



Commentary The Brain–Intestinal Mucosa–Appendix– Microbiome–Brain Loop

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Abstract: The brain and the gut are connected from early fetal life. The mother's exposure to microbial molecules is thought to exert in utero developmental effects on the fetus. These effects could importantly underpin the groundwork for subsequent pathophysiological mechanisms for achieving immunological tolerance and metabolic equilibrium post birth, events that continue through to 3-4 years of age. Furthermore, it is understood that the microbiome promotes cues that instruct the neonate's mucosal tissues and skin in the language of molecular and cellular biology. Post birth mucosal lymphoid tissue formation and maturation (most probably including the vermiform appendix) is microbiota-encouraged co-establishing the intestinal microbiome with a developing immune system. Intestinal mucosal tissue maturation loops the brain-gut-brain and is postulated to influence mood dispositions via shifts in the intestinal microbiome phyla. A plausible appreciation is that dysregulated pro-inflammatory signals from intestinal resident macrophages could breach the loop by providing adverse mood signals via vagus nerve afferents to the brain. In this commentary, we further suggest that the intestinal resident macrophages act as an upstream traffic controller of translocated microbes and metabolites in order to maintain local neuro-endocrine-immunological equilibrium. When macrophages are overwhelmed through intestinal microbiome and intestinal epithelial cell dysbiosis, pro-inflammatory signals are sustained, which may then lead to mood disorders. The administration of probiotics as an adjunctive medicine co-administered with antidepressant medications in improving depressed mood may have biological and clinical standing.

Keywords: brain-intestinal-brain axis; intestinal epithelia; macrophages; vagus nerve; microbiome; dysbiosis; vermiform appendix; probiotics

1. Commentary

1.1. Macrophages and Intestinal Tissue

Both the intestines and the brain develop from the same cluster of embryonic tissue that can be traced back to the primitive streak in early vertebrate fetal growth. As such, this grooved structure that forms on day 15 of human development, along the caudal midline of the bilaminar embryonic disc is the first visible sign of gastrulation that will give rise to the ectoderm, mesoderm and endoderm [1]. When the tissue divides, a portion develops into the Central Nervous System (CNS) (i.e., brain and spinal cord) and the other into the Enteric Nervous System (ENS). Immune molecules are intimately related to the development of the CNS as they are for the intestinal tract. The intestinal tract is a complex multi-dimensional structure that is derived from a simple tubular structure [2]. Such is the contain absorptive enterocytes and secretory tuft, goblet, Paneth, entero-endocrine cell types; while also

accommodating resident intestinal stem cells and rapidly dividing progenitor cells. Adjacent to the intestinal epithelium is an additional complex structure comprising the lamina propria, submucosa, and muscular layers.

Immune molecules have been reported to participate in integral functions in the CNS throughout various stages of neural development, including affecting neurogenesis, neuronal migration, axon guidance, synapse formation, activity-dependent refinement of circuits, and synaptic plasticity [3]. For example, the innate-like, evolutionarily conserved MR1-restricted mucosa-associated invariant T cells (MAIT) from humans represent the most abundant T-cell subset that quickly respond to a wide variety of bacteria [4]. MAIT cells in humans are prevalent and are distributed throughout the blood and mucosal sites [5]. Studies of MAIT cells in human fetal development are important for the consideration of the fundamental immune-biological characteristics expressed, as well as for the role these cells play in the fetus and the newborn. In humans T cells are enriched in mucosal tissues [6].

In the intestines there is recognized a tripartite co-operation in order to maintain intestinal steady state homeostasis. This occurs between the intestinal epithelium barrier, the intestinal microbiome and gut mucosal immune cells such as macrophages [2,7,8].

Macrophages (mononuclear phagocytes) are distributed throughout the body in tissue sites that include for example the intestines, the lung, the liver, and the brain [9]. Macrophages are established from prenatal signals and from circulating monocytes post birth [9]. The appreciation that macrophages have multiple actions in order to maintain the overall immunological efficiency in the gut is an idea that reflects macrophage functional diversity in specific tissues, where the tissue site provides instructive signals for local macrophage differentiation [10].

In the gut, intestinal resident macrophages (CX3CR1^{hi}) are specialized cells that are involved in antigen presentation to T cells that in turn shape the T cell responses generated [11]. As such intestinal resident macrophages are important participants in contributing and maintaining the steady state equilibrium of mucosal immunity. Macrophages shape mucosal immune equilibrium through the action of phagocytosis eliciting protection from pathobiont translocations across the intestinal epithelium [12] (Figure 1).

