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# PANoptosis opens new treatment options for allergic bronchopulmonary aspergillosis

Dalan Smallwood, MD, Richard F. Lockey, MD, and Narasaiah Kolliputi, PhD Tampa, Fla

Background: Allergic bronchopulmonary aspergillosis (ABPA) is a rare airway disorder primarily affecting patients with asthma and cystic fibrosis. Persistent airway inflammation brought on by Aspergillus fumigatus exacerbates the underlying condition and can cause significant respiratory damage. Treatments center on reducing inflammation with the use of corticosteroids and antifungals. PANoptosis is a new concept in the field of cell death and inflammation that posits the existence of cross talk and a master control system for the 3 programmed cell death (PCD) pathways, namely, apoptosis, pyroptosis, and necroptosis. This concept has revolutionized the understanding of PCD and opened new avenues for its exploration. Studies show that Aspergillus is one of the pathogens that is capable of activating PANoptosis via the Z-DNA binding protein 1 (ZBP1) pathway and plays an active role in the inflammation caused by this organism.

Objective: This article explores the nature of inflammation in ABPA and ways in which PCD could lead to novel treatment options.

Method: PubMed was used to review the literature surrounding *Aspergillus* infection–related inflammation and PANoptosis.

Results: There is evidence that apoptosis and pyroptosis protect against *Aspergillus*-induced inflammation, whereas necroptosis promotes inflammation.

Conclusion: Experimental medications, in particular, necroptosis inhibitors such as necrosulfonamide and necrostatin-1, should be studied for use in the treatment of ABPA. (J Allergy Clin Immunol Global 2024;3:100298.)

Key words: ABPA, allergic bronchopulmonary aspergillosis, apoptosis, Aspergillus fumigatus, necroptosis, PANoptosis, programmed cell death, pyroptosis, TAK1, ZBP1

Allergic bronchopulmonary aspergillosis (ABPA) is a rare airway disorder characterized by persistent inflammation in response to infection by fungi from the genus *Aspergillus*.

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Abbreviati	ions used		
ABPA:	Allergic bronchopulmonary aspergillosis		
AIM2:	Absent in melanoma 2		
CASP8:	Caspase-8		
GSDMD:	Gasdermin D		
MLKL:	Mixed-lineage kinase domain-like		
Nec-1:	Necrostatin-1		
NLRP3:	Nucleotide-binding oligomerization domain-like receptor		
	family pyrin domain-containing 3		
NSA:	Necrosulfonamide		
PCD:	Programmed cell death		
PRR:	Pattern recognition receptor		
RIPK1:	RIPK1: Receptor-interacting serine/threonine-protein kinase-1		
RIPK3:	Receptor-interacting serine/threonine-protein kinase-3		
TAK1:	TGF-β-activated kinase-1		
ZBP1:	Z-DNA binding protein-1		

Although other fungi, such as *Candida albicans* and *Shizophyl*lum commune, are capable of causing ABPM, Aspergillus is the most common and best-studied fungus associated with this disease.<sup>1,2</sup> ABPA has an estimated prevalence of 2.5% in subjects with asthma and 8.9% in subjects with cystic fibrosis. These numbers may be as high as 22.4% and 25%, respectively, depending on the diagnostic criteria used, and underdiagnosis of the disease is speculated.<sup>3-6</sup> ABPA primarily affects patients with chronic airway diseases, although cases without underlying respiratory problems do occur.<sup>7-9</sup> ABPA exacerbates airway symptoms and causes hemoptysis, a productive cough with mucus plugs, and dyspnea.<sup>5</sup> Its diagnosis is based on laboratory and radiologic findings, including blood eosinophilia, elevated level of specific IgG to A fumigatus, elevated total and specific levels of IgE, and bronchiectasis on computed tomography.<sup>5,6,10</sup> These criteria make ABPA significantly easier to diagnose than the other ABPMs. Treatment strategies include systemic glucocorticosteroids and antifungals, which reduce inflammation and fungal load, with targeted biologics emerging as alternative therapeutic options.<sup>11,1</sup>

ABPA-associated inflammation occurs via the  $T_{H2}$  cell inflammatory pathway, in which  $T_{H2}$  cells produce IL-4, IL-5, and IL-13, which in turn activate downstream pathways.<sup>13,14</sup> IL-33, which is located in the nuclei of epithelial cells, activates cytokines and enhances production of IL-4, IL-5, and IL-13 in conjunction with the other critical epithelial-derived  $T_{H2}$  inflammatory cytokines, IL-25 and thymic stromal lympopoietin (TSLP).<sup>14-17</sup> IL-33 binds to IL-1 receptor-like 1 (IL1RL1 [also known as ST2]) during allergic inflammation, signaling the formation of a heterodimer between ST2L, the transmembrane isoform of ST2, and IL-1 receptor accessory protein (IL-1 RAcP). The result is a signaling cascade that activates the transcription of proinflammatory molecules, including nuclear factor-κB.<sup>18</sup> This leads to the activation of eosinophils and other cells involved in allergic inflammation.

