Deb R, Latheef SK, Abdul Samad H, Tiwari R, Verma AK, Kumar A, Dhama K (2012) Immunomodulatory and



- therapeutic potentials of herbal, traditional/indigenous and ethnoveterinary medicines. *Pakistan J Biol Sci PJBs* 15:754–774. <https://doi.org/10.3923/pjbs.2012.754.774>
- Makhahlela NB, Liebenberg D, Van Hamburg H, Taioe MO, Onyiche TG, Ramatla T, Thekisoe OMM (2022) Detection of pathogens of veterinary importance harboured by *Stomoxys calcitrans* in South African feedlots. *Sci African* 15:e01112. <https://doi.org/10.1016/j.sciaf.2022.e01112>
- Martin RSH, Chaudhry AS (2024) The effects of garlic as a feed additive on ruminal fermentability and ruminant performance: a meta-analysis. *J Agric Food Res* 18:101531. <https://doi.org/10.1016/j.jafr.2024.101531>
- Mattioli RC, Pandey VS, Murray M, Fitzpatrick JL (2000) Immunogenetic influences on tick resistance in African cattle with particular reference to trypanotolerant N'Dama (*Bos taurus*) and trypanosusceptible Gobra zebu (*Bos indicus*) cattle. *Acta Trop* 75:263–277. [https://doi.org/10.1016/S0001-706X\(00\)00063-2](https://doi.org/10.1016/S0001-706X(00)00063-2)
- Melander AL (1914) Can insects become resistant to sprays? I. *J Econ Entomol* 7:167
- Melguizo-Rodríguez L, García-Recio E, Ruiz C, De Luna-Bertos E, Illescas-Montes R, Costela-Ruiz VJ (2022) Biological properties and therapeutic applications of garlic and its components. *Food Funct* 13:2415–2426. <https://doi.org/10.1039/d1fo03180e>
- Miekus N, Marszałek K, Podlacha M, Iqbal A, Puchalski C, Swiergiel AH (2020) Health benefits of plant-derived sulfur compounds, glucosinolates, and organosulfur compounds. *Molecules* 25:3804. <https://doi.org/10.3390/molecules25173804>
- Mierziak J, Kostyn K, Boba A, Czemplik M, Kulma A, Wojtasik W (2021) Influence of the bioactive diet components on the gene expression regulation. *Nutrients* 13:1–33. <https://doi.org/10.3390/nu13113673>
- Mireji PO, Mangera CM, Bwana BK, Hassanali A (2022) Perspectives on odor-based control of tsetse flies in Africa. *Front Physiol* 13:1–7. <https://doi.org/10.3389/fphys.2022.831618>
- Miron T, Rabinkov A, Mirelman D, Wilchek M, Weiner L (2000) The mode of action of allicin: Its ready permeability through phospholipid membranes may contribute to its biological activity. *Biochim Biophys Acta - Biomembr* 1463:20–30. [https://doi.org/10.1016/S0005-2736\(99\)00174-1](https://doi.org/10.1016/S0005-2736(99)00174-1)
- Mohammed AN, Abdel Azeem NM, Abdel Latif GK (2016) Field trial using combined treatment of garlic and organic spray formula for fly's control and animal defensive behavior alleviation in cattle farm. *Asian J Anim Sci* 10:280–289. <https://doi.org/10.3923/ajas.2016.280.289>
- Morales-González JA, Madrigal-Bujaidar E, Sánchez-Gutiérrez M, Izquierdo-Vega JA, Del Carmen Valadez-Vega M, Álvarez-González I, Morales-González Á, Madrigal-Santillán E (2019) Garlic (*Allium sativum* L.): a brief review of its antigenotoxic effects. *Foods* 8:1–17. <https://doi.org/10.3390/foods8080343>
- Moreki JC, Tjinyeka K, Makore J, Tlotleng K, Moseki MI (2022) The impact of stable flies (*Stomoxys calcitrans* L) on small stock production in Bodibeng, Bothatogo and Sehithwa in the north west district, Botswana; a survey study. *Online J Anim Feed Res* 12:73–80. <https://doi.org/10.51227/ojafr.2022.10>
