# LETTER TO THE EDITOR

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# Molecular evidence of *Rickettsia raoultii*, "Candidatus Rickettsia barbariae" and a novel Babesia genotype in marbled polecats (Vormela peregusna) at the China-Kazakhstan border

Xiafei Liu<sup>1+</sup>, Meihua Yang<sup>2+</sup>, Guangyuan Liu<sup>3+</sup>, Shanshan Zhao<sup>1</sup>, Wumei Yuan<sup>1</sup>, Ronghai Xiao<sup>4</sup>, Wurelihazi Hazihan<sup>5</sup>, Sándor Hornok<sup>6</sup> and Yuanzhi Wang<sup>1\*</sup>

# Abstract

In this study, two road-killed marbled polecats (*Vormela peregusna*) were molecularly analysed for tick-borne pathogens. *Rickettsia raoultii, "Candidatus* Rickettsia barbariae" and a novel *Babesia* genotype have been identified, for the first time in marbled polecat. DNA of this *Babesia* sp. genotype was also present in four out of 15 *Haemaphysalis erinacei* ticks collected from the *Babesia* PCR-positive marbled polecat. Results of this study suggest that marbled polecats may serve as reservoirs for these bacteria and protozoans.

**Keywords:** Babesia, "Candidatus Rickettsia barbariae", China-Kazakhstan border, Haemaphysalis erinacei, Marbled polecat, Rickettsia raoultii

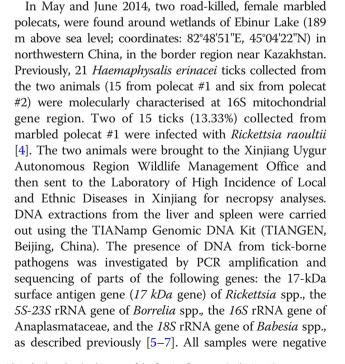
# Letter to the Editor

The marbled polecat (*Vormela peregusna*) is a small carnivorous mammal (Carnivora: Mustelidae) with a broad geographical range, extending from southeast Europe, through southwest and central Asia, to Mongolia and northern China. This species is the only member of the genus and has been listed as globally vulnerable, due to substantially declining populations [1]. Apart from the loss of steppe habitats and desertification, infection with various pathogens may contribute to its decreasing numbers. At the same time, data are very limited on the epidemiological role of the marbled polecat as a reservoir of pathogens with veterinary or medical significance [2, 3]. Therefore, molecular investigations for pathogens in this endangered species have multifold importance.

\* Correspondence: wangyuanzhi621@126.com

<sup>1</sup>School of Medicine, Shihezi University, Shihezi, Xinjiang Uygur Autonomous Region 832002, People's Republic of China

Full list of author information is available at the end of the article

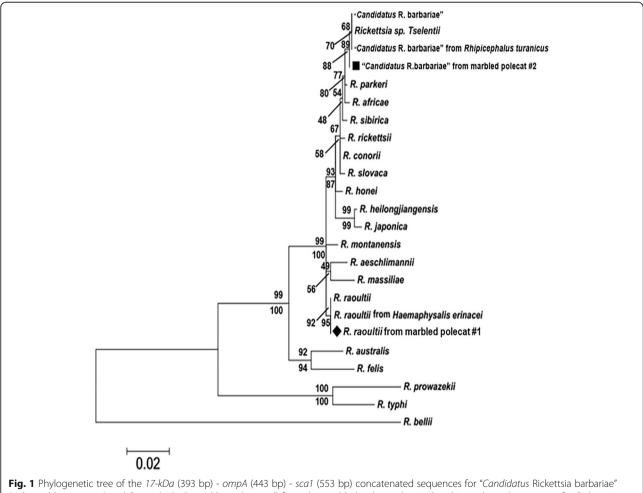


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 $<sup>^{\</sup>dagger}\mathrm{Xiafei}$  Liu, Meihua Yang and Guangyuan Liu contributed equally to this work.

for Borrelia spp. and Anaplasmataceae. To confirm the results and to compare additional genetic markers, further PCR and sequencing were performed, to detect the outer membrane protein A (*ompA*) and cell surface antigen 1 (sca1) genes of Rickettsia spp. [5] and the cytochrome b (cytb) gene of Babesia spp. [8]. All PCRs were performed including double distilled water (ddH<sub>2</sub>O) as a negative control. Sequences were compared with GenBank data using the nucleotide BLAST program (http://www.ncbi.nlm.nih. gov/BLAST/). All obtained sequences were deposited in the GenBank database [17-kDa: MG674917 (R. raoultii) and MG674918 ("Candidatus Rickettsia barbariae"); ompA: MG662380 (R. raoultii) and MG662381 ("Candidatus Rickettsia barbariae"); sca1: MG662382 (R. raoultii) and MG662383 ("Candidatus Rickettsia barbariae"); 18S rRNA: MG799848 and MG813565; cytb: MG832590 and MG832591 (Babesia sp. from marbled polecat and H. erinacei, respectively)].