From the Division of Allergy and Immunology, Department of Internal Medicine, Morsani College of Medicine, University of South Florida, Tampa.

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Corresponding author: Narasaiah Kolliputi, PhD, Division of Allergy and Immunology, Department of Internal Medicine, Morsani College of Medicine, University of South Florida, 12901 Bruce B. Downs Blvd, Tampa, FL 33612. E-mail: nkollipu@usf.edu.

<sup>2772-8293</sup> 

*Aspergillus* and other fungal allergens contain proteases capable of cleaving IL-33 into a more mature form, increasing its bioactivity compared with that of the full-length form.<sup>19</sup> Increased levels of IL-33 in ABPA and other *Aspergillus* spp infections make it a potential target for future ABPA therapies.<sup>20,21</sup> Inhibition of the IL-33 signaling pathway in a mouse model of ABPA significantly reduces inflammation in invasive aspergillosis and *Aspergillus*–sensitive asthma, making it likely that the increased bioactivity from IL-33 cleavage is the source of this inflammation.<sup>22,23</sup> Thus, it can reasonably be concluded that inhibition of this signaling pathway may similarly reduce ABPA inflammation.

# METHODS PCD pathways

Apoptosis is the best-known and best-understood programmed cell death (PCD) pathway. It proceeds through 2 methods: intrinsic and extrinsic. The intrinsic pathway is activated when there is an absence of prosurvival molecules, such as growth factors, or the presence of proapoptotic molecules, such as reactive oxygen species. These molecules trigger the release of mitochondrial proteins into the cytoplasm capable of activating apoptotic protease activating factor 1 (APAF1), which assembles into an oligomer known as the apoptosome. The apoptosome activates procaspase-9, which subsequently activates procaspase-3, the final executioner of apoptosis. The extrinsic pathway involves death signals released from natural killer cells and macrophages, such as TNF- $\alpha$  and Fas ligand (Fas-L). Binding of these death ligands to a death receptor allows the recruitment and activation of procaspase-8, which is able to dimerize and activate procaspase-3.24

Pyroptosis is governed by inflammasomes, which form when pattern-associated molecular patterns (PAMPs) and damage-associated molecular patterns (DAMPs) are detected by NOD-like receptors (NLRs), absent in melanoma 2 (AIM2), and other pattern recognition receptors (PRRs).<sup>25</sup> Interaction of these PRRs with adapter apoptosis-associated speck-like protein containing a caspase recruitment domain (ASC) and procaspase-1 generates an inflammasome. The inflammasome can produce activated caspase-1 molecules, which cleave gasdermin D (GSDMD). The N-terminal of GSDMD inserts pores into the cellular membrane, causing secretion of the proinflammatory molecules IL-1 $\beta$  and IL-18, thereby activating pyroptosis.<sup>26</sup>

Necroptosis is the least-understood cell death pathway. Initiation is similar to the extrinsic pathway of apoptosis in that it relies on the death signal, TNF, to activate TNF receptor 1 (TNFR1). This causes the formation of a complex that includes the molecules TNFR-associated factors (TRAF), receptor-interacting serine/threonine-protein kinase-1 (RIPK1), cellular inhibitor of apoptosis protein 1 (cIAP1), cellular inhibitor of apoptosis protein 2 (cIAP2), Fas-associated via death domain (FADD), and TNFR superfamily member 1A -associated via death domain (TRADD). The cIAPs and TRAF work together to ubiquitinate RIPK1. FADD recruits procespase-8, and RIPK1 recruits receptor-interacting serine/threonine-protein kinase-3 (RIPK3).<sup>26</sup> The ubiquitination process is opposed by the deubiquitinating molecule, cylindromatosis (CYLD). If RIPK1 is deubiquitinated and caspase-8 is not present, RIPK1 and

RIPK3 will complex to activate mixed-lineage kinase domainlike (MLKL), which permeabilizes the cell membrane and causes cell death.<sup>26</sup> The requirement that caspase-8 (CASP8) be absent has led to the hypothesis that necroptosis is meant to be a backup cell death pathway for cells incapable of undergoing apoptosis. Table I summarizes the molecules involved in apoptosis, pyroptosis, and necroptosis.