The activated macrophage phenotype is known to present antigens to T lymphocytes, which initiate the controlled and appropriate immune response that is elicited by a recognition signal that responds to microbial proteins [13–15] (Figure 1). Studies report that antigen presentation triggers macrophage activities that activate T cells; the activity of macrophages is linked to up regulation of a sequence of cytokines that includes interleukin 1 (IL-1), interferon-alpha (IFN-a) and other cytotoxic proteins [13]. have an important role in maintaining immunological equilibrium [15].

Maintaining immunological equilibrium also involves the phagocytosis of exogenous antigens, cellular debris, insoluble particles and activated clotting factors [15]. It is also noted that intestinal resident macrophage populations sustain mucosal tolerance by contributing to the survival and expanding T lymphocytes already primed toward immunological defense of pathogens [8]. Further, the colitogenic T lymphocyte inflammatory response that occurs in the intestinal mucosa is suppressed by the anti-inflammatory cytokine IL-10 elaborated by intestinal resident macrophages; an activity promoting intestinal mucosa immunological tolerance. The macrophage can be hence envisaged to promote regulatory T cell activities [8].

The intestinal epithelia produce soluble protein factors (e.g., thymic stromal lymphopoietin, transforming growth factor-b, and retinoic acid) and also express the integrin ligand semaphorin 7A that undergoes contact-dependent interactions with intestinal macrophages. This activity induces the expression of IL-10 that in turn further promotes intestinal homeostasis. Therefore these cumulative actions combine components of the local innate immune system (i.e., macrophages and dendritic cells and others) and the intestinal epithelia in an interaction that preserves a tolerogenic functional steady state [13].

A recent review by Roman and colleagues [16] has progressed the view that macrophages can influence inflammatory disease outcomes and that sustained inflammatory responses can lead to

depressive moods. Furthermore it was postulated that antidepressive medications can influence peripheral and brain macrophages skewing them toward anti-inflammatory activities that then can improve cognitive functions [16].

1.2. The Brain-Intestinal Microbiome/Epithelia-Mucosa-Brain Loop

There are multiple mechanisms in the gut including a mucus layer, antimicrobial peptides and a tight junction protein network that cooperate to continuously preserve local homeostasis and hence ensure that the intestinal epithelial barrier integrity is maintained. Goblet cells secrete mucin to provide a protective coating, provide structural integrity and regulate macrophage and adaptive T cell responses during inflammation [8,17–29] (Figure 1).

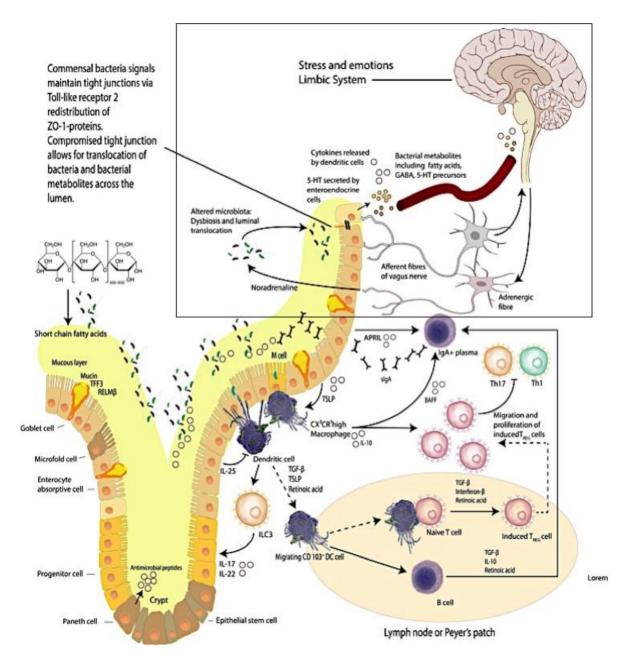


Figure 1. The Brain–Intestinal Microbiome–Intestinal Epithelia–Neuroendocrine-immune–Vagus Nerve–Brain Loop. Mucus production and immune activities delineate the complexity of the intestinal mucosa site. (This figure was adapted from selected reviews [8,30]).