However, the liver and spleen samples of both marbled polecats were PCR-positive for rickettsiae. In animal #1, the ompA and sca1 sequences showed 100% identities with R. raoultii strain Khabarovsk<sup>T</sup> (GenBank: CP010969). This result is in line with previous findings, i.e. two out of the fifteen ticks (i.e. 13.33%) collected from animal #1 harboured R. raoultii. The phylogenetic analysis showed that the genetic pattern was the same between R. raoultii from a marbled polecat #1 and R. raoultii previously found in H. erinacei ticks collected from marbled polecat #1 (Fig. 1) [4]. In animal #2, "Candidatus Rickettsia barbariae" was identified, with 99.8% (552/553 bp) and 100% (443/443 bp) similarities to sequences available in GenBank [from Vermipsylla alakurt in China: KT284718 (sca1) and KU645284 (ompA)]. Results of sequence alignments were confirmed by phylogenetic analyses, placing rickettsiae detected here into the cluster of either R. raoultii or "Candidatus Rickettsia barbariae" (Fig. 1 and Additional file 1: Table S1).



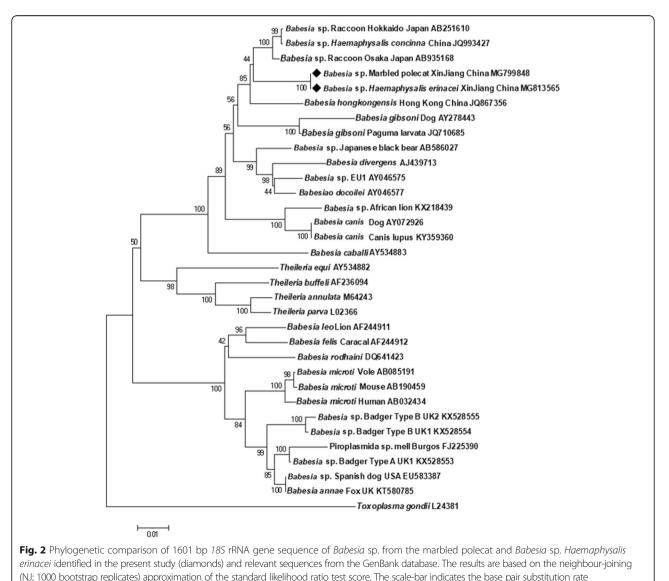
(indicated by a square) and *R. raoultii* (indicated by a diamond) from the marbled polecat obtained in this study, and sequences for *Rickettsia* species retrieved from the GenBank database. The tree was constructed using the neighbour-joining method (NJ; 1000 bootstrap replicates) and maximum-likelihood (ML; 1000 bootstrap replicates) analyses using MEGA6. The scale-bar represents the inferred substitutions per nucleotide site. The relative support for clades in the tree was produced from the NJ and ML analyses

Both Rickettsia spp. were detected for the first time in marbled polecats. Spotted fever group rickettsiae (Proteobacteria: Rickettsiales) are obligatory intracellular, Gram-negative bacteria, which can cause zoonotic disease after transmission by a blood-sucking arthropod [9]. Rickettsia raoultii is one of the causative agents of human rickettsioses (SENLAT/TIBOLA/DEBONEL [10]). Until recently, R. raoultii has not been reported from mammals other than humans [11], but during the past years, this species was detected in Mongolian gazelle [12] and dogs [13]. The most important tick vectors of *R. raoultii* appear to be Dermacentor spp., with transstadial and transovarial maintenance of this agent [14]. Accordingly, a previous study revealed R. raoultii as the predominant species of Rickettsia found in D. nuttalli from Mongolian regions and in D. silvarum ticks in the border region between China and Russia [15, 16]. Therefore, simultaneous detection of R.

*raoultii* in a marbled polecat (as shown here) and in *H. erinacei* removed from the same animal justify further studies on the vector role of this tick species, which is outside the typical vector range of *R. raoultii*.

"Candidatus Rickettsia barbariae" has been detected and described from ticks of domestic and wild animals [17–20] and humans [21] in the Mediterranean region. Our previous work also showed the presence of DNA of "Candidatus Rickettsia barbariae" in the tick *Rhipicephalus* turanicus [22] and the flea Vermipsylla alakurt [5] collected from sheep around the Taklamakan Desert in Xinjiang, northwestern China. To the best of our knowledge, "Candidatus Rickettsia barbariae" was not detected in any vertebrate hosts of the above ectoparasites, further increasing the significance of the present findings.