#### The PANoptosome

The term *PANoptosis*, which was first coined in 2019 (with the initial data dating back to as early as 2016), is an emerging concept in understanding cell death pathways.<sup>27-29</sup> Underlying this concept is the hypothesis that pyroptosis, apoptosis, and necroptosis involve cross talk regulated by a master controlling complex, the PANoptosome.<sup>29</sup> Research on PANoptosis has opened up many avenues in cell death related–research, in particular, infectious organisms such as *A fumigatus*.<sup>30,31</sup>

Two critical molecules at the top of the PANoptosis pathway are Z-DNA binding protein 1 (ZBP1) and TGF- $\beta$ -activated kinase 1 (TAK1).<sup>29</sup> A 2020 study demonstrates that the ZBP1 molecule plays a role in inflammation related to *Aspergillus* infections.<sup>32</sup>

ZBP1 contains a  $Z\alpha$  domain capable of detecting Z-nucleic acids derived from infectious pathogens.<sup>29</sup> This domain is rarely found in mammals and is shared only endogenously, with the molecule adenosine deaminase acting on ribonucleic acid (RNA) 1 (ADAR1), an inhibitor of ZBP1.33 Activation of this domain in concert with type 1 interferon signaling is essential for its activation.<sup>34</sup> After activation, ZBP1 can then interact via its RIP homotypic activation motif (RHIM) with RIPK3, which forms a complex with CASP8 to induce downstream PCD pathways.<sup>29</sup> RIPK3 interacts with MLKL to induce necroptosis, CASP8 induces apoptosis, and the complex interacting with nucleotide-binding oligomerization domain-like receptor family pyrin domain-containing 3 (NLRP3) induces pyroptosis.<sup>29</sup> Evidence suggests that deletion of the ZBP1 gene prevents the activation of PCD pathways in influenza A-infected cells.<sup>34</sup> In contrast, deletion of the gene in the presence of C albicans and Aspergillus fumigatus infection results in reduced, but not absent, PCD.<sup>32</sup> Therefore, either alternative PANoptosis pathways are involved in activating the PANoptosome or activation of the individual PCD pathways by variance virulence factors plays a significant role in fungal infections.

TAK1 has an effect opposite that of ZBP1 by inhibiting the PANoptosis pathway. Activity of this molecule prevents formation of RIPK1-FADD-CASP8 complexes, which function as a PANoptosome similar to ZBP1-RIPK3-CASP8, activating PCD pathways in the same way.<sup>29</sup> Inactivation of this molecule results in unregulated activity of PCD, allowing exploitation by infectious organisms, such as *Yersinia*. *Yersinia* spp produce toxins capable of blocking TAK1 activity to promote their pathogenesis.<sup>29</sup> Experiments in a *Drosophila* model indicate that *Aspergillus* may produce a cyclopentanediol analog capable of acting on this pathway (Fig 1). However, further investigation is necessary to confirm this fact.<sup>35</sup> If such a virulence factor were discovered, it could become a target for future therapeutic agents with the benefit of having a low side effect profile because of its target specificity.

#### TABLE I. Molecules involved in PCD

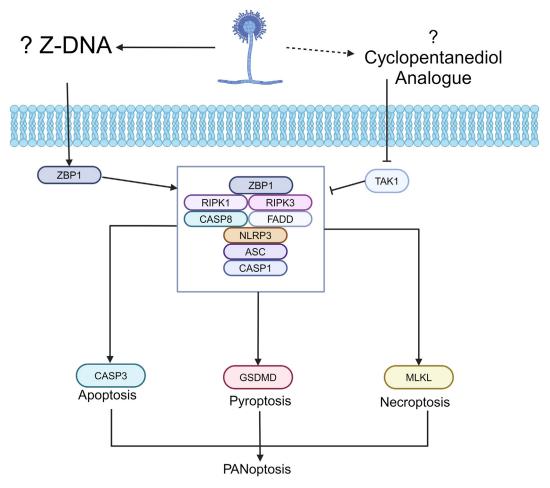
Molecule name	Abbreviation	Pathway	Function
Absent in melanoma 2	AIM2	Pyroptosis	Pattern recognition receptor that starts the pyroptosis cascade
Apoptotic protease activating factor 1	APAF1	Apoptosis (intrinsic)	Activator of procaspase-9
Apoptosis-associated speck-like protein containing a caspase recruitment domain	ASC	Pyroptosis	Recruits procaspase-1 to the inflammasome
Caspase-1	CASP1	Pyroptosis	Cleaves GSDMD
Caspase-3	CASP3	Apoptosis	Executioner caspase that triggers apoptosis
Caspase-8	CASP8	Apoptosis (extrinsic), necroptosis	Initiator caspase of the extrinsic apoptosis pathway; activates procaspase-3; its presence inhibits necroptosis
Caspase-9	CASP9	Apoptosis (intrinsic)	Initiator caspase of intrinsic apoptosis pathway; activates procaspase-3
Cellular inhibitor of apoptosis protein 1	cIAP1	Apoptosis (extrinsic), necroptosis	Ubiquitinates caspase-8, allowing necroptosis to proceed
Cellular inhibitor of apoptosis protein 2	cIAP2	Apoptosis (extrinsic), necroptosis	Ubiquitinates caspase-8, allowing necroptosis to proceed
Cylindromatosis	CYLD	Apoptosis (extrinsic), necroptosis	Deubiquitinates caspase-8 to inhibit necroptosis
Fas-associated via death domain	FADD	Apoptosis (extrinsic), necroptosis	Recruits procaspase-8
Fas ligand	Fas-L	Apoptosis (extrinsic), necroptosis	Binds death receptor to initiate extrinsic apoptosis or necroptosis
Gasdermin D	GSDMD	Pyroptosis	Perforates cell membrane to allow release of IL-1 $\beta$ and IL-18
Mixed-lineage kinase domain like	MLKL	Necroptosis	Final executor of necroptosis; permeabilizes cell membrane
Receptor-interacting serine/ threonine-protein kinase 1	RIPK1	Apoptosis (extrinsic), necroptosis	Recruits RIPK3; can form apoptotic complex
Receptor-interacting serine/ threonine-protein kinase 3	RIPK3	Necroptosis	Activates MLKL
TNF-α	TNF-α	Apoptosis (extrinsic), necroptosis	Binds death receptors to trigger extrinsic apoptosis or necroptosis
TNF receptor 1	TNFR1	Apoptosis (extrinsic), necroptosis	Activated by TNF- $\alpha$ to initiate extrinsic apoptosis or necroptosis
TNF receptor-associated factor	TRAF	Apoptosis (extrinsic), necroptosis	Ubiquitinates caspase-8, allowing necroptosis to proceed
TNF receptor superfamily member 1A-associated via death domain	TRADD	Necroptosis	Recruits procaspase-8