Mental health is very much intricately linked to physical health [31]. Gastrointestinal dysbiosis is associated with an intestinal epithelial cell barrier dysfunction that can be due to environmental and nutritional triggers, which can be further progressed by the intestinal pathobiont cohort that exacerbates and maintains intestinal barrier in a dysfunctional pro-inflammatory state. It has been recently reported that depression is linked to the exacerbation in gap junction integrity between intestinal epithelial cells (i.e., also termed a leaky gut) is a contentious posit. A recent study has reported that approximately 35 % of depressed individuals exhibited evidence of a leaky gut [32,33].

The importance exhibited by the brain–intestine–brain axis is that it provides a bidirectional flow of neuroendocrine-immunological equilibrium control. The intestinal microbiome is thought to exert effects on this axis that significantly impacts the biochemistry of the central and peripheral nervous systems and in turn behavior [34] (Figure 1 (see within the rectangular area)). Studies reporting that depression is accompanied by activation of immune–inflammatory pathways [35] with increased IgM/IgA responses to lipopolysaccharides (LPS) from gram-negative commensal bacteria, indicate that at least in part adverse mood is supported by commensal microbes and or metabolite translocations across the intestinal epithelial barrier [32,33]. We have reported [36] (as have others [37]) that resistant depression can be accompanied by systemic inflammatory states that are posited to originate from intestinal inflammation and the resultant intestinal dysbiosis.

1.3. The Vermiform Appendix

The vermiform appendix is characterized as a diverticulum of the cecum and delineates the beginning of the colon in the confluence of tanias [38]. The appendix is posterior-medially attached to the cecum, approximately 2 cm below the ileocecal junction. The histological structure of the appendix reflects that of the intestinal wall of the large bowel, in particular with appendiceal structures such as the mucosa, submucosa and lymphoid follicles (Figure 2).

It has only been recently recognized that the human vermiform appendix is not just a rudimentary part of the intestine, but rather as suggested by numerous studies an organ of immunological importance for the development and preservation of the intestinal immune system [39,40]. Furthermore the importance of the vermiform appendix has been demonstrated to have a direct functional interaction with the intestinal microbiome [41,42].

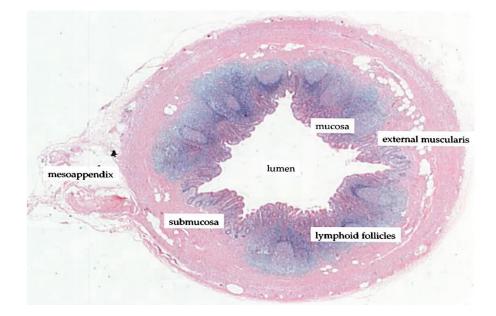


Figure 2. Histological transverse section of the vermiform appendix (adapted/modified from Kooij et al., 2016 [43]).

Reports present the vermiform appendix as an important participant in intestinal ecological microbial contributions and maintenance [41]. This is especially relevant to the ascending colon and the distal small bowel where much of the microbiome is posited to have high metabolic activities. Reports have documented key activities that through co-evolution the human host and the microbial cohort have shaped a tolerant relationship of benefits. These beneficial activities include (i) the intestinal based immune system sustains microbial biofilms in the intestines and is a key constituent of the mutualistic connection between mammals and bacteria [44,45] (ii) biofilms defined as mucilage layers in the gut have been reported to be safe bacterial zones [46], where bacteria and other entities (e.g., fungi) form communal relationships for safeguarding their activities and survival; (iii) the vermiform appendix is very much recognized as an immune tissue [47], with concentrated gut-associated lymphoid tissue; (iv) a recent reported concentrated biofilms contribute to the overall epithelial structure of the vermiform appendix [41,48]; (v) research also shows that biofilms in the intestines are subject to continual turnover an activity that helps to limit bacterial translocations across the intestinal epithelial cells and Peyer's patches [48], where this mucin turnover activity is rapid (approximately 1–2 h) of any biofilms that adhere to the intestinal epithelia [49].

Early reports through culture dependent studies have demonstrated the presence of several Gram-negative bacilli such as Klebsiella, Enterobacter and *Escherichia coli* whereas Gram-positive cocci were less frequently observed [50]. In a more recent study with patients following an appendectomy showed a comprehensive view of the microbial population within the biofilm of the appendix, this can be determined by high-throughput DNA sequencing [51]. The human appendix was demonstrated to contain members of some fifteen bacterial phyla [51], including Firmicutes (the most dominant), Proteobacteria, Bacteroidetes, Actinobacteria, and Fusobacteria. Moreover, certain oral pathogens not associated with the intestines were also detected in the appendix samples (i.e., Gemella, Parvimonas, and Fusobacterium). This report presents an immunological organ with a significant associated microbial diversity which in part supports the posit that the appendix microbiota has important functions in human health; the biofilm in the appendix acts as a safe house for commensal bacteria, therefore, facilitating re-inoculation of the intestines post a gastrointestinal tract infection as the appendiceal lumen is spared from diarrhoeal clearance [40].