- Mwai O, Hanotte O, Kwon YJ, Cho S (2015) African indigenous cattle: Unique genetic resources in a rapidly changing world. *Asian-Australasian J Anim Sci* 28:911–921. <https://doi.org/10.5713/ajas.15.0002R>
- Naessens J, Mwangi DM, Buza J, Moloo SK (2003) Local skin reaction (chancre) induced following inoculation of metacyclic trypanosomes in cattle by tsetse flies is dependent on CD4 T lymphocytes. *Parasite Immunol* 25:413–419. <https://doi.org/10.1111/j.1365-3024.2003.00649.x>
- Nisar MS, Ismail MA, Ramzan H, Maqbool MM, Ahmed T, Ghramh HA, Khalofah A, Kmet J, Horvát M, Farooq S (2021) The impact of different plant extracts on biological parameters of housefly [*Musca domestica* (Diptera: Muscidae)]: implications for management. *Saudi J Biol Sci* 28:3880–3885. <https://doi.org/10.1016/j.sjbs.2021.03.070>
- Ogolla KO, Onyango T, Bwana BK, Otiende MY, Mang'era CM, Ochieng B, Omolo MO, Mugambi JM, Hassanali A, Omondi P, Mireji PO (2023) Bloodmeal host identities among sympatric *Glossina austeni* and *Glossina pallidipes* tsetse flies in Shimba Hills National Reserve, Kwale, Kenya. *Front. Trop. Dis.* 4:1–7. <https://doi.org/10.3389/fttd.2023.1145993>
- Olaide OY, Tchouassi DP, Yusuf AA, Pirk CWW, Masiga DK, Saini RK, Torto B (2019) Zebra skin odor repels the savannah tsetse fly, *Glossina pallidipes* (Diptera: Glossinidae). *PLoS Negl Trop Dis* 13:e0007460. <https://doi.org/10.1371/journal.pntd.0007460>
- Oyarzún MP, Quiroz A, Birkett MA (2008) Insecticide resistance in the horn fly: alternative control strategies. *Med Vet Entomol* 22:188–202. <https://doi.org/10.1111/j.1365-2915.2008.00733.x>
- Pal A, Chakravarty AK (2020) Disease resistance for different livestock species. *Genet Breed Dis Resist Livest* 25:271–296. <https://doi.org/10.1016/B978-0-12-816406-8.00019-X>
- Panei CJ, Larsen AE, Fuentealba NA, Metz GE, Echeverría MG, Galosi CM, Valera AR (2019) Study of horn flies as vectors of bovine leukemia virus. *Open Vet J* 9:33–37. <https://doi.org/10.4314/ovj.v9i1.6>
- Parra L, Rojas C, Catrileo A, Galdames R, Mutis A, Birkett MA, Quiroz A (2013) Differences in the fly-load of *Haematobia irritans* (Diptera: Muscidae) on cattle is modified by endophyte infection of pastures. *Electron J Biotechnol* 16:4
- Pavela R, Canale A, Mehlhorn H, Benelli G (2016) Application of ethnobotanical repellents and acaricides in prevention, control and management of livestock ticks: a review. *Res Vet Sci* 109:1–9. <https://doi.org/10.1016/j.rvsc.2016.09.001>
- Plata-Rueda A, Martínez LC, Dos Santos MH, Fernandes FL, Wilcken CF, Soares MA, Serrão JE, Zanuncio JC (2017) Insecticidal activity of garlic essential oil and their constituents against the mealworm beetle, *Tenebrio molitor* Linnaeus (Coleoptera: Tenebrionidae). *Sci Rep* 7:1–11. <https://doi.org/10.1038/srep46406>
- Rabee AE, El Shereef AA, Nassar MS, El-Rayes MAH, Mohammed RS, Bakr SA (2025) Effect of garlic powder supplementation on rumen microbiota and histology, and blood metabolites in Barki lambs. *BMC Vet Res* 21:116. <https://doi.org/10.1186/s12917-025-04521-5>