# RESULTS

# PCD and A fumigatus

A fumigatus contains multiple virulence factors involved in inducing PCD. The 2 major players in apoptosis are dihydroxynapthalene-melanin and gliotoxin.<sup>31</sup> Dihydroxynapth alene-melanin activates protein kinase B (PKB/Akt), an inhibitor of caspase-9 and other molecules involved in the extrinsic apoptosis pathway, allowing Aspergillus to produce conidia. Once they germinate and grow hyphae, A fumigatus produces gliotoxin, a molecule capable of activating the intrinsic pathway via c-Jun N-terminal kinase (JNK).<sup>31</sup> The pyroptosis pathway is induced via interaction of the fungal cell wall component  $\beta$ glucan with NLRP3 accompanied by activation of the PRR dectin-1<sup>31,36</sup> Toll-like receptor 2 (TLR2) associates with dectin-1 in A fumigatus recognition and influences Treg cell differentiation.<sup>37</sup> Chitin is a common component of fungal cell walls and binds Toll-like receptor 2 to trigger inflammation.<sup>38</sup> Because this antigen is also found in A fumigatus, it is most likely an important component of the pyroptosis pathway in ABPA.<sup>39</sup> On the basis of the mechanisms of inflammasome activation seen in other fungi, it is also suspected that ergosterol is involved in activating pyroptosis.<sup>31,40</sup> On the basis of studies with *Candida*, it is hypothesized that dectin-1 activation plays a role in necroptosis that is similar to its role in pyroptosis (Fig 2).<sup>31,41</sup>

The varying activation of the PCD pathways by A fumigatus influences the level of inflammation that the organism causes, particularly through varying amounts of IL-33 released. Death by apoptosis results in decreased IL-33 signaling owing to caspase 3- and caspase 7-induced cleavage of the C-terminal portion of IL-33, rendering it inert.<sup>17</sup> Alternatively, necroptosis induces the release of uncleaved IL-33.42 Although less active than the mature form seen following fungal protease cleavage, the fulllength IL-33 can produce an immune response. Finally, although a direct connection between IL-33 release in Aspergillus infection and pyroptosis is not established, there is evidence that it plays a protective role. There is increased susceptibility to invasive aspergillosis in AIM2 and NLRP3 knockout mice (AIM2 and NLRP3 being 2 genes involved in the pyroptosis pathway).<sup>43</sup> These findings suggest that whereas pyroptosis and apoptosis promote clearance of Aspergillus infection, necroptosis plays a pivotal role in promoting Aspergillus infections, and thus ABPA. Whether targeting the PANoptosis pathway would be beneficial or deleterious is unclear. Today, there is no evidence that deletion of ZBP1 increases or decreases the incidence of aspergillosis. It is possible that inhibition of PANoptosis would block most inflammation and thus allow asymptomatic colonization, given the fact that Aspergillus, a ubiquitous organism, often colonizes the respiratory tract without inducing any pathology. However, reduced



**FIG 1.** PANoptosis is suspected to be induced by *A fumigatus*. Evidence suggests there may be a cyclopentanediol analog produced by *A fumigatus* that serves as an inhibitor of TAK1. In addition, direct activation can occur through an unknown Z-DNA ligand binding to ZBP1. The PANoptosome complex formed from either pathway can then go on to induce the 3 modalities of cell death, namely, apoptosis, pyroptosis, and necroptosis. *ASC*, Adapter apoptosis-associated speck-like protein containing a caspase recruitment domain; *CASPB*, caspase-8; *FADD*, FAS-associated via death domain.

ability to clear the pathogen may allow for unchecked proliferation, eventually causing more severe disease. One thing is clear: therapies directed toward inhibition of the necroptosis pathway should be considered.