Health interventions that may disrupt the microbiome ecology of the large bowel may lead to disease. A recent study has highlighted that the vermiform appendix may have a significant association with chronic diseases in particular in the development of large bowel cancer following the surgical removal of the appendix [52]. Recently a study of patients clinically treated with antibiotics for appendicitis were reported to have a significant increased risk for large bowel cancer [53]. Moreover, although there is a paucity of studies on the relationship of appendicectomy to mood disorders, an early study has postulated that an appendicectomy may lead to psychological disturbances [54]. An interesting corollary from another study with subjects undergoing an appendicectomy showed that 80% of the participants with inflammatory problems of the appendix were designated as positive for depressive symptoms [55]. Moreover psychological depressive symptoms have been reported to continue post an appendicectomy [56]. These studies overall tend to support the notion that severe disruptions of the intestinal microbiome and the loss of the appendix biofilm may increase the risk of disease including adverse mood dispositions.

1.4. Probiotics

Current research continues to draw connections between the intestinal microbiome and environmental factors to sensitivities to the host's emotional states [57]. In animal models, dysbiosis has been demonstrated to impair vagus signaling which results in reduced protein synthesis in the hippocampus, corrected by rescuing the intestinal microbiome with either specific strains of probiotics [58,59]. Our group has recently demonstrated in a small pilot study [60] with treatment resistant depression (while on SSRI medications) that the administration of a probiotic formulation improved depressive symptoms in a small cohort. An anti-inflammatory response was suggested.

A meta-analysis by Ng and colleagues [61] investigating the administration of probiotics to alleviate depressive symptoms was largely inconclusive with an insignificant effect on mood.

An interesting recent study [62] in a healthy murine model demonstrated that oral administration of probiotic bacteria cell walls stimulated the immune system. This study although preliminary, showed that probiotic bacteria and their cell walls have important immunoregulatory effects on intestinal epithelial cells without an adverse effect on the local metabolic and environment equilibrium. The study by Lemme-Dumit and colleagues [62] showed that cell walls from probiotic bacteria increased immunoglobulin A secreting cells in the intestines and in innate immune cells as well as other tissues such as those of the spleen and peritoneum [62]. This study also demonstrated the capacity that probiotic bacteria provide to stimulate important cellular and immune elements such as the intestinal epithelia and intestinal resident macrophages in the gut [62].

2. Reprise

We have previously postulated that depressed mood [36] is linked to antagonistic immuno-endocrine control of intestinal homeostasis. The scientific rationale posited suggests that inflammation of the intestines relevant to depressed mood is an effect associated with the intestinal mucosa. Intestinal microbiome adverse shifts that maintain low-level pro-inflammatory activity and intestinal dysbiosis that overwhelm intestinal resident macrophages is posited to play a role in depressed mood. Clinical studies show that the vermiform appendix re-inoculates the proximal colon following excessive pro-inflammatory triggers (e.g., gastrointestinal inflammations, dysentery, antibiotic administration). Reports of a surgically removed chronically inflamed vermiform appendix have been linked to chronic disease developed including depression.

We postulate that intestinal inflammations may provoke an increased risk for adverse mood disorders in those without a vermiform appendix. In depression the loss of keystone intestinal bacterial species and the incapacity to restore microbial diversity and stability in the proximal colon (i.e., due to an appendectomy) could be an important factor that disrupts the steady state of neuro-endocrine-immunological equilibrium in the intestinal mucosa, especially following the over prescription of antibiotics. We advance the idea that an in situ normal functioning vermiform appendix continually contributes to diversity and stability to the intestinal bacterial cohort over a lifetime. Probiotics that can regulate the functioning of the immune system in the gut may have important plausible implications in improving depressive mood states; dedicated studies are warranted.

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Conflicts of Interest: L.V. and S.H. participate in research on probiotics in Medlab Clinical's research laboratory facility in Sydney, Australia. The authors declare no other conflict of interest.