- Rahman MS (2007) Allicin and other functional active components in garlic: health benefits and bioavailability. *Int J Food Prop* 10:245–268. <https://doi.org/10.1080/10942910601113327>
- Rao P, Midde NM, Miller DD, Chauhan S, Kumar A, Kumar S (2015) Diallyl sulfide: potential use in novel therapeutic interventions in alcohol, drugs, and disease mediated cellular toxicity by targeting cytochrome P450 2E1 HHS Public Access
- Redoy MRA, Shuvo AAS, Cheng L, Al-Mamun M (2020) Effect of herbal supplementation on growth, immunity, rumen histology, serum antioxidants and meat quality of sheep. *Animal* 14:2433–2441. <https://doi.org/10.1017/S1751731120001196>
- Reinhart KM, Talati R, White CM, Coleman CI (2009) The impact of garlic on lipid parameters: a systematic review and meta-analysis. *Nutr Res Rev* 22:39–48. <https://doi.org/10.1017/S0954422409350003>
- Rivlin RS (2001) Historical perspective on the use of garlic. *J Nutr* 131:951S–954S. <https://doi.org/10.1093/jn/131.3.951S>
- Rochon K, Hogsette JA, Kaufman PE, Olafson PU, Swiger SL, Taylor DB (2021) Stable fly (Diptera: Muscidae) - biology, management, and research needs. *J Integr Pest Manag* 12:38. <https://doi.org/10.1093/jipm/pmab029>
- Rossi G, Schiavon S, Lomolino G, Cipolat-Gotet C, Simonetto A, Bitante G, Tagliapietra F (2018) Garlic (*Allium sativum* L.) fed to dairy cows does not modify the cheese-making properties of milk

- but affects the color, texture, and flavor of ripened cheese. *J Dairy Sci* 101:2005–2015. <https://doi.org/10.3168/jds.2017-13884>
- Saastamoinen M, Särkijärvi S, Hyypä S (2019) Garlic (*Allium sativum*) supplementation improves respiratory health but has increased risk of lower hematologic values in horses. *Animals* 9:38. <https://doi.org/10.3390/ani9010013>
- Saini RK, Orindi BO, Mbahin N, Andoke JA, Muasa PN, Mbuvi DM, Muya CM, Pickett JA, Borgemeister CW (2017) Protecting cows in small holder farms in East Africa from tsetse flies by mimicking the odor profile of a non-host bovid. *Physiol Ecol* 11(10):e0005977. <https://doi.org/10.1371/journal.pntd.0005977>
- Sheoran N, Kumar R, Kumar A, Batra K, Sihag S, Maan S, Maan NS (2017) Nutrigenomic evaluation of garlic (*Allium sativum*) and holy basil (*Ocimum sanctum*) leaf powder supplementation on growth performance and immune characteristics in broilers. *Vet. World* 10:121–129. <https://doi.org/10.14202/vetworld.2017.121-129>
- Shokrollahi B, Hesami SM, Baneh H (2016) The effect of garlic extract on growth, haematology and cell-mediated immune response of newborn goat kids. *J Agric Rural Dev Trop Subtrop* 117:225–232
- Showler AT (2017) Botanically based repellent and insecticidal effects against horn flies and stable flies (Diptera: Muscidae). *J Ofintegrated PestManagement* 8:1–11. <https://doi.org/10.1093/jipm/pmx010>
- Siddiqui JA, Fan R, Naz H, Bamisile BS, Hafeez M, Ghani MI, Wei Y, Xu Y, Chen X (2023) Insights into insecticide-resistance mechanisms in invasive species: challenges and control strategies. *Front Physiol* 13:1–18. <https://doi.org/10.3389/fphys.2022.1112278>
- Sorge US, Moon RD, Stromberg BE, Schroth SL, Michels L, Wolff LJ, Kelton DF, Heins BJ (2015) Parasites and parasite management practices of organic and conventional dairy herds in Minnesota. *J Dairy Sci* 98(5):3143–3151. <https://doi.org/10.3168/jds.2014-9031>