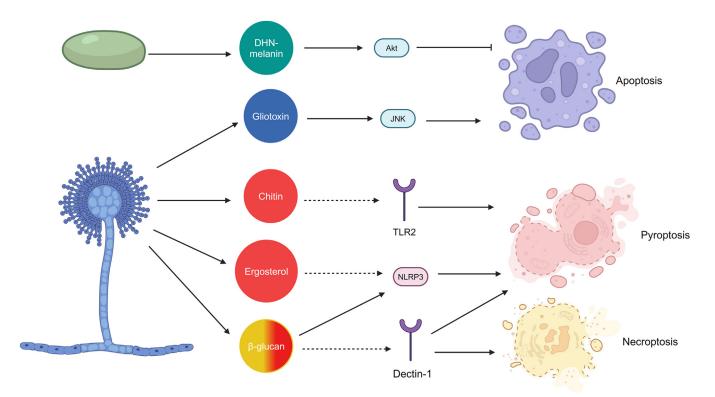
# DISCUSSION Necroptosis inhibitors as novel therapeutics to treat ABPA

Therapeutic agents that target the necroptosis pathway are available and are being evaluated to treat cancer, stroke, acute coronary syndrome, and other diseases.<sup>30,44,45</sup> Necrosulfonamide (NSA) is an inhibitor of MLKL that demonstrates efficacy in slowing breast tumor growth in xenografted mice.<sup>46</sup> Because MLKL is involved only in necroptosis and not in apoptosis or pyroptosis, it would be an ideal target for therapy. However, the preclinical animal model studies required before human testing would be difficult to perform because NSA does not bind to the mouse variant of MLKL.<sup>47</sup> Additionally, NSA may also bind GSDMD from the pyroptosis pathway, inhibiting its activation.<sup>48-50</sup> There are also data to suggest that NSA has an inhibitory

effect on apoptosis, further reducing the potential clinical significance of this compound.<sup>51</sup>

Necrostatin-1 (Nec-1) is a drug that targets necroptosis via inhibiting autophosphorylation of RIPK1. RIPK1 dimerizes and autophosphorylates to form a complex with RIPK3 to activate MLKL.<sup>52</sup> Because it is upstream in the necroptosis pathway, targeting RIPK1 over MLKL has its own issues. Namely, RIPK1 also serves as a driving force in the apoptosis pathway, although there is an RIPK1-independent apoptotic pathway.<sup>52</sup> The inhibitory effect of Nec-1 may be a promising area for clinical trials in the treatment of ABPA. There is evidence to suggest that in the absence of TAK1, necroptosis can also be activated in an RIPK1-independent manner.<sup>53</sup> This pathway may reduce the potential efficacy of Nec-1 as a treatment option for ABPA.

Although these 2 compounds are not the only PANoptosis regulators that may treat ABPA, they demonstrate the potential of this pathway and its complexity. Because of the cross talk among the 3 PCD pathways seen in PANoptosis, even drugs targeting a particular PCD pathway may show unexpected effects, as demonstrated by NSA potentially influencing



**FIG 2.** *A fumigatus*-induced cell death. Solid lines represent known relationships, dashed lines represent suspected relationships based on studies in *C albicans. A fumigatus* conidia inhibits apoptosis through the secretion of dihydroxynapthalene-melanin (DHN-melanin). Once it has germinated, *A fumigatus* has multiple virulence factors capable of inducing the 3 different cell death pathways. Gliotoxin serves as an activator of apoptosis. Ergosterol and chitin are known virulence factors of *A fumigatus* but have not been directly shown to activate the pyroptosis pathway in aspergillosis. Activation of dectin-1 via β-glucan has been demonstrated to play a role in the pyroptosis pathway but is currently only suspected in necroptosis.

apoptosis despite targeting a relatively downstream necroptosis molecule.<sup>49</sup> These cascading effects spilling over into the other PCD pathways also highlights the importance of targeted localization of these drugs. Although this article highlights necroptosis inhibitors as potential treatments, apoptosis and pyroptosis inhibit the release of IL-33 and could therefore be targeted by activators rather than by inhibitors. This would be associated with a greater risk of triggering cell death in otherwise healthy cells if sufficient drug localization could not be achieved. mAbs are also becoming treatment options for managing ABPA.<sup>54,55</sup> The mAb targeting IL-33 tozorakimab may be another promising candidate to treat ABPA.<sup>56</sup> Advancements in research open the door to development of new therapeutic options, and PANoptosis may be the next step in research on ABPA.