References

- 1. Williams, M.L.; Solnica-Krezel, L. Regulation of gastrulation movements by emergent cell and tissue interactions. *Curr. Opin. Cell Biol.* **2017**, *48*, 33–39. [CrossRef] [PubMed]
- 2. De Santa Barbara, P.; van den Brink, G.R.; Roberts, D.J. Development and differentiation of the intestinal epithelium. *Cell. Mol. Life Sci. CMLS* **2003**, *60*, 1322–1332. [CrossRef] [PubMed]
- Garay, P.A.; McAllister, A.K. Novel roles for immune molecules in neural development: Implications for neurodevelopmental disorders. *Front. Synaptic Neurosci.* 2010, 2, 136. [CrossRef] [PubMed]
- 4. Lantz, O.; Legoux, F. Mait cells: An historical and evolutionary perspective. *Immunol. Cell Biol.* **2017**. [CrossRef] [PubMed]
- 5. Napier, R.J.; Adams, E.J.; Gold, M.C.; Lewinsohn, D.M. The role of mucosal associated invariant t cells in antimicrobial immunity. *Front. Immunol.* **2015**, *6*, 344. [CrossRef] [PubMed]

- Wong, E.B.; Ndung'u, T.; Kasprowicz, V.O. The role of mucosal-associated invariant t cells in infectious diseases. *Immunology* 2017, 150, 45–54. [CrossRef] [PubMed]
- 7. Caricilli, A.M.; Castoldi, A.; Camara, N.O. Intestinal barrier: A gentlemen's agreement between microbiota and immunity. *World J. Gastrointest. Pathophysiol.* **2014**, *5*, 18–32. [CrossRef] [PubMed]
- 8. Peterson, L.W.; Artis, D. Intestinal epithelial cells: Regulators of barrier function and immune homeostasis. *Nat. Rev. Immunol.* **2014**, *14*, 141–153. [CrossRef] [PubMed]
- 9. Epelman, S.; Lavine, K.J.; Randolph, G.J. Origin and functions of tissue macrophages. *Immunity* **2014**, *41*, 21–35. [CrossRef] [PubMed]
- Gautier, E.L.; Shay, T.; Miller, J.; Greter, M.; Jakubzick, C.; Ivanov, S.; Helft, J.; Chow, A.; Elpek, K.G.; Gordonov, S.; et al. Gene-expression profiles and transcriptional regulatory pathways that underlie the identity and diversity of mouse tissue macrophages. *Nat. Immunol.* 2012, *13*, 1118–1128. [CrossRef] [PubMed]
- Zhou, D.; Huang, C.; Lin, Z.; Zhan, S.; Kong, L.; Fang, C.; Li, J. Macrophage polarization and function with emphasis on the evolving roles of coordinated regulation of cellular signaling pathways. *Cell. Signal.* 2014, 26, 192–197. [CrossRef] [PubMed]
- 12. Grainger, J.R.; Konkel, J.E.; Zangerle-Murray, T.; Shaw, T.N. Macrophages in gastrointestinal homeostasis and inflammation. *Pflug. Arch. Eur. J. Physiol.* **2017**, *469*, 527–539. [CrossRef] [PubMed]
- 13. Sansonetti, P. Host-pathogen interactions: The seduction of molecular cross talk. *Gut* **2002**, *50* (Suppl. 3), iii2–iii8. [CrossRef] [PubMed]
- 14. Weber, B.; Saurer, L.; Mueller, C. Intestinal macrophages: Differentiation and involvement in intestinal immunopathologies. *Semin. Immunopathol.* **2009**, *31*, 171–184. [CrossRef] [PubMed]