In addition to the above results, *Babesia* sp. DNA was detected both in four *H. erinacei* ticks and the organs of



their parasitised host (marbled polecat #1). The 18S rRNA gene sequences were 100% identical between the marbled polecat and its PCR-positive ticks. However, this *Babesia* genotype had only 97.12% (1517/1562 bp) 18S rRNA gene similarity to the closest genotype in GenBank, detected in the blood of a racoon in Japan (GenBank: AB935168). The cytb gene sequence comparisons supported the uniqueness of this Babesia sp. from marbled polecat because the gene sequence showed 85.74% (433/505 bp) similarity to that of Babesia gibsoni (no corresponding cytb gene sequence was found for Babesia sp. from a racoon in GenBank). The phylogenetic analysis confirmed these results: this novel Babesia genotype clustered separately from the above racoon-associated piroplasm (GenBank: AB935168) and belonged to a phylogenetic group including Babesia spp. from Caniformia or tick species (i.e. H. concinna and from the present study H. erinacei) frequently infesting Caniformia (Fig. 2). The separation of this phylogenetic group from a cat-associated piroplasm (B. hongkongensis) was strongly (85%) supported (Fig. 2).

To the best of our knowledge, this is the first report on the presence of Babesia DNA in marbled polecat and H. erinacei ticks. Babesia spp. (Apicomplexa: Piroplasmida) are intraerythrocytic parasites, which have been reported from birds and mammals (including wild carnivores) worldwide [7, 23] and have ixodid ticks as their principal vectors [24]. The 18S rRNA gene is the most widely used genetic marker for the identification of babesiae [24]. Based on 18S rRNA gene sequence data and the topology of the phylogenetic tree (Fig. 2), the Babesia sp. detected here in the marbled polecat, and the attendant ticks was most closely related to a Babesia sp. from a racoon (GenBank: AB935168: reported from Osaka, Japan) and differed from all other piroplasmid species. Taking into account that the 18S rRNA gene may have very few nucleotide substitutions between closely related species [e.g. only 0.2% difference delineating B. divergens (Gen-Bank: FJ944825) and B. capreoli (GenBank: AY726009) [25]], this new *Babesia* genotype most likely represents a species different from the one detected in raccoon in Japan. Unfortunately, morphological characterisation of this novel Babesia genotype was not possible because samples were obtained from road-killed (i.e. not freshly dead) animals, and Babesia are known to undergo quick degradation and morphological changes post-mortem [26].

In conclusion, results of this study suggest that marbled polecats may serve as reservoirs for *R. raoultii*, *"Candidatus* Rickettsia barbariae" and a novel *Babesia* genotype. Further studies are needed to evaluate if rickettsemia in this host species is of sufficient magnitude and duration to infect ticks, which is a known prerequisite for effective transmission of other tick-borne rickettsiae [27].

# **Additional file**

Additional file 1: Table S1. Information for the sequences from the GenBank database used in Fig. 1. (DOCX 18 kb)

# Abbreviations

cytb: cytochrome b; DEBONEL: *Dermacentor*-borne necrosis, eschar and lymphadenopathy; HGA: human granulocytic anaplasmosis; IUCN: International Union for Conservation of Nature; ompA: outer membrane protein A; sca1: surface cell antigen 1; TIBOLA: tick-borne lymphadenopathy

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#### Availability of data and materials

The sequences obtained and analysed during the present study are deposited in the GenBank database, under the accession numbers MG674917, MG662380 and MG662382 (*R. raoultii*); MG674918, MG662381 and MG662383 (*"Candidatus* Rickettsia barbariae"); MG799848, MG813565, MG832590 and MG832591 (*Babesia* sp.). All other relevant data are included in the article.

#### Authors' contributions

XL, MY, GL and YW conceived and designed the study. SZ, WY, RX and WH processed the samples and performed molecular and phylogenetic analyses. SH contributed to the study design and the manuscript. All authors read and approved the final manuscript.

#### **Ethics** approval

This study was approved by the Animal Ethics Committee of Shihezi University (Approval No. AECSU2014-03).

#### Consent for publication

Not applicable.

#### Competing interests

The authors declare that they have no competing interests.

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## Author details

<sup>1</sup>School of Medicine, Shihezi University, Shihezi, Xinjiang Uygur Autonomous Region 832002, People's Republic of China. <sup>2</sup>School of Agriculture, Shihezi University, Shihezi, Xinjiang Uygur Autonomous Region 832000, People's Republic of China. <sup>3</sup>State Key Laboratory of Veterinary Etiological Biology, Key Laboratory of Veterinary Parasitology of Gansu Province, Lanzhou Veterinary Research Institute, Chinese Academy of Agricultural Science, Xujiaping 1#, Lanzhou, Gansu 730046, People's Republic of China. <sup>4</sup>Inspection and Comprehensive Technology Center of Ruili Entry-Exit Inspection and Quarantine Bureau, No.75, Ruihong Road, Ruili 678600, Yunnan, People's Republic of China. <sup>5</sup>School of Animal Science and Technology, Shihezi University, Shihezi, Xinjiang Uygur Autonomous Region 832000, People's Republic of China. <sup>6</sup>Department of Parasitology and Zoology, University of Veterinary Medicine, Budapest, Hungary.

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