#### Conclusion

ABPA is a rare disease of the airways primarily affecting subjects with asthma and cystic fibrosis. Research demonstrates that PCD pathways play a role in ABPA; however, the emerging concept of PANoptosis and cross talk between these pathways provides a new avenue for understanding and possibly treating this disease. Studies show that pyroptosis and apoptosis reduce disease burden whereas necroptosis promotes pathology. Novel drugs that target elements of these pathways are under study and show promise as new treatment options for ABPA. The drugs NSA and Nec-1 are being explored to treat of other diseases, such as intervertebral disk degeneration and ischemia-reperfusion injury; they demonstrate how necroptosis inhibitors could be effective to treat ABPA. Additionally, other elements in these pathways, such as *Aspergillus*-produced virulence factors that influence TAK1 activity, are worth exploring. Blocking a potential TAK1 inhibitor may be a way of targeting PANoptosis as a whole order to inhibit the capacity of *Aspergillus* to induce inflammation. For this reason, depending on what future experiments in ZBP1-deficient mice show, ZBP1 also may be a worthy therapeutic target. As research on PANoptosis progresses, even more options for targetable pathways may be discovered.

### DISCLOSURE STATEMENT

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Clinical implications: Necroptosis inhibitors and other therapeutic agents targeting PANoptosis should be explored as treatment options for ABPA.

#### REFERENCES

- Harada T, Inui G, Ishikawa H, Kato R, Sueda Y, Funaki Y, et al. The clinical characteristics of allergic bronchopulmonary mycosis differ among pathogenic fungi. Yonago Acta Med 2023;66:257-62.
- Asano K, Hebisawa A, Ishiguro T, Takayanagi N, Nakamura Y, Suzuki J, et al. New clinical diagnostic criteria for allergic bronchopulmonary aspergillosis/ mycosis and its validation. J Allergy Clin Immunol 2021;147:1261-8.e5.
- **3.** Denning DW, Pleuvry A, Cole DC. Global burden of allergic bronchopulmonary aspergillosis with asthma and its complication chronic pulmonary aspergillosis in adults. Med Mycol 2013;51:361-70.
- Maturu VN, Agarwal R. Prevalence of Aspergillus sensitization and allergic bronchopulmonary aspergillosis in cystic fibrosis: systematic review and meta-analysis. Clin Exp Allergy 2015;45:1765-78.
- Roboubi A, Audousset C, Frealle E, Brun AL, Laurent F, Vitte J, et al. Allergic bronchopulmonary aspergillosis: a multidisciplinary review. J Mycol Med 2023; 33:101392.
- Greenberger PA. Allergic bronchopulmonary aspergillosis. J Allergy Clin Immunol 2002;110:685-92.
- Shah A, Maurya V, Panjabi C, Khanna P. Allergic bronchopulmonary aspergillosis without clinical asthma caused by Aspergillus niger. Allergy 2004;59:236-7.
- Koh WJ, Han J, Kim TS, Lee KS, Jang HW, Kwon OJ. Allergic bronchopulmonary aspergillosis coupled with broncholithiasis in a non-asthmatic patient. J Korean Med Sci 2007;22:365-8.
- Dong YT, Greco A, Garimella BV, Kumley B. Case report of allergic bronchopulmonary aspergillosis in a patient without asthma or bronchiectasis. Chest 2022; 162:18a-a.
- Sisodia J, Bajaj T. Allergic bronchopulmonary aspergillosis. Treasure Island, FL. StatPearls Publishing; 2024.
- Greenberger PA. Defining the outcome markers and therapeutic role for omalizumab in treatment of allergic bronchopulmonary aspergillosis. J Allergy Clin Immunol Pract 2023;11:906-7.
- Jin ML, Douglass JA, Elborn JS, Agarwal R, Calhoun WJ, Lazarewicz S, et al. Omalizumab in allergic bronchopulmonary aspergillosis: a systematic review and meta-analysis. J Allergy Clin Immunol 2023;11:896-905.
- Latge JP. Aspergillus fumigatus and aspergillosis. Clin Microbiol Rev 1999;12: 310-50.
- Kokubo K, Onodera A, Kiuchi M, Tsuji K, Hirahara K, Nakayama T. Conventional and pathogenic Th2 cells in inflammation, tissue repair, and fibrosis. Front Immunol 2022;13:945063.
- Saenz SA, Taylor BC, Artis D. Welcome to the neighborhood: epithelial cellderived cytokines license innate and adaptive immune responses at mucosal sites. Immunol Rev 2008;226:172-90.
- Hiraishi Y, Yamaguchi S, Yoshizaki T, Nambu A, Shimura E, Takamori A, et al. IL-33, IL-25 and TSLP contribute to development of fungal-associated proteaseinduced innate-type airway inflammation. Sci Rep 2018;8:18052.
- Cayrol C, Girard JP. Interleukin-33 (IL-33): a critical review of its biology and the mechanisms involved in its release as a potent extracellular cytokine. Cytokine 2022;156:155891.
- Pusceddu I, Dieplinger B, Mueller T. ST2 and the ST2/IL-33 signalling pathwaybiochemistry and pathophysiology in animal models and humans. Clin Chim Acta 2019;495:493-500.
- Cayrol C, Duval A, Schmitt P, Roga S, Camus M, Stella A, et al. Environmental allergens induce allergic inflammation through proteolytic maturation of IL-33. Nat Immunol 2018;19:375-85.
- 20. Ito Y, Takazono T, Obase Y, Fukahori S, Ashizawa N, Hirayama T, et al. Serum cytokines usefulness for understanding the pathology in allergic bronchopulmonary aspergillosis and chronic pulmonary aspergillosis. J Fungi (Basel) 2022;8:436.
- Bojanowski CM, Bitoun JP, Kolls JK. An ALARMINg Type 2 Response in cystic fibrosis-the key to understanding ABPA? Am J Respir Crit Care Med 2023;207: 1418-9.
- Ramaprakash H, Shibata T, Duffy KE, Ismailoglu UB, Bredernitz RM, Moreira AP, et al. Targeting ST2L potentiates CpG-mediated therapeutic effects in a chronic fungal asthma model. Am J Pathol 2011;179:104-15.
- 23. Garth JM, Reeder KM, Godwin MS, Mackel JJ, Dunaway CW, Blackburn JP, Steele C. IL-33 Signaling regulates innate IL-17A and IL-22 production via suppression of prostaglandin E(2) during lung fungal infection. J Immunol 2017;199:2140-8.
- D'Arcy MS. Cell death: a review of the major forms of apoptosis, necrosis and autophagy. Cell Biol Int 2019;43:582-92.