- 15. De Schepper, S.; Stakenborg, N.; Matteoli, G.; Verheijden, S.; Boeckxstaens, G.E. Muscularis macrophages: Key players in intestinal homeostasis and disease. *Cell. Immunol.* **2017**, in press. [CrossRef] [PubMed]
- 16. Roman, A.; Kreiner, G.; Nalepa, I. Macrophages and depression—A misalliance or well-arranged marriage? *Pharmacol. Rep.* **2013**, *65*, 1663–1672. [CrossRef]
- 17. Robinson, K.; Deng, Z.; Hou, Y.; Zhang, G. Regulation of the intestinal barrier function by host defense peptides. *Front. Vet. Sci.* 2015, 2, 57. [CrossRef] [PubMed]
- Ai, T.L.; Solomon, B.D.; Hsieh, C.S. T-cell selection and intestinal homeostasis. *Immunol. Rev.* 2014, 259, 60–74. [CrossRef] [PubMed]
- Guilliams, M.; Ginhoux, F.; Jakubzick, C.; Naik, S.H.; Onai, N.; Schraml, B.U.; Segura, E.; Tussiwand, R.; Yona, S. Dendritic cells, monocytes and macrophages: A unified nomenclature based on ontogeny. *Nat. Rev. Immunol.* 2014, 14, 571–578. [CrossRef] [PubMed]
- Gottschalk, C.; Kurts, C. The debate about dendritic cells and macrophages in the kidney. *Front. Immunol.* 2015, *6*, 435. [CrossRef] [PubMed]
- 21. Hume, D.A. Macrophages as apc and the dendritic cell myth. *J. Immunol.* **2008**, *181*, 5829–5835. [CrossRef] [PubMed]
- 22. Kayama, H.; Ueda, Y.; Sawa, Y.; Jeon, S.G.; Ma, J.S.; Okumura, R.; Kubo, A.; Ishii, M.; Okazaki, T.; Murakami, M.; et al. Intestinal CX3C chemokine receptor 1(high) (CX3CR1(high)) myeloid cells prevent t-cell-dependent colitis. *Proc. Natl. Acad. Sci. USA* **2012**, *109*, 5010–5015. [CrossRef] [PubMed]
- 23. Kidd, P. Th1/Th2 balance: The hypothesis, its limitations, and implications for health and disease. *Altern. Med. Rev.* 2003, *8*, 223–246. [PubMed]
- 24. Zheng, S.G. Regulatory T cells vs Th17: Differentiation of Th17 versus treg, are the mutually exclusive? *Am. J. Clin. Exp. Immunol.* **2013**, *2*, 94–106. [PubMed]
- 25. Wahl, S.M. Transforming growth factor-beta: Innately bipolar. *Curr. Opin. Immunol.* **2007**, *19*, 55–62. [CrossRef] [PubMed]
- 26. Nakahashi-Oda, C.; Udayanga, K.G.; Nakamura, Y.; Nakazawa, Y.; Totsuka, N.; Miki, H.; Iino, S.; Tahara-Hanaoka, S.; Honda, S.; Shibuya, K.; et al. Apoptotic epithelial cells control the abundance of treg cells at barrier surfaces. *Nat. Immunol.* **2016**, *17*, 441–450. [CrossRef] [PubMed]
- 27. Ogino, H.; Nakamura, K.; Ihara, E.; Akiho, H.; Takayanagi, R. CD4⁺CD25⁺ regulatory T cells suppress Th17-responses in an experimental colitis model. *Dig. Dis. Sci.* **2011**, *56*, 376–386. [CrossRef] [PubMed]
- 28. Smith, P.M.; Howitt, M.R.; Panikov, N.; Michaud, M.; Gallini, C.A.; Bohlooly, Y.M.; Glickman, J.N.; Garrett, W.S. The microbial metabolites, short-chain fatty acids, regulate colonic treg cell homeostasis. *Science* **2013**, *341*, 569–573. [CrossRef] [PubMed]