- Sprygin A, Pestova Y, Wallace DB, Tuppurainen E, Kononov AV (2019) Transmission of lumpy skin disease virus: a short review. *Virus Res* 269:197637. <https://doi.org/10.1016/j.virusres.2019.05.015>
- Steelman CD, Gbur EE, Tolley G, Brown AHJ (1993) Individual variation within breeds of beef cattle in resistance to horn fly (Diptera: Muscidae). *J Med Entomol* 30:414–420. <https://doi.org/10.1093/jmedent/30.2.414>
- Steelman CD, McNew RW, Simpson RB, Rorie RW, Phillips JM, Rosenkrans CF Jr (2003) Evaluation of alternative tactics for management of insecticide-resistant horn flies (Diptera: Muscidae). *J Econ Entomol* 96:892–901. <https://doi.org/10.1093/jee/96.3.892>
- Takemura S, Minamiyama Y, Kodai S, Shinkawa H, Tsukioka T, Okada S, Azuma H, Kubo S (2013) S-Allyl cysteine improves nonalcoholic fatty liver disease in type 2 diabetes Otsuka Long-Evans Tokushima Fatty rats via regulation of hepatic lipogenesis and glucose metabolism. *J Clin Biochem Nutr* 53:94–101. <https://doi.org/10.3164/jcbs.13-1>
- Taylor DB, Moon RD, Mark DR (2012) Economic impact of stable flies (Diptera: Muscidae) on dairy and beef cattle production. *J Med Entomol* 49:198–209. <https://doi.org/10.1603/ME10050>
- Vilcinskas A (2013) Evolutionary plasticity of insect immunity. *J Insect Physiol* 59:123–129. <https://doi.org/10.1016/j.jinsphys.2012.08.018>
- Warner A, Ling A, Krause T, Heins B, Hinkle N, Pringle D, Aggrey SE, Rekaya R (2022) Thrombin as a potential proxy to select for horn fly abundance in beef cattle. *Animals* 12:2982. <https://doi.org/10.3390/ani12212982>
- Weyrich A, Lenz D, Fickel J (2019) Environmental change-dependent inherited epigenetic response. *Genes (Basel)*. 10:4. <https://doi.org/10.3390/genes10010004>
- Xu Y, Yi M, Sun S, Wang L, Zhang Z, Ling Y, Cao H (2024) The regulatory mechanism of garlic skin improving the growth performance of fattening sheep through metabolism and immunity. *Front Vet Sci* 11:1409518. <https://doi.org/10.3389/fvets.2024.1409518>
- Zeng Y, Li Y, Yang J, Pu X, Du J, Yang X, Yang T, Yang S (2017) Therapeutic role of functional components in alliums for preventive chronic disease in human being. Evidence-based Complement. *Altern Med* 2017:9402849. <https://doi.org/10.1155/2017/9402849>
- Zhang D, Zhang X, Li F, Li C, La Y, Mo F, Li G, Zhang Y, Li X, Song Q, Zhao Y, Wang W (2019) Transcriptome analysis identifies candidate genes and pathways associated with feed efficiency in hu sheep. *Front Genet* 10:1–9. <https://doi.org/10.3389/fgene.2019.01183>
- Zhao NN, Zhang H, Zhang XC, Luan XB, Zhou C, Liu QZ, Shi WP, Liu ZL (2013) Evaluation of acute toxicity of essential oil of garlic (*Allium sativum*) and its selected major constituent compounds against overwintering *Cacopsylla chinensis* (Hemiptera: Psyllidae). *J Econ Entomol* 106:1349–1354. <https://doi.org/10.1603/EC12191>
- Zhong R, Xiang H, Cheng L, Zhao C, Wang F, Zhao X, Fang Y (2019) Effects of feeding garlic powder on growth performance, rumen fermentation, and the health status of lambs infected by gastrointestinal nematodes. *Animals* 9:1–10. <https://doi.org/10.3390/ani9030102>

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