- Yu P, Zhang X, Liu N, Tang L, Peng C, Chen X. Pyroptosis: mechanisms and diseases. Signal Transduct Target Ther 2021;6:128.
- Ketelut-Carneiro N, Fitzgerald KA. Apoptosis, pyroptosis, and necroptosis-oh my! The many ways a cell can die. J Mol Biol 2022;434:167378.
- Kuriakose T, Man SM, Malireddi RK, Karki R, Kesavardhana S, Place DE, et al. ZBP1/DAI is an innate sensor of influenza virus triggering the NLRP3 inflammasome and programmed cell death pathways. Sci Immunol 2016;1:aag2045.
- Zhu P, Ke ZR, Chen JX, Li SJ, Ma TL, Fan XL. Advances in mechanism and regulation of PANoptosis: prospects in disease treatment. Front Immunol 2023;14:1120034.
- Malireddi RKS, Kesavardhana S, Kanneganti TD. ZBP1 and TAK1: master regulators of NLRP3 inflammasome/pyroptosis, apoptosis, and necroptosis (PAN-optosis). Front Cell Infect Microbiol 2019;9:406.
- Yan WT, Zhao WJ, Hu XM, Ban XX, Ning WY, Wan H, et al. PANoptosis-like cell death in ischemia/reperfusion injury of retinal neurons. Neural Regen Res 2023;18:357-63.
- Williams TJ, Gonzales-Huerta LE, Armstrong-James D. Fungal-induced programmed cell death. J Fungi (Basel) 2021;7:231.
- 32. Banoth B, Tuladhar S, Karki R, Sharma BR, Briard B, Kesavardhana S, et al. ZBP1 promotes fungi-induced inflammasome activation and pyroptosis, apoptosis, and necroptosis (PANoptosis). J Biol Chem 2020;295:18276-83.
- Karki R, Kanneganti TD. ADAR1 and ZBP1 in innate immunity, cell death, and disease. Trends Immunol 2023;44:201-16.
- 34. Samir P, Malireddi RKS, Kanneganti TD. The PANoptosome: a deadly protein complex driving pyroptosis, apoptosis, and necroptosis (PANoptosis). Front Cell Infect Microbiol 2020;10:238.
- 35. Sekiya M, Ueda K, Okazaki K, Kikuchi H, Kurata S, Oshima Y. A cyclopentanediol analogue selectively suppresses the conserved innate immunity pathways, Drosophila IMD and TNF-alpha pathways. Biochem Pharmacol 2008;75:2165-74.
- 36. Briard B, Karki R, Malireddi RKS, Bhattacharya A, Place DE, Mavuluri J, et al. Fungal ligands released by innate immune effectors promote inflammasome activation during Aspergillus fumigatus infection. Nat Microbiol 2019;4:316-27.
- 37. Yan W, Zhao YS, Xie K, Xing Y, Xu F. Aspergillus fumigatus influences gasdermin-D-dependent pyroptosis of the lung via regulating Toll-like receptor 2-mediated regulatory T cell differentiation. J Immunol Res 2021;2021: 5538612.
- Fuchs K, Cardona Gloria Y, Wolz OO, Herster F, Sharma L, Dillen CA, et al. The fungal ligand chitin directly binds TLR2 and triggers inflammation dependent on oligomer size. EMBO Rep 2018;19:e46065.
- **39.** Abad A, Fernandez-Molina JV, Bikandi J, Ramirez A, Margareto J, Sendino J, et al. What makes Aspergillus fumigatus a successful pathogen? Genes and molecules involved in invasive aspergillosis. Rev Iberoam Micol 2010;27:155-82.
- 40. Rodrigues ML. The multifunctional fungal ergosterol. mBio 2018;9:e01755.
- Cao M, Wu Z, Lou Q, Lu W, Zhang J, Li Q, et al. Dectin-1-induced RIPK1 and RIPK3 activation protects host against Candida albicans infection. Cell Death Differ 2019;26:2622-36.
- 42. Shlomovitz I, Erlich Z, Speir M, Zargarian S, Baram N, Engler M, et al. Necroptosis directly induces the release of full-length biologically active IL-33 in vitro and in an inflammatory disease model. FEBS J 2019;286:507-22.
- 43. Karki R, Man SM, Malireddi RKS, Gurung P, Vogel P, Lamkanfi M, Kanneganti TD. Concerted activation of the AIM2 and NLRP3 inflammasomes orchestrates host protection against Aspergillus infection. Cell Host Microbe 2015;17:357-68.
- Khoury MK, Gupta K, Franco SR, Liu B. Necroptosis in the pathophysiology of disease. Am J Pathol 2020;190:272-85.
- 45. Tong X, Tang R, Xiao M, Xu J, Wang W, Zhang B, et al. Targeting cell death pathways for cancer therapy: recent developments in necroptosis, pyroptosis, ferroptosis, and cuproptosis research. J Hematol Oncol 2022;15:174.
- 46. Liu X, Zhou M, Mei L, Ruan J, Hu Q, Peng J, et al. Key roles of necroptotic factors in promoting tumor growth. Oncotarget 2016;7:22219-33.
- 47. Sun L, Wang H, Wang Z, He S, Chen S, Liao D, et al. Mixed lineage kinase domain-like protein mediates necrosis signaling downstream of RIP3 kinase. Cell 2012;148:213-27.
- 48. Rathkey JK, Zhao J, Liu Z, Chen Y, Yang J, Kondolf HC, et al. Chemical disruption of the pyroptotic pore-forming protein gasdermin D inhibits inflammatory cell death and sepsis. Sci Immunol 2018;3:eaat2738.
- 49. Zhang J, Wei K. Necrosulfonamide reverses pyroptosis-induced inhibition of proliferation and differentiation of osteoblasts through the NLRP3/caspase-1/GSDMD pathway. Exp Cell Res 2021;405:112648.
- 50. Yang W, Tao K, Wang Y, Huang Y, Duan C, Wang T, et al. Necrosulfonamide ameliorates intestinal inflammation via inhibiting GSDMD-medicated pyroptosis and MLKL-mediated necroptosis. Biochem Pharmacol 2022;206:115338.
- Zhang QX, Guo D, Wang FC, Ding WY. Necrosulfonamide (NSA) protects intervertebral disc degeneration via necroptosis and apoptosis inhibition. Eur Rev Med Pharmacol Sci 2020;24:2683-91.
- Cao L, Mu W. Necrostatin-1 and necroptosis inhibition: pathophysiology and therapeutic implications. Pharmacol Res 2021;163:105297.

- 53. Malireddi RKS, Gurung P, Kesavardhana S, Samir P, Burton A, Mummareddy H, et al. Innate immune priming in the absence of TAK1 drives RIPK1 kinase activityindependent pyroptosis, apoptosis, necroptosis, and inflammatory disease. J Exp Med 2020;217:jem.20191644.
- 54. Lewington-Gower E, Chan L, Shah A. Review of current and future therapeutics in ABPA. Ther Adv Chronic Dis 2021;12:20406223211047003.
- O'Reilly A, Dunican E. The use of targeted monoclonal antibodies in the treatment of ABPA-a case series. Medicina (Kaunas) 2021;58:53.
- 56. England E, Rees DG, Scott IC, Carmen S, Chan DTY, Chaillan Huntington CE, et al. Tozorakimab (MEDI3506): an anti-IL-33 antibody that inhibits IL-33 signalling via ST2 and RAGE/EGFR to reduce inflammation and epithelial dysfunction. Sci Rep 2023;13:9825.