- 29. Abbas, A.K.; Benoist, C.; Bluestone, J.A.; Campbell, D.J.; Ghosh, S.; Hori, S.; Jiang, S.; Kuchroo, V.K.; Mathis, D.; Roncarolo, M.G.; et al. Regulatory T cells: Recommendations to simplify the nomenclature. *Nat. Immunol.* **2013**, *14*, 307–308. [CrossRef] [PubMed]
- 30. Collins, S.M.; Surette, M.; Bercik, P. The interplay between the intestinal microbiota and the brain. *Nat. Rev. Microbiol.* **2012**, *10*, 735–742. [CrossRef] [PubMed]
- 31. Essential exercise: Physical and mental health "inextricably" linked. Major disease prevention is shown to be an additional benefit. *DukeMed. Healthnews* **2010**, *16*, 4–5.
- 32. Maes, M.; Kubera, M.; Leunis, J.C.; Berk, M.; Geffard, M.; Bosmans, E. In depression, bacterial translocation may drive inflammatory responses, oxidative and nitrosative stress (O&NS), and autoimmune responses directed against O&NS-damaged neoepitopes. *Acta Psychiatr. Scand.* **2013**, *127*, 344–354. [PubMed]
- Slyepchenko, A.; Maes, M.; Jacka, F.N.; Kohler, C.A.; Barichello, T.; McIntyre, R.S.; Berk, M.; Grande, I.; Foster, J.A.; Vieta, E.; et al. Gut microbiota, bacterial translocation, and interactions with diet: Pathophysiological links between major depressive disorder and non-communicable medical comorbidities. *Psychother. Psychosom.* 2017, *86*, 31–46. [CrossRef] [PubMed]
- Dinan, T.G.; Cryan, J.F. The microbiome-gut-brain axis in health and disease. *Gastroenterol. Cl. N. Am.* 2017, 46, 77–89. [CrossRef] [PubMed]
- 35. Morris, G.; Reiche, E.M.V.; Murru, A.; Carvalho, A.F.; Maes, M.; Berk, M.; Puri, B.K. Multiple immune-inflammatory and oxidative and nitrosative stress pathways explain the frequent presence of depression in multiple sclerosis. *Mol. Neurobiol.* **2018**. [CrossRef] [PubMed]
- 36. Vitetta, L.; Bambling, M.; Alford, H. The gastrointestinal tract microbiome, probiotics, and mood. *Inflammopharmacology* **2014**, *22*, 333–339. [CrossRef] [PubMed]
- 37. Clapp, M.; Aurora, N.; Herrera, L.; Bhatia, M.; Wilen, E.; Wakefield, S. Gut microbiota's effect on mental health: The gut-brain axis. *Clin. Pract.* **2017**, *7*, 987. [CrossRef] [PubMed]
- Wakeley, C.P. The position of the vermiform appendix as ascertained by an analysis of 10,000 cases. *J. Anat.* 1933, 67, 277–283. [PubMed]
- 39. Gebbers, J.O.; Laissue, J.A. Bacterial translocation in the normal human appendix parallels the development of the local immune system. *Ann. N. Y. Acad. Sci.* **2004**, *1029*, 337–343. [CrossRef] [PubMed]
- Im, G.Y.; Modayil, R.J.; Lin, C.T.; Geier, S.J.; Katz, D.S.; Feuerman, M.; Grendell, J.H. The appendix may protect against clostridium difficile recurrence. *Clin. Gastroenterol. Hepatol.* 2011, *9*, 1072–1077. [CrossRef] [PubMed]
- 41. Randal Bollinger, R.; Barbas, A.S.; Bush, E.L.; Lin, S.S.; Parker, W. Biofilms in the large bowel suggest an apparent function of the human vermiform appendix. *J. Theor. Biol.* **2007**, 249, 826–831. [CrossRef] [PubMed]
- Van Wey, A.S.; Cookson, A.L.; Roy, N.C.; McNabb, W.C.; Soboleva, T.K.; Shorten, P.R. Bacterial biofilms associated with food particles in the human large bowel. *Mol. Nutr. Food Res.* 2011, 55, 969–978. [CrossRef] [PubMed]
- 43. Kooij, I.A.; Sahami, S.; Meijer, S.L.; Buskens, C.J.; Te Velde, A.A. The immunology of the vermiform appendix: A review of the literature. *Clin. Exp. Immunol.* **2016**, *186*, 1–9. [CrossRef] [PubMed]
- 44. Sonnenburg, J.L.; Angenent, L.T.; Gordon, J.I. Getting a grip on things: How do communities of bacterial symbionts become established in our intestine? *Nat. Immunol.* **2004**, *5*, 569–573. [CrossRef] [PubMed]
- 45. Bollinger, R.R.; Everett, M.L.; Wahl, S.D.; Lee, Y.H.; Orndorff, P.E.; Parker, W. Secretory iga and mucin-mediated biofilm formation by environmental strains of escherichia coli: Role of type 1 pili. *Mol. Immunol.* **2006**, *43*, 378–387. [CrossRef] [PubMed]
- 46. Davies, D. Understanding biofilm resistance to antibacterial agents. *Nature reviews. Drug Discov.* **2003**, *2*, 114–122. [CrossRef] [PubMed]
- 47. Bazar, K.A.; Lee, P.Y.; Joon Yun, A. An "eye" in the gut: The appendix as a sentinel sensory organ of the immune intelligence network. *Med. Hypotheses* **2004**, *63*, 752–758. [CrossRef] [PubMed]
- 48. Pelaseyed, T.; Bergstrom, J.H.; Gustafsson, J.K.; Ermund, A.; Birchenough, G.M.; Schutte, A.; van der Post, S.; Svensson, F.; Rodriguez-Pineiro, A.M.; Nystrom, E.E.; et al. The mucus and mucins of the goblet cells and enterocytes provide the first defense line of the gastrointestinal tract and interact with the immune system. *Immunol. Rev.* 2014, 260, 8–20. [CrossRef] [PubMed]
- 49. Johansson, M.E. Fast renewal of the distal colonic mucus layers by the surface goblet cells as measured by in vivo labeling of mucin glycoproteins. *PLoS ONE* **2012**, *7*, e41009. [CrossRef] [PubMed]

- 50. Leigh, D.A.; Simmons, K.; Norman, E. Bacterial flora of the appendix fossa in appendicitis and postoperative wound infection. *J. Clin. Pathol.* **1974**, 27, 997–1000. [CrossRef] [PubMed]
- Guinane, C.M.; Tadrous, A.; Fouhy, F.; Ryan, C.A.; Dempsey, E.M.; Murphy, B.; Andrews, E.; Cotter, P.D.; Stanton, C.; Ross, R.P. Microbial composition of human appendices from patients following appendectomy. *MBio* 2013, 4, e00366-12. [CrossRef] [PubMed]
- 52. Wu, S.C.; Chen, W.T.; Muo, C.H.; Ke, T.W.; Fang, C.W.; Sung, F.C. Association between appendectomy and subsequent colorectal cancer development: An asian population study. *PLoS ONE* **2015**, *10*, e0118411. [CrossRef] [PubMed]
- 53. Enblad, M.; Birgisson, H.; Ekbom, A.; Sandin, F.; Graf, W. Increased incidence of bowel cancer after non-surgical treatment of appendicitis. *Eur. J. Surg. Oncol.* **2017**, *43*, 2067–2075. [CrossRef] [PubMed]
- 54. Canton, G.; Santonastaso, P.; Fraccon, I.G. Life events, abnormal illness behavior, and appendectomy. *Gen. Hosp. Psychiatry* **1984**, *6*, 191–195. [CrossRef]
- 55. Beaurepaire, J.E.; Jones, M.; Eckstein, R.P.; Smith, R.C.; Piper, D.W.; Tennant, C. The acute appendicitis syndrome: Psychological aspects of the inflamed and non-inflamed appendix. *J. Psychosom. Res.* **1992**, *36*, 425–437. [CrossRef]
- 56. Creed, F. Life events and appendicectomy. Lancet 1981, 1, 1381–1385. [CrossRef]
- 57. Fond, G.; Boukouaci, W.; Chevalier, G.; Regnault, A.; Eberl, G.; Hamdani, N.; Dickerson, F.; Macgregor, A.; Boyer, L.; Dargel, A.; et al. The "psychomicrobiotic": Targeting microbiota in major psychiatric disorders: A systematic review. *Pathol. Biol.* **2015**, *63*, 35–42. [CrossRef] [PubMed]
- 58. Dinan, T.G.; Cryan, J.F. Melancholic microbes: A link between gut microbiota and depression? *Neurogastroenterol. Motil.* **2013**, *25*, 713–719. [CrossRef] [PubMed]
- 59. Dinan, T.G.; Stanton, C.; Cryan, J.F. Psychobiotics: A novel class of psychotropic. *Biol. Psychiatry* **2013**, 74, 720–726. [CrossRef] [PubMed]
- 60. Bambling, M.; Edwards, S.C.; Hall, S.; Vitetta, L. A combination of probiotics and magnesium orotate attenuate depression in a small ssri resistant cohort: An intestinal anti-inflammatory response is suggested. *Inflammopharmacology* **2017**, *25*, 271–274. [CrossRef] [PubMed]
- 61. Ng, Q.X.; Peters, C.; Ho, C.Y.X.; Lim, D.Y.; Yeo, W.S. A meta-analysis of the use of probiotics to alleviate depressive symptoms. *J. Affect. Disord.* **2018**, *228*, 13–19. [CrossRef] [PubMed]
- 62. Lemme-Dumit, J.M.; Polti, M.A.; Perdigon, G.; Galdeano, C.M. Probiotic bacteria cell walls stimulate the activity of the intestinal epithelial cells and macrophage functionality. *Benefic. Microbes* **2018**, *9*, 153–164. [CrossRef] [PubMed